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J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.0c00703 • Publication Date (Web): 07 May 2020

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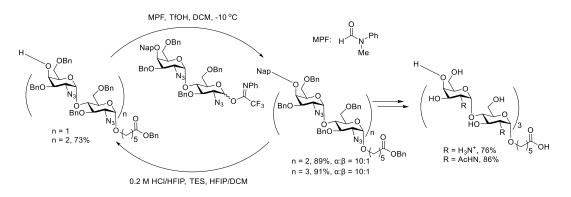
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Reagent Controlled Glycosylations for the Assembly of Well-defined Pel Oligosaccharides

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ABSTRACT: A new additive, methyl(phenyl)formamide (MPF), is introduced for the glycosylation of 2-azido-2-deoxyglucose building blocks. A linear α -(1,4)-glucosamine tetrasaccharide was assembled to prove the utility of MPF. Next, a hexasaccharide fragment of the *Pseudomonas aeruginosa* exopolysaccharide Pel was assembled using a [2+2+2] strategy modulated by MPF. The used [galactosazide- α -(1,4)-glucosazide] disaccharide building blocks were synthesized using a 4,6-*O*-DTBS protected galactosyl azide donor.

Introduction

Pel is one of the exopolysaccharides that is involved in the biofilm formation of Pseudomonas aeruginosa, an opportunistic Gram-positive pathogen that is the major cause of morbidity and mortality in cystic fibrosis patients.¹ Pel is a linear polysaccharide composed of 1,4-linked α -N-acetyl galactosamine (GalNAc) and α -*N*-acetyl glucosamine (GlcNAc) residues, present in a ±6:1 ratio, of which some of the residues have been de-acetylated to generate positively charged galactosamine (GalN) and glucosamine (GlcN) moieties (Figure 1).^{1b} Well-defined Pel fragments can be used to unravel their role in biofilm formation, to study their biosynthesis and possibly as synthetic antigens in the development of a (semi)-synthetic vaccine against P. aeruginosa. Because of the random distribution of the monosaccharides in Pel, it is impossible to isolate well-defined oligosaccharides from natural sources and therefore organic synthesis is necessary to provide these structures.

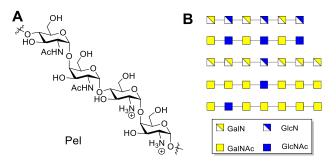


Figure 1. The repeating structures of Pel.

The key challenge in the generation of these oligosaccharides is the stereoselective construction of the 1,2-*cis*-glycosidic linkages. Four kinds of *cis*-glycosidic linkages, namely α -D-GlcN-(1 \rightarrow 4)-D-GlcN, α -D-GlcN-(1 \rightarrow 4)-D-GalN, α -D-GalN-(1 \rightarrow 4)-D-GlcN and α -D-GalN-(1 \rightarrow 4)-D-GalN, have to be constructed. Zhang *et al.* have recently reported an effective synthetic strategy to assembly galactosaminogalactans (GAGs), fungal polysaccharides composed of 1,4-linked α -D-Gal, α -D-GalN and α -D-GalNAc moieties.² For the formation of the 1,2-*cis*-linkages in these structures, 4,6-di-*tert*-

butylsilylene (4,6-O-DTBS) protected GalN-donors were used to control the selectivity.3 This strategy allowed the use of galactosamine donors bearing differently masked amine functionalities. Galactosazide and trichloroacetyl protected GalN donors were used to combine GalN and Gal-NAc at pre-determined sites in the target GAG oligosaccharides. Of note, the stereodirecting capacity of the DTBS group in GalN donors effectively overrides the neighboring group participation by C2-participating functionalities such as the trichloroacetamide. Thus, DTBS-GalN donors also represent attractive building blocks for Pel-assembly. For the stereoselective introduction of α-D-GlcN linkages no general solution exists, even though the construction of this type of glycosidic linkage has attracted significant attention,^{4,5} as it is present in many important natural polysaccharides and glycoconjugates, such as heparin, heparan sulfate,6 GPI anchors and various bacterial polysaccharides.7

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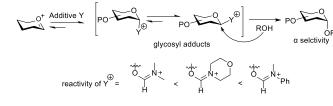
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16 Additive controlled glycosylations are gaining increasing 17 interest for the stereoselective construction of glycosidic 18 linkages.8 In these approaches the nature of the additive de-19 termines the reactivity of in situ formed glycosylating species and the influence of the additive can be tuned to match 20 the reactivity of the glycosyl donor⁹ and acceptor¹⁰ building 21 blocks. We have recently reported on the fully stereoselec-22 tive assembly of a branched α -glucan with an α -(1 \rightarrow 4)-23 linked backbone from *Mycobacterium tuberculosis*, α -(1,3)-24 glucans from the Aspergillus fumigatus fungal cell wall as 25 well as the assembly of α -(1,3)-glucans found attached to 26 lipoteichoic acids of Enterococcus faecalis.11 The synthetic 27 strategy used in these approaches hinged on the use of ad-28 ditive controlled glycosylation reactions in combination 29 with the use of a single benzyl-type protecting group (Bn, 30 PMB, Nap). For glycosylations with relatively reactive pri-31 mary alcohol acceptors, the trimethylsilyliodide (TMSI)-32 triphenylphosphine oxide (Ph₃P=0) activator couple was 33 used, while the condensations with less reactive secondary 34 alcohols required the use of the trifluoromethanesulfonic acid (TfOH)-dimethylformamide (DMF) pair. The successful 35 construction of multiple $1,4-\alpha$ -glucosidic linkages was an 36 incentive to explore this strategy for the assembly of the Pel 37 oligasaccharides. Mong and co-workers have previously de-38 scribed how formamide additives can be used for the con-39 struction of 1,2-cis-GalN3 and GlcN3 linkages. They intro-40 duced N-formyl-morpholine (NFM) to modulate the reactiv-41 ity of tri-O-benzyl GlcN₃ and 4,6-benzylidene-GalN₃ donors 42 and showed that glycosylations mediated by NFM pro-43 ceeded with higher stereoselectivity than the correspond-44 ing DMF-modulated condensations.7c Because of the 45 stronger electron withdrawing effect of the azide group 46 with respect to a benzyl ether, 2-azido donors are generally 47 less reactive than their 2-0-benzyl counterparts. This lower reactivity can be counterbalanced by the use of a somewhat 48 less nucleophilic additive, resulting in a better leaving group 49 Y, thereby explaining why NFM outperforms DMF in these 50 glycosylations (See Scheme 1). 51



Scheme 1. The relative reactivity of DMF and NFM glycosyl adducts.

We here describe a strategy to synthesize Pel oligosaccharides using additive-controlled glycosylations to match the reactivities of the GlcN₃-donor and the Pel acceptors. Because of the relatively low nucleophilicity of the GlcN3-C4-OH and especially the GalN3-C4-OH, a new additive is introduced that generates intermediates that are more reactive than the previously introduced DMF and NFM-imidinium ions.

Results and discussion

First, we paid attention to the formation of α -D-GlcN-(1 \rightarrow 4)-D-GlcN linkages. In line with previous work, solely benzyl type protecting group (PMB, Nap, Bn) were used -besides the azide at C2- to generate orthogonally protected building blocks of uniform reactivity. With donor 1 and acceptor 4 (See SI for the syntheses of these building blocks), DMF was investigated as an additive to control the selectivity according to previous successful experiments. Thus, donor α -D-GlcN 1, acceptor 4 and the additive were mixed in DCM with molecular sieves and cooled to -78 °C. Next, TfOH was added and after stirring for 0.5h, the mixture was placed at 0 °C and allowed to stir for 24h. As shown in Table 1, this produced the desired disaccharide product **8** with complete α selectivity, but the yield was only 32% (entry 1). Performing the reaction at room temperature did not lead to erosion of stereoselectivity but only marginally improved the yield (entry 2). Likely, the low reactivity of the donor and acceptor led to the observed poor yield and therefore NFM was probed as additive.6c Use of this additive provided complete α -selectivity, and raised the yield of the condensation to 55% yield. To further improve the reaction, a slightly less nucleophilic additive was sought and N-methyl-N-phenylformamide (MPF) was explored. It was expected that the imidinium ion formed from this additive would be more reactive because the aniline-type nitrogen would be less capable of supporting the (partial) positive charge in the ion (See Scheme 1). The reaction of donor 1 and acceptor 4 proceeded with excellent yield (91%) when performed at 0 °C, and the disaccharide **8** was obtained with 15:1 α : β -selectivity (entry 4). Although the stereoselectivity of this condensation is somewhat less than the DMF or NFM mediated glycosylations, the improved yield allows for an overall more productive reaction.12

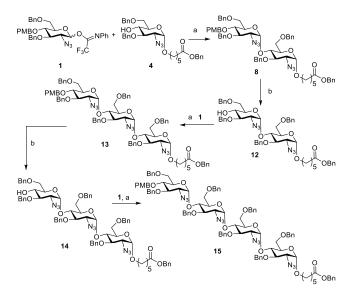
Next, our attention was turned to the formation of the α -GlcN- $(1 \rightarrow 4)$ -GalN-linkage exploring the additives as described above. First donor 2 was coupled with acceptor 5 using DMF to provide product 9 in low yield and poor selectivity (Table 1, entry 5). The use of NFM instead of DMF did not improve the outcome of this glycosylation (entry 6). Likely the poor reactivity of the GalN₃-C4-OH hampers the union of the two carbohydrate building blocks. Next, the use of MPF was explored. At 0 °C, disaccharide 9 was obtained in high yield (83%), but with moderate α : β -selectivity (5:1). Performing the same reaction at -10°C increased the α -selectivity (α : β = 10:1), but led to a relatively low yield (43%, entry 8). To increase the yield of the reaction, the concentration was raised from 0.1 M to 0.2 M (entry 9). This led to the formation of the desired compound 9 with a yield of 88% and a 10:1 α : β ratio. Having defined adequate conditions for

the construction of α -GlcN-(1→4)-GlcN and α -GlcN-(1→4)-GalN linkages, the use of MPF in combination with galactosazide donor **3** was explored for the construction of the target α -GalN-(1→4)-GlcN and α -GalN-(1→4)-GalN linkages. Under the conditions established above, donor **3** was coupled with glucosyl acceptor **6** to give the disaccharide **10** in excellent yield and 8:1 α/β -stereo selectivity (entry 10). Contrary, disaccharide **11**, formed from donor **3** and galactosyl acceptor **7**, was obtained with relatively poor selectivity (α : β = 4:1, entry 11). From these model reactions, it can be concluded that three out of four Pel-type linkages can effectively be installed using the MPF-mediated glycosylations. For the α -GalN-(1→4)-GalN linkages, the previously reported approach using 4,6-*0*-DTBS galactosamine donors is clearly superior. Next we probed the robustness of the MPF-mediated protocol in the synthesis of Pel-type oligosaccharides. First the assembly of an all-1,2-*cis* linked tetraglucosamine was explored as depicted in Scheme 2. Thus, donor **1** and acceptor **4** were coupled under the above identified reaction conditions to provide the desired disaccharide **8**. The PMB was removed using a catalytic amount of HCl to give disaccharide acceptor **12** in 88% yield.¹³ Next, compound **12** was glycosylated with donor **1** under the MPF-conditions to form the desired trisaccharide **13** in 83% yield and excellent stereoselectivity (α : $\beta > 19$:1). Repetition of the deprotection and glycosylation reactions then uneventfully provided tetrasaccharide **15**. The successful assembly of this tetrasaccharide indicates that the yield and stereoselectivity do not decrease with the growing of the sugar chain.

Table 1. Glycosylation between 2-azido Glu/Gal donors and 4-OH-2-azido Glu/Gal acceptors.

donors:	acceptors:								
<)Bn	OBn	BnO OB	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		но	OBn HO OBn HO		
		Ph BnO		-√~ ^O ∕~ NPh	nO N ₃	O BnO		N ₃ E	BnO N ₃
1	F ₃ C	2	F ₃ C	N ₃ / F ₃ C	40	¹ ∕ ₅ OBn		6	7
				BnO—		OBn	BnO		·····
		<	20	BnO DO	I	BnO OBII	BnO-	20	
donor	TfOH, addi	PMBO BnO	OBn N ₃	NL I	OBn Br	Na Na	OBn	N ₃ OBn	
+ acceptor	DCM, 2		BnO	BnO	-0		10 million	BnO	
	DCIM, 2	411	8 N ₃ O	0.0	N ₃ 0	l ₃ 10	N ₃	11 N ₃	\sim
			8 (~)5	OBN 6	, ,	l ₃ 10		11	
entry	donor	acceptor	c(mmol/ml)	additive	eq	T(°C)	product	yield ^a	α : β^{b}
1	1	4	0.1	DMF	16	0	8	32%	>20:1
2	1	4	0.1	DMF	16	rt	8	38%	>20:1
3	1	4	0.1	NFM	16	rt	8	55%	>20:1
4	1	4	0.1	MPF	16	0	8	91%	~15:1
5	2	5	0.1	DMF	16	0	8	23%	6:1
6	2	5	0.1	NFM	16	0	9	24%	6:1
7	2	5	0.1	MPF	16	0	9	83%	5:1
8	2	5	0.1	MPF	16	-10	9	43%	10:1
9	2	5	0.2	MPF	16	-10	9	88%	10:1
10	3	6	0.1	MPF	16	-10	10	88%	8:1
11	3	7	0.1	MPF	16	-10	11	80%	4:1
lsolated yie	ld. ^b The	α:β ratio was	determined by	¹ H NMR.					



