Three Types of Products Obtained Unexpectedly from the Reaction of **Dimedone with Chalcones**

Guan-Wu Wang,*^[a] Qing-Quan Lu,^[a] and Jing-Jing Xia*^[a,b]

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The apparently simple Michael reaction of dimedone with chalcones afforded three types of products. The reaction of dimedone with chalcones in water at reflux, mediated by cetyltrimethylammonium bromide, gave the expected Michael adducts in good to excellent yields after purification by column chromatography. The attempted purification of the Michael adducts by recrystallization from ethanol in air unexpectedly generated hydroperoxidated and hydroxylated Michael adducts in low yields. The efficient synthesis of hydroperoxidated Michael adducts could be achieved in nearly

quantitative yields by autoxidation of the Michael adducts in 1,2-dichloroethane at room temperature under aerobic conditions. The conversion of the hydroperoxidated Michael adducts to hydroxylated Michael adducts was facilitated by reduction with triphenylphosphane in excellent yields. Some of the reported Michael adducts in the literature need to be reassigned. Particular caution must be exerted to assign the Michael products generated from cyclic β -diketones, such as dimedone, because different purification processes may give different types of products.

Introduction

The Michael reaction stands as one of the most important processes for the formation of C-C bonds in organic synthesis, because the formed Michael adducts are versatile precursors for many biologically active compounds.^[1] A survey of the literature indicated that, among the numerous reported Michael reactions, the addition of cyclic β-diketones to α,β -conjugated compounds has been relatively scarce,^[2,3] because the reaction tends to continue to subsequent cyclization steps in most cases.^[2]

On the other hand, water in place of harmful organic solvents as the environmentally friendly and benign reaction medium has gained increasing popularity in organic synthesis.^[4] As a continuation of our interest in aqueous organic reactions,^[2c,5] we recently investigated the Michael reaction of 5,5-dimethylcyclohexane-1,3-dione (dimedone, 1) with 1,3-diaryl-2-propen-1-ones (chalcones, 2) with water as the reaction medium. We unexpectedly isolated three types of products during the workup processes, which were identified as simple Michael adducts, hydroperoxidated and hydroxylated Michael adducts. Herein, we disclose these re-

E-mail: xiajj@mail.ustc.edu.cn

sults and further manipulation of the Michael adducts for the efficient synthesis of hydroperoxidated and hydroxylated Michael adducts.

Results and Discussion

In an attempt to extend the methodology of synthesizing 1.4-dihydropyridine derivatives with water as the reaction medium,^[6] we recently explored the reaction of dimedone with chalcones and ammonium acetate in refluxing water. The expected products should be 1,4-dihydropyridine derivatives formed through Michael addition followed by cyclization.^[6] However, we found that ammonium acetate was not involved, and the reaction could not proceed beyond the Michael addition step. We thus focused on the aqueous Michael addition. The Michael addition of 1 to the simplest chalcone, namely, 1,3-diphenyl-2-propen-1-one (2a), was chosen as the model reaction for investigation. The Michael reaction is conventionally catalyzed by strong bases, such as NaOH, KOH, and NaOEt, which often lead to undesirable side reactions, such as bis(addition) and subsequent condensation. We intended to use mild conditions to avoid such side reactions. At the onset, the reaction of 1 with 2a in water at reflux (120 °C in an oil bath) for 3 h gave Michael adduct 3a in 28% yield (Table 1, Entry 1). Benzyl triethylammonium chloride (BTEAC) as a phase-transfer catalyst (PTC) was previously applied by us to the aqueous aminochlorination of chalcones.^[5] We found that BTEAC could increase the product yield to 49% (Table 1, Entry 2), and tetrabutylammonium bromide (TBAB) further improved the product yield to 72% (Table 1, Entry 3). Cetyltrimethylammonium bromide (CTAB) was superior to

[[]a] Hefei National Laboratory for Physical Sciences at Microscale, CAS Key Laboratory of Soft Matter Chemistry, Joint Laboratory of Green Synthetic Chemistry, and Department of Chemistry, University of Science and Technology of China, Hefei, Anhui 230026, P. R. China Fax: +86-551-360-7864 E-mail: gwang@ustc.edu.cn

[[]b] School of Materials and Chemical Engineering, Anhui University of Architecture, Hefei, Anhui 230601, P. R. China

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other ammonium salts and afforded 3a in 76% yield (Table 1, Entry 4). Decreasing the temperature of the oil bath from 120 to 110 °C reduced the yield to 70% (Table 1, Entry 5). Prolonging the reaction time from 3 to 5 h enhanced the yield to 89% (Table 1, Entry 6). However, anionic surfactants, such as sodium dodecyl sulfate (SDS) and sodium dodecylbenzenesulfonate (ABS-Na), reduced the formation of 3a, giving yields even lower than that without any additive (Table 1, Entries 7 and 8 vs. Entry 1). Therefore, equimolar amounts of 1 and 2a with 10 mol-% of CTAB were selected as the optimal conditions for the aqueous Michael reaction of 1 and 2a.

Table 1. Optimization of reaction conditions for the aqueous Michael reaction of 1 with chalcone $2a.^{\rm [a]}$

o	- + ()		Iditive (10 mol-%) H ₂ O, reflux	
1	2a		/	3a
Entry	Additive	Time [h]	Temp. [°C] ^[b]	Yield [%] ^[c]
1	none	3	120	28
2	BTEAC	3	120	49
3	TBAB	3	120	72
4	CTAB	3	120	76
5	CTAB	3	110	70
6	CTAB	5	120	89
7	SDS	3	120	17
8	ABS-Na	3	120	24

[a] All reactions were performed with 1 (0.2 mmol), chalcone 2a (0.2 mmol), and additive (10 mol-%) in water (2 mL). [b] Temperature of the oil bath. [c] Isolated yield after chromatographic purification.

With the optimized conditions in hands, we then extended the substrate to other chalcones. A variety of chalcones with electron-withdrawing or -donating groups on Table 2. Aqueous Michael reaction of 1 with 2 mediated by $\mbox{CTAB}.^{[a]}$



[a] All reactions were performed with 1 (0.2 mmol), 2 (0.2 mmol), and CTAB (10 mol-%) in water (2 mL) at 120 °C (oil bath temperature). [b] Isolated yield after chromatographic purification.

either phenyl ring performed well and afforded Michael adducts **3a–i** in good to excellent yields (Table 2, Entries 1–9).

Michael adducts **3a**–i were fully characterized by HRMS, as well as by ¹H and ¹³C NMR and FTIR spectroscopy. The HR mass spectra of products **3a–i** gave the correct molecular ion masses. The ¹H NMR spectra in [D₆]DMSO showed the expected signals from the chalcone moiety, including two doublets and one pseudo-triplet for the newly formed –CHCH₂– fragment, as well as signals of the dimedone moiety. The ¹³C NMR spectra of **3a–i** displayed signals for the two phenyl rings; one coincidently overlapped the signal at $\delta \approx 199$ ppm for the carbonyl groups, one signal at $\delta \approx 115$ ppm, and four signals in the range of $\delta = 28$ – 41 ppm for aliphatic carbon atoms. The dimedone moiety



Figure 1. HMQC spectrum of 3a. Inset: partial assignment of 3a.



should exist in an enol form, since one broad signal at δ = 10.42–10.86 ppm due to the enolic hydroxy group and a signal at $\delta = 114.6$ –115.7 ppm for the enolic carbon atom were observed in the ¹H and ¹³C NMR spectra, respectively. The dimedone moiety as an enol form was also observed previously by us in the nonreductive addition products from the manganese(III) acetate mediated reaction of 1,3-cyclohexanediones with imines.^[7] Surprisingly, in the ¹³C NMR spectra of 3, only four signals for the seven sp³-carbon atoms were observed. Among the four signals, one overlapped signal at $\delta = 27.7 - 27.9$ ppm corresponds to two carbon atoms, leaving two of the seven sp³-carbon atoms for which no ¹³C NMR signals could be identified. The heteronuclear multiple quantum correlation (HMQC) spectrum of 3a (Figure 1) was measured to help assign its molecular structure. Two protons [signals at δ = 3.65 (dd, J = 16.8, 6.8 Hz) and 3.87 (dd, J = 16.8, 8.4 Hz) ppm] are attached to the same carbon atom (signal at $\delta = 40.9$ ppm), whereas one proton [signal at δ = 4.78 (t, J = 7.5 Hz) ppm] is bonded to the carbon atom with signal at $\delta = 34.4$ ppm. Unexpectedly, the broad signal for the methylene protons of the dimedone moiety showed no cross-peak in the HMOC spectrum, indicating that fast ring flipping of the dimedone fragment resulted in the signal disappearance of the methylene carbon atoms in the ¹³C NMR spectrum of **3a**. The same effect was responsible for the occurrence of only one signal at δ = 199.2 ppm for the two carbonyl carbon atoms and one signal at $\delta = 27.8$ ppm for the two methyl carbon atoms of the dimedone moiety.

To further confirm that the integrity of the dimedone fragment was preserved in the Michael adduct, compound **3a** was treated with acetyl chloride in the presence of triethylamine at room temperature for 40 min, and acetoxylated product **4a** was obtained in 89% yield (Scheme 1).



Scheme 1. Acetoxylation of Michael adduct 3a.

