Highly Efficient Allylation of Aldehydes Promoted by Maleic Acid in Aqueous Media

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Abstract: A highly efficient promoter for allylation of aldehydes in aqueous media was developed. Under the promotion of maleic acid, the allylation of various aldehydes can be finished in a short period of reaction time to afford the corresponding homoallylic alcohols in high to quantitative yields.

Key words: aldehydes, allylation, aqueous media, allyltributyltin, maleic acid

The allylation of aldehydes, as a very important method for forming synthetically useful homoallylic alcohols, has subjected extensive investigation.¹ In recent years, there has been an upsurge of interest in using water as the solvent in terms of both economical and ecological aspects.² Yamamoto et al. described that using 5 mol% of tetraallyltin or SnCl₄ could catalyze the allylation of aldehydes and ketones with allyltributyltin in aqueous HCl solution and THF.³ Since then, there have been several examples reported in this field.⁴

Recently, during our studies of allylation of carbonyl compounds, we found that carboxylic acids with appropriate acidity and solubility are general, practical and highly efficient promoters for the allylation of aldehydes with allyltributyltin in acetonitrile (Equation 1).⁵ In this allylation, *p*-nitrobenzoic acid gave nearly quantitative yields, other stronger carboxylic acids were found much faster than *p*-nitro benzoic acid, but lower yields were obtained due to the decomposition of allyltributyltin during the allylation. Moreover, when maleic acid was used as a promoter, the allylation was finished within one hour in a yield of 81%. Herein, we report that maleic acid can be used as a highly efficient promoter for the allylation of aldehydes with allyltributyltin in aqueous media under mild reaction conditions.

We have reported originally that the allylation, which was mediated by *p*-nitrobenzoic acid (insoluble in water), was very sluggish in water⁵ (entry 1, Table 1). So, in order to carry out this allylation in aqueous media, two crucial factors (the solubility and acidity of the carboxylic acids) are taken into consideration. In the first step, several watersoluble carboxylic acids were screened according to their pK_a values. The reaction of benzaldehyde (0.5 mmol) and





allyltributyltin (0.6 mmol) was promoted under various carboxylic acids (0.5 mmol) in water (1.0 mL) at room temperature. The results were summarized in Table 1. The weaker carboxylic acids ($pK_a \ge 2.85$) only gave trace amounts of allylation product of 3a (entries 2-5). Comparatively, with the decrease of pK_a values, the stronger carboxylic acids (pK_a \leq 2.83) provided good to high yield of the desired product (entries 6-8). Among these carboxylic acids, maleic acid gave the best result with a yield of 95% within a short period of reaction time (1.25 h, entry 7). The further decrease of pK_a value of the carboxylic acid, for example, using trifluoroacetic acid as a promoter $(pK_a = 0.23, entry 8)$, resulted in a lower allylation yield (89%) due to the fact that allyltributyltin was partially decomposed under such a strong acidic reaction condition. Reducing the amount of maleic acid to 0.5 equivalents only a yield of 66% (3a) was obtained (entry 9). It is noteworthy that compared with the fact that the tin reagent was totally decomposed in acetonitrile within one hour in the presence of maleic acid (monitrored by TLC); only little amount of allyltributyltin was decomposed in deuterium oxide within 23 hours under the promotion of maleic acid (monitored by ¹H NMR in deuterium oxide). And the reaction in aqueous phase is slower than that in MeCN.⁵

Next, a variety of aldehydes were examined. As shown in Table 2, apparently, the electronic nature of the substitutes of aromatic aldehydes has little effect on the high yields (entries 1-6). Salicylaldehyde is soluble in water, which may benefit the allylation (entry 6). As for the aliphatic aldehydes, the allylation promoted under maleic acid in aqueous medium also exhibited high efficiency. For example, the allylation of cyclohexanecarboxaldehyde afforded 3h in a yield of 84% within 110 minutes (entry 7). Nonyl aldehyde needed a longer period of reaction time to complete the allylation (83%, entry 8) and this can be rationalized on the basis of its hydrophobic property. In the mixed solvent (THF– $H_2O = 2:8$, v/v), the allylation of water-insoluble *p*-nitrobenzaldehyde could proceed smoothly to produce the corresponding product in quantitative yield (entry 9). When a reaction using acetophenone as a substrate was performed, no reaction