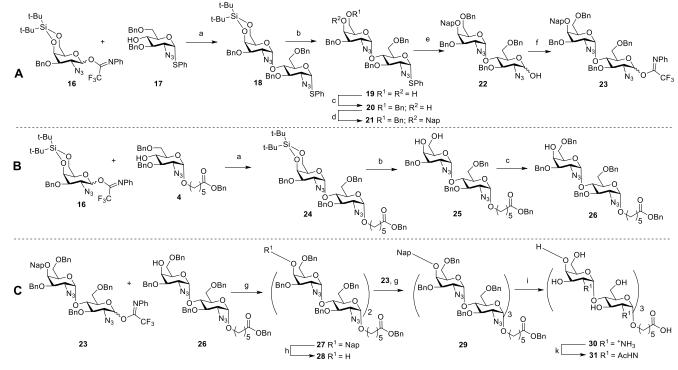


(a) MPF, TfOH, DCM, -78-0 °C, **8**: 91%, α : β = 15:1; **13**: 83%, α : β > 19:1; **15**: 90%, α : β > 20:1. (b) 0.2 M HCl/HFIP, TES, HFIP/DCM, **12**: 88%; **14**: 78%.

Scheme 2. Assembly of an α -glucosazide tetrasaccharide using MPF mediated glycosylations.

Next the synthesis of a Pel hexasaccharide, featuring both GalN and GlcN residues, was undertaken. A [2+2+2] strategy was designed to streamline the assembly of the structures, building on MFP-mediated glycosylations of the GalN₃-GlcN₃ donor **23**. The procedure for the synthesis of the required building blocks **23** and **26** is depicted in

Scheme 3 A and B. First donor 16 was coupled with glucoazide 17 in a chemoselective glycosylation to form disaccharide 18 as a single anomer. Next, the silvlidene ketal was cleaved with HF-pyridine, after which a benzyl ether was regioselectively introduced under the aegis of Taylor's borinic acid catalyst.¹⁴ Protection of the remaining C4'-OH with a naphthyl group delivered compound **21**. Next the anomeric thiophenol group was removed using N-iodosuccinimide in acetone/water, and the resulting hydroxyl group turned into the desired N-phenyltrifluoroimidate functionality to provide donor 23. Acceptor 26 was obtained from donor 16 and acceptor 4. These two building blocks were united to stereoselectively provide disaccharide 24. Removal of the silylidene ketal and introduction of the C6'-O-benzyl ether as described above provided 26. With building blocks 23 and 26 in hand, the assembly of the target hexasaccharides was undertaken (Scheme 3C). First, donor 23 was glycosylated with acceptor 26 using MPF as additive at -10 °C at a 0.2 M concentration to form tetrasaccharide 27 in 89% yield as a 10:1 α/β -mixture. Then the Nap ether was removed using HCl and triethylsilane in DCM/HFIP to give the tetrasaccharide acceptor 28. Compound 28 was treated with donor 23 under the optimal MPF-mediated glycosylation conditions to deliver hexasaccharide 29 in high yield and stereoselectivity. Reduction of the six azides and removal of the benzyl ester and ethers were accomplished in a one-step reduction to give the compound 30, of which the amino groups were acetylated with acetic anhydride to afford the Pel-structure **31**.



(a) TfOH, DCM, **18**: 70%; **24**: 92%. (b) HF-pyridine, THF, **19**: 98%, **25**: 91%. (c) BnBr, borinic acid-catalyzed, K₂CO₃, KI, CH₃CN, 60 °C, **20**: 96%; **26**: 95%. (d) NapBr, NaH, DMF, **21**: 93%. (e) NIS, acetone, H₂O. (f) 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride, Cs₂CO₃, acetone, **23**: 83% over two steps. (g) MPF, TfOH, DCM, -10 °C, 48 h, **27**: 89%, α:β = 10:1; **29**: 91%, α:β = 10:1. (h) 0.2 M HCl/HFIP, TES, HFIP/DCM, **28**: 73%. (i) H₂, Pd(OH)₂/C, CH₃COOH, THF/H₂O/t-BuOH, **30**: 76%. (k) Ac₂O, NaHCO₃, H₂O, **31**: 86%.

Scheme 3. A: Synthesis of donor 23. B: synthesis of acceptor 26. C: Assembly of Pel fragment 31.

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Conclusion

In conclusion, MPF is here reported for the first time as a moderator to enable the stereoselective construction of α -GlcN₃ linkages. This additive complements previously introduced glycosylation additives, such as DMF and NFM and expands the "nucleophilic additive toolbox", that can be used to match the reactivity of glycosyl donor-acceptor pairs. The applicability of the MPF-mediated glycosylations in oligosaccharide synthesis has been demonstrated by the hand of the assembly of Pel-type oligosaccharides. A linear 10 glucosazide tetrasaccharide was assembled, through highly 11 stereoselective glycosylation reactions, using building 12 blocks, solely equipped with benzyl type (Bn and PMB) hy-13 droxyl protecting groups. A [2+2+2] strategy was developed for the assembly of a (GalN-GlcN)₃ hexasaccharide in 14 which the α -GlcN linkages were constructed in glycosyla-15 tion reactions using MPF as an additive. 16

Experiment section

General experimental procedures

19 All reagents were of commercial grade and used as received. 20 All moisture sensitive reactions were performed under an 21 argon atmosphere. DCM used in the glycosylation reactions 22 was dried with flamed 4Å molecular sieves before being 23 used. Reactions were monitored by TLC analysis with detec-24 tion by UV (254 nm) and where applicable by spraying with 25 20% sulfuric acid in EtOH or with a solution of 26 $(NH_4)_6Mo_7O_{24} \cdot 4H_2O$ (25 g/L) and $(NH_4)_4Ce(SO_4)_4 \cdot 2H_2O$ (10 27 g/L) in 10% sulfuric acid (aq.) followed by charring at 28 ~150 °C. Column chromatography was carried out using sil-29 ica gel (0.040-0.063 mm). Size-exclusion chromatography 30 was carried out using Sephadex LH-20. ¹H and ¹³C spectra 31 were recorded on a Bruker AV 400 and Bruker AV 500 in CDCl₃ or D₂O. Chemical shifts (δ) are given in ppm relative 32 to tetramethylsilane as internal standard (¹H NMR in CDCl₃) 33 or the residual signal of the deuterated solvent. Coupling 34 constants (J) are given in Hz. All ¹³C spectra are proton de-35 coupled. NMR peak assignments were made using COSY and 36 HSQC experiments, where applicable Clean TOCSY, HMBC 37 and GATED experiments were used to further elucidate the 38 structure. The anomeric product ratios were analyzed 39 through integration of proton NMR signals.

40 Procedure A for the glycosylation of secondary alcohols: 41 A mixture of donor (1.0 eq), acceptor (0.7 eq) (donors and 42 acceptors co-evaporated with toluene three times), MPF 43 (16 eq) in dry DCM were stirred over fresh flame-dried mo-44 lecular sieves 3A under nitrogen. The solution was cooled 45 to -78 °C, after which TfOH (1.0 eq) was added. After 30 min, 46 the reaction was stirred at 0 or -10 $^{\circ}$ C until TLC-analysis 47 showed complete conversion of the acceptor. The reaction 48 was quenched with Et₃N, filtered and concentrated *in vacuo*. 49 The products were purified by size exclusion and silica gel 50 column chromatography.

Procedure B for the glycosylation of primary alcohols:

52 A mixture of donor (1.0 eq), acceptor (0.7 eq) (donors and 53 acceptors co-evaporated with toluene three times), Ph₃P=O 54 (6 eq) in dry DCM were stirred over fresh flame-dried mo-55 lecular sieves 3A under nitrogen. Then TMSI (1.0 eq) was 56 added slowly in the mixture. The reaction was stirred at 57 room temperature until TLC-analysis indicated the reaction 58

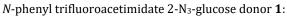
to be complete. The solution was diluted and the reaction quenched with saturated Na₂S₂O₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The products were purified by size exclusion and silica gel column chromatography.

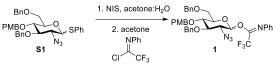
Procedure C for deprotection of the PMB and Nap protecting group:13

The starting material (1 eq) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (2.0 eq) and 0.2M HCl/HFIP (0.1-1eq) were added to the mixture. The reaction stirred until TLCanalysis indicated full consumption of the starting material (15min-2h). Then the mixture was diluted with DCM and the reaction quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The product was purified by silica gel column chromatography.

Experimental Procedures and Characterization Data of Products

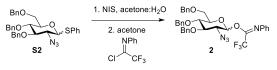
For the synthesis procedure and data of known compounds 915a, S115a, S215b, S315c and S105e see references. We used "a", "b", "c", "d", "e", "f", "g", "h" and "i" to specify the H-1 and C-13 NMR signals of sugar rings from the "reducing" to the "non-reducing" end and "°" to specify the H-1 and C-13 NMR signals of the spacer.





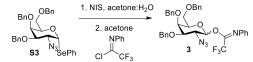
Compound S1 (9.1 g, 15.2 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). N-Iodosuccinimide (NIS) (6.9 g, 30.5 mmol) was added in one portion and the reaction was stirred at room temperature for 2 hours. The solution was diluted with DCM and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered and concentrated *in vacu*o, and the product purified by column chromatography (pentane : ethyl acetate (EA) = 3:1). The lactol (7.2 g, 14.3 mmol) was obtained as colourless syrup. Next, the lactol was dissolved in acetone (150 mL). Cs₂CO₃ (7.0 g, 21.3 mmol) and 2,2,2trifluoro-N-phenylacetimidoyl chloride (3.4 mL, 21.3 mmol) were added to the solution respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by column chromatography (pentane:EA = 40:1-20:1). Compound 1 (8.5 g, 79% over two steps, pentane:EA = 10:1, Rf = 0.45-0.55) was obtained as yellow syrup. IR (neat, cm⁻¹) v 697, 737, 1029, 1082, 1119, 1210, 1251, 1312, 1514, 1720, 2112 (N₃), 2872, 2912. ¹H-NMR (CDCl₃, 500 MHz, 60 °C) δ 7.38-7.20 (m, aromatic *H*), 7.11-7.06 (m, aromatic *H*), 6.82-6.78 (m, aromatic H), 6.37 (bs, 1 H), 5.41 (bs, 1 H), 4.92-4.80 (m), 4.74-4.69 (m), 4.60-4.48 (m), 3.96 (t, J = 10.0 Hz, 1 H), 3.90 (bd, 1 H), 3.77-3.58 (m), 3.43 (t), 3.33 (bs, 1 H). ¹³ C-APT (CDCl₃, 125 MHz, 60[°]C) δ 159.8, 159.8, 143.6, 143.5, 138.3, 138.2, 138.1, 130.3 (aromatic C), 129.7, 128.9, 128.6, 128.6, 128.5, 128.1, 128.0, 1278.0, 127.9, 127.9, 127.8, 124.7, 124.6, 119.6, 114.2, 114.2 (aromatic CH), 96.2 (C-1), 94.4 (C-1), 83.3, 80.5, 77.7, 77.3, 76.4, 75.7, 75.0, 74.8, 73.9, 73.8, 73.7, 68.5, 65.8, 63.5, 55.4. HRMS (ESI) m/z: Calculated for [M-[O(C=NPh)CF₃]+OH+Na]⁺ C₂₈H₃₁O₆N₃Na: 582.21051, found: 582.20943.

Synthesis of *N*-phenyl trifluoroacetimidate 2-N₃-glucose donor **2**:



Compound S2 (8.5 g, 15 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). N-Iodosuccinimide (NIS) (6.7 g, 30 mmol) was added in one portion and the reaction was stirred at room temperature for 2 hours. The solution was diluted with DCM and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered and concentrated *in vacuo*, and the product purified by column chromatography (pentane : ethyl acetate (EA) = 3:1). The lactol (6.1 g, 13 mmol) was obtained as colorless syrup. Next, the lactol was dissolved in acetone (150 mL). Cs₂CO₃ (6.4 g, 19.6 mmol) and 2,2,2trifluoro-*N*-phenylacetimidoyl chloride (3.4 mL, 21.3 mmol) were added to the solution respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered and concentrated *in vacuo*. The product was purified by column chromatography (pentane:EA = 40:1-20:1). Compound 2 (7.3 g, 87%) was obtained as yellow syrup. IR (neat, cm⁻¹) v 694, 734, 1027, 1073, 1116, 1150, 1208, 1312, 1361, 1452, 1490, 1497, 1598, 1717, 2110 (N₃), 2869, 3032. ¹H-NMR (CDCl₃, 500 MHz, 60[°]C) δ 7.52-6.81 (m, aromatic *H*), 6.37 (bs, 1 H, H-1α), 5.43 (bs, 1 H, H-1β), 4.89-4.76 (m, CHH), 4.60-4.48 (m, CHH), 3.98 (t, J = 9.5 Hz, 1 H), 3.91 (bd, 1 H), 3.80-3.59 (m), 3.46 (t), 3.36 (bs, 1 H). 13 C-APT (CDCl₃, 125 MHz, 60[°]C) δ 143.6, 143.5, 138.2, 138.2, 138.1, 138.1, 138.1 (aromatic C), 129.5, 128.9, 128.8, 128.6, 128.6, 128.5, 128.2, 128.1, 128.1, 128.0, 128.0, 127.97, 127.95, 127.91, 127.9, 126.5, 124.7, 124.6, 120.8, 119.6 (aromatic CH), 96.2 (C-1), 94.4 (C-1), 83.3, 80.5, 78.0, 77.6, 76.3, 75.7, 75.7, 75.4, 75.2, 73.9, 73.8, 73.7, 68.5, 65.8, 63.5. HRMS (ESI) m/z: Calculated for [M-[O(C=NPh)CF3]+OH+Na]+C27H29O5N3Na: 498.19994, found: 498.19848.