The HR mass spectrum of **4a** gave a signal at m/z = 390.1829, corresponding to the molecular ion. The ¹H and ¹³C NMR spectra showed all expected signals, including those at $\delta = 42.8$, 51.8, and 167.5 ppm, which correspond to two methylene carbon atoms and one enolic carbon atom in the dimedone moiety, but were absent in the ¹³C NMR spectrum of **3a**. The broad signals at $\delta = 2.02$ and 2.29 ppm for the two CH₂ groups in the ¹H NMR spectrum of **3a** became a singlet at $\delta = 2.24$ ppm and two doublets at $\delta = 2.42$ and 2.53 ppm with a coupling constant of 17.7 Hz in the ¹H NMR spectrum of **4a**.

In initial attempts to purify Michael adducts **3**, the products were recrystallized from ethanol. Some other products, including those identified later as hydroperoxides **5** and hydroxylated Michael adducts **6**, were obtained in low yields. During the NMR spectroscopy experiments, we found that adducts 3 were stable in $[D_6]DMSO$ and $[D_6]acetone$, but unstable in CDCl₃. We then screened different solvents with or without an additive for the efficient conversion of 3a. We found that Michael adduct 3a was converted to hydroperoxide 5a in low yields at room temperature in air for 4 h when toluene, ethanol, or N,N-dimethylformamide (DMF) were employed as the solvents (Table 3, Entries 1–3). When chloroform was used, a moderate yield could be obtained (Table 3, Entry 4). The yield was significantly increased to 88% when the reaction was performed in dichloromethane (Table 3, Entry 5). Further improvement of the yield to 91% was attained if 1,2-dichloroethane (DCE) was utilized as the solvent (Table 3, Entry 6). However, when acetic acid was added, the yield decreased from 91 to 68% (Table 3, Entry 6 vs. 7). Nearly quantitative yields were achieved when the reaction in DCE was conducted under 1 atm of O₂ or prolonged from 4 to 6 h (Table 3, Entries 8 and 9), indicating that this transformation involved oxygen.

Table 3. Solvent screening for the conversion of Michael adduct 3a into hydroperoxide 5a.^[a]



[a] Michael adduct **3a** (0.2 mmol), solvent (2 mL), and additive were stirred in the presence of air at room temperature for the indicated time. [b] Isolated yield after chromatographic purification. [c] Mixture of **5a** and **6a** (see below) in a ratio of 1:1. [d] AcOH (0.2 mL) as the additive. [e] Under 1 atm of O_2 .

It is more convenient to operate in air than under oxygen. Therefore, the conversion of Michael adducts 3b-i to hydroperoxides 5b-i was performed in air at room temperature with DCE as the solvent. The results are shown in Table 4 and demonstrate that autoxidation of 3a-i in DCE is very efficient and affords 5a-i in 95-98% yields.

In attempts to purify products **5** by recrystallization from ethanol, a third type of products, **6**, was identified. Products **6** showed similar ¹H and ¹³C NMR spectra to those of hydroperoxides **5** with the same numbers of protons and carbon atoms (see below). This puzzle was eventually solved by obtaining the single-crystal X-ray structure of **6d** (see below). The third type of products was identified as the hydroxylated Michael adducts, which were likely to be generated by the reduction of hydroperoxides **5** with ethanol. The yield from **5** to **6** by recrystallization from ethanol was Table 4. Autoxidation of Michael adducts 3 to hydroperoxides 5.^[a]



[a] All reactions were performed by stirring **3** (0.2 mmol) in DCE (2 mL) in air at room temperature for the indicated time. [b] Isolated yield after chromatographic purification.

low to moderate. Therefore, triphenylphosphane was added to reduce the –OOH group to the –OH group.^[8] We found that after the conversion of Michael adducts **3** into hydroperoxides **5** was complete in air at room temperature, subsequent treatment of the reaction mixture with Ph_3P (1 equiv.) gave hydroxylated products **6** in 90–95% yields (Table 5).

Table 5. One-pot conversion of Michael adducts 3 into hydroxylated products $6.^{\left[a\right] }$



[a] The reaction conditions for the first step were the same as those given in Table 4. After the completion of the first step, Ph_3P (0.2 mmol) was added to the reaction mixture, which was then stirred at 60 °C for 30 min. [b] Isolated yield after chromatographic purification.

Hydroperoxides **5a–i** and hydroxylated products **6a–i** were characterized by HRMS, as well as ¹H and ¹³C NMR and FTIR spectroscopy. The HR mass spectra of **5** exhibited mass signals that correspond to [M-O], indicating that

the hydroperoxides could easily lose an oxygen atom during mass spectrometry. While different from Michael adducts 3, products 5 and 6 displayed similar ¹H and ¹³C NMR spectra, and showed the expected signals for all protons and carbon atoms. The only significant differences in the NMR spectra of 5 and 6 were the signal shifts from $\delta = 9.38$ -9.46 ppm for the –OOH group to $\delta = 4.04$ –4.13 ppm for the –OH group and from δ = 99.5–100.7 to 91.2–91.9 ppm for the carbon atoms connected to the -OOH and -OH groups, respectively. The broad signal(s) for the two CH₂ groups of the dimedone moiety in the ¹H NMR spectra of 3 became two doublets and two double doublets in the ¹H NMR spectra of both 5 and 6. In addition, the missing signals for the two CH₂ groups of the dimedone moiety in the ¹³C NMR spectra of **3** could be observed at $\delta = 52.4$ – 52.6 and 53.3–53.5 ppm for **5**, and δ = 51.6–51.7 and 51.7– 51.9 ppm for 6. Furthermore, the identity of product 6d was unequivocally established by its single-crystal X-ray structure, which is shown in Figure 2.



Figure 2. Single-crystal X-ray structure of 6d. Ellipsoids are drawn at the 40% probability level.

The α -hydroxylation of β -dicarbonyl compounds by using various oxidants, such as peracids, dimethyldioxirane, or molecular oxygen, has been reported.^[9] Oxidation by O₂ of compounds containing a tertiary carbon atom to give hydroperoxides is known in the literature. For example, the autoxidation of cyclic β -diketones with a tertiary α -C atom to give hydroperoxides was described over 50 years ago.^[10] A large number of Michael adducts have been synthesized.^[1] Nevertheless, there has been no report on the peroxidation of Michael adducts from dimedone and chalcones.^[3] By comparing the spectroscopic data in ref.^[3a] with ours, we found that the reported spectroscopic data of 3a, 3d, and 3f in ref.^[3a] match more closely with those of 6a, 5d, and 6f, respectively. Therefore, the assumed Michael adducts in ref.^[3a] should be corrected and reassigned. We repeated the KF/Al₂O₃-catalyzed reaction of dimedone and chalcones in DMF at 80 °C,^[3a] and indeed obtained the expected Michael adducts with the same workup procedure for our aqueous Michael reaction. The hydroperoxidated



Michael adduct (5d) and hydroxylated Michael adducts (6a and 6f) in ref.^[3a] should result from further transformation of the Michael adducts during recrystallization from ethanol. This is consistent with our observation that both hydroperoxidated and hydroxylated Michael adducts could be generated by the treatment of 3 in ethanol (see above). Deduced from the reported spectroscopic data, it is likely that all of the reported Michael adducts in ref.^[3a] are in fact either the corresponding hydroperoxidated or hydroxylated products after treatment with ethanol. It is taken for granted that only Michael products can be isolated from the addition of β -diketones to α,β -conjugated compounds.^[3a] In our case, three types of products were obtained from the Michael reaction of dimedone with chalcones, depending on the purification processes. The present work clearly demonstrates that special caution must be exerted for the purification procedure and product assignment of Michael adducts from cyclic β -diketones, such as 1.

Conclusions

Three types of products, that is, Michael adducts (3), hydroperoxidated (5) and hydroxylated (6) Michael adducts, were unexpectedly obtained from the reaction of 1 with 2 depending on different purification processes. The reaction of 1 with 2 in water at reflux mediated by CTAB gave the expected Michael adducts 3 in good to excellent yields (74-93%) after purification by column chromatography. When our Michael adducts 3 were treated in ethanol both hydroperoxidated (5) and hydroxylated (6) Michael adducts were generated in low yields. Hydroxylated Michael adducts 6 could also be formed by recrystallization of 5 from ethanol in low to moderate yields. Gratifyingly, hydroperoxidated Michael adducts 5 were easily produced in nearly quantitative yields (95–98%) by autoxidation of **3** in 1,2-dichlorethane at room temperature under aerobic conditions. The conversion of hydroperoxidated Michael adducts 5 to hydroxylated Michael adducts 6 could be facilitated by reduction with triphenylphosphane in >90% yields. Some of the reported Michael adducts in the literature may need to be revised. Particular care must be exerted to determine what type of product is generated from cyclic β -diketones, such as 1, because purification processes have substantial effects on the type of products.

Experimental Section

General Methods: Flash chromatographic purification of products was performed on silica gel (200–300 mesh). Thin-layer chromatography was visualized with a UV light (254 and 365 nm). IR spectra were recorded with a Bruker Vector-22 spectrometer as KBr pellets and are reported in cm⁻¹. ¹H NMR spectra were recorded at 300 MHz with a Bruker Avance-300 spectrometer in CDCl₃ with chemical shifts (δ) given in ppm relative to TMS as an internal standard. ¹³C NMR spectra were recorded with a Bruker Avance-300 (75.5 MHz) spectrometer with complete proton decoupling; chemical shifts are reported in ppm relative to the solvent resonance as the internal standard (CDCl₃: δ = 77.16 ppm). All inten-

sities in the ¹³C NMR spectroscopic data are 1 C, except where indicated. Single-crystal X-ray analysis was carried out with an Oxford Diffraction Xcalibur diffractometer equipped with a Sapphire 3 CCD detector and graphite-monochromated Mo-K_{α} radiation (λ = 0.71073 Å).

