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Bioorganic & Medicinal Chemistry Letters xxx (2017) xxx-xxx

Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Synthesis, conformational study and antiviral activity of L-like neplanocin derivatives

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ARTICLE INFO

Article history: Received 13 June 2017 Revised 31 July 2017 Accepted 5 August 2017 Available online xxxx

Keywords: Carbocyclic nucleosides L-like nucleosides Nucleoside conformation Neplanocin Antiviral

ABSTRACT

The L-like enantiomer of 9-(trans-2', trans-3'-dihydroxycyclopent-4'-enyl)-3-deazaadenine (DHCDA) (1), its 3-deaza-3-bromo derivative (3), and the conformational restricted methanocarba (MC) nucleoside analogues (2 and 4) were synthesized. X-ray crystal structures showed the L isomer MC analogue 4 adopts a similar North-like locked conformation as conventional D-MC nucleosides, while the DHCDA analogue 3 preferred south-like conformer. Compounds 1 and 4 showed potent antiviral activity against norovirus, while compound 2 and 3 were less potent or inactive. The conformational behavior of "sugar" puckering (north/south) and nucleobase orientation (syn /anti) may contribute to the antiviral activity differences. For compound 3, antiviral activity was also found against Ebola virus.

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Introduction

Nucleoside analogues play a vital role in chemotherapy of viral infectious diseases.¹ D-like carbocyclic² nucleosides, such as abacavir³ and entecavir⁴ (Fig. 1) with the similar configuration as naturally occurring nucleosides, effectively interact with target enzymes and, thus, possess interesting biological activities.^{3,4} Despite acting as conventional polymerase inhibitors, the antiviral properties of adenine-derived carbocyclic nucleosides can act by the inhibition of S-adenosylhomocysteine (AdoHcy) hydrolase, a cellular enzyme involved in controlling viral transcription by regulating the mRNA cap structures.^{2,5} However, the development of such carbocyclic nucleosides as therapeutics is limited by cytotoxicity that commonly arises from its C-5' phosphate metabolites arising from various cellular enzymes.⁵ In order to seek ways to circumvent this metabolic pathway, and, hence decrease unwanted toxicity, the uncommon L-like carbocyclic enantiomeric nucleosides arose.⁶

Recently, we reported the D-like (**5**) and L-like (**6**) enantiomers of 3-deaza- and 3-deaza-3-bomo-1', 6'-isoneplanocin (Fig. 2), which have been found to possess potent and broad spectrum antiviral activities. Interestingly, the L-isomers (**6**) were equally or more potent than the D-isomers against human cytomegalovirus, hepatitis B virus (HBV), norovirus, measles and Ebola virus.^{7,8}

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http://dx.doi.org/10.1016/j.bmcl.2017.08.009 0960-894X/© 2017 Elsevier Ltd. All rights reserved. To further investigate L-like carbocyclic nucleosides, based on compound **6**, we sought the analogues of DHCDA (that is **1** and **3**).⁹ DHCDA is a neplanocin derivative whose broad spectrum antiviral activities were attributed to the inhibition of AdoHcy hydrolase, and reduced cytotoxicity due to lacking the 5' hydroxyl group necessary for the 5'-phosphorylation and circumventing antiviral activity through polymerase inhibition.

It has been well recognized that the conformational behavior of ribonucleosides and modified nucleosides, including those in the category carbocyclic, owe their biological properties to sugar puckering and the orientation of the heterocyclic base. Among these properties are their metabolic pathways, enzyme binding affinity, and therapeutic effects and toxicity.¹⁰ The sugar puckering of ribo-furanosyl nucleosides is known to exist in dynamic equilibrium between north (3'-endo) and south (2'-endo) conformations. In most the cases, binding affinity to a pharmacological target favors one conformer over the other. Furthermore, nucleosides adopt either an anti or a syn nucleobase orientation. It has been reported¹¹ that nucleoside analogues with a syn orientation



Fig. 1. Examples for carbocyclic nucleosides.

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Fig. 2. D and L-like 1', 6'-isoneplanocin derivatives and designed targer compounds.

exhibited much lower binding affinities to adenosine based receptors. Incorporating these parameters in our study, the 3-bromo substitution in **2** is envisioned to increase the stereo hindrance significantly and force the nucleobase to adopt an anti conformation (Fig. 3). In that direction, methanocarba (MC) nucleoside analogues (**2** and **4**, Fig. 2) were also included as target compounds in this study. The bicyclic[3.1.0]hexane ring in **2** and **4** was expected to constrain the pseudo sugar component and result in locked north-like conformers,¹² which are expected to show the different antiviral profile comparing with conformers **1** and **3**. Further Structure-Activity Relationship (SAR) study can be expected to provide valuable information for exploring the mechanism of carbocyclic nucleosides as antiviral agents.