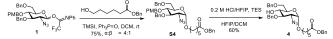
Synthesis of *N*-phenyl trifluoroacetimidate 2-N₃-galactose donor **3**:



Compound **S3** (3.7 g, 6.0 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). N-Iodosuccinimide (NIS) (2.7 g, 12 mmol) was added in one portion and the reaction was stirred at room temperature for 2 hours. The solution was diluted with DCM and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered and concentrated *in vacu*o, and the product purified by column chromatography (pentane:EA = 3:1). The lactol was obtained as colourless syrup. Next, the lactol was dissolved in acetone. Cs₂CO₃ (3.0 g, 9 mmol) and

2,2,2-trifluoro-N-phenylacetimidoyl chloride (1.5 mL, 9 mmol) were added to the solution respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by column chromatography (pentane:EA = 40:1-20:1). Compound 3 (3.3 g, 86%) was obtained as yellow syrup. IR (neat, cm⁻¹) v 695, 734, 751, 986, 1027, 1153, 1316, 1364, 1454, 1490, 1497, 1590, 1717, 2114 (N₃), 2870, 2915. ¹H-NMR (CDCl₃, 400 MHz) δ 7.56-6.79 (m, aromatic *H*), 6.35 (bs, 1 H, H-1), 5.49 (bs, 1 H, H-1), 5.28 (d), 4.90-4.84 (m, CHH), 4.78-4.31 (m), 4.15-3.83 (m), 3.76 (dd), 3.65-3.31 (m). 13 C-APT (CDCl₃, 100 MHz) δ 143.5, 143.4, 138.5, 138.3, 138.3, 138.2, 138.1, 137.7, 137.7, 137.6, 137.6, 137.4, 137.3, 135.2 (aromatic *C*), 129.5, 128.8, 128.7, 128.6, 128.6, 128.6, 128.5, 128.5, 128.43, 128.37, 128.3, 128.19, 128.17, 128.15, 128.14, 128.07, 128.03, 128.00, 127.95, 127.9, 126.48, 124.46, 120.6, 119.4 (aromatic CH), 96.5 (C-1), 92.5 (C-1), 80.9, 80.7, 77.4, 75.1, 74.9, 74.8, 74.7, 74.6, 73.8, 73.67, 73.65, 73.62, 73.5, 72.9, 72.7, 72.6, 72.5, 72.4, 72.3, 72.2, 71.9, 69.7, 69.3, 68.7, 68.3, 68.1, 64.7, 62.2, 60.4, 59.2. HRMS (ESI) m/z: [M+Na]⁺ Calculated for C₂₇H₂₉O₅N₃Na: 669.22953, found: 669.22913.

Synthesis of monosaccharide 4:



The reaction was carried out according to the standard procedure B. A mixture of donor 1 (1.0 g, 1.5 mmol), benzyl 6hydroxyhexanoate (520 mg) (donors and acceptors coevaporated with toluene three times), $Ph_3P=0$ (2.6 g, 9.3 mmol) in dry DCM (15 mL) were stirred over fresh flamedried molecular sieves 3A under nitrogen. Then TMSI (222 μL, 1.5 mmol) was added slowly in the mixture. The reaction was stirred at room temperature until TLC-analysis indicated the reaction to be complete. The solution was diluted and the reaction quenched with saturated Na₂S₂O₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The products were purified by silica gel column chromatography (pentane:EA = 8:1, Rf = 0.63). Compound **S4** (800 mg, 75% yield, $\alpha:\beta = 5:1$) was obtained as a colorless syrup. IR (neat, cm⁻¹) v 697, 736, 1002, 1029, 1037, 1075, 1150, 1248, 1358, 1454, 1611, 1733 (C=O), 2105 (N₃), 2866, 2933. ¹H-NMR (CDCl₃, 400 MHz) δ 7.40-7.21 (m, 15 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.09 (s, 2 H, PhCH₂), 4.90 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (s, 2 H, PhCH₂),4.71 (d, J = 10.4 Hz, 1 H, CHH), 4.63 (d, J = 12.4 Hz, 1 H, CH*H*), 4.49 (d, *J* = 12.4 Hz, 1 H, C*H*H), 4.43 (d, *J* = 10.4 Hz, 1 H, CHH), 3.975 (t, t, J = 9.6 Hz, 1 H, H-3a), 3.79-3.63 (m, 5 H, H-2a, H-4a, H-5a, H-6a, H-1^o_a), 3.47-3.37 (m, 1 H, H-1^o_b), 3.33 (dd, 1 H, I_1 = 10.0 Hz, I_2 = 2.0 Hz, H-2a), 2.36 (t, I = 7.6 Hz, 2H, H-5°), 1.70-1.58 (m, 4 H, H-2°, H-4°), 1.43-1.36 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.4 (C=O), 159.4, 138.1, 137.9, 130.1 (aromatic *C*), 129.6, 128.6, 128.5, 128.5, 128.2, 127.99, 127.96, 127.9, 127.8, 113.9 (aromatic CH), 97.9 (C-1a), 80.2 (C-3a), 78.0 (C-4a), 75.3, 74.8, 73.6 (CH₂), 70.7 (C-5a), 68.3 (C-6a), 68.0 (C-1^o), 66.1 (PhCH₂), 63.4 (C-2a), 55.3 (OCH₃), 34.2 (C-5^o), 29.1 (C-2^o), 25.7 (C-3^o), 24.7 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₄₁H₅₁N₄O₈: 727.37014, found: 727.37015.

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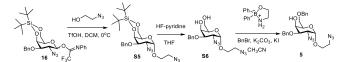
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Then the reaction was carried out according to the standard procedure C. The starting material S4 (700 mg, 0.99 m mol) 2 was dissolved in DCM:HFIP (1:1, 0.1 M). TES (314 mL) and 0.2M HCl/HFIP (0.5 mL) were added to the mixture. The reaction stirred until TLC-analysis indicated full consumption of the starting material (15min). Then the mixture was diluted with DCM and the reaction quenched with saturated 6 NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO4, filtered and concen-8 trated in vacuo. The product was purified by silica gel col-9 umn chromatography (pentane:EA = 4:1, Rf = 0.34). Com-10 pound 4 (350 mg, 60% yield) was obtained as a colorless 11 syrup. [α]_{D²⁰} +59.3 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 737, 12 1050, 1147, 1455, 1734 (C=O), 2105 (N₃), 2866, 2926, 3478. 13 ¹H-NMR (CDCl₃, 400 MHz) δ 7.41-7.23 (m, 15 H, aromatic *H*), 14 5.10 (s, 2 H, PhC H_2), 4.90 (d, J = 11.2 Hz, 1 H, CHH), 4.87 (d, 15 J = 3.6 Hz, 1 H, H-1a), 4.81 (d, J = 11.2 Hz, 1 H, CHH), 4.59 (d, / = 12.0 Hz, 1 H, CHH), 4.53 (d, / = 12.0 Hz, 1 H, CHH), 3.86-16 17 3.64 (m, 6 H, H-2a, H-3a, H-4a, H-5a, H-6a, H-1^o_a), 3.47-3.41 (m, 1 H, H-1^{\circ}_b), 3.25 (dd, 1 H, J_1 = 10.0 Hz, J_2 = 2.0 Hz, H-2a), 18 2.37 (t, J = 7.6 Hz, 2H, H-5°), 1.72-1.61 (m, 4 H, H-2°, H-4°), 19 1.47-1.37 (m, 2 H, H-3²). ¹³C-APT (CDCl₃, 100 MHz) δ 173.6 20 (C=0), 138.2, 137.9, 136.1 (aromatic C), 128.7, 128.6, 128.5, 21 128.3, 128.3, 128.1, 128.05, 127.9, 127.7, 127.5 (aromatic 22 CH), 98.0 (C-1a), 79.8 (C-3a), 75.0 (C-6a), 73.7 (CH₂), 72.2 23 (c-4a), 70.2 (c-5a), 69.8 (PhCH₂), 68.1 (C-1^o), 66.2 (PhCH₂), 24 62.8 (C-2a), 34.2 (C-5^o), 29.1 (C-2^o), 25.7 (C-3^o), 24.7 (C-4^o). 25 HRMS (ESI) m/z: $[M+NH_4]^+$ Calculated for $C_{33}H_{43}O_7N_4$: 26 607.31263, found: 607.31238. 27

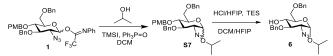
Synthesis of acceptor 5



Donor 16 (620 mg, 1.0 mmol) and 2-azidoethanol (178 mg, 2.0 mmol) were dissolved in DCM, cooled to 0 °C and TfOH (15 μ L, 0.1 mmol) was added. The reaction was stirred at 0 $^{\circ}$ C until TLC-analysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completed checking by TLC, filtered and concentrated in vacuo. Compound **S5** (370 mg, 73%) was obtained with full α -selectivity. Then compound **S5** was dissolved in THF. HF-pyridine was added to the solution. After TLC-analysis showed complete consumption of the starting material, the reaction was quenched with saturated NaHCO₃. The mixture was diluted with ethyl acetate, washed with H₂O and brine, dried with anhydrous MgSO₄, filtered, concentrated in vacuo. Crude compound S6, K₂CO₃, KI, and borinic acid-catalyzed were mixed in CH₃CN, and then BnBr was added in the solution. The reaction was stirred at 60 °C until TLC-analysis showed complete conversion of the starting material. The reaction was quenched with H₂O after completed checking by TLC, filtered and concentrated in vacuo, purified by column chromatography (pentane:EA = 5:1). Compound 5 (280 mg, 84%) yield over two steps) was obtained as colorless syrup. $[\alpha]_D^{20}$ +89.9 (c=1, CHCl₃). IR (neat, cm⁻¹) v 698, 738, 1052, 1096, 1146, 1454, 2108 (N₃), 2873, 2923, 3483. ¹H-NMR (CDCl₃, 500 MHz) δ 7.40-7.28 (m, 10 H, aromatic H), 4.95 (d, I = 3.5Hz, 1 H, H-1a), 4.71 (d, J = 11.5 Hz, 1 H, CHH), 4.68 (d, J = 11.5 Hz, 1 H, CHH), 4.60 (d, J = 12.0 Hz, 1 H, CHH), 4.57 (d, J = 12.0

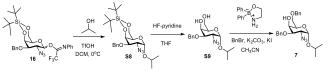
Hz, 1 H, CHH), 4.12 (t, I = 1.5 Hz, 1 H, H-4a), 3.98 (t, I = 6.0Hz, 1 H, H-5a), 3.93 (dd, 1 H, $I_1 = 10.5$ Hz, $I_2 = 3.0$ Hz, H-3a), 3.90-3.86 (m, 1 H, H-1ºa), 3.77-3.63 (m, 4 H, H-2a, H-6ª, H- 1°_{b}), 3.57-3.52 (m, 1 H, H- 2°_{a}), 3.37-3.33 (m, 1 H, H- 2°_{b}), 2.61 (bt, 1 H, OH), 1.21-1.18 (bt, 6 H, 2 CH₃). ¹³C-APT (CDCl₃, 125 MHz) δ 137.9, 137.3 (aromatic C), 128.8, 128.6, 128.4, 128.2, 127.9, 127.8 (aromatic CH), 98.5 (C-1a), 76.0 (C-3a), 73.8, 72.1 (CH₂), 69.6 (C-6a), 69.2 (C-5a), 67.2 (C-1^o), 66.8 (C-4a), 59.0 (C-2a), 50.8 (C-2^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₂₂H₃₀O₅N₇: 472.23029, found: 472.23003.

Synthesis of acceptor 6:



Donor 1 (820 mg, 1.2 mmol), isopropanol (200 µL, 2.6 mmol) and Ph₃P=O (2 g) were dissolved in DCM (12 mL), and TMSI (173 µL) was added at room temperature. The reaction was stirred at rt until TLC-analysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completed checking by TLC, filtered and concentrated in *vacuo*, purified by column chromatography. Compound **S7** was obtained with α : β = 5:1. Then compound **S7** was dissolved in DCM/HFIP (1.5 mL: 1.5 mL). TES (380 µL) and 0.2M HCl/HFIP (600 µL) were added to the mixture. The reaction stirred until TLC-analysis indicated full consumption of the starting material (30min). Then the mixture was diluted with DCM and the reaction quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:EA = 5:1). Compound 6 (240 mg, 47% yield over two steps) was obtained as colorless syrup. $[\alpha]_D^{20}$ +83.4 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 735, 1029, 1047, 1120, 1454, 2105 (N₃), 2920, 2974, 3476. ¹H-NMR (CDCl₃, 400 MHz) δ 7.40-7.20 (m, 10 H, aromatic *H*), 4.97 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, J = 11.2 Hz, 1 H, CHH), 4.78 (d, J = 11.2 Hz, 1 H, CHH), 4.57 (d, J = 12.0 Hz, 1 H, CHH), 4.50 (d, J = 12.0 Hz, 1 H, CHH), 3.93-3.82 (m, 3 H, H-3a, H-5a, H-1°), 3.73-3.61 (m, 3 H, H-4a, H-6a), 3.18 (dd, 1 H, J₁ = 10.0 Hz, J₂ = 3.6 Hz, H-2a), 2.76 (bs, 1 H, 0H), 1.23, (d, J = 8.4 Hz, 3 H, CH₃), 1.21 (d, J = 8.4 Hz, 3 H, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 138.2, 137.8 (aromatic C), 128.5, 128.4, 128.0, 127.9, 127.7, 127.6 (aromatic CH), 96.4 (C-1a), 79.6 (C-3a), 74.9, 73.6 (CH2), 72.1 (C-4a), 70.8 (C-1º), 70.1 (C-5a), 69.7 (C-6a), 62.5 (C-2a), 23.2 (CH₃), 21.5 (CH₃). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₂₃H₃₃O₅N₄: 445.24455, found: 445.24441.