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 Table 1
 Screening of Carbonic Acid Promoters for Allylation of Benzaldehyde in Aqueous Media^a

	O H _ SnBu ₃ .	carboxylic a (1 equiv) H ₂ O, r.t.	cid	OH
~ 1a	2		~	3a
Entry	Carboxylic acid	pK _a ⁶	Time (h)	Yield (%) ^b
1	p-Nitrobenzoic acid	3.42	23	Trace
2	Succinic acid	4.21	23	Trace
3	L-(+)-Tartaric acid	3.22	23	Trace
4	Citric acid	3.17	8	Trace
5	Chloroacetic acid	2.85	1.25	Trace
6	Malonic acid	2.83	8	53
7	Maleic acid	1.92	1.25	95
8	Trifluoroacetic acid	0.23	1.25	89
9	Maleic acid	1.92	5	66 ^c

^a Ratio: 1a:2 = 1 equiv (0.5 mmol):1.2 equiv (0.6 mmol).

^b Isolated yields.

^c 0.5 Equiv (0.25 mmol) of maleic acid was used as promoter.

occurred. In addition, the maleic acid was also efficient for allylation of α , β -unsaturated aldehyde (quantitative yield, entry 10). Finally, the reaction of the crotylation of benzaldehyde using maleic acid as a promoter in water was carried out. The yield was got in 69%; however, the regioselectivity was poor (α -adduct: γ -adduct = 1.77:1.98).

In summary, we have developed a highly efficient promoter for the allylation of aldehydes in aqueous media. Under the promotion of maleic acid, the allylation of various aldehydes was very fast and afforded high to quantitative yields of desired products in a short period of reaction time under mild conditions.

All the ¹H NMR and ¹³C NMR spectra were recorded with TMS as an internal standard on a Bruker AM-300 or Varian EM-360 (300 and 75 MHz, respectively). IR spectra were recorded on a Nicolet AV360 spectrometer. EI Mass spectra were determined on an HP-5989A spectrometer. Elemental analysis for carbon and hydrogen were performed on a Rapid CHN-O analyzer.

Allyltributyltin was prepared by the reaction of allyl Grignard with *n*-Bu₃SnCl. All carboxylic acids, aldehydes and solvents are commercially available and were used without further purification.

Typical Procedure for the Allylation of Aldehydes

To a solution of maleic acid (57 mg, 0.5 mmol) in $\rm H_2O$ (1.0 mL) were added successively 4-n-butoxybenzaldehyde (89 mg, 0.5