General Procedure A. Michael Addition of Dimedone 1 to Chalcones 2: To a mixture of dimedone (1; 28.0 mg, 0.2 mmol), chalcone 2a (2b-i, 0.2 mmol), and CTAB (7.3 mg, 0.02 mmol) in a 25 mL round-bottomed flask was added H_2O (2 mL). This reaction mixture was stirred vigorously in an oil bath preheated at 120 °C for the desired time (monitored by TLC). The residue remaining after concentration was separated by column chromatography on silica gel with petroleum ether/ethyl acetate (3:1) as the eluent to give Michael product 3a (3b-i).

3-Hydroxy-5,5-dimethyl-2-(3-oxo-1,3-diphenylpropyl)cyclohex-2-enone (3a): According to general procedure A, chalcone 2a (41.6 mg, 0.2 mmol) was employed. Column chromatography gave 3a as a white solid (62.0 mg, 89%). ¹H NMR (300 MHz, [D₆]DMSO): δ = 0.90 (s, 6 H, CH₃), 2.02 (br. s, 2 H, CH₂), 2.29 (br. s, 2 H, CH₂), $3.65 (dd, J = 16.8, 6.8 Hz, 1 H, CH_2), 3.87 (dd, J = 16.8, 8.4 Hz,$ 1 H, CH₂), 4.78 (t, J = 7.5 Hz, 1 H, CH), 7.08 (t, J = 7.2 Hz, 1 H, ArH), 7.19 (t, J = 7.5 Hz, 2 H, ArH), 7.26 (d, J = 7.5 Hz, 2 H, ArH), 7.50 (t, J = 7.4 Hz, 2 H, ArH), 7.61 (t, J = 7.3 Hz, 1 H, ArH), 7.90 (d, J = 7.4 Hz, 2 H, ArH), 10.48 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): δ = 199.2 (2 C), 144.7, 137.1, 132.8, 128.6 (2 C), 127.8 (2 C), 127.7 (2 C), 127.5 (2 C), 125.2, 115.4, 40.9, 34.4, 31.5, 27.8 (2 C) ppm. FTIR (KBr): $\tilde{v} = 3426$, 2957, 1685, 1599, 1493, 1449, 1378, 1253, 1210, 1151, 1076, 1008, 752, 696 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{23}H_{24}O_3$ [M]⁺ 348.1725; found 348.1732.

3-Hydroxy-5,5-dimethyl-2-(3-oxo-3-phenyl-1-p-tolylpropyl)cyclohex-2-enone (3b): According to general procedure A, chalcone 2b (44.5 mg, 0.2 mmol) was employed. Column chromatography gave **3b** as a white solid (61.0 mg, 84%). ¹H NMR (300 MHz, $[D_6]$ -DMSO): $\delta = 0.90$ (s, 6 H, CH₃), 1.99–2.35 (m, 4 H, CH₂), 2.21 (s, 3 H, CH₃), 3.62 (dd, J = 16.6, 6.9 Hz, 1 H, CH₂), 3.83 (dd, J =16.6, 8.3 Hz, 1 H, CH₂), 4.73 (t, J = 7.5 Hz, 1 H, CH), 6.99 (d, J = 7.8 Hz, 2 H, ArH), 7.13 (d, J = 7.8 Hz, 2 H, ArH), 7.50 (t, J = 7.5 Hz, 2 H, ArH), 7.60 (t, J = 7.3 Hz, 1 H, ArH), 7.89 (d, J =8.0 Hz, 2 H, ArH), 10.42 (br. s, 1 H, OH) ppm. ¹³C NMR $(75 \text{ MHz}, [D_6]DMSO): \delta = 199.2 (2 \text{ C}), 141.7, 137.1, 134.0, 132.8,$ 128.6 (2 C), 128.3 (2 C), 127.7 (2 C), 127.4 (2 C), 115.6, 40.9, 34.0, 31.5, 27.8 (2 C), 20.5 ppm. FTIR (KBr): $\tilde{v} = 3428$, 2957, 2924, 1686, 1597, 1513, 1449, 1378, 1314, 1254, 1210, 1151, 1057, 1008, 814, 765, 691 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₄H₂₆O₃ [M]⁺ 362.1882; found 362.1879.

3-Hydroxy-2-[1-(4-methoxyphenyl)-3-oxo-3-phenylpropyl]-5,5-dimethylcyclohex-2-enone (3c): According to general procedure A, chalcone 2c (47.7 mg, 0.2 mmol) was employed. Column chromatography gave 3c as a white solid (56.0 mg, 74%). ¹H NMR (300 MHz, $[D_6]DMSO$): $\delta = 0.90$ (s, 6 H, CH₃), 2.00 (br. s, 2 H, CH₂), 2.28 (br. s, 2 H, CH₂), 3.64 (dd, *J* = 16.7, 6.9 Hz, 1 H, CH₂), 3.68 (s, 3 H, OCH₃), 3.84 (dd, J = 16.7, 8.4 Hz, 1 H, CH₂), 4.72 (t, J = 7.5 Hz, 1 H, CH), 6.76 (d, J = 8.6 Hz, 2 H, ArH), 7.17 (d, J = 8.6 Hz, 2 H, ArH), 7.50 (t, J = 7.3 Hz, 2 H, ArH), 7.61 (t, J = 7.3 Hz, 1 H, ArH), 7.89 (d, J = 7.3 Hz, 2 H, ArH), 10.46 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, $[D_6]DMSO$): δ = 199.3 (2 C), 157.0, 137.1, 136.7, 132.8, 128.6 (2 C), 128.4 (2 C), 127.7 (2 C), 115.7, 113.1 (2 C), 54.9, 41.2, 33.7, 31.5, 27.8 (2 C) ppm. FTIR (KBr): $\tilde{v} = 3437$, 2956, 2927, 1684, 1607, 1511, 1450, 1378, 1308, 1248, 1178, 1035, 830, 765, 691 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₄H₂₆O₄ [M]⁺, 378.1831; found 378.1840.

2-[1-(4-Chlorophenyl)-3-oxo-3-phenylpropyl]-3-hydroxy-5,5-dimethylcyclohex-2-enone (3d): According to general procedure A, chalcone **2d** (48.6 mg, 0.2 mmol) was employed. Column chromatography gave **3d** as a white solid (68.2 mg, 89%). ¹H NMR (300 MHz, [D₆]DMSO): δ = 0.90 (s, 6 H, CH₃), 2.03 (br. s, 2 H, CH₂), 2.30 (br. s, 2 H, CH₂), 3.68 (dd, *J* = 17.0, 7.1 Hz, 1 H, CH₂), 3.83 (dd, *J* = 17.0, 8.0 Hz, 1 H, CH₂), 4.77 (t, *J* = 7.5 Hz, 1 H, CH), 7.26 (s, 4 H, ArH), 7.50 (t, *J* = 7.6 Hz, 2 H, ArH), 7.61 (t, *J* = 7.2 Hz, 1 H, ArH), 7.90 (d, *J* = 7.6 Hz, 2 H, ArH), 10.63 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): δ = 199.0 (2 C), 143.7, 137.0, 132.9, 129.8, 129.3 (2 C), 128.6 (2 C), 127.7 (2 C), 127.6 (2 C), 115.2, 40.6, 33.8, 31.5, 27.7 (2 C) ppm. FTIR (KBr): \tilde{v} = 3064, 2957, 1687, 1596, 1580, 1491, 1449, 1377, 1315, 1254, 1211, 1150, 1092, 1056, 1014, 825, 760, 690 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃O₃³⁷Cl [M]⁺ 384.1306; found 384.1299.

2-[1-(3,4-Dichlorophenyl)-3-oxo-3-phenylpropyl]-3-hydroxy-5,5-dimethylcyclohex-2-enone (3e): According to general procedure A, chalcone **2e** (55.4 mg, 0.2 mmol) was employed. Column chromatography gave **3e** as a white solid (70.9 mg, 85%). ¹H NMR (300 MHz, [D₆]DMSO): δ = 0.90 (s, 6 H, CH₃), 1.99–2.35 (br., 4 H, CH₂), 3.71 (dd, *J* = 17.3, 7.1 Hz, 1 H, CH₂), 3.85 (dd, *J* = 17.3, 8.0 Hz, 1 H, CH₂), 4.76 (t, *J* = 7.4 Hz, 1 H, CH), 7.23 (d, *J* = 8.4 Hz, 1 H, ArH), 7.46 (s, 1 H, ArH), 7.46–7.53 (m, 3 H, ArH), 7.62 (t, *J* = 7.3 Hz, 1 H, ArH), 7.92 (d, *J* = 8.2 Hz, 2 H, ArH), 10.77 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): δ = 198.7 (2 C), 146.0, 136.9, 133.0, 130.2, 129.9, 129.4, 128.7 (2 C), 128.0, 127.8 (2 C), 127.7, 114.7, 40.4, 33.7, 31.5, 27.7 (2 C) ppm. FTIR (KBr): \tilde{v} = 3083, 2958, 1686, 1596, 1580, 1470, 1449, 1375, 1252, 1211, 1149, 1057, 1029, 818, 761, 689 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₂O₃³⁵Cl₂ [M]⁺ 416.0946; found 416.0932.