Synthesis of acceptor 7:



Donor 16 (2.77 g, 4.6 mmol) and isopropanol were dissolved in DCM (40 mL), cooled to 0 °C and TfOH (40 µL) was added. The reaction was stirred at 0 $\,^\circ\!\mathrm{C}\,$ until TLCanalysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completed checking by

TLC, filtered and concentrated *in vacuo*. Compound **S8** was obtained with full α -selectivity. Then compound S8 was dissolved in THF (20 mL). HF-pyridine (1 mL) was added to the solution. After TLC-analysis showed complete consumption of the starting material, the reaction was quenched with saturated NaHCO₃. The mixture was diluted with ethyl acetate, washed with H₂O and brine, dried with anhydrous MgSO₄, filtered, concentrated in vacuo, purified by column chromatography (pentane:EA = 3:1). Compound S9 (1.45 g) was obtained with 94% yield over two steps. Then compound S9 (665 mg, 1.97 mmol), K₂CO₃ (293 mg), KI (327 mg), and bo-10 rinic acid-catalyzed (44 mg) were mixed in CH₃CN (20 mL), 11 and then BnBr was added in the solution. The reaction was 12 stirred at 60 °C in oil bath until TLC-analysis showed com-13 plete conversion of the starting material. The reaction was 14 quenched with H₂O after completed checking by TLC, fil-15 tered and concentrated in vacuo, purified by column chro-16 matography (pentane: EA = 5:1). Compound 7 (745 mg, 80%) 17 yield) was obtained as colorless syrup. $[\alpha]_{D^{20}}$ +102.7 (c=1, CHCl₃). IR (neat, cm⁻¹) v 698, 737, 1052, 1454, 2108 (N₃), 18 2892, 2926. 2972. ¹H-NMR (CDCl₃, 400 MHz) δ 7.42-7.27 (m, 19 10 H, aromatic H), 5.02 (d, I = 3.6 Hz, 1 H, H-1a), 4.71 (bs, 2 20 H, PhC H_2), 4.58 (bs, 2 H, PhC H_2), 4.15 (t, I = 1.6 Hz, 1 H, H-21 4a), 4.01 (bt, 1 H, H-5a), 3.95-3.89 (m, 2 H, H-3a, H-1º), 3.76 22 $(dd, 1 H, I_1 = 10.0 Hz, I_2 = 6.0 Hz, H-6a_a), 3.70-3.62 (m, 2 H, 10.0 Hz, I_2 = 0.0 Hz, H-6a_a), 3.70-3.62 (m, 2 Hz, I_2 = 0.0 Hz, I_2 = 0$ 23 $H-6a_b$, H-2a), 2.60 (bs, 1 H, OH), 1.23 (d, 3 H, I = 10.4 Hz, CH_3), 24 1.21 (d, 3 H, J = 10.4 Hz, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 25 138.0, 137.5 (aromatic C), 128.8, 128.6, 128.3, 128.1, 127.9, 26 127.8 (aromatic CH), 96.7 (C-1a), 76.1 (C-3a), 73.8, 72.0 27 (CH₂), 70.9 (C-1^o), 69.6 (C-6a), 68.7 (C-5a), 66.8 (C-4a), 59.0 28 (C-2a), 23.3 (CH₃), 21.6 (CH₃). HRMS (ESI) m/z: [M+NH₄]⁺ 29 Calculated for C₂₃H₃₃O₅N₄: 445.24455, found: 445.24455.

30 Synthesis of disaccharide 8: The reaction was carried out 31 according to the standard procedure A. A mixture of donor 32 1 (320 mg, 0.47 mmol), acceptor 4 (185 mg, 0.31 mmol) 33 (donors and acceptors co-evaporated with toluene three 34 times), MPF (610 µL) in dry DCM (3 mL) were stirred over 35 fresh flame-dried molecular sieves 3A under nitrogen. The 36 solution was cooled to -78 $^{\circ}$ C, after which TfOH (42 μ L) was 37 added. After 30 min, the reaction was stirred at -10 $^\circ \! \mathbb{C}$ until 38 TLC-analysis showed complete conversion of the acceptor. 39 The reaction was quenched with Et₃N, filtered and concen-40 trated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **8** (304 mg, 88% yield, α : β = 41 15:1, PE:EA = 4:1, Rf = 0.51) was obtained as a colorless 42 syrup. IR (neat, cm⁻¹) v 697, 736, 1027, 1147, 1249, 1358, 43 1454, 1514, 1734 (C=O), 2103 (N₃), 2866, 2928. ¹H-NMR 44 $(CDCl_3, 400 \text{ MHz}) \delta 7.39-7.21 \text{ (m, 25 H, aromatic H)}, 7.00 \text{ (bd,}$ 45 2 H, aromatic *H*), 6.79 (bd, 2 H, aromatic *H*), 5.66 (d, *J* = 4.0 46 Hz, 1 H, H-1b), 5.11 (s, 2 H, PhCH₂), 4.98(d, J = 10.4 Hz, 1 H, 47 CHH), 4.93 (d, / = 4.0 Hz, 1 H, H-1a), 4.89-4.82 (m, 3 H, 3 48 CHH), 4.66 (d, J = 10.0 Hz, 1 H, CHH), 4.54-4.47 (m, 3 H, 3 49 CHH), 4.37 (d, / = 10.4 Hz, 1 H, CHH), 4.23 (d, / = 10.4 Hz, 1 50 H, CHH), 4.07 (t, J = 9.2 Hz, 1 H, H-3a), 3.98 (t, J = 9.2 Hz, 1 H, 51 H-4a), 3.87-3.61 (m, 10 H, H-3b, H-4b, H-5a, H-5b, H-6a, H-52 6b_a, OCH₃), 3.54-3.44 (m, 2 H, H-6b_b, H-1^o_a), 3.35-3.29 (m, 3 53 H, H-2a, H-2b, H-1^ob), 2.38 (t, J = 7.6 Hz, 2H, H-5^o), 1.73-1.63 (m, 4 H, H-2°, H-4°), 1.46-1.38 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 54 100 MHz) & 173.5 (C=0), 159.4, 138.2, 138.0, 137.84, 137.82, 55 136.2, 130.2 (aromatic C), 129.7, 128.7, 128.5, 128.4, 128.2, 56 128.1, 128.0, 127.9, 127.84, 127.78, 127.6, 127.4, 113.9 57

(aromatic CH), 97.8 (C-1b), 97.7 (C-1a), 80.9 (C-3a), 80.3 (C-3b), 77.8 (C-4b), 75.5, 74.7, 74.5, 73.6, 73.5 (PhCH₂), 73.4 (c-4a), 71.6 (c-5b), 70.2 (C-5a), 69.1 (C-6a), 68.2 (C-6b), 67.9 (C-1^o), 66.2 (PhCH₂), 63.8 (C-2), 63.4 (C-2), 55.4 (OCH₃), 34.2 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₆₁H₇₂N₇O₁₂: 1094.52335, found: 1094.52388.

Synthesis of disaccharide 9: The reaction was carried out according to the standard procedure A. A mixture of donor 2 (146 mg, 0.22 mmol), acceptor 5 (50 mg, 0.11 mmol) (donors and acceptors co-evaporated with toluene three times), MPF (216 µL) in dry DCM were stirred over fresh flamedried molecular sieves 3A under nitrogen. The solution was cooled to -78 $^{\circ}$ C, after which TfOH (19 μ L) was added. After 30 min, the reaction was stirred at -10 $^{\circ}$ C until TLC-analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound 9 (86 mg, 87%, α : β = 10:1) was obtained as a colorless syrup. IR (neat, cm⁻¹) v 697, 736, 1027, 1046, 1093, 1127, 1150, 1259, 1359, 1454, 2105 (N₃), 2869, 2923. ¹H-NMR (CDCl₃, 400 MHz) δ 7.40-7.05 (m, 25 H, aromatic H), 4.99 (bt, 2 H, H-1a and H-1b), 4.90-4.76 (m, 3 H, 3 CHH), 4.69 (d, I = 10.8 Hz, 1 H, CHH), 4.63 (d, I = 10.8 Hz, 1 H, CHH),4.59-4.53 (m, 2 H, 2 CHH), 4.39 (bt, 2 H, 2 CHH), 4.31 (d, J = 2.4 Hz, 1 H), 4.13-3.49 (m, 13 H), 3.39-3.29 (m, 2 H), 3.22 $(dd, J_1 = 12.4 Hz, J_2 = 2.0 Hz, 1 H), 2.96 (dd, J_1 = 10.8 Hz, J_2 =$ 2.0 Hz, 1 H), 4.48 (d, J₁ = 10.8 Hz, J₂ = 1.6 Hz, 1 H). ¹³C-APT (CDCl₃, 100 MHz) δ 138.1, 137.8, 137.7, 137.5 (aromatic *C*), 128.6, 128.5, 128.4, 128.4, 128.2, 128.07, 128.06, 127.9, 127.82, 127.78, 127.75, 127.7, 127.2 (aromatic CH), 98.9 (C-1), 98.5 (C-1), 80.2, 78.1, 75.6, 75.4, 74.9, 73.7, 73.3, 73.3, 72.0, 70.9, 69.6, 67.3, 67.3, 67.0, 64.0, 59.4, 50.7. HRMS (ESI) m/z: [M+NH₄]⁺: Calculated for C₄₉H₅₇O₉N₁₀: 929.43045, found: 929.43039.

Synthesis of disaccharide 10: The reaction was carried out according to the standard procedure A. A mixture of donor 3 (77 mg, 0.12 mmol), acceptor 6 (34 mg, 0.08 mmol) (donors and acceptors co-evaporated with toluene three times), MPF (156 µL) in dry DCM were stirred over fresh flamedried molecular sieves 3A under nitrogen. The solution was cooled to -78 $^{\circ}$ C, after which TfOH (8 μ L) was added. After 30 min, the reaction was stirred at -10 $^{\circ}$ C until TLC-analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **10** (56 mg, 88% yield, α : β = 8:1) was obtained as a colorless syrup. IR (neat, cm^{-1}) v 697, 737, 1050, 1097, 1122, 1258, 1454, 2108 (N₃), 2869, 2928. ¹H-NMR (CDCl₃, 400 MHz) δ 7.41-7.19 (m, 25 H, aromatic *H*), 5.64 (d, I = 3.6 Hz, 1 H, H-1a), 5.03 (d, I = 3.6 Hz, 1 H, H-1b), 4.96 (d, *J* = 10.0 Hz, 1 H, CHH), 4.91 (d, *J* = 10.0 Hz, 1 H, CHH), 4.81 (d, J = 11.2 Hz, 1 H, CHH), 4.67 (d, J = 11.2 Hz, 1 H, CHH), 4.61 (d, J = 11.2 Hz, 1 H, CHH), 4.56 (d, J = 12.4 Hz, 1 H, CHH), 4.48 (d, / = 11.2 Hz, 1 H, CHH), 4.44 (d, / = 12.4 Hz, 1 H, CHH), 4.29 (d, / = 11.6 Hz, 1 H, CHH), 4.22 (d, / = 11.6 Hz, 1 H, CHH), 4.07 (dd, J = 8.0, 10.0 Hz, 1 H, H-3b), 3.98-3.78 (m, 7 H), 3.72-3.63 (m, 2 H, H-6), 3.48-3.37 (m, 2 H, H-6), 3.29 (dd, J = 3.6, 10.0 Hz, 1H, H-2b), 1.28 (d, J = 6.4 Hz, 1 H, CH₃), 1.24 (d, J = 6.4

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Hz, 1 H, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 138.4, 138.2, 137.9, 137.6 (aromatic *C*), 128.62, 128.57, 128.5, 128.42, 128.37, 128.3, 128.0, 127.92, 127.89, 127.85, 127.8, 127.5, 127.4 (aromatic *C*H), 98.0 (C-1a), 96.2 (C-1b), 80.8 (C-3b), 77.6 (C-3a), 74.9, 74.5 (PhCH₂), 74.0 (C-2a), 73.6, 73.2 (PhCH₂), 72.9 (C-4b), 72.2 (PhCH₂), 71.1 (C-4a), 70.2 (C-5b), 70.1 (C-5a), 69.5 (C-6), 68.5 (C-6), 63.6 (C-2b), 59.8 (C-1°), 23.4 (*C*H₃), 21.7 (*C*H₃). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for $C_{50}H_{60}O_9N_7$: 902.44470, found: 902.44467.