We envisioned a versatile retrosynthetic plan (Scheme 1) to the proposed targets. The plan involved a coupling reaction between two main components: a cyclopentenol pseudo-sugar potion and a modified purine base. In the latter regard, synthesis of the 3-deazapurine base **7** was accomplished by adapting known procedures using 2-chloro-3,4-diaminopyridine (**11**) as the starting material (Scheme 2).^{13,14} Construction of the carbocyclic coupling



Fig. 3. Syn and Anti conformers of compound 3.



Scheme 1. Retrosynthesis design for trager compounds.



Scheme 3. Synthesis of cyclopentenol derivatives. Reaction and conditions: a. LiAlH₄, CeCl₃, THF, 95%; b. Diethylzinc, CH₂l₂, DCM, 70%.



Fig. 4. Stereochemistry of bicycle[3.1.0]hexanol 9.

precursors **8** and **9** was achieved from L-cyclopentenone **10**¹⁵ via a Luche condition reduction (for **8**), and a Simmons Smith reaction (for **9**) (Scheme 3).^{16,17} The stereo chemistry of compound **9**¹⁸ was confirmed by ROSEY (Fig. 4¹⁸) and X-ray crystallography of compound **4**.

Mitsunobu coupling reaction was preformed between **7** and L-form cyclopentenol **8** (Scheme 4) or **9** (Scheme 5) to provide coupled products **15** and **16**, which, upon deprotection, gave 1^{19} and **2**.²⁰ 3-Bromo functionalized targets 3^{21} and 4^{22} were accessible from **1** and **2** via suitable bromination conditions.

In addition to NMR data, X-ray crystallography structures²³ of **3** and **4** further confirmed the regiospecific bromination at the C-3 position of **1** and **2**.

Crystal structures also showed **3** adopting a south (3'-exo) conformation when posed with similar orientation with D-nucleosides, while more structural rigid bicyclo[3.1.0] hexane locked **4** into an north (2'-exo) conformation in the solid state (Fig. 5). As expected, the bromo group at the C-3 position forced both **3** and **4** to adopt the less congested anti conformation.

Compounds **1–4** were assayed for their antiviral potential.^{24,25} Compounds **1** and **4** displayed activity against norovirus. Com-



Scheme 2. 13,14Synthesis of 3-deazapurine base. Reaction and conditions: a. CH(OMe)₃, 85%; b. i)NH₂NH₂ ii) Raney Ni, H₂O, 70%; c. (Boc)₂O, THF, 90%; d. TBAF, THF, 88%.

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Scheme 4. Synthesis of compound 3. Reaction and conditions: a. DIAD, PPh3, THF, 77%; b. 1 M HCI/MeOH, 87%; c. NBS, DCM/DMF, 74%.



Scheme 5. Synthesis of compound 4. Reaction and conditions: a. DIAD, PPh₃, THF, 56%; b. 1 M HCl/MeOH, 81%; c. NBS, DCM/DMF, 71%.



Fig. 5. North and South conformation for compound 3 and 4.

pound **1** (EC₅₀ = 4.2 μ M) was more potent than its bromo analogue **3** (EC₅₀ = 33 μ M). On the other hand, the antiviral profile against norovirus for MC nucleosides **2** and **4** were reversed. In contrast to **1**, compound **2** (EC₅₀ > 100 μ M) lost anti-norovirus activity, the bromo derivative **4** (EC₅₀ = 3.0 μ M) showed stronger viral inhibition than **3**. As evidenced in the crystal structures, **3** and **4** are locked into the anti-conformation in solid state, while **1** and **2** are more flexible. The most active norovirus **4** represents the anti-north L-like nucleosides and suggests more extensive study is in order. In comparison with the D-like nucleosides, compound DHCDA (**1**) showed antiviral activity against vaccinia virus (EC₅₀ = 0.7 μ g/mL)²⁶ and vesicular stomatitis virus (EC₅₀ = 0.2 μ g/mL)²⁶, but no anti-norovirus activity was reported to our knowledge.