 Table 2
 The Scope of Aldehydes in the Allylation Promoted by Maleic Acid^a

maleic acid (1 equiv) maleic acid (1 equiv) 1a-i 2 3a-j Entry Aldehydes Time (min) Product ^b Yield (%) 1 C_6H_3CHO 75 3a 95 2 m -MeOC ₆ H ₄ CHO 80 3b 97 3 $3,4,5$ - $F_3C_6H_2CHO$ 70 3c 86 4 p - n -BuOC ₆ H ₄ CHO 120 3d 97 5 p - FC_6H_4CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c - $C_6H_{11}CHO$ 110 3g 84 8 n - $C_8H_{17}CHO$ 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.			OH			
RCHO + 2 H_2O , r.t. R $X \ll X$ 1a-i 2 $3a-j$ Entry Aldehydes Time (min) Product ^b Yield (%) 1 C_6H_5CHO 75 $3a$ 95 2 m -MeOC ₆ H ₄ CHO 80 $3b$ 97 3 $3,4,5$ - $F_3C_6H_2CHO$ 70 $3c$ 86 4 p - n -BuOC ₆ H ₄ CHO 120 $3d$ 97 5 p -FC ₆ H ₄ CHO 75 $3e$ 94 6 Salicylaldehyde 110 $3f$ Quant. 7 c - $C_6H_{11}CHO$ 110 $3g$ 84 8 n - $C_8H_{17}CHO$ 6 h $3h$ 83 9 p -NO ₂ C ₆ H ₄ CHO 150 $3i$ Quant. ^d 10 E -PhCH=CHCHO 120 $3j$ Quant.		s SpBu	maleic acid (1 equ	iiv)		
1a-i 2 3a-j Entry Aldehydes Time (min) Product ^b Yield (%) 1 C_6H_3CHO 75 3a 95 2 m -MeOC ₆ H ₄ CHO 80 3b 97 3 $3,4,5$ - $F_3C_6H_2CHO$ 70 3c 86 4 p - n -BuOC ₆ H ₄ CHO 120 3d 97 5 p -FC ₆ H ₄ CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c -C ₆ H ₁₁ CHO 110 3g 84 8 n -C ₈ H ₁₇ CHO 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.	RCHO	+	H ₂ O, r.t.	—► R.	~ ~	
Entry Aldehydes Time (min) Product ^b Yield (%) 1 C_6H_3CHO 75 3a 95 2 m -MeOC ₆ H ₄ CHO 80 3b 97 3 $3,4,5$ - $F_3C_6H_2CHO$ 70 3c 86 4 p - n -BuOC ₆ H ₄ CHO 120 3d 97 5 p - FC_6H_4CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c - $C_6H_{11}CHO$ 110 3g 84 8 n - $C_8H_{17}CHO$ 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant. ^d	1a–i	2		3a–j		
1 C_6H_5CHO 75 3a 95 2 $m-MeOC_6H_4CHO$ 80 3b 97 3 $3,4,5-F_3C_6H_2CHO$ 70 3c 86 4 $p-n-BuOC_6H_4CHO$ 120 3d 97 5 $p-FC_6H_4CHO$ 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 $c-C_6H_{11}CHO$ 110 3g 84 8 $n-C_8H_{17}CHO$ 6 h 3h 83 9 $p-NO_2C_6H_4CHO$ 150 3i Quant. ^d 10 $E-PhCH=CHCHO$ 120 3j Quant.	Entry	Aldehydes	Time (min)	Product ^b	Yield (%) ^c	
2 m -MeOC ₆ H ₄ CHO 80 3b 97 3 $3,4,5$ - $F_3C_6H_2CHO$ 70 3c 86 4 p - n -BuOC ₆ H ₄ CHO 120 3d 97 5 p -FC ₆ H ₄ CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c -C ₆ H ₁₁ CHO 110 3g 84 8 n -C ₈ H ₁₇ CHO 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.	1	C ₆ H ₅ CHO	75	3a	95	
3 $3,4,5-F_3C_6H_2CHO$ 70 $3c$ 86 4 $p-n$ -BuOC ₆ H ₄ CHO 120 $3d$ 97 5 $p-FC_6H_4CHO$ 75 $3e$ 94 6 Salicylaldehyde 110 $3f$ Quant. 7 $c-C_6H_{11}CHO$ 110 $3g$ 84 8 $n-C_8H_{17}CHO$ 6 h $3h$ 83 9 $p-NO_2C_6H_4CHO$ 150 $3i$ Quant. ^d 10 E -PhCH=CHCHO 120 $3j$ Quant.	2	<i>m</i> -MeOC ₆ H ₄ CHO	80	3b	97	
4 p - n -BuOC ₆ H ₄ CHO 120 3d 97 5 p -FC ₆ H ₄ CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c -C ₆ H ₁₁ CHO 110 3g 84 8 n -C ₈ H ₁₇ CHO 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.	3	3,4,5-F ₃ C ₆ H ₂ CHO	70	3c	86	
5 p -FC ₆ H ₄ CHO 75 3e 94 6 Salicylaldehyde 110 3f Quant. 7 c -C ₆ H ₁₁ CHO 110 3g 84 8 n -C ₈ H ₁₇ CHO 6 h 3h 83 9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.	4	<i>p-n-</i> BuOC ₆ H ₄ CHO	120	3d	97	
6 Salicylaldehyde 110 3f Quant. 7 c - C_6H_{11} CHO 110 3g 84 8 n - C_8H_{17} CHO 6 h 3h 83 9 p -NO ₂ C_6H_4 CHO 150 3i Quant. ^d 10 E -PhCH=CHCHO 120 3j Quant.	5	<i>p</i> -FC ₆ H ₄ CHO	75	3e	94	
7 $c-C_6H_{11}CHO$ 110 3g 84 8 $n-C_8H_{17}CHO$ 6 h 3h 83 9 $p-NO_2C_6H_4CHO$ 150 3i Quant. ^d 10 $E-PhCH=CHCHO$ 120 3j Quant.	6	Salicylaldehyde	110	3f	Quant.	
8 $n-C_8H_{17}CHO$ 6 h 3h 83 9 $p-NO_2C_6H_4CHO$ 150 3i Quant. ^d 10 <i>E</i> -PhCH=CHCHO 120 3j Quant.	7	<i>c</i> -C ₆ H ₁₁ CHO	110	3g	84	
9 p -NO ₂ C ₆ H ₄ CHO 150 3i Quant. ^d 10 <i>E</i> -PhCH=CHCHO 120 3j Quant.	8	<i>n</i> -C ₈ H ₁₇ CHO	6 h	3h	83	
10 <i>E</i> -PhCH=CHCHO 120 3j Quant.	9	<i>p</i> -NO ₂ C ₆ H ₄ CHO	150	3i	Quant. ^d	
	10	E-PhCH=CHCHO	120	3j	Quant.	