9 Synthesis of disaccharide 11: The reaction was carried out 10 according to the standard procedure A. A mixture of donor 11 3 (77 mg, 0.12 mmol), acceptor 7 (34 mg, 0.08 mmol) (do-12 nors and acceptors co-evaporated with toluene three times), 13 MPF (156 µL) in dry DCM were stirred over fresh flame-14 dried molecular sieves 3A under nitrogen. The solution was 15 cooled to -78 °C, after which TfOH (8 µL) was added. After 16 30 min, the reaction was stirred at -10 °C until TLC-analysis 17 showed complete conversion of the acceptor. The reaction 18 was guenched with Et₃N, filtered and concentrated *in vacuo*. 19 The product was purified by size exclusion (DCM:MeOH = 20 1:1). Compound **11** (62 mg, 80% yield, α : β = 4:1) was ob-21 tained as a colorless syrup. $[\alpha]_D^{20}$ +85.8 (c=1, CHCl₃). IR 22 (neat, cm⁻¹) v 697, 736, 986, 1037, 1117, 1209, 1261, 1454, 23 2106 (N₃), 2870, 2925. ¹H-NMR (CDCl₃, 400 MHz) δ 7.43-24 7.12 (m, 25 H, aromatic H), 5.05 (d, J = 3.6 Hz, 1 H, H-1a), 25 4.98 (d, J = 3.6 Hz, 1 H, H-1b), 4.88 (d, J = 12.0 Hz, 1 H, CHH), 4.80 (d, I = 10.8 Hz, 1 H, CHH), 4.72 (d, I = 11.2 Hz, 1 H, CHH),26 4.63 (d, J = 11.2 Hz, 1 H, CHH), 4.54 (bd, 3 H, 3 CHH), 4.47 (d, 27 *J* = 10.8 Hz, 1 H, CH*H*), 4.36 (dd, *J* = 5.2, 9.2 Hz, 1 H, H-5a), 28 4.28 (d, / = 2.8 Hz, 1 H, H-4a), 4.10 (s, 1 H, H-4b), 4.03-3.85 29 (m, 8 H, H-6b_a, H-5b, H-3b, H-3a, H-2b, H-2a, H-1^o), 3.60 (dd, 30 *J* = 3.6, 11.2 Hz, 1H, H-2a), 3.56-3.49 (m, 2 H, H-6b_b, H-6a_a), 31 $3.14-3.09 (m, 2 H, H-6a_b), 1.20 (d, I = 6.0 Hz, 1 H, CH_3), 1.19$ 32 (d, J = 6.0 Hz, 1 H, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 138.7, 33 138.0, 137.7, 137.6 (aromatic C), 128.64, 128.58, 128.55, 34 128.4, 128.3, 128.2, 128.12, 128.09, 128.0, 127.90, 127.87, 35 127.75, 127.74, 127.6, 127.3 (aromatic CH), 98.2 (C-1b), 36 96.8 (C-1a), 77.4 (C-3b), 75.9 (C-3a), 75.0, 73.7, 73.2 37 (PhCH2), 73.0 (C-4b), 72.9 (C-4a), 71.9, 71.9 (PhCH2), 71.0 (C-1º), 69.4 (C-5b), 69.2 (C-5a), 67.7 (C-6a), 67.2 (C-6b), 38 60.4 (C-2b), 59.5 (C-2a), 23.4 (CH₃), 21.7 (CH₃). HRMS (ESI) 39 m/z: [M+NH₄]⁺ Calculated for C₅₀H₆₀O₉N₇: 902.44470, found: 40 902.44482. 41

43 Synthesis of disaccharide 12: The reaction was carried out according to the standard procedure C. Compound 8 (200 44 mg, 0.18 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES 45 (60 μ L) and 0.2M HCl/HFIP (100 μ L) were added to the mix-46 ture. The reaction stirred until TLC-analysis indicated full 47 consumption of the starting material (30 min). Then the 48 mixture was diluted with DCM and the reaction quenched 49 with saturated NaHCO₃. The organic phase was washed 50 with water and brine, dried with anhydrous MgSO₄, filtered 51 and concentrated in vacuo. The product was purified by sil-52 ica gel column chromatography (pentane:EA = 5:1, Rf = 53 0.22). Compound 12 (152 mg, 88% yield) was obtained as a 54 colorless syrup. $[\alpha]_D^{20}$ +62.9 (c=1, CHCl₃). IR (neat, cm⁻¹) v 55 697, 736, 1029, 1043, 1146, 1261, 1454, 1734 (C=O), 2105 56 (N₃), 2868, 2926, 3491. ¹H-NMR (CDCl₃, 400 MHz) δ 7.42-57 7.20 (m, 25 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-1b),

5.11 (s, 2 H, PhC H_2), 4.98 (d, I = 10.4 Hz, 1 H, CHH), 4.93 (d, / = 3.6 Hz, 1 H, H-1a), 4.89-4.82 (m, 3 H, 3 CHH), 4.55 (d, / = 12.0 Hz, 1 H, CHH), 4.51 (d, J = 12.0 Hz, 1 H, CHH), 4.08 (dd, *J*₁ = 8.8 Hz, *J*₂ = 10.0 Hz, 1 H, H-3a), 3.99 (t, *J* = 8.8 Hz, 1 H, H-4a), 3.86-3.65 (m, 7 H, H-3b, H-4b, H-5a, H-5b, H-6b, H-6aa), 3.53-3.44 (m, 2 H, H-6ab, H-1ºa), 3.40-3.33 (m, 2 H, H-2a, H- 1°_{b}), 3.24 (dd, J_{1} = 3.6 Hz, J_{2} = 10.0 Hz, 1 H, H-2b), 2.68 (bs, 1 H, OH), 2.38 (t, J = 7.6 Hz, 2H, H-5°), 1.73-1.64 (m, 4 H, H-2°, H-4^o), 1.47-1.39 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.56 (C=0), 138.23, 138.17, 137.8, 137.7, 136.2 (aromatic C), 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.83, 127.79, 127.6, 127.4 (aromatic CH), 97.7 (C-1a), 97.6 (C-1b), 80.9 (C-3a), 79.7 (C-3b), 75.2, 74.5, 73.7, 73.4 (PhCH₂), 73.1 (C-4b), 72.8 (c-4a), 70.6 (c-5b), 70.2 (C-5a), 69.9 (C-6a), 69.0 (C-6b), 68.2 (C-1^o), 66.3 (PhCH₂), 63.8 (C-2a), 62.8 (C-2b), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₅₃H₆₄N₇O₁₁: 974.46583, found: 974.46576.

Synthesis of trisaccharide 13: The reaction was carried out according to the standard procedure A. A mixture of donor **1** (160 mg, 0.24 mmol), acceptor **12** (150 mg, 0.16 mmol) (donors and acceptors co-evaporated with toluene three times), MPF (307 mL) in dry DCM (1.5 mL) were stirred over fresh flame-dried molecular sieves 3A under nitrogen. The solution was cooled to -78 $^{\circ}$ C, after which TfOH (300 μ L) was added. After 30 min, the reaction was stirred at -10 $\,^\circ\!\!\mathbb{C}$ until TLC-analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound 13 (186 mg, 81% yield, $\alpha:\beta > 19:1$, PE:EA = 4:1, Rf = 0.40) was obtained as a colorless syrup. $[\alpha]_D^{20}$ +75.8 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 736, 1029, 1147, 1249, 1359, 1454, 1514, 1734 (C=O), 2106 (N₃), 2866, 2932. ¹H-NMR (CDCl₃, 400 MHz) δ 7.39-7.21 (m, 35 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic *H*), 5.69 (d, *J* = 3.6 Hz, 1 H, H-1), 5.67 (d, *J* = 3.6 Hz, 1 H, H-1), 5.11 (s, 2 H, PhCH₂), 5.02-4.82 (m, 7 H, 6 CHH, H-1a), 4.66 (d, J = 10.0 Hz, 1 H, CHH), 4.56-4.46 (m, 3 H, 3 CHH), 4.39-4.33 (m, 2 H, 2 CHH), 4.26 (d, J = 12.0 Hz, 1 H, CHH), 4.18 (d, J = 12.0 Hz, 1 H, CHH), 4.14-3.98 (m, 4 H), 3.90-3.59 (m, 11 H), 3.56-3.44 (m, 3 H), 3.37-3.24 (m, 3 H), 2.38 (t, / = 7.6 Hz, 2H, H-5°), 1.73-1.63 (m, 4 H, H-2°, H-4°), 1.47-1.39 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=O), 159.4, 138.3, 138.2, 138.0, 137.8, 137.7, 137.6, 136.1, 130.3 (aromatic C), 129.6, 128.6, 128.57, 128.55, 128.4, 128.3, 128.26, 128.1, 127.9, 127.86, 127.8, 127.7, 127.6, 127.5, 127.4, 127.3, 113.8 (aromatic CH), 97.8, 97.7, 97.4 (C-1a, 1b and 1c), 81.0, 80.7, 79.9 (C-3a, 3b and 3c), 77.7 (C-4c), 75.3, 74.7, 74.6, 74.2, 73.6, 73.5 (PhCH₂), 73.0, 72.5 (C-4a and 4b), 71.5, 71.1, 70.2 (c-5a, 5b and 5c), 68.9, 68.7 (2 C-6), 68.2 (C-1^o), 67.7 (C-6), 66.2 (PhCH₂), 63.9, 63.6, 63.1 (C-2a, 2b and 2c), 55.3 (OCH₃), 34.2 (C-5^o), 29.1 (C-2^o), 25.7 (C-3^o), 24.7 (C-4º). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₈₁H₉₃N₁₀O₁₆: 1461.67655, found: 1461.67594.

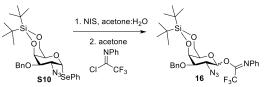
Synthesis of trisaccharide acceptor **14**: The reaction was carried out according to the standard procedure C. The starting material **13** (320 mg, 0.22 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (71 μ L) and 0.2M HCl/HFIP (110 μ L) were added to the mixture. The reaction stirred

until TLC-analysis indicated full consumption of the starting material (15min). Then the mixture was diluted with DCM and the reaction quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The product was purified by silica gel column chromatography (pentane:EA = 4:1). Compound 14 (230 mg, 78% yield) was obtained as a colorless syrup. $[\alpha]_{D^{20}}$ +51.0 (c=3mg/mL, CHCl₃). IR (neat, cm⁻¹) v 697, 737, 1028, 1148, 1454, 1736 (C=O), 2106 (N₃), 2866, 2926. ¹H-NMR (CDCl₃, 400 MHz) δ 7.42-7.17 (m, 35 H, aromatic H), 5.67-5.65 (m, 2 H, H-1b and 10 H-1c), 5.12 (s, 2 H, PhCH2), 5.01-4.85 (m, 7 H, 6 CHH, H-1a), 11 4.56 (d, J = 12.0 Hz, 1 H, CHH), 4.50 (d, J = 12.0 Hz, 1 H, CHH), 12 4.37-4.32 (m, 3 H, 3 CHH), 4.22 (d, J = 12.0 Hz, 1 H, CHH), 13 4.14-3.99 (m, 4 H), 3.87-3.62 (m, 8 H), 3.56-3.43 (m, 3 H), 14 3.37-3.30 (m, 4 H), 3.18 (dd, J₁ = 3.6 Hz, J₂ = 10.0 Hz, 1 H, H-15 2c), 2.76 (bs, 1 H, OH), 2.39 (t, J = 7.6 Hz, 2H, H-5°), 1.74-1.64 16 (m, 4 H, H-2^o, H-4^o), 1.47-1.41 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 17 100 MHz) δ 173.6 (*C*=0), 138.3, 138.20, 138.17, 137.8, 137.6, 137.5, 136.2 (aromatic C), 138.7, 128.7, 128.6, 128.55, 128.5, 18 128.4, 128.3, 128.13, 128.09, 128.0, 127.95, 127.9, 127.8, 19 127.7, 127.5, 127.48, 127.3 (aromatic CH), 97.8, 97.7, 97.4 20 (C-1a, 1b and 1c), 81.1, 80.8, 79.1 (C-3a, 3b and 3c), 75.0, 21 74.6, 74.3, 73.7, 73.5, 73.4 (PhCH2), 73.0, 72.9, 72.3 (C-4a, 4b 22 and 4c), 71.1, 70.3, 70.2 (c-5a, 5b and 5c), 70.0, 68.9, 68.6 23 (C-6a, 6b and 6c), 68.3 (C-1º), 66.26 (PhCH2), 63.9, 63.7, 24 62.5 (C-2a, 2b and 2c), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 25 24.8 (C-4^{\circ}). HRMS (ESI) m/z: Calculated for C₇₃H₈₅N₁₀O₁₅: 26 1341.61904. found: 1341.61923. 27

28 Synthesis of tetrasaccharide 15: The reaction was carried 29 out according to the standard procedure A. A mixture of do-30 nor **1** (40 mg, 0.06 mmol), acceptor **14** (35 mg, 0.03 mmol) 31 (donors and acceptors co-evaporated with toluene three 32 times), MPF (52 µL) in dry DCM (0.3 mL) were stirred over 33 fresh flame-dried molecular sieves 3A under nitrogen. The 34 solution was cooled to -78 $^{\circ}$ C, after which TfOH (5 μ L) was 35 added. After 30 min, the reaction was stirred at -10 $^\circ \!\!\! \mathbb{C}$ until 36 TLC-analysis showed complete conversion of the acceptor. 37 The reaction was quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by size exclusion 38 (DCM:MeOH = 1:1). Compound **15** (43 mg, 87% yield, α : β > 39 20:1) was obtained as a colorless syrup. $[\alpha]_D^{20}$ +94.8 (c=1, 40 CHCl₃). IR (neat, cm⁻¹) v 697, 737, 1029, 1148, 1251, 1359, 41 1454, 1514, 1735 (C=O), 2106 (N₃), 2868, 2928. ¹H-NMR 42 (CDCl₃, 400 MHz) 87.42-7.15 (m, 45 H, aromatic H), 7.00 (bd, 43 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.70-5.67 (m, 3 44 H, H-1b, 1c, 1d), 5.11 (s, 2 H, PhCH₂), 5.02-4.87 (m, 8 H, 7 45 CHH, H-1a), 4.81 (d, J = 10.4 Hz, 1 H, CHH), 4.66 (d, J = 10.4 46 Hz, 1 H, CHH), 4.54 (s, 2 H, PhCH₂), 4.65 (d, J = 12.0 Hz, 1 H, 47 CHH), 4.38-4.28 (m, 4 H, 4 CHH), 4.22-4.00 (m, 8 H), 3.90-48 3.59 (m, 15 H), 3.52-3.44 (m, 3 H), 3.39-3.34 (m, 7 H), 2.38 49 (t, J = 7.6 Hz, 2H, H-5°), 1.73-1.64 (m, 4 H, H-2°, H-4°), 1.47-50 1.38 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=0), 159.4, 138.3, 138.2, 138.0, 137.84, 137.78, 137.6, 136.2, 51 52 130.3 (aromatic C), 129.7, 128.7, 128.65, 128.61, 128.59, 128.48, 128.45, 128.4, 128.33, 128.30, 128.2, 128.02, 53 127.98, 127.93, 127.87, 127.8, 127.74, 127.71, 127.6, 127.5, 54 127.4, 127.3, 113.9 (aromatic CH), 97.9, 97.8, 97.5, 97.48 (C-55 1a, 1b, 1c and 1d), 81.0, 80.9, 80.8, 80.0 (C-3a, 3b, 3c and 3d), 56 77.8 (C-4), 75.3, 74.7, 74.4, 74.3, 73.6, 73.6, 73.54, 73.51 57

(PhCH₂), 73.1 (C-4), 72.4 (C-4), 72.1 (C-4), 71.5, 71.2, 71.1, 70.2 (c-5a, 5b, 5c and 5d), 68.9, 68.6, 68.3 (3 C-6), 68.29 (C-1º), 67.8 (C-6), 66.3 (PhCH₂), 63.8, 63.7, 63.6, 63.2 (C-2a, 2b, 2c and 2d), 55.4 (OCH₃), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 24.8 (C-4º).