3-Hydroxy-5,5-dimethyl-2-[1-(3-nitrophenyl)-3-oxo-3-phenylpropyl]cyclohex-2-enone (3f): According to general procedure A, chalcone 2f (50.7 mg, 0.2 mmol) was employed. Column chromatography gave 3f as a yellow solid (73.2 mg, 93%). ¹H NMR (300 MHz, $[D_6]DMSO$): $\delta = 0.91$ (s, 6 H, CH₃), 2.07 (br. s, 2 H, CH₂), 2.30 (br. s, 2 H, CH₂), 3.81 (dd, J = 17.4, 7.4 Hz, 1 H, CH₂), $3.90 (dd, J = 17.4, 7.9 Hz, 1 H, CH_2), 4.90 (t, J = 7.4 Hz, 1 H, T)$ CH), 7.49–7.55 (m, 3 H, ArH), 7.63 (t, J = 7.3 Hz, 1 H, ArH), 7.73 (d, J = 7.7 Hz, 1 H, ArH), 7.94 (d, J = 8.2 Hz, 2 H, ArH), 7.99 (d, J = 8.1 Hz, 1 H, ArH), 8.11 (s, 1 H, ArH), 10.86 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): δ = 198.7 (2 C), 147.5, 147.0, 136.9, 134.5, 133.0, 129.2, 128.7 (2 C), 127.8 (2 C), 122.0, 120.5, 114.7, 40.4, 34.1, 31.6, 27.7 (2 C) ppm. FTIR (KBr): \tilde{v} = 3425, 2958, 2925, 1686, 1598, 1527, 1449, 1378, 1349, 1255, 1213, 1150, 1081, 1011, 805, 748, 690 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃NO₅ [M]⁺ 393.1576; found 393.1595.

3-Hydroxy-5,5-dimethyl-2-[1-(4-nitrophenyl)-3-oxo-3-phenylpropyl]cyclohex-2-enone (3g): According to general procedure A, chalcone **2g** (50.7 mg, 0.2 mmol) was employed. Column chromatography gave **3g** as a yellow solid (68.5 mg, 87%). ¹H NMR (300 MHz, [D₆]DMSO): δ = 0.91 (s, 6 H, CH₃), 2.05 (br. s, 2 H, CH₂), 2.31 (br. s, 2 H, CH₂), 3.83 (d, *J* = 7.5 Hz, 2 H, CH₂), 4.91 (t, *J* = 7.5 Hz, 1 H, CH), 7.51 (d, *J* = 8.7 Hz, 2 H, ArH), 7.51 (t, *J* = 7.5 Hz, 2 H, ArH), 7.63 (t, *J* = 7.2 Hz, 1 H, ArH), 7.93 (d, *J* = 7.2 Hz, 2 H, ArH), 8.09 (d, *J* = 8.7 Hz, 2 H, ArH), 10.79 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): δ = 198.6 (2 C), 153.0, 145.4, 136.8, 133.0, 128.6 (4 C), 127.8 (2 C), 122.9 (2 C), 114.6, 39.6, 34.3, 31.5, 27.7 (2 C) ppm. FTIR (KBr): \tilde{v} = 3424, 3180, 2957, 2923, 1677, 1602, 1507, 1449, 1378, 1346, 1313, 1270, 1152, 1109, 1057, 1011, 855, 753, 698 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃NO₅ [M]⁺ 393.1576; found 393.1566.

2-[3-(4-Chlorophenyl)-1-(4-methoxyphenyl)-3-oxopropyl]-3-hydroxy-5,5-dimethylcyclohex-2-enone (3h): According to general procedure A, chalcone **2h** (54.6 mg, 0.2 mmol) was employed. Column chromatography gave **3h** as a white solid (62.0 mg, 75%). ¹H NMR (400 MHz, [D₆]DMSO): δ = 0.88 (s, 6 H, CH₃), 2.15 (br. s, 4 H, CH₂), 3.61 (dd, *J* = 16.7, 7.2 Hz, 1 H, CH₂), 3.71 (dd, *J* = 16.7, 7.9 Hz, 1 H, CH₂), 3.81 (s, 3 H, OCH₃), 4.76 (t, *J* = 7.6 Hz, 1 H, CH), 7.00 (d, *J* = 8.9 Hz, 2 H, ArH), 7.22 (d, *J* = 9.0 Hz, 2 H, ArH), 7.25 (d, *J* = 9.0 Hz, 2 H, ArH), 7.87 (d, *J* = 8.9 Hz, 2 H, ArH), 10.85 (br. s, 1 H, OH) ppm. ¹³C NMR (100 MHz, [D₆]-DMSO): δ = 197.6 (2 C), 163.1, 143.9, 130.2 (2 C), 130.0, 129.9, 129.4 (2 C), 127.7 (2 C), 115.4, 114.0 (2 C), 55.6, 40.3, 34.0, 31.6, 27.9 (2 C) ppm. FTIR (KBr): \tilde{v} = 3079, 2957, 2928, 1676, 1600, 1575, 1509, 1491, 1376, 1313, 1256, 1169, 1091, 1056, 1014, 833 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₄H₂₅O₄³⁵C1 [M]⁺ 412.1441; found 412.1450.

2-[1,3-Bis(4-chlorophenyl)-3-oxopropyl]-3-hydroxy-5,5-dimethylcyclohex-2-enone (3i): According to general procedure A, chalcone **2i** (55.4 mg, 0.2 mmol) was employed. Column chromatography gave **3i** as a white solid (70.1 mg, 84%, white solid). ¹H NMR (400 MHz, [D₆]DMSO): δ = 0.88 (s, 6 H, CH₃), 2.15 (br. s, 4 H, CH₂), 3.65 (dd, *J* = 16.9, 6.9 Hz, 1 H, CH₂), 3.82 (dd, *J* = 16.9, 8.4 Hz, 1 H, CH₂), 4.74 (t, *J* = 7.6 Hz, 1 H, CH), 7.23 (d, *J* = 8.9 Hz, 2 H, ArH), 7.26 (d, *J* = 8.9 Hz, 2 H, ArH), 7.55 (d, *J* = 8.5 Hz, 2 H, ArH), 7.90 (d, *J* = 8.5 Hz, 2 H, ArH), 10.57 (br. s, 1 H, OH) ppm. ¹³C NMR (100 MHz, [D₆]DMSO): δ = 198.2 (2 C), 143.6, 138.0, 135.7, 130.0, 129.8 (2 C), 129.4 (2 C), 128.9 (2 C), 127.7 (2 C), 115.1, 40.8, 33.9, 31.6, 27.8 (2 C) ppm. FTIR (KBr): \tilde{v} = 3087, 2958, 2928, 1687, 1589, 1490, 1377, 1314, 1254, 1092, 1150, 1011, 1055, 890, 823 cm⁻¹. HRMS (EI–TOF): calcd. for C₂₃H₂₂O₃³⁵Cl₂ [M]⁺ 416.0946; found 416.0955.

Procedure for the Acetoxylation of Michael Adduct 3a To Give 5,5-Dimethyl-3-oxo-2-(3-oxo-1,3-diphenylpropyl)cyclohex-1-enyl Acetate (4a): Triethylamine (117 µL, 0.84 mmol) was added dropwise at room temperature to a solution of Michael adduct 3a (104.4 mg, 0.30 mmol) in dichloromethane (3 mL). After stirring for 10 min, acetyl chloride (53.1 µL, 0.75 mmol) was added, and stirring was maintained for 40 min. The crude product obtained was separated by chromatography on silica gel with petroleum ether/acetyl acetate (4:1) as the eluent to give 4a (104.0 mg, 89%). ¹H NMR (300 MHz, CDCl₃): δ = 1.01 (s, 3 H, CH₃), 1.03 (s, 3 H, CH₃), 2.16 (s, 3 H, CH₃), 2.24 (s, 2 H, CH₂), 2.42 (d, J = 17.7 Hz, 1 H, CH₂), 2.53 (d, J = 17.7 Hz, 1 H, CH₂), 3.74 (dd, J = 17.1, 6.9 Hz, 1 H, CH₂), 3.82 (dd, J = 17.1, 8.1 Hz, 1 H, CH₂) 4.79 (t, J = 7.4 Hz, 1 H, CH), 7.12–7.18 (m, 1 H, ArH), 7.21–7.29 (m, 4 H, ArH), 7.44 (t, J = 7.5 Hz, 2 H, ArH), 7.54 (t, J = 7.4 Hz, 1 H, ArH), 7.87 (d, J =7.5 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 198.9, 198.8, 167.5, 164.0, 142.2, 137.0, 133.1, 129.1, 128.6 (2 C), 128.3 (2 C), 128.2 (2 C), 127.6 (2 C), 126.3, 51.8, 42.8, 40.9, 35.9, 32.7, 28.0 (2 C), 21.0 ppm. FTIR (KBr): $\tilde{v} = 2960$, 1768, 1684, 1598, 1494, 1450, 1358, 1272, 1183, 1142, 1078, 1015, 850, 753, 699 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₅H₂₆O₄ [M]⁺ 390.1831; found 390.1829.

General Procedure B. Conversion of Michael Adducts 3 to Hydroperoxides 5: Michael adduct 3a (3b–i, 0.2 mmol) was stirred in DCE (2 mL) in air at room temperature for the indicated time (monitored by TLC). The reaction mixture was separated on a silica gel column (pretreated with AcOH) with petroleum ether/ethyl acetate (3:1) as the eluent to afford hydroperoxide 5a (5b–i).

