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Aryl-Containing Esters Of Triphosphoric Acid As Substrates Of Terminal Deoxynucleotidyl Transferase

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ARYL-CONTAINING ESTERS OF TRIPHOSPHORIC ACID AS SUBSTRATES OF TERMINAL DEOXYNUCLEOTIDYL TRANSFERASE

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□ *A new group of terminal deoxynucleotidyltransferase (TDT) substrates, namely, non-nucleoside triphosphates (NNTP) bearing 5-substituted 2,4-dinitrophenyl fragments instead of nucleoside residues was synthesized.*

Keywords Non-nucleoside triphosphates; terminal deoxynucleotidyltransferase

INTRODUCTION

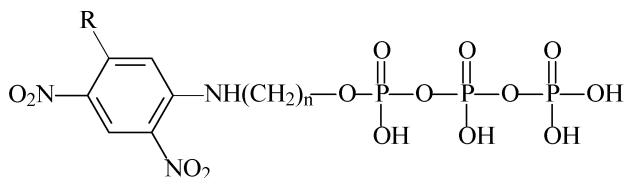
Terminal deoxynucleotidyltransferase (TDT, EC 2.7.7.31) is a unique template-independent DNA polymerase. TDT belongs to the X polymerase family, a subclass of an ancient nucleotidyltransferase superfamily, which includes nucleic acid polymerases such as DNA polymerases β , λ , μ , and some others. Unlike any other DNA polymerases, TDT incorporates both ribo- and deoxyribonucleotides *in vitro* with the equal efficacy as well as a large array of unnatural nucleoside triphosphates. It was shown in our laboratory that triphosphate analogues bearing different bulky alkyl and aryl groups instead of a nucleoside residue can serve as substrates for calf thymus TDT, thus, demonstrating that the presence of a nucleic base in the substrate molecule is not a determining factor for the binding to the enzyme active site and incorporation into the growing DNA chain.^[1–3] The efficacy of these compounds depends on substituent and linker structures and length.^[4,5] It was shown that the affinities of some of the compounds of this series towards TDT were similar to those of natural substrates. Moreover, non-nucleoside triphosphates (NNTP) were demonstrated to be inhibitors and/or substrate terminators of other polymerases of the X family, particularly, of human β and λ polymerases.^[6]

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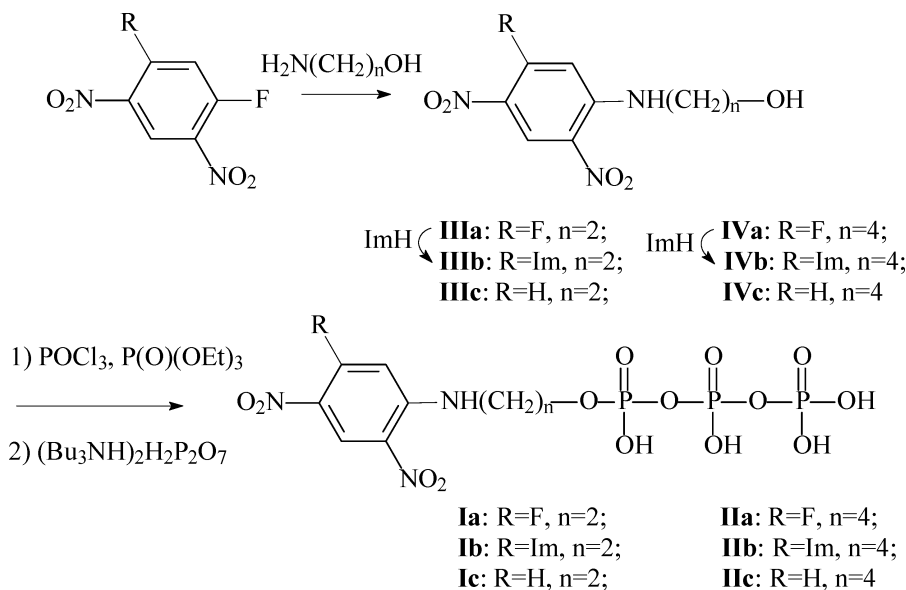
RESULTS AND DISCUSSION

We present in this work the synthesis of new NNTPs of the following structures:



Ia: $n=2$, $R=F$; **Ib:** $n=2$, $R=Im$; **Ic:** $n=2$, $R=H$; **IIa:** $n=4$, $R=F$; **IIb:** $n=4$, $R=Im$; **IIc:** $n=4$, $R=H$

Compounds (**IIIa**, **IIIc**, **IVa**, and **IVc**) were obtained by the reaction of the corresponding aminoalcohol with 1,3-difluoro-4,6-dinitrobenzene (for **IIIa** and **IVa**) or 1-fluoro-2,4-dinitrobenzene (for **IIIc** and **IVc**). For the preparation of the imidazole derivative (**IIIb** and **IVb**), the corresponding fluoro-containing precursors were treated with a solution of imidazole in DMF. The Ludwig triphosphorylation^[7] carried out without the isolation of intermediate monophosphates gave the target triphosphates (Scheme).