Synthesis of N-phenyl trifluoroacetimidate 2-N₃-galactose donor **16**:



NIS (9.15 g, 40.68 mmol) was added to the solution of compound **S3** (18 g, 31.3 mmol) in Acetone/H₂O (210 ml/72ml) at 0 °C. The reaction was slowly warmed to room temperature and stirred until TLC-analysis indicated full consumption of the starting material (± 1h). Then the mixture was diluted with DCM and washed with saturated Na₂S₂O₃ and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The lactol was purified by silica gel column chromatography (pentane: EA = 4:1). Cs₂CO₃ was added to the solution of The lactol (10.59g, 24.33 mmol) in 140 ml acetone. The mixture was stirred at 0 °C for 15 minutes. Then CF₃C(=NPh)Cl (6.06 g, 29.2 mmol) was added to the solution. which was slowly warmed to room temperature and stirred overnight. The reaction was quenched with Et₃N and concentrated *in vacuo*. The product **16** was purified by silica gel column chromatography (pentane: $Et_2O = 30:1 -$ 10:1). Compound **16** (13.3 g, a/b = 2:1, 90% yield, PE: Et₂O = 10:1, Rf = 0.45-0.55) was obtained as white solid. α isomer: ¹H-NMR (CDCl₃, 400 MHz) δ 7.50 – 7.24 (m, 7H, aromatic H), 7.15 - 7.05 (m, 1H, aromatic H), 6.84 (d, I = 7.7 Hz, 2H, aromatic H), 6.47 (bs, 1H, H-1), 4.78 (d, J = 11.4 Hz, 1H, CH₂Ph), 4.69 (d, J = 11.4 Hz, 1H, CH₂Ph), 4.63 (s, 1H, H-4), 4.22 (q, J = 12.8 Hz, 2H. H-6), 4.10 (t, J = 6.3 Hz, 1H, H-2), 3.89 (d, J = 9.5 Hz, 1H, H-3), 3.76 (s, 1H, H-5), 1.09-1.02 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 143.29, 137.45, 128.74, 128.56, 128.01, 127.91, 124.40, 119.35 (aromatic C/CH), 94.73 (C-1), 76.04 (C-3), 70.71 (CH₂Ph), 69.89 (C-5), 69.16 (C-4), 66.76 (C-6), 57.71 (C-2), 27.59 (CH₃), 27.23 (CH₃), 23.38 (C-Si), 20.73 (C-Si). β isomer: ¹H-NMR (CDCl₃, 400 MHz) δ 7.48 – 7.25 (m, 7H, aromatic H), 7.14 – 7.04 (m, 1H, aromatic H), 6.85 (d, J = 7.7 Hz, 2H, aromatic H), 5.50 (bs, 1H, H-1), 4.77 $(d, J = 11.9 \text{ Hz}, 1\text{H}, C\text{H}_2Ph), 4.66 (d, J = 11.9 \text{ Hz}, 1\text{H}, C\text{H}_2Ph),$ 4.43 (s, 1H, H-5), 4.19 (s, 2H, H-6), 4.02 (s, 1H, H-4), 3.30 (s, 2H, H-2, 3), 1.15 - 1.00 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 143.5, 137.5, 128.8, 128.7, 128.2, 128.0, 124.5, 119.4 (aromatic C/CH), 95.8 (C-1), 79.6 (C-3), 72.2 (C-2), 71.0 (CH₂Ph), 68.6 (C-5), 66.8 (C-6), 60.8 (C-4), 27.7 (CH₃), 27.4 (CH3), 23.6 (C-Si), 20.9 (C-Si). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₂₁H₃₇N₃O₅Si: 629.2383, found: 629.2376.

Synthesis of disaccharide 18: Donor 16 (5 g, 8.2 mmol) and acceptor 17 (3.32 g, 6.95 mmol) (donors and acceptors coevaporated with toluene three times) were dissolved in DCM (65 mL), cooled to 0 °C and TfOH (60 µL) was added. The reaction was stirred at 0 $\,^\circ C$ until TLC-analysis showed complete conversion of the donor. The reaction was

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quenched with Et₃N after completed checking by TLC, fil-1 tered and concentrated *in vacuo*. The product **16** was puri-2 fied by silica gel column chromatography (pentane: $Et_2O =$ 3 10:1). Compound 18 (4.36g, 70% yield) was obtained with full α -selectivity as a colorless syrup. [α]_D²⁰ +153.3 (c=1, 4 CHCl₃). IR (neat, cm⁻¹) v 651, 698, 738, 797, 826, 984, 1043, 5 1066, 1100, 1171, 1364, 1473, 2107 (N₃), 2859, 2933. ¹H-6 NMR (CDCl₃, 400 MHz) δ 7.53-7.51 (m, 2 H, aromatic H), 7 7.44-7.22 (m, 18 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-8 1b), 5.59 (d, / = 5.2 Hz, 1 H, H-1a), 5.00 (d, / = 10.4 Hz, 1 H, 9 CHH), 4.91 (d, J = 10.4 Hz, 1 H, CHH), 4.73 (d, J = 11.6 Hz, 1 10 H, CHH), 4.63 (d, J = 11.6 Hz, 1 H, CHH), 4.43-4.37 (m, 4 H, 2 11 CHH, H-4b, H-5a), 3.96-3.77 (m, 6 H, H-2a, H-2b, H-3a, H-4a, 12 H-6), 3.72-3.64 (m, 2 H, H-3b, H-6a), 3.53 (dd, $J_1 = 2.0$ Hz, J_2 13 = 10.8 Hz, 1 H, H-6b), 3.42 (s, 1 H, H-5b), 1.03 (s, 9 H, 3 CH₃), 14 0.97 (s, 9 H, 3 CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 137.9, 137.8, 15 137.4, 133.5 (aromatic C), 132.1, 129.2, 128.62, 128.58, 16 128.51, 128.48, 128.00. 127.98, 127.8, 127.5 (aromatic CH), 17 97.7 (C-1b), 87.1 (C-1a), 82.3 (C-3a), 75.5 (C-3b), 75.0, 73.3 (PhCH₂), 72.8 (c-4a), 71.3 (c-5a), 70.5 (PhCH₂), 69.6 (C-4b), 18 68.9 (C-6), 68.0 (C-5b), 66.9 (C-6), 64.6 (C-2a), 58.1 (C-2b), 19 27.7 (3 CH₃), 27.3 (3 CH₃), 23.4, 20.7. HRMS (ESI) m/z: 20 $[M+NH_4]$ Calculated for C₄₇H₆₂N₇O₈SSi: 912.41444, found: 21 912.41409. 22

23 Synthesis of disaccharide 20: Compound 18 (4.1 g, 4.6 24 mmol) was dissoveld in THF (40 mL) in a round flusk. Then 25 HF-pyridine (1.2 mL) was added in the solution. The reac-26 tion stirred until TLC-analysis indicated full consumption of 27 the starting material (30 min). Then the mixture was diluted 28 with DCM and the reaction quenched with saturated Na-29 HCO₃. The organic phase was washed with water and brine, 30 dried with anhydrous MgSO₄, filtered and concentrated in 31 vacuo. The crude compound 19 was dissloved in CH₃CN (47 32 mL). Then BnBr (880 µL), borinic acid-catalyzed (110 mg), 33 K₂CO₃ (710 mg), KI (800 mg) were added in the mixture. 34 The reaction stirred at 60 °C in oil bath until TLC-analysis 35 indicated full consumption of the starting material (24 h). Then the mixture was diluted with ethyl acetate and the re-36 action quenched with saturated NaHCO₃. The organic phase 37 was washed with water and brine, dried with anhydrous 38 MgSO₄, filtered and concentrated *in vacuo*. The product was 39 purified by silica gel column chromatography (pen-40 tane:Et₂O = 5:1). Compound **20** (3.6 g, 94% yield with two 41 steps) was obtained as a colorless syrup. $[\alpha]_D^{20}$ +11.7 (c=1, 42 CHCl₃). IR (neat, cm⁻¹) v 697, 737, 1029, 1046, 1077, 1266, 43 2106 (N₃), 2870, 2919, 3493. ¹H-NMR (CDCl₃, 400 MHz) δ 44 7.54-7.51 (m, 2 H, aromatic H), 7.41-7.23 (m, 23 H, aromatic 45 *H*), 5.61 (d, *J* = 3.6 Hz, 1 H, H-1b), 5.60 (d, *J* = 5.2 Hz, 1 H, H-46 1a), 5.00 (d, J = 10.4 Hz, 1 H, CHH), 4.94 (d, J = 10.4 Hz, 1 H, 47 CHH), 4.66 (s, 2 H, PhCH₂), 4.50-4.37 (m, 3 H, 2 CHH, 5a), 4.08 (s, 1 H, H-4b), 3.97-3.91 (m, 2 H, H-2b, H-4a), 3.85-3.37 48 (m, 5 H), 3.66 (dd, J₁ = 2.4 Hz, J₂ = 10.8 Hz, 1 H, H-6_b), 3.59 49 (dd, J₁ = 5.6 Hz, J₂ = 9.6 Hz, 1 H), 3.51 (dd, J₁ = 5.6 Hz, J₂ = 9.6 50 Hz, 1 H), 2.63 (s, 1 H, OH). ¹³C-APT (CDCl₃, 100 MHz) δ 138.3, 51 137.8, 137.6, 137.2, 133.5 (aromatic C), 132.3, 128.8, 128.6, 52 128.5, 128.4, 128.3, 128.1, 128.0, 127.9, 127.86, 127.85, 53 127.6, 127.55 (aromatic CH), 98.2 (C-1b), 87.1 (C-1a), 82.1 54 (C-3a), 76.3 (C-3b), 75.1 (PhCH2), 74.4 (c-4a), 73.8, 73.2, 55 71.8 (PhCH₂), 71.3 (c-5a), 69.5 (C-6), 69.4 (C-6), 69.3 (C-5b), 56 66.5 (C-4b), 64.8 (C-2b), 59.0 (C-2a). HRMS (ESI) m/z: 57

 $[M\!+\!NH_4]^*$ Calculated for $C_{46}H_{52}N_7O_8S$: 862.35926, found: 862.35895.

Synthesis of thio-disaccharide 21: The compound 20 (3.83 g, 4.53 mmol) was dissloved in DMF (10 mL). Then NaH (544 mg) and NapBr (1.5 g) were added in the mixture. The reaction stirred until TLC-analysis indicated full consumption of the starting material (2 h). Then the mixture was diluted with ethyl acetate and the reaction quenched with ice water. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The product was purified by silica gel column chromatography (pentane: $Et_20 = 10:1$). Compound **21** (4.15 g, 93% yield) was obtained as a colorless syrup. $[\alpha]_{D^{20}}$ +208.6 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 737, 1028, 1049, 1093, 1123, 1362, 1454, 2106 (N₃), 2870. 2914. ¹H-NMR (CDCl₃, 400 MHz) δ 7.80-7.11 (m, 32 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-1b), 5.59 (d, J = 5.2 Hz, 1 H, H-1a), 5.01-4.89 (m, 3 H, 3 CHH), 4.70-4.62 (m, 3 H, 3 CHH), 4.50 (d, J = 12.0 Hz, 1 H, CHH), 4.44-4.40 (m, 2 H, CHH, 5b), 4.27 (d, J = 11.6 Hz, 1 H, CHH), 4.17 (d, J = 11.6 Hz, 1 H, CHH), 4.04 (s, 1 H, H-4b), 3.97-3.81 (m, 6 H, H-2a, H-2b, H-3a, H-3b, H-4a, H-5a), 3.77-3.65 (m, 2 H, H-6), 3.51-3.40 (m, 2 H, H-6). ¹³C-APT (CDCl₃, 100 MHz) & 138.3, 137.8, 137.60, 137.58, 135.6, 133.6, 133.2, 133.1 (aromatic C), 133.08, 129.2, 128.7, 128.6, 128.5, 128.4, 128.3, 128.1, 127.99, 127.97, 127.93, 127.80, 127.6, 127.4, 127.2, 126.5, 126.2, 126.1 (aromatic CH), 98.3 (C-1b), 87.1 (C-1a), 82.2 (C-3a), 77.5 (C-3b), 75.2, 74.9 (PhCH₂), 74.1 (c-4a), 73.6, 73.1 (PhCH₂), 72.8 (C-4b), 72.3 (PhCH₂), 71.3 (C-5b), 70.4 (C-5a), 69.4 (C-6), 68.6 (C-6), 64.8 (C-2a), 69.8 (C-2b). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₅₇H₆₀N₇O₈S: 1002.42186, found: 1002.42125.