2-Hydroperoxy-5,5-dimethyl-2-(3-oxo-1,3-diphenylpropyl)cyclohexane-1,3-dione (5a): According to general procedure B, Michael adduct **3a** (69.7 mg, 0.2 mmol) was employed. Column chromatography gave **5a** as a white solid (73.8 mg, 97%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.87$ (s, 3 H, CH₃), 1.24 (s, 3 H, CH₃), 2.42 (dd, J =



14.1, 3.0 Hz, 1 H, CH₂), 2.56 (dd, J = 15.3, 3.0 Hz, 1 H, CH₂), 2.92 (d, J = 14.1 Hz, 1 H, CH₂), 3.17 (d, J = 15.3 Hz, 1 H, CH₂), 3.37 (dd, J = 18.8, 7.3 Hz, 1 H, CH₂), 3.66 (dd, J = 18.8, 4.5 Hz, 1 H, CH₂), 4.39 (dd, J = 7.3, 4.5 Hz, 1 H, CH), 7.22–7.29 (m, 3 H, ArH), 7.34 (d, J = 6.3 Hz, 2 H, ArH), 7.41 (t, J = 7.5 Hz, 2 H, ArH), 7.54 (t, J = 7.5 Hz, 1 H, ArH), 7.87 (d, J = 7.5 Hz, 2 H, ArH), 9.43 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta =$ 204.0, 202.9, 197.2, 137.1, 136.4, 133.6, 129.3 (2 C), 128.8 (2 C), 128.7 (2 C), 128.3 (3 C), 100.5, 53.5, 52.6, 45.8, 39.1, 30.8, 30.7, 26.2 ppm. FTIR (KBr): $\tilde{v} = 3334$, 3064, 2966, 2927, 1731, 1703, 1449, 1357, 1239, 1086, 1010, 748, 703, 634 cm⁻¹ HRMS (EI-TOF): calcd. for C₂₃H₂₄O₄ [M – O]⁺ 364.1675; found 364.1670.

2-Hydroperoxy-5,5-dimethyl-2-(3-oxo-3-phenyl-1-p-tolylpropyl)cyclohexane-1,3-dione (5b): According to general procedure B, Michael adduct 3b (72.5 mg, 0.2 mmol) was employed. Column chromatography gave **5b** as a white solid (75.7 mg, 96%). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3): \delta = 0.86 \text{ (s, 3 H, CH}_3), 1.24 \text{ (s, 3 H, CH}_3), 2.27$ (s, 3 H, CH₃), 2.41 (dd, J = 14.0, 3.2 Hz, 1 H, CH₂), 2.55 (dd, J = 15.3, 3.2 Hz, 1 H, CH₂), 2.92 (d, J = 14.0 Hz, 1 H, CH₂), 3.16 (d, $J = 15.3 \text{ Hz}, 1 \text{ H}, \text{ CH}_2$, 3.35 (dd, $J = 18.7, 7.4 \text{ Hz}, 1 \text{ H}, \text{ CH}_2$), $3.62 (dd, J = 18.7, 4.4 Hz, 1 H, CH_2), 4.35 (dd, J = 7.4, 4.4 Hz, 1$ H, CH), 7.06 (d, J = 8.0 Hz, 2 H, ArH), 7.21 (d, J = 8.0 Hz, 2 H, ArH), 7.41 (t, J = 7.6 Hz, 2 H, ArH), 7.52 (t, J = 7.4 Hz, 1 H, ArH), 7.87 (d, J = 7.6 Hz, 2 H, ArH), 9.38 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 204.1, 203.0, 197.2, 137.9, 136.4, 134.0, 133.5, 129.5 (2 C), 129.1 (2 C), 128.7 (2 C), 128.2 (2 C), 100.6, 53.5, 52.6, 45.5, 39.1, 30.7, 30.6, 26.2, 21.2 ppm. FTIR (KBr): $\tilde{v} = 3420, 2955, 1736, 1705, 1449, 1360, 1240, 761, 687,$ 623 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{24}H_{26}O_4$ [M – O]⁺ 378.1831; found 378.1827.

2-Hydroperoxy-2-[1-(4-methoxyphenyl)-3-oxo-3-phenylpropyl]-5,5dimethylcyclohexane-1,3-dione (5c): According to general procedure B, Michael adduct 3c (75.7 mg, 0.2 mmol) was employed. Column chromatography gave 5c as a white solid (78.8 mg, 96%). ¹H NMR (300 MHz, CDCl₃): δ = 0.87 (s, 3 H, CH₃), 1.24 (s, 3 H, CH_3), 2.41 (dd, J = 13.9, 3.2 Hz, 1 H, CH_2), 2.55 (dd, J = 15.3, 3.2 Hz, 1 H, CH₂), 2.90 (d, J = 13.9 Hz, 1 H, CH₂), 3.15 (d, J = 15.3 Hz, 1 H, CH₂), 3.35 (dd, J = 18.7, 7.6 Hz, 1 H, CH₂), 3.59 $(dd, J = 18.7, 4.3 Hz, 1 H, CH_2), 3.74 (s, 3 H, OCH_3), 4.35 (dd, J)$ = 7.6, 4.3 Hz, 1 H, CH), 6.78 (d, J = 8.7 Hz, 2 H, ArH), 7.25 (d, J = 8.7 Hz, 2 H, ArH), 7.41 (t, J = 7.6 Hz, 2 H, ArH), 7.53 (t, J= 7.4 Hz, 1 H, ArH), 7.87 (d, J = 7.6 Hz, 2 H, ArH), 9.45 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 204.0, 202.9, 197.3, 159.3, 136.4, 133.6, 130.4 (2 C), 129.0, 128.7 (2 C), 128.2 (2 C), 114.2 (2 C), 100.7, 55.2, 53.5, 52.6, 45.2, 39.2, 30.7, 30.6, 26.2 ppm. FTIR (KBr): $\tilde{v} = 3400, 2957, 1737, 1704, 1686, 1611, 1514, 1449,$ 1371, 1298, 1249, 1181, 1072, 1033, 836, 762, 735, 690, 623 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{24}H_{26}O_5 [M - O]^+$ 394.1780; found 394.1791.

2-[1-(4-Chlorophenyl)-3-oxo-3-phenylpropyl]-2-hydroperoxy-5,5-dimethylcyclohexane-1,3-dione (5d): According to general procedure B, Michael adduct **3d** (76.6 mg, 0.2 mmol) was employed. Column chromatography gave **5d** as a white solid (78.9 mg, 95%). ¹H NMR (400 MHz, CDCl₃): $\delta = 0.87$ (s, 3 H, CH₃), 1.24 (s, 3 H, CH₃), 2.43 (dd, J = 14.0, 3.3 Hz, 1 H, CH₂), 2.57 (dd, J = 15.3, 3.3 Hz, 1 H, CH₂), 2.86 (d, J = 14.0 Hz, 1 H, CH₂), 3.15 (d, J = 15.3 Hz, 1 H, CH₂), 3.33 (dd, J = 18.8, 7.6 Hz, 1 H, CH₂), 3.62 (dd, J = 18.8, 4.3 Hz, 1 H, CH₂), 4.36 (dd, J = 7.6, 4.3 Hz, 1 H, CH), 7.23 (d, J = 8.7 Hz, 2 H, ArH), 7.29 (d, J = 8.7 Hz, 2 H, ArH), 7.42 (t, J = 7.7 Hz, 2 H, ArH), 7.54 (t, J = 7.4 Hz, 1 H, ArH), 7.86 (dd, J = 8.4 Hz, 2 H, ArH), 9.46 (s, 1 H, OOH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 203.5$, 202.5, 196.8, 136.0, 135.5

134.1, 133.7, 130.6 (2 C), 128.9 (2 C), 128.7 (2 C), 128.1 (2 C), 100.1, 53.3, 52.4, 44.9, 38.9, 30.7, 30.5, 26.0 ppm. FTIR (KBr): $\tilde{v} = 3403, 2957, 1735, 1704, 1595, 1491, 1449, 1417, 1358, 1237, 1204, 1093, 1074, 1014, 754, 690, 618 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃O₄³⁵Cl [M - O]⁺ 398.1285; found 398.1290.$

2-[1-(3,4-Dichlorophenyl)-3-oxo-3-phenylpropyl]-2-hydroperoxy-5,5-dimethylcyclohexane-1,3-dione (5e): According to general procedure B, Michael adduct 3e (83.5 mg, 0.2 mmol) was employed. Column chromatography gave **5e** as a white solid (85.4 mg, 95%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.88$ (s, 3 H, CH₃), 1.26 (s, 3 H, CH_3 , 2.47 (dd, J = 14.0, 3.3 Hz, 1 H, CH_2), 2.58 (dd, J = 15.3, $3.3 \text{ Hz}, 1 \text{ H}, \text{CH}_2$, $2.85 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), $3.14 \text{ (d}, J = 14.0 \text{ Hz}, 1 \text{ H}, \text{CH}_2$), 3.14 Hz, 1 Hz, 115.3 Hz, 1 H, CH₂), 3.29 (dd, J = 18.9, 7.5 Hz, 1 H, CH₂), 3.63 $(dd, J = 18.9, 4.2 Hz, 1 H, CH_2), 4.34 (dd, J = 7.5, 4.2 Hz, 1 H,$ CH), 7.22 (dd, J = 8.4, 2.1 Hz, 1 H, ArH), 7.34 (d, J = 8.3 Hz, 1 H, ArH), 7.43 (d, J = 7.5 Hz, 2 H, ArH), 7.44 (d, J = 2.1 Hz, 1 H, ArH), 7.56 (t, J = 7.4 Hz, 1 H, ArH), 7.87 (d, J = 7.5 Hz, 2 H, ArH), 9.42 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 203.3, 202.5, 196.7, 137.4, 136.0, 133.9, 132.8, 132.6, 131.3, 130.8, 128.9 (2 C), 128.8, 128.3 (2 C), 99.9, 53.5, 52.6, 44.6, 39.0, 30.8, 30.6, 26.2 ppm. FTIR (KBr): $\tilde{v} = 3414$, 2957, 1737, 1707, 1686, 1597, 1471, 1449, 1359, 1236, 1209, 1134, 1076, 1031, 1003, 758, 688, 634 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{23}H_{22}O_4{}^{35}Cl_2$ $[M - O]^+$ 432.0895; found 432.0890.