^a Ratio: 1:2 = 1 equiv (0.5 mmol):1.2 equiv (0.6 mmol).

^b All products were identified by ¹H NMR, IR, MS.

^c Isolated yields.

^d Solvent: THF– $H_2O = 0.2:0.8$ (mL).

mmol) and allyltributyltin (198 mg, 0.6 mmol) at r.t. After its completion (120 min, monitored by TLC), the reaction was quenched with aq NaOH (2 M, 2 mL). Then extracted with CH_2Cl_2 (3 × 5 mL). The combined organic layer was dried over Na_2SO_4 and concentrated under reduced pressure to furnish the crude product, which was purified by silica gel chromatography [typical eluent: petroleum ether–EtOAc = 15:1(v/v)] to afford 1-(4-*n*-butoxyphenyl)-3-buten-1-ol (**3d**).

1-(4-n-butoxyphenyl)-3-buten-1-ol (3d)

Yield 107 mg, 97%; $R_f = 0.81$ (petroleum ether–EtOAc = 7:1). ¹H NMR (300 MHz, CDCl₃): δ = 7.25 (d, 2 H, J = 8.3 Hz), 6.86 (d, 2 H, J = 8.5 Hz), 5.84–5.75 (m, 1 H), 5.18–5.11 (m, 2 H), 4.66 (t, 1 H, J = 6.4 Hz), 3.93 (t, 2 H, J = 6.3 Hz), 2.47 (t, 2 H, J = 6.2, 6.8 Hz), 1.97 (s, 1 H), 1.81–1.72 (m, 2 H), 1.53–1.45 (m, 2 H), 0.95 (t, 3 H, J = 7.8, 6.9 Hz). ¹³C NMR (75 MHz, CDCl₃): δ = 158.7, 135.9, 134.6, 127.0, 118.0, 114.4, 73.0, 67.8, 43.6, 31.3, 19.2, 13.7. IR (KBr, film): v = 3415, 3075, 2959, 2873, 1640, 1612, 1512, 1240 cm⁻¹. MS (EI, 70 eV): m/z = 220 [M⁺], 179, 123, 95, 77, 41. Anal. Calcd for C₁₄H₂₀O₂ (220.31): C, 76.33; H, 9.15. Found: C, 75.94; H, 9.31.

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