N-phenyl trifluoroacetimidate disaccharide donor **23**: Compound **21** (4.15 g, 4.21 mmol) was dissolved in acetone:H₂O (10:1, 44 mL). N-Iodosuccinimide (NIS) (2.0 g, 8.8 mmol) was added in one portion and the reaction was stirred at room temperature for 2 hours. The solution was diluted with DCM and the reaction was guenched with saturated aqueous Na₂S₂O₃. Then the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered and concentrated in vacuo, and the product purified by column chromatography (pentane:EA = 3:1). The lactol 22 was obtained as colourless syrup. Next, the lactol was dissolved in acetone (40 mL). Cs_2CO_3 (1.9 g) and 2,2,2-trifluoro-N-phenylacetimidoyl chloride (960 μ L) were added to the solution respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by column chromatography (pentane:EA = 40:1-20:1). Compound 23 (3.7 g, 81% over two steps) was obtained as yellow syrup. IR (neat, cm⁻¹) v 695, 734, 818, 1027, 1116, 1209, 1312, 1454, 1489, 1497, 1717, 2107, 2870, 2918. ¹H-NMR (CDCl₃, 500 MHz) δ 7.80-7.09 (m, aromatic H), 6.81 (bt, 1 H), 5.65 (dd, 1 H), 5.01-4.87 (m), 4.68-4.54 (m), 4.45-4.42 (m), 4.33-4.18 (m), 4.03-3.41 (m). ¹³ C-APT (CDCl₃, 125 MHz) δ 143.4, 143.2, 138.20, 138.18, 137.8, 137.6, 137.55, 137.5, 135.6, 133.3, 133.2 (aromatic C), 128.9, 128.7, 128.53, 128.52, 128.44, 128.43, 128.3, 128.08, 128.05, 128.0, 127.97, 127.9, 127.84, 127.81, 127.69, 127.66, 127.65, 127.6, 127.2, 127.17, 124.7, 124.6 (aromatic CH), 119.4 (C-1), 98.3 (C-1),

98.2 (C-1), 83.6, 81.0, 77.6, 77.3, 75.5, 75.18, 75.16, 75.0, 74.9. 73.64, 73.60, 73.30, 73.2, 73.1, 73.0, 72.8, 72.7, 72.3, 72.2, 70.4, 70.3, 69.0, 68.5, 65.8, 63.7, 59.7, 59.6. HRMS (ESI) $[M-[O(C=NPh)CF3]+OH+Na]^+$ Calculated m/z: for C₅₉H₅₆F₃N₇O₉Na: 910.41340, found: 910.41374.

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Synthesis of disaccharide 24: Donor 16 (1.09 g) and acceptor 4 (790 mg) (donors and acceptors co-evaporated with toluene three times) were dissolved in DCM (12 mL), cooled to 0 °C and TfOH (12 µL) was added. The reaction 10 was stirred at 0 °C until TLC-analysis showed complete conversion of the donor. The reaction was quenched with Et₃N 11 after completed checking by TLC, filtered and concentrated 12 in vacuo. The product was purified by size exclusion 13 (DCM:MeOH = 1:1). Compound 24 (1.24 g, 92% yield) was 14 obtained with full α -selectivity as a colorless syrup. $[\alpha]_{D^{20}}$ 15 +95.9 (c=1, CHCl₃). IR (neat, cm⁻¹) v 651, 698, 737, 765, 797, 16 826, 984, 1004, 1040, 1130, 1144, 1171, 1455, 1474, 1735 17 (C=O), 2106 (N₃), 2860, 2933. ¹H-NMR (CDCl₃, 400 MHz) δ 18 7.44-7.24 (m, 20 H, aromatic H), 5.67 (d, J = 3.6 Hz, 1 H, H-19 1b), 5.12 (s, 2 H, PhCH₂), 4.96 (d, J = 10.4 Hz, 1 H, CHH), 4.92 20 (d, l = 3.6 Hz, 1 H, H-1a), 4.86 (d, l = 10.4 Hz, 1 H, CHH), 4.7121 (d, / = 11.6 Hz, 1 H, CHH), 4.61 (d, / = 11.6 Hz, 1 H, CHH), 4.48 22 (s, 2 H, PhCH₂), 4.36 (d, J = 2.0 Hz, 1 H, H-4b), 4.06 (dd, J₁ = 23 10 Hz, J₂ = 8.4 Hz, 1 H, H-3a), 3.91-3.78 (m, 4 H), 3.74-3.45 24 (m, 6 H), 3.34-3.30 (m, 2 H, H-2a, H-1^o_b), 2.39 (t, J = 7.6 Hz, 2H, H-5°), 1.74-1.64 (m, 4 H, H-2°, H-4°), 1.48-1.42 (m, 2 H, 25 H-3^o), 1.03 (s, 9 H, 3 CH₃), 0.95 (s, 9 H, 3 CH₃). ¹³C-APT (CDCl₃, 26 100 MHz) & 173.5 (C=O), 138.0, 137.9, 137.7, 136.2 27 (aromatic C), 128.7, 128.6, 128.3, 128.0, 127.96, 127.9, 28 127.86, 127.6 (aromatic CH), 97.9 (C-1b), 97.5 (C-1a), 81.0 29 (C-3a), 75.5 (C-3b), 74.3, 73.5 (PhCH₂), 72.4 (c-4a), 70.5 30 (PhCH₂), 70.1 (c-5a), 69.6 (C-4b), 69.1 (C-6), 68.3 (C-1^o), 31 67.9 (C-5b), 66.9 (C-6), 66.3 (PhCH2), 63.6 (C-2a), 58.1 (C-32 2b), 34.3 (C-5^o), 29.2 (C-2^o), 27.7 (3 CH₃), 27.3 (3 CH₃), 25.8 33 (C-3^o), 24.8 (C-4^o), 23.4, 20.7. HRMS (ESI) m/z: [M+NH₄]⁺ 34 Calculated for $C_{54}H_{74}N_7O_{11}Si: 1024.52101$, found: 35 1024.52157.

37 Synthesis of disaccharide 25: Compound 24 (1.16 g, 1.15 38 mmol) was dissoveld in THF (11 mL) in a round flusk. Then 39 HF-pyridine (300 µL) was added in the solution. The reac-40 tion stirred until TLC-analysis indicated full consumption of 41 the starting material (30 min). Then the mixture was diluted 42 with DCM and the reaction quenched with saturated Na-HCO₃. The organic phase was washed with water and brine, 43 dried with anhydrous MgSO4, filtered and concentrated in 44 vacuo. The product was purified by silica gel column chro-45 matography (pentane: $Et_20 = 3:1$). Compound **25** (910 mg, 46 91% yield) was obtained as a colorless syrup. $[\alpha]_D^{20}$ +80.6 47 (c=1, CHCl₃). IR (neat, cm⁻¹) v 698, 738, 1040, 1145, 1262, 48 1354, 1455, 1733 (C=O), 2106 (N₃), 2872, 2932, 3461. ¹H-49 NMR (CDCl₃, 400 MHz) δ 7.38-7.25 (m, 20 H, aromatic H), 50 5.65 (d, J = 3.6 Hz, 1 H, H-1b), 5.12 (s, 1 H, PhCH₂), 4.97 (d, J 51 = 10.4 Hz, 1 H, CHH), 4.92 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, J 52 = 10.4 Hz, 1 H, CHH), 4.67-4.54 (m, 4 H, 4 CHH), 4.05 (dd, J1 53 = 10.0 Hz, J₂ = 8.4 Hz, 1 H, H-3a), 3.91-3.80 (m, 2 H, H-4a, H-54 5), 3.74-3.57 (m, 8 H), 3.51-3.44 (m, 1 H, H-1^o_b), 3.31 (dd, *J*₁ 55 = 10.0 Hz, J₂ = 3.6 Hz, 1 H, H-2a), 2.65 (s, 1 H, OH), 2.39 (t, J = 7.6 Hz, 2H, H-5°), 2.29 (s, 1 H, OH), 1.74-1.64 (m, 4 H, H-2°, 56 H-4^o), 1.47-1.40 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 57

173.60 (C=0), 138.1, 137.80, 137.1, 136.1 (aromatic C), 128.8, 128.7, 128.6, 128.5, 128.4, 128.31, 128.29, 128.1. 127.9, 127.8, 127.7 (aromatic CH), 97.9 (C-1b), 97.7 (C-1a), 80.8 (C-3a), 76.1 (C-3b), 74.5 (PhCH₂), 73.8 (c-4a), 73.7 (PhCH₂), 71.9 (PhCH₂), 70.3 (c-5b), 70.2 (C-5a), 69.7 (C-6), 68.3 (C-1º), 67.2 (C-4b), 66.3 (PhCH2), 63.8 (C-2a), 62.9 (C-6), 58.8 (C-2b), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₄₆H₆₀N₇O₁₁: 884.41888, found: 884.41942.

Synthesis of disaccharide acceptor 26: The compound 25 (865 mg, 1.0 mmol) was dissloved in CH₃CN (10 mL). Then BnBr (182 µL), borinic acid-catalyzed (22 mg), K₂CO₃ (148 mg), KI (166 mg), were added in the mixture. The reaction stirred at 60 °C in oil bath until TLC-analysis indicated full consumption of the starting material (24 h). Then the mixture was diluted with ethyl acetate and the reaction quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered and concentrated in vacuo. The product was purified by silica gel column chromatography (pentane: $Et_2O =$ 5:1). Compound 26 (910 mg, 95%) was obtained as a colorless syrup. $[\alpha]_D^{20}$ +66.6 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 736, 1040, 1096, 1259, 1455, 1734 (C=O), 2106 (N₃), 2869, 2928. ¹H-NMR (CDCl₃, 400 MHz) δ 7.40-7.21 (m, 25 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-1b), 5.11 (s, 1 H, PhCH₂), 4.96 (d, J = 10.4 Hz, 1 H, CHH), 4.91 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, I = 10.4 Hz, 1 H, CHH), 4.63 (bs, 2 H, 2 CHH), 4.55 (d, I = 12.0 Hz, 1 H, CHH), 4.45 (d, I = 12.0 Hz, 1 H, CHH),4.47-4.36 (m, 3 H, 3 CHH), 4.07-4.03 (m, 2 H, H-3a, H-5a), 3.91-3.44 (m, 12 H), 3.32 (dd, J₁ = 11.2 Hz, J₂ = 3.6 Hz, 1 H, H-2a), 2.65 (s, 1 H, OH), 2.38 (t, I = 7.6 Hz, 2H, H-5°), 1.73-1.63 (m, 4 H, H-2°, H-4°), 1.47-1.39 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=0), 138.3, 137.8, 137.3, 136.1 (aromatic C), 129.1, 128.7, 128.65, 128.54, 128.50, 128.4, 128.3, 128.0, 127.9, 127.8, 127.6, 127.6, 127.3 (aromatic CH), 97.9 (C-1b), 97.7 (C-1a), 80.8 (C-3a), 76.3 (C-3b), 74.5 (PhCH₂), 73.9 (C-4a), 73.8, 73.3, 71.7 (PhCH₂), 70.1 (C-4b), 69.5, 69.4 (C-6), 69.2 (C-5b), 68.2 (C-1^o), 66.4 (C-5a), 66.2 (PhCH₂), 63.7 (C-2a), 58.9 (C-2b), 34.2 (C-5^o), 29.1 (C-2^o), 25.7 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for C₅₃H₆₄N₇O₁₁: 974.46583, found: 974.46660.