2-Hydroperoxy-5,5-dimethyl-2-[1-(3-nitrophenyl)-3-oxo-3-phenylpropyllcyclohexane-1,3-dione (5f): According to general procedure B, Michael adduct 3f (78.7 mg, 0.2 mmol) was employed. Column chromatography gave 5f as a yellow solid (82.5 mg, 97%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.89$ (s, 3 H, CH₃), 1.27 (s, 3 H, CH_3), 2.47 (dd, J = 14.1, 3.3 Hz, 1 H, CH_2), 2.62 (dd, J = 15.3, 3.3 Hz, 1 H, CH₂), 2.88 (d, J = 14.1 Hz, 1 H, CH₂), 3.17 (d, J = 15.3 Hz, 1 H, CH₂), 3.37 (dd, J = 18.9, 7.5 Hz, 1 H, CH₂), 3.71 $(dd, J = 18.9, 4.4 Hz, 1 H, CH_2), 4.49 (dd, J = 7.5, 4.4 Hz, 1 H,$ CH), 7.41–7.49 (m, 3 H, ArH), 7.56 (t, J = 7.5 Hz, 1 H, ArH), 7.73 (d, *J* = 7.8 Hz, 1 H, ArH), 7.88 (d, *J* = 7.3 Hz, 2 H, ArH), 8.11 (d, J = 7.8 Hz, 1 H, ArH), 8.23 (s, 1 H, ArH), 9.39 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 203.4, 202.6, 196.5, 148.3, 139.2, 135.9, 135.5, 133.9, 129.7, 128.8 (2 C), 128.3 (2 C), 124.3, 123.2, 99.5, 53.4, 52.5, 44.9, 38.9, 30.8, 30.4, 26.1 ppm. FTIR (KBr): $\tilde{v} = 3413$, 2956, 1739, 1710, 1682, 1597, 1525, 1447, 1352, 1257, 1208, 1074, 1004, 744, 698, 685 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{23}H_{23}NO_6 [M - O]^+$ 409.1525; found 409.1523.

2-Hydroperoxy-5,5-dimethyl-2-[1-(4-nitrophenyl)-3-oxo-3-phenylpropyl]cyclohexane-1,3-dione (5g): According to general procedure B, Michael adduct 3g (78.7 mg, 0.2 mmol) was employed. Column chromatography gave 5g as a yellow solid (81.7 mg, 96%). ¹H NMR (300 MHz, CDCl₃): δ = 0.89 (s, 3 H, CH₃), 1.26 (s, 3 H, CH₃), 2.47 (dd, J = 14.1, 3.0 Hz, 1 H, CH₂), 2.62 (dd, J = 15.3, 3.0 Hz, 1 H, CH₂), 2.85 (d, J = 14.1 Hz, 1 H, CH₂), 3.16 (d, J = 15.3 Hz, 1 H, CH₂), 3.36 (dd, J = 18.9, 7.8 Hz, 1 H, CH₂), 3.68 $(dd, J = 18.9, 4.2 Hz, 1 H, CH_2), 4.48 (dd, J = 7.8, 4.2 Hz, 1 H,$ CH), 7.43 (t, J = 7.8 Hz, 2 H, ArH), 7.53–7.58 (m, 3 H, ArH), 7.87 (d, J = 7.2 Hz, 2 H, ArH), 8.13 (d, J = 8.7 Hz, 2 H, ArH), 9.43 (s, 1 H, OOH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 203.3, 202.4, 196.5, 147.7, 144.4, 135.9, 134.0, 130.4 (2 C), 128.9 (2 C), 128.2 (2 C), 123.9 (2 C), 99.5, 53.4, 52.5, 45.0, 38.8, 30.8, 30.5, 26.1 ppm. FTIR (KBr): $\tilde{v} = 3405, 2955, 1735, 1707, 1680, 1598, 1515, 1450,$ 1349, 1236, 1211, 1111, 1074, 1003, 857, 754, 706, 690 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃NO₆ [M – O]⁺ 409.1525; found 409.1530.

2-[3-(4-Chlorophenyl)-1-(4-methoxyphenyl)-3-oxopropyl]-2-hydroperoxy-5,5-dimethylcyclohexane-1,3-dione (5h): According to general procedure B, Michael adduct **3h** (82.6 mg, 0.2 mmol) was employed. Column chromatography gave **5h** as a white solid (87.2 mg, 98%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.87$ (s, 3 H, CH₃), 1.24 (s, 3 H, CH₃), 2.42 (dd, J = 13.8, 3.0 Hz, 1 H, CH₂), 2.56 (dd, J =15.3, 3.0 Hz, 1 H, CH₂), 2.86 (d, J = 13.8 Hz, 1 H, CH₂), 3.16 (d, J = 15.3 Hz, 1 H, CH₂), 3.28 (dd, J = 18.6, 7.5 Hz, 1 H, CH₂), $3.53 (dd, J = 18.6, 4.2 Hz, 1 H, CH_2), 3.85 (s, 3 H, OCH_3), 4.37$ (dd, J = 7.5, 4.2 Hz, 1 H, CH), 6.88 (d, J = 8.9 Hz, 2 H, ArH),7.23 (d, J = 8.6 Hz, 2 H, ArH), 7.28 (d, J = 8.6 Hz, 2 H, ArH), 7.85 (d, J = 8.9 Hz, 2 H, ArH), 9.40 (s, 1 H, OOH) ppm. ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3)$: $\delta = 203.6, 202.7, 195.3, 164.1, 135.7, 134.1,$ 130.7 (2 C), 130.6 (2 C), 129.2, 129.0 (2 C), 113.9 (2 C), 100.4, 55.6, 53.4, 52.5, 45.2, 38.6, 30.8, 30.6, 26.2 ppm. FTIR (KBr): $\tilde{v} = 3417$, 2959, 2925, 1733, 1704, 1688, 1603, 1574, 1510, 1492, 1420, 1358, 1241, 1170, 1093, 1031, 1014, 838, 822, 600 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{24}H_{23}O_5{}^{35}Cl [M - H_2O]^+$ 426.1234; found 426.1237.

2-[1,3-Bis(4-chlorophenyl)-3-oxopropyl]-2-hydroperoxy-5,5-dimethylcyclohexane-1,3-dione (5i): According to general procedure B, Michael adduct 3i (83.5 mg, 0.2 mmol) was employed. Column chromatography gave 5i as a white solid (87.2 mg, 97%). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.87$ (s, 3 H, CH₃), 1.25 (s, 3 H, CH₃), 2.44 $(dd, J = 13.5, 3.2 Hz, 1 H, CH_2), 2.57 (dd, J = 15.3, 3.2 Hz, 1 H,$ CH_2), 2.84 (d, J = 13.5 Hz, 1 H, CH_2), 3.13 (d, J = 15.3 Hz, 1 H, CH_2), 3.25 (dd, J = 18.8, 7.2 Hz, 1 H, CH_2), 3.59 (dd, J = 18.8, 4.5 Hz, 1 H, CH₂), 4.33 (dd, J = 7.2, 4.5 Hz, 1 H, CH), 7.26–7.33 (m, 4 H, ArH), 7.40 (d, J = 8.4 Hz, 2 H, ArH), 7.80 (d, J = 8.4 Hz, 2 H, ArH), 9.38 (s, 1 H, OOH) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 203.7, 202.6, 195.8, 140.3, 135.4, 134.3, 134.1, 130.6 (2 C), 129.7$ (2 C), 129.1 (2 C), 129.0 (2 C), 100.0, 53.4, 52.6, 45.0, 39.0, 30.8, 30.6, 26.1 ppm. FTIR (KBr): $\tilde{v} = 3412$, 2958, 2920, 1737, 1705, 1688, 1590, 1491, 1400, 1237, 1093, 1013, 831, 778, 724 $\rm cm^{-1}.$ HRMS (EI-TOF): calcd. for $C_{23}H_{22}O_4{}^{35}Cl_2$ [M - O]⁺ 432.0895; found 432.0899.

General Procedure C. One-Pot Conversion of Michael Adducts 3 to Products 6: Michael adduct 3a (3b–i, 0.2 mmol) was stirred in DCE (2 mL) in air at room temperature for the desired time until the starting Michael adduct had disappeared (monitored by TLC). Then, Ph₃P (0.2 mmol) was added to the reaction mixture, which was stirred at 60 °C for 30 min. The reaction mixture was separated on a silica gel column with petroleum ether/ethyl acetate (3:1) as the eluent to give product **6a** (**6b–i**).