The synthesized fluoro-containing triphosphates proved to be rather labile under the conditions of ion-exchange chromatography on DEAE (HCO_3^- -form), and, therefore, were isolated in the following way. The reaction mixture was diluted with water and passed through a Dowex 50^(H+)

column. The resulting solution was concentrated, and the target product was purified on a reversed-phase LiChroprep RP-18 column. Total yields of the synthesized triphosphates achieved 15–30% from the starting amino alcohol.

All the synthesized compounds demonstrated potent substrate properties in cell-free experiments with TDT.^[8]

EXPERIMENTAL SECTION

N-(2,4-Dinitro-5-fluorophenyl)-2-aminoethyl triphosphate Ia. Yield 21%. UV (H₂O, pH 6): λ_{\max} 265 nm (ϵ 9000), 349 nm (ϵ 15900). ¹H-NMR (D₂O): 9.10 (1H, d, *J* 8.1, H₃), 7.04 (1H, d, *J* 14.6, H₆), 4.22 (2H, dt, CH₂), 3.75 (2H, t, *J* 5.3, CH₂N). ³¹P-NMR (D₂): -5.72 (1P, d, *J* 20.3, P_γ), -10.39 (1P, d, *J* 19.3, P_α), -21.50 (1P, dd, P_β). Mass (m/e): 484.1 [M⁺-1].

N-(2,4-Dinitro-5-imidazolylphenyl)-2-aminoethyl triphosphate Ib. Yield 17%. UV (H₂O, pH 6): λ_{\max} 272 nm (ϵ 5800), 365 nm (ϵ 7000). ¹H-NMR (D₂O): 9.24 (1H, s, H₃), 8.96, 7.67 and 7.55 (3H, 3 br.s, Im), 7.50 (1H, s, H₆), 4.40 (2H, m, CH₂), 3.83 (2H, t, *J* 5.3, CH₂N). ³¹P-NMR (D₂): -10.24 (1P, d, *J* 19.3, P_γ), -10.83 (1P, d, *J* 20.3, P_α), -22.61 (1P, dd, P_β). Mass (m/e): 532.2 [M⁺-1].

N-(2,4-Dinitrophenyl)-2-aminoethyl triphosphate Ic. Yield 24%. UV-VIS (H₂O, pH 6): λ_{\max} 265 nm (ϵ 8300), 363 nm (ϵ 17500). ¹H-NMR (D₂O): 8.95 (1H, d, *J* 2.5, H₃), 8.23 (3H, dd, -5), 7.21 (1H, d, *J* 9.65, H₆), 4.01 (2H, m, CH₂), 3.72 (2H, t, *J* 5.9, CH₂N). ³¹P NMR (D₂O): δ -9.88 (d, 1P, *J* 21.4, P_γ), -10.32 (d, 1P, *J* 19.3, P_α), -21.77 (dd, 1P, P_β). Mass (m/e): 466.1 [M⁺-1].

N-(2,4-Dinitro-5-fluorophenyl)-4-aminobutyl triphosphate IIa. Yield 15%. UV (H₂O, pH 6): λ_{\max} 265 nm (ϵ 9100), 349 nm (ϵ 16000). ¹H-NMR (D₂O): 9.10 (1H, d, *J* 8.1, H₃), 6.94 (1H, d, *J* 15.6, H₆), 4.01 (2H, m, CH₂), 3.50 (2H, t, *J* 6.8, CH₂N), 1.77 (4, m, (CH₂)₂). ³¹P-NMR (D₂): -10.12 (1P, d, *J* 19.3, P_γ), -10.32 (1P, d, *J* 20.3, P_α), -22.65 (1P, dd, P_β). Mass (m/e): 512.1 [M⁺-1].

N-(2,4-Dinitro-5-imidazolylphenyl)-4-aminobutyl triphosphate IIb. Yield 23%. UV (H₂O, pH 6): λ_{\max} 272 nm (ϵ 5800), 365 nm (ϵ 7000). ¹H-NMR (D₂O): 9.19 (1H, s, H₃), 8.30, 7.45 and 7.33 (3H, 3 br.s, Im), 7.19 (1H, s, H₆), 3.99 (2H, m, CH₂), 3.54 (2H, t, *J* 6.8, CH₂N), 1.82-1.72 (4, m, (CH₂)₂). ³¹P-NMR (D₂): -10.10 (1P, d, *J* 19.3, P_γ), -10.31 (1P, d, *J* 20.3, P_α), -22.64 (1P, dd, P_β). Mass (m/e): 560.2 [M⁺-1].

N-(2,4-Dinitrophenyl)-4-aminobutyl triphosphate IIc. Yield 30%. UV-VIS (H₂O, pH 6): λ_{\max} 265 nm (ϵ 8300), 363 nm (ϵ 17500). ¹H-NMR (D₂O): 9.19 (1H, d, *J* 2.8, H₃), 8.30 (3H, dd, -5), 7.19 (1H, d, *J* 9.65, H₆), 3.99 (2H, m, CH₂), 3.54 (2H, t, *J* 6.8, CH₂N), 1.82-1.72 (4, m, (CH₂)₂). ³¹P-NMR (D₂): -10.10 (1P, d, *J* 19.3, P_γ), -10.31 (1P, d, *J* 20.3, P_α), -22.64 (1P, dd, P_β). Mass (m/e): 494.2 [M⁺-1].

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