Synthesis of tetrasaccharide 27: The reaction was carried out according to the standard procedure A. A mixture of donor 23 (520 mg, 0.49 mmol), acceptor 26 (238 mg, 0.25 mmol) (donors and acceptors co-evaporated with toluene three times), MPF (490 µL) in dry DCM (1 mL) were stirred over fresh flame-dried molecular sieves 3A under nitrogen. The solution was cooled to -78 $^{\circ}$ C, after which TfOH (40 μ L) was added. After 30 min, the reaction was stirred at -10 $\,^\circ\!\mathrm{C}$ until TLC-analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound 27 (280 mg, 89%, $\alpha:\beta = 10:1$) was obtained as a colorless syrup. IR (neat, cm⁻ ¹) ν 697, 736, 1028, 1096, 1258, 1319, 1356, 1454, 1497, 1731 (C=O), 2105 (N₃), 2868, 2925. ¹H-NMR (CDCl₃, 400 MHz) & 7.77-7.69 (m, 3 H, aromatic H), 7.60 (bs, 1 H, aromatic H), 7.42-7.17 (m, 48 H, aromatic H), 5.73 (d, I = 3.6Hz, 1 H, H-1d), 5.65 (d, J = 3.6 Hz, 1 H, H-1b), 5.10 (s, 1 H,

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PhCH₂), 4.96-4.90 (m, 7 H), 4.74-4.48 (m, 7 H), 4.36-3.61 (m, 22 H), 3.51-3.26 (m, 6 H), 3.20 (d, / = 10.0 Hz, 1 H), 3.03 (d. *J* = 10.0 Hz, 1 H), 2.36 (t, *J* = 7.6 Hz, 2H, H-5°), 1.71-1.61 (m, 4 H, H-2°, H-4°), 1.45-1.37 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 100 MHz) δ 173.4 (C=O), 138.4, 138.2, 137.7, 137.68, 137.6, 137.54, 137.47, 136.1, 135.6, 133.1, 133.0 (aromatic C), 128.6, 128.5, 128.4, 128.38, 128.35, 128.27, 128.25, 128.2, 128.1, 128.07, 128.0, 127.9, 127.88, 127.86, 127.78, 127.76, 127.66, 127.6, 127.4, 127.3, 127.1, 127.0, 126.4, 126.0, 125.9 (aromatic CH), 98.7 (C-1), 98.1 (C-1), 97.8 (C-1), 97.7 (C-1), 81.0 (C-3), 80.7 (C-3), 76.8 (C-3), 75.1 (C-3), 74.9, 74.5, 10 74.4 (CH2), 73.5 (C-4), 73.5, 73.3 (CH2), 73.2 (C-4), 72.9 (C-11 4), 72.8 (C-4), 72.8, 72.2, 71.7 (CH2), 70.6 (C-5), 70.1 (C-5), 12 69.9 (C-5), 69.8 (C-5), 69.2, 68.4, 68.1, 67.9, 66.8, 66.1 (CH₂), 13 64.7 (C-2), 63.7 (C-2), 59.5 (C-2), 59.4 (C-2), 34.1 (C-5^o), 14 29.0 (C-2º), 25.6 (C-3º), 24.7 (C-4º). HRMS (ESI) m/z: 15 [M+NH₄]⁺ Calculated for C₁₀₄H₁₁₄N₁₃O₁₉: 1848.83484, found: 16 1848.83541. 17

18 Synthesis of tetrasaccharide acceptor 28: The reaction was 19 carried out according to the standard procedure C. Com-20 pound 27 (700 mg, 0.38 mmol) was dissolved in DCM:HFIP 21 (1:1, 0.1 M). TES (304 µL, 1.91 mmol) and 0.2M HCl/HFIP 22 (1.9 mL) were added to the mixture. The reaction stirred until TLC-analysis indicated full consumption of the starting 23 material (15min). Then the mixture was diluted with DCM 24 and the reaction quenched with saturated NaHCO₃. The or-25 ganic phase was washed with water and brine, dried with 26 anhydrous MgSO₄, filtered and concentrated in vacuo. The 27 product was purified by silica gel column chromatography 28 (pentane:Et₂O = 5:1). Compound **28** (297mg, 73% yield) 29 was obtained as a colorless syrup. $[\alpha]_{D^{20}}$ +106.7 (c=1, CHCl₃). 30 IR (neat, cm⁻¹) v 696, 737, 1040, 1100, 1261, 1454, 1735 31 (C=O), 2106 (N₃), 2869, 2926. ¹H-NMR (CDCl₃, 500 MHz) δ 32 7.42-7.17 (m, 45 H, aromatic H), 5.71 (d, J = 3.5 Hz, 1 H, H-33 1d), 5.65 (d, / = 3.5 Hz, 1 H, H-1b), 5.10 (s, 1 H, PhCH₂), 4.95-34 4.89 (m, 6 H, H-1a, H-1c, 4 CHH), 4.72 (d, J = 12.0 Hz, 1 H, 35 CHH), 4.68 (s, 2 H, 2 CHH), 4.57-4.48 (m, 3 H), 4.36-4.17 (m, 7 H), 4.09-3.58 (m, 17 H), 3.51-3.30 (m, 3 H), 3.36-3.30 (m, 36 3 H), 3.91-3.44 (m, 12 H), 3.17 (dd, J1 = 11.5 Hz, J2 = 2.5 Hz, 37 1 H), 3.02 (dd, J_1 = 11.5 Hz, J_2 = 2.5 Hz, 1 H), 2.66 (s, 1 H, OH), 38 2.37 (t, J = 7.5 Hz, 2H, H-5°), 1.71-1.63 (m, 4 H, H-2°, H-4°), 39 1.45-1.39 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 125 MHz) δ 173.5 40 (C=0), 138.5, 138.3, 137.8, 137.7, 137.68, 137.6, 137.4, 41 136.2 (aromatic C), 128.8, 128.7, 128.6, 128.6, 128.5, 128.3, 42 128.2, 128.18, 128.0, 127.9, 127.9, 127.86, 127.8, 127.7, 43 127.69, 127.5, 127.5, 127.3, 127.2 (aromatic CH), 98.8 (C-1), 44 98.1 (C-1), 97.9 (C-1), 97.8 (C-1), 80.9 (C-3), 80.8 (C-3), 76.1 45 (C-3), 75.8 (C-3), 74.54, 74.46, 73.7 (CH2), 73.6 (C-4), 73.56, 46 73.4 (CH2), 73.1 (2 C-4), 73.07 71.8, 71.75 (CH2), 70.7 (C-4), 47 70.2 (C-5), 69.9 (C-5), 69.3, 69.1 (CH2), 68.8 (C-5), 68.5, 68.3, 68.9 (CH₂), 66.5 (C-5), 66.3 (PhCH₂), 64.7 (C-2), 63.8 (C-2), 48 59.6 (C-2), 58.8 (C-2), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 49 24.8 (C-4^o). HRMS (ESI) m/z: [M+NH₄]⁺ Calculated for 50 C₉₃H₁₀₆N₁₃O₁₉: 1708.77224, found: 1708.77299. 51

Synthesis of hexasaccharide 29: The reaction was carried out according to the standard procedure A. A mixture of donor 23 (540 mg, 0.5 mmol), acceptor 28 (360 mg, 0.21 mmol) (donors and acceptors co-evaporated with toluene three times), MPF (400 µL) in dry DCM (1 mL) were stirred over

fresh flame-dried molecular sieves 3A under nitrogen. The solution was cooled to -78 $^{\circ}$ C, after which TfOH (44 μ L) was added. After 30 min, the reaction was stirred at -10 °C until TLC-analysis showed complete conversion of the acceptor (48 h). The reaction was quenched with Et₃N, filtered and concentrated in vacuo. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound 29 (500 mg, 91%, $\alpha:\beta = 10:1$) was obtained as a colorless syrup. [α]_D²⁰ +123.0 (c=1, CHCl₃). IR (neat, cm⁻¹) v 697, 736, 1039, 1099, 1261, 1319, 1359, 1454, 1734 (C=O), 2106 (N₃), 2870, 2926. ¹H-NMR (CDCl₃, 400 MHz) δ 7.77-7.69 (m, 3 H, aromatic H), 7.60 (bs, 1 H, aromatic H), 7.44-7.07 (m, 68 H, aromatic H), 5.73 (d, J = 3.6 Hz, 1 H, H-1), 5.71 (d, J = 3.2 Hz, 1 H, H-1), 5.65 (d, J = 3.6 Hz, 1 H, H-1), 5.09 (s, 1 H, PhCH₂), 4.96-4.91 (m, 9 H), 4.77-4.65 (m, 5 H), 4.58-4.48 (m, 4 H), 4.33-3.62 (m, 37 H), 3.52-3.16 (m, 10 H), 3.05 (d, / = 10.0 Hz, 1 H), 2.99 (d, J = 10.0 Hz, 1 H), 2.36 (t, J = 7.6 Hz, 2H, H-5°), 1.71-1.61 (m, 4 H, H-2^o, H-4^o), 1.45-1.37 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.4 (*C*=0), 138.5, 138.4, 138.2, 137.7, 137.7, 137.6, 137.59, 137.57, 137.5, 137.4, 136.1, 135.7, 133.2, 133.0 (aromatic C), 128.6, 128.5, 128.49, 128.48, 128.45, 128.39, 128.36, 128.3, 128.2, 128.1, 128.05, 128.1, 128.0, 127.94, 127.90, 127.86, 127.82, 127.78, 127.77, 127.72, 127.67, 127.6, 127.42, 127.35, 127.3, 127.12, 127.11, 127.03, 127.0, 126.4, 126.0, 125.9 (aromatic CH), 98.8 (C-1), 98.7 (C-1), 98.1 (C-1), 97.9 (C-1), 97.8 (C-1), 97.7 (C-1), 80.9 (C-3), 80.8 (C-3), 80.7 (C-3), 76.9 (C-3), 76.1 (C-3), 75.7 (C-3), 74.9, 74.5, 74.4, 73.5, 73.3 (CH₂), 73.3 (C-4), 73.2 (C-4), 73.0 (CH₂), 72.83 (C-4), 72.80 (CH₂), 72.7 (C-4), 72.2, 71.9, 71.7 (CH₂), 70.6 (C-5), 70.5 (C-5), 70.1 (C-5), 69.9 (C-5), 69.8 (C-5), 69.6 (C-5), 69.2, 68.3, 68.1, 67.9, 66.8, 66.5, 66.1 (CH₂), 64.7 (2 C-2), 63.7 (C-2), 59.5 (C-2), 59.47 (C-2), 59.4 (C-2), 34.2 (C-5^o), 29.1 (C-2^o), 2575 (C-3^o), 24.7 (C-4^o).

Synthesis of hexasaccharide 30: Compound 29 (20 mg, 0.0078 mmol) was dissolved in THF/H₂O/tert-BuOH (2 mL/2 mL/1 mL) before a catalytic amount of Pd(OH)₂/C was added. The reaction mixture was stirred for 3 days under a H₂ atmosphere, filtered and concentrated in vacuo. A white powder 30 (6.7 mg, 76%) was obtained after purification by gel filtration (HW-40, 0.15M NH₄OAc in H₂O). ¹H-NMR (D₂O, 500 MHz) δ 5.40-5.35 (m, 3 H, 3 H-1), 4.85-4.81 (m, 3 H, 3 H-1), 4.13-4.06 (m, 2 H), 3.97-3.50 (m, 35 H), 3.41-3.37 (m, 1 H), 3.15-3.08 (m, 2 H), 2.81 (dd, 2 H), 2.72-2.70 (m, 2 H), 2.06 (t, 2 H), 1.55-1.43 (m, 5 H), 1.29-1.23 (m, 2 H). ¹³C-APT (CDCl₃, 125 MHz) δ 99.6 (C-1), 99.5 (C-1), 99.5 (C-1), 99.2 (2 C-1), 97.4 (C-1), 76.9, 76.7, 76.6, 76.7, 73.7, 73.3, 72.4, 72.3, 71.9, 71.1, 71.0, 70.5, 69.2, 68.8, 68.4, 68.3, 61.2, 60.6, 60.4, 55.3, 55.3, 54.6, 51.1, 51.1, 51.0, 37.5, 28.3, 25.7, 25.4. HRMS (ESI) m/z: [M+2H]+/2 Calculated for C₄₂H₈₀N₆O₂₇: 550.25302; found: 550.25247.

Synthesis of hexasaccharide **31**: Compound **30** (5 mg) was dissoled in H₂O. Then Ac₂O and NaHCO₃ were added in the solution. The reaction mixture was stirred for 3 days until TLC-analysis showed complete conversion of the starting martials. The product was purified by gel filtration (HW-40, 0.15M NH₄OAc in H₂O). Compound **31** (5.5 mg, 86%) was obtained as a white solid. ¹H-NMR (D₂O, 500 MHz) δ 5.36-5.34 (m, 4 H, 4 H-1), 5.27 (d, J = 4.0 Hz, 1 H, H-1), 4.79 (bt, 2 H, 2 H-1), 4.73 (d, J = 3.0 Hz, 1 H, H-1), 4.21-4.06 (m, 5 H), 3.97-3.57 (m, 37 H), 3.40-3.35 (m, 1 H), 2.24 (bt, 2 H), 1.97-1.91 (m, 18 H, 6 CH₃), 1.80-1.74 (m, 2 H), 1.54-1.46 (m, 4 H), 1.32-1.26 (m, 2 H). ¹³C-APT (CDCl₃, 125 MHz) δ 180.2, 174.7, 174.69, 174.6, 174.5, 174.46 (6 C=0), 98.2, 98.15, 96.6 (6 C-1), 77.4, 77.2, 76.1, 75.7, 75.4, 72.5, 72.4, 71.8, 71.7, 71.3, 71.2, 70.7, 70.7, 70.4, 68.5, 68.1, 67.9, 67.6, 66.9, 61.6, 60.7, 60.3, 60.0, 54.4, 54.2, 49.9, 34.6, 28.2, 25.0, 24.4, 22.1 (CH₃), 22.0 (CH₃), 22.0 (CH₃), 21.9 (CH₃), 21.9 (2 CH₃). HRMS (ESI) m/z: [M+2H]+/2 Calculated for C₅₄H₉₂O₃₃N₆: 676.28472; found: 676.28489.

Supporting Information

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The Supporting Information is available free of charge on the ACS Publications website.

Full experimental details and characterization of all new compounds (PDF). NMR spectra of all new compounds (PDF)

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Author Contributions

The manuscript was written through contributions of all authors. / All authors have given approval to the final version of the manuscript.

Notes

Any additional relevant notes should be placed here.

ACKNOWLEDGMENT

This work was supported by the Chinese Scholarship Council (CSC grant to L.W.) and the European Research Council (ERC-CoG-726072-'GLYCONTROL', to J.D.C.C.).

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