2-Hydroxy-5,5-dimethyl-2-(3-oxo-1,3-diphenylpropyl)cyclohexane-1,3-dione (6a): According to general procedure C, Michael adduct **3a** (69.7 mg, 0.2 mmol) was employed. Column chromatography gave 6a as a white solid (67.0 mg, 92%, white solid). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3): \delta = 0.77 \text{ (s, 3 H, CH}_3), 1.31 \text{ (s, 3 H, CH}_3), 2.40$ $(dd, J = 13.6, 2.7 Hz, 1 H, CH_2), 2.53 (dd, J = 14.5, 2.7 Hz, 1 H,$ CH₂), 3.09 (d, J = 13.6 Hz, 1 H, CH₂), 3.42 (dd, J = 18.7, 6.4 Hz, 1 H, CH₂), 3.46 (d, J = 14.5 Hz, 1 H, CH₂), 3.58 (dd, J = 18.7, 5.5 Hz, 1 H, CH₂), 4.06 (s, 1 H, OH), 4.39 (t, J = 5.9 Hz, 1 H, CH), 7.22-7.29 (m, 3 H, ArH), 7.37-7.44 (m, 4 H, ArH), 7.54 (t, J = 7.2 Hz, 1 H, ArH), 7.89 (d, J = 7.4 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 205.7, 204.6, 197.9, 138.2, 136.5, 133.6, 128.9 (2 C), 128.74 (2 C), 128.68 (2 C), 128.3 (2 C), 128.1, 91.8, 51.84, 51.80, 47.3, 40.2, 31.6, 30.9, 26.5 ppm. FTIR (KBr): v = 3460, 3062, 2951, 1730, 1698, 1595, 1450, 1354, 1323, 1238, 1141, 1079, 1003, 748, 703, 686, 633, 600 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₄O₄ [M]⁺ 364.1675; found 364.1680.

2-Hydroxy-5,5-dimethyl-2-(3-oxo-3-phenyl-1-*p***-tolylpropyl)cyclohex-ane-1,3-dione (6b):** According to general procedure C, Michael adduct **3b** (72.5 mg, 0.2 mmol) was employed. Column chromatog-

raphy gave **6b** as a white solid (71.2 mg, 94%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.77$ (s, 3 H, CH₃), 1.31 (s, 3 H, CH₃), 2.27 (s, 3 H, CH₃), 2.40 (d, J = 13.4 Hz, 1 H, CH₂), 2.52 (d, J = 14.5 Hz, 1 H, CH₂), 3.09 (d, J = 13.4 Hz, 1 H, CH₂), 3.40 (dd, J = 18.7, 6.4 Hz, 1 H, CH₂), 3.43 (d, J = 14.5 Hz, 1 H, CH₂), 3.54 (dd, J = 18.7, 6.4 Hz, 1 H, CH₂), 3.43 (d, J = 14.5 Hz, 1 H, CH₂), 3.54 (dd, J = 18.7, 6.4 Hz, 1 H, CH₂), 7.07 (d, J = 7.7 Hz, 2 H, ArH), 7.25 (d, J = 7.7 Hz, 2 H, ArH), 7.41 (t, J = 7.4 Hz, 2 H, ArH), 7.53 (t, J = 7.3 Hz, 1 H, ArH), 7.88 (d, J = 7.2 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = 205.7$, 204.7, 197.9, 137.8, 136.6, 135.1, 133.5, 129.6 (2 C), 128.7 (2 C), 128.5 (2 C), 128.2 (2 C), 91.9, 51.82, 51.79, 47.0, 40.2, 31.6, 30.8, 26.5, 21.2 ppm. FTIR (KBr): $\tilde{v} = 3491, 2957, 2924, 1734, 1694, 1515, 1450, 1365, 1238, 1142, 1079, 1000, 814, 764, 689, 529 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₄H₂₆O₄ [M]⁺ 378.1831; found 378.1815.$

2-Hydroxy-2-[1-(4-methoxyphenyl)-3-oxo-3-phenylpropyl]-5,5dimethylcyclohexane-1,3-dione (6c): According to general procedure C, Michael adduct 3c (75.7 mg, 0.2 mmol) was employed. Column chromatography gave 6c as a white solid (71.8 mg, 91%). ¹H NMR (400 MHz, CDCl₃): $\delta = 0.77$ (s, 3 H, CH₃), 1.30 (s, 3 H, CH_3), 2.39 (dd, J = 13.5, 2.8 Hz, 1 H, CH_2), 2.52 (dd, J = 14.4, 2.8 Hz, 1 H, CH₂), 3.07 (d, J = 13.5 Hz, 1 H, CH₂), 3.40 (dd, J =18.6, 6.7 Hz, 1 H, CH₂), 3.42 (d, J = 14.4 Hz, 1 H, CH₂), 3.50 (dd, $J = 18.6, 5.0 \text{ Hz}, 1 \text{ H}, \text{ CH}_2$, 3.74 (s, 3 H, OCH₃), 4.05 (s, 1 H, OH), 4.35 (t, *J* = 6.0 Hz, 1 H, CH), 6.79 (d, *J* = 8.7 Hz, 2 H, ArH), 7.30 (d, J = 8.7 Hz, 2 H, ArH), 7.41 (t, J = 7.7 Hz, 2 H, ArH), 7.53 (t, J = 7.4 Hz, 1 H, ArH), 7.88 (d, J = 7.2 Hz, 2 H, ArH) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 205.7, 204.6, 198.0, 159.2, 136.5, 133.5, 130.1, 129.6 (2 C), 128.7 (2 C), 128.2 (2 C), 114.2 (2 C), 91.9, 55.2, 51.7 (2 C), 46.6, 40.2, 31.5, 30.8, 26.5 ppm. FTIR (KBr): $\tilde{v} = 3462, 2951, 2936, 1728, 1698, 1611, 1514, 1448, 1358,$ 1299, 1251, 1179, 1141, 1078, 1032, 820, 764, 733, 690 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₄H₂₆O₅ [M]⁺ 394.1780; found 394.1775.

2-[1-(4-Chlorophenyl)-3-oxo-3-phenylpropyl]-2-hydroxy-5,5-dimethylcyclohexane-1,3-dione (6d): According to general procedure C, Michael adduct 3d (76.6 mg, 0.2 mmol) was employed. Column chromatography gave 6d as a white solid (71.8 mg, 90%). ¹H NMR (300 MHz, CDCl₃): δ = 0.78 (s, 3 H, CH₃), 1.32 (s, 3 H, CH₃), 2.41 (dd, J = 13.5, 2.6 Hz, 1 H, CH₂), 2.54 (dd, J = 14.4, 2.6 Hz, 1 H, CH₂), 3.03 (d, J = 13.5 Hz, 1 H, CH₂), 3.35–3.43 (m, 2 H, CH₂), 3.50 (dd, J = 18.6, 5.4 Hz, 1 H, CH₂), 4.07 (s, 1 H, OH), 4.36 (t, J = 5.9 Hz, 1 H, CH), 7.24 (d, J = 8.4 Hz, 2 H, ArH), 7.34 (d, J = 8.4 Hz, 2 H, ArH), 7.43 (t, J = 7.5 Hz, 2 H, ArH), 7.55 (t, J = 7.5 Hz, 1 H, ArH), 7.88 (d, J = 7.5 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 205.5, 204.3, 197.6, 136.7, 136.4, 134.1, 133.7, 130.0 (2 C), 129.1 (2 C), 128.8 (2 C), 128.2 (2 C), 91.6, 51.8, 51.7, 46.6, 40.0, 31.6, 30.8, 26.5 ppm. FTIR (KBr): $\tilde{v} = 3447, 2959, 2923, 1733, 1693, 1597, 1491, 1449, 1237, 1142,$ 1080, 1016, 822, 753, 689 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃O₄Cl [M]⁺ 398.1285; found 398.1286.

2-[1-(3,4-Dichlorophenyl)-3-oxo-3-phenylpropyl]-2-hydroxy-5,5-dimethylcyclohexane-1,3-dione (6e): According to general procedure C, Michael adduct **3e** (83.5 mg, 0.2 mmol) was employed. Column chromatography gave **6e** as a white solid (80.6 mg, 93%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.78$ (s, 3 H, CH₃), 1.32 (s, 3 H, CH₃), 2.44 (dd, J = 13.5, 2.9 Hz, 1 H, CH₂), 2.54 (dd, J = 14.4, 2.9 Hz, 1 H, CH₂), 3.00 (d, J = 13.5 Hz, 1 H, CH₂), 3.36 (dd, J = 18.8, 6.6 Hz, 1 H, CH₂), 3.37 (d, J = 14.4 Hz, 1 H, CH₂), 3.50 (dd, J = 18.8, 5.3 Hz, 1 H, CH₂), 4.08 (s, 1 H, OH), 4.34 (t, J = 6.0 Hz, 1 H, CH), 7.27 (dd, J = 8.4, 2.0 Hz, 1 H, ArH), 7.50 (d, J = 2.0 Hz, 1 H, ArH), 7.66 (t, J = 7.5 Hz, 1 H, ArH), 7.88 (d, J = 7.2 Hz, 2 H,



ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 205.2, 204.2, 197.2, 138.4, 136.2, 133.8, 132.9, 132.4, 130.8, 130.6, 128.8 (2 C), 128.2 (2 C), 128.0, 91.3, 51.8, 51.6, 46.2, 39.9, 31.7, 30.8, 26.4 ppm. FTIR (KBr): \tilde{v} = 3492, 2960, 1738, 1700, 1597, 1471, 1450, 1358, 1237, 1143, 1080, 1030, 1002, 829, 757, 688, 629 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₂O₄³⁵Cl₂ [M]⁺ 432.0895; found 432.0890.

2-Hydroxy-5,5-dimethyl-2-[1-(3-nitrophenyl)-3-oxo-3-phenylpropyllcyclohexane-1,3-dione (6f): According to general procedure C, Michael adduct 3f (78.7 mg, 0.2 mmol) was employed. Column chromatography gave **6f** as a yellow solid (73.7 mg, 90%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.78$ (s, 3 H, CH₃), 1.34 (s, 3 H, CH_3), 2.44 (dd, J = 13.6, 2.8 Hz, 1 H, CH_2), 2.57 (dd, J = 14.5, 2.8 Hz, 1 H, CH₂), 3.06 (d, J = 13.6 Hz, 1 H, CH₂), 3.40 (d, J = 14.5 Hz, 1 H, CH₂), 3.45 (dd, J = 18.7, 6.9 Hz, 1 H, CH₂), 3.56 (dd, J = 18.7, 5.2 Hz, 1 H, CH₂), 4.13 (s, 1 H, OH), 4.51 (t, J =6.0 Hz, 1 H, CH), 7.41–7.50 (m, 3 H, ArH), 7.56 (t, J = 7.4 Hz, 1 H, ArH), 7.79 (d, J = 7.7 Hz, 1 H, ArH), 7.89 (d, J = 7.3 Hz, 2 H, ArH), 8.10 (d, J = 8.1 Hz, 1 H, ArH), 8.28 (s, 1 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 205.1, 204.2, 197.0, 148.4, 140.2, 136.1, 134.8, 133.9, 129.8, 128.9 (2 C), 128.3 (2 C), 123.8, 123.2, 91.2, 51.9, 51.6, 46.7, 39.8, 31.7, 30.8, 26.4 ppm. FTIR (KBr): v = 3481, 3431, 2957, 2917, 1738, 1692, 1529, 1348, 1234, 1143, 1079, 817, 754, 737, 693 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{23}H_{23}NO_6$ [M]⁺ 409.1525; found 409.1518.

2-Hydroxy-5,5-dimethyl-2-[1-(4-nitrophenyl)-3-oxo-3-phenylpropyl]cyclohexane-1,3-dione (6g): According to general procedure C, Michael adduct 3g (78.7 mg, 0.2 mmol) was employed. Column chromatography gave **6g** as a vellow solid (76.2 mg, 93%). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.79$ (s, 3 H, CH₃), 1.34 (s, 3 H, CH_3), 2.44 (dd, J = 13.6, 2.8 Hz, 1 H, CH_2), 2.58 (dd, J = 14.4, 2.8 Hz, 1 H, CH₂), 3.02 (d, J = 13.6 Hz, 1 H, CH₂), 3.39 (d, J = 14.4 Hz, 1 H, CH₂), 3.43 (dd, J = 18.8, 6.9 Hz, 1 H, CH₂), 3.54 (dd, J = 18.8, 5.2 Hz, 1 H, CH₂), 4.12 (s, 1 H, OH), 4.48 (t, J = 6.0 Hz, 1 H, CH), 7.44 (t, J = 7.6 Hz, 2 H, ArH), 7.57 (t, J = 7.8 Hz, 1 H, ArH), 7.61 (d, J = 8.8 Hz, 2 H, ArH), 7.88 (t, J =7.8 Hz, 2 H, ArH), 8.14 (d, J = 8.8, Hz, 2 H, ArH) ppm. ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3): \delta = 205.1, 204.0, 197.0, 147.7, 145.5, 134.8,$ 134.0, 129.7 (2 C), 128.9 (2 C), 128.2 (2 C), 124.0 (2 C), 91.2, 51.9, 51.6, 46.8, 39.8, 31.7, 30.8, 26.4 ppm. FTIR (KBr): v = 3490, 2952, 1732, 1693, 1596, 1515, 1449, 1353, 1237, 1143, 1080, 1002, 970, 873, 859, 752, 686 cm⁻¹. HRMS (EI-TOF): calcd. for C₂₃H₂₃NO₆ [M]⁺ 409.1525; found 409.1530.

2-[3-(4-Chlorophenyl)-1-(4-methoxyphenyl)-3-oxopropyl]-2-hydroxy-5,5-dimethylcyclohexane-1,3-dione (6h): According to general procedure C, Michael adduct 3h (82.6 mg, 0.2 mmol) was employed. Column chromatography gave **6h** as a white solid (81.5 mg, 95%). ¹H NMR (300 MHz, CDCl₃): δ = 0.77 (s, 3 H, CH₃), 1.31 (s, 3 H, CH₃), 2.40 (dd, J = 13.5, 2.7 Hz, 1 H, CH₂), 2.53 (dd, J = 14.4, 2.7 Hz, 1 H, CH₂), 3.03 (d, J = 13.5 Hz, 1 H, CH₂), 3.34 (dd, J = 18.3, 7.5 Hz, 1 H, CH₂), 3.40 (d, J = 14.4 Hz, 1 H, CH₂), 3.41 (dd, J = 18.3, 4.2 Hz, 1 H, CH₂), 3.85 (s, 3 H, OCH₃), 4.07 (s, 1 H, OH), 4.36 (t, J = 5.9 Hz, 1 H, CH), 6.89 (d, J = 8.6 Hz, 2 H, ArH), 7.24 (d, J = 8.7 Hz, 2 H, ArH), 7.33 (d, J = 8.6 Hz, 2 H, ArH), 7.85 (d, J = 8.7 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = 205.5, 204.4, 195.9, 164.0, 136.8, 134.0, 130.6$ (2 C), 130.0 (2 C), 129.5, 129.0 (2 C), 113.9 (2 C), 91.7, 55.6, 51.8, 51.7, 46.7, 39.6, 31.6, 30.8, 26.5 ppm. FTIR (KBr): $\tilde{v} = 3493$, 2962, 2929, 1732, 1694, 1605, 1574, 1510, 1493, 1422, 1359, 1324, 1261, 1240, 1169, 1143, 1093, 1079, 1034, 996, 838, 823, 677, 604, 522 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{24}H_{25}O_5^{35}Cl [M]^+ 428.1391$; found 428.1391.

2-[1,3-Bis(4-chlorophenyl)-3-oxopropyl]-2-hydroxy-5,5-dimethylcyclohexane-1,3-dione (6i): According to general procedure C, Michael adduct 3i (83.5 mg, 0.2 mmol) was employed. Column chromatography gave 6i as a white solid (81.5 mg, 94%). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 0.77$ (s, 3 H, CH₃), 1.31 (s, 3 H, CH₃), 2.41 $(dd, J = 13.5, 2.9 Hz, 1 H, CH_2), 2.53 (dd, J = 14.4, 2.9 Hz, 1 H,$ CH₂), 3.00 (d, *J* = 13.5 Hz, 1 H, CH₂), 3.32 (dd, *J* = 18.6, 6.6 Hz, 1 H, CH₂), 3.37 (d, J = 14.4 Hz, 1 H, CH₂), 3.49 (dd, J = 18.6, 5.4 Hz, 1 H, CH₂), 4.05 (s, 1 H, OOH), 4.33 (t, J = 6.0 Hz, 1 H, CH), 7.24 (d, J = 8.4 Hz, 2 H, ArH), 7.32 (d, J = 8.4 Hz, 2 H, ArH), 7.40 (d, J = 8.7 Hz, 2 H, ArH), 7.82 (d, J = 8.7 Hz, 2 H, ArH) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 205.5, 204.3, 196.4, 140.3, 136.5, 134.6, 134.1, 129.9 (2 C), 129.6 (2 C), 129.1 (4C), 91.4, 51.8, 51.7, 46.5, 40.0, 31.6, 30.8, 26.5 ppm. FTIR (KBr): $\tilde{v} = 3500$, 2962, 2948, 1734, 1692, 1590, 1491, 1404, 1357, 1342, 1236, 1203, 1173, 1144, 1095, 1080, 1016, 999, 970, 830, 760, 674, 621, 520 cm⁻¹. HRMS (EI-TOF): calcd. for $C_{23}H_{22}O_4^{35}Cl_2$ [M]⁺ 432.0866; found 432.0890.

X-ray Crystallographic Analysis of 6d:[11-13] Compound 6d was crystallized as white crystals from EtOAc with the diffusion of petroleum ether at 0 °C in a refrigerator for two weeks. C23H22Cl2O4, $M_{\rm r}$ = 433.31, T = 291(2) K, triclinic, space group $P\bar{1}$, a = 10.9377(5) Å, b = 10.9571(5) Å, c = 11.0854(6) Å, $a = 67.781(5)^{\circ}$, $\beta = 61.343(5)^\circ$, $\gamma = 86.701(6)^\circ$, $V = 1066.63(9) \text{ Å}^3$, Z = 2, $\rho_{\text{calcd.}} =$ 1.349 g cm^{-3} , crystal size: $0.50 \times 0.40 \times 0.40 \text{ mm}$, F(000) = 452, μ (Mo- K_{α}) = 0.331 mm⁻¹, min. transmission 0.8521, max. transmission: 0.8791, $2\theta_{\text{max.}} = 52.04^\circ$, 8824/4198 measured/independent reflections [R(int) = 0.0184], 3196/266 reflections used/number of parameters, refinement against |F|, GOF = 1.067, final R indices were $R_1 = 0.0333 \ [I > 2\sigma(I)]$ and $wR_2 = 0.0877 \ [I > 2\sigma(I)]$. Mo- K_{α} radiation ($\lambda = 0.71073$). The data collection routine, unit-cell refinement, and data processing were carried out with the program CrysAlis.^[11] The structure was solved by direct methods and refined by full-matrix least-squares methods with the SHELXL-97 programs.^[12]

Supporting Information (see footnote on the first page of this article): Copies of ¹H and ¹³C NMR spectra of compounds **3a–i**, **4a**, **5a–i**, and **6a–i**.

Acknowledgments

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