Additionally, **3** (EC₅₀ = 8.3 μ g/mL) showed stronger activity against Ebola virus than **1** (EC₅₀ = 29.5 μ g/mL), while MC analogues **2** and **4** were completely inactive. Compound **3** displayed toxicity related mild active towards the flavivirus family in vero cell line.²⁷ Compounds **1**,**2** and **4** showed no apparent toxicity towards the testing cell lines.

No effects were found for **1–4** towards AdoHcy hydrolase $(IC_{50} > 10^5 \text{ mM} \text{ for all compounds})$ which most likely correlated with the activity of p-carbocyclic nucleosides.²

Acknowledgments

We are grateful to the Slippery Rock University, Faculty/Student Research Grants and to Stewart W. Schneller and Chong Liu at Auburn University for support of this research. We are indebted to the NIAID in vitro assay team for the viral data presented herein: Don Smee, Utah State University; Brent Korba, Georgetown University; Mark Prichard, University of Alabama – Birmingham; Michael Murray, Southern Research Institute and to We also appreciate the assistance of Steven Cardinale and Terry Bowlin of Microbiotix, Inc. for providing the S-adenosylhomocysteine hydrolase data and John Gorden at Auburn University for X-ray crystallography analysis.

References

- 1. De Clercq E. Nat Rev Drug Discovery. 2002;1:13.
- 2. De Clercq E. Nucleosides Nucleotides. 1998;17:625.
- 3. Curran A, Ribera E. Expert Opin Drug Saf. 2011;10:389.
- 4. Campian M, Putala M, Sebesta R. Curr Org Chem. 2014;18:2808.
- 5. De Clarcq E. Nucleosides Nucleotides. 2005;24:1395.
- 6. Schneller SW. Rec Adv Nucleosides. 2002;291–297.
- 7. Liu C, Chen Q, Schneller SW. Bioorg Med Chem Lett. 2016;26:928.
- 8. Schneller SW, Liu C, Chen Q, Ye W. PCT Int. Appl. 2016, WO 2016022563 A1; US 9,657,048.
- 9. Andrei G, De Clercq E. Antiviral Res. 1990;14:287.
- 10. Gagneron J, Gosselin G, Mathe C. J Org Chem. 2005;70:6891.
- 11. Yu J, Zhao LX, Park J, et al. J Med Chem. 2017;60:3422.
- 12. Comin MJ, Agbaria R, Ben-Kasus T, et al. J Am Chem Soc. 2007;129:6216.
- 13. Liu C, Chen Q, Schneller SW. Bioorg Med Chem Lett. 2012;22:5182.
- 14. Crey-Desbiolles C, Kotera M. Bioorg Med Chem. 2006;14:1935.
- 15. Smith ABIII, Han Q, Dreslin PA, Beauchamp GK. Org Lett. 2005;7:5075.
- 16. Nayak A, Chandra G, Hwang I, et al. J Med Chem. 2014;57:1344.
- 17. Choi MJ, Chandra G, Lee HW, et al. Org Biomol Chem. 2011;9:695
- 18. Structural assignments for **9** were accomplished by ¹H COSY and the stereochemistry was assigned by a ROSEY, which shows strong NOE correlations between H-1 and H-5, H-3 and H-4, H4 and H-5 protons (Fig. 4).
- 19. Compound **1**, white solid; ¹H NMR (600 MHz, MeOD): δ 8.07 (s, 1H), 7.68 (d, J = 6.1 Hz, 1H), 6.90 (d, J = 6.1 Hz, 1H), 6.32 (dt, J = 6.2, 2.52 Hz, 1H), 6.20 (dd, J = 6.2, 1.7 Hz, 1H), 5.41 (m, 1H), 4.66 (dd, J = 2.7, 1.3 Hz, 1H), 4.65 (dd, J = 2.6, 1.3 Hz, 1H); ¹³C NMR (150.9 MHz, MeOD): δ 153.5, 142.3, 141.0, 140.2, 137.9, 133.7, 128.4, 99.5, 78.9, 74.5, 68.3; HRMS Calcd for C₁₁H₁₃N₄O₂ [M+H]⁺ 233.1039, found 233.1035. Anal. Calcd for C₁₁H₁₂N₄O₂. 0.3H₂O: C, 55.60; H, 5.34; N, 23.58. found: C, 55.83; H, 5.32; N, 23.34.
- 20. Compound **2**, white solid; ¹H NMR (600 MHz, MeOD): δ 8.24 (s, 1H), 7.73 (d, J = 6.0 Hz, 1H), 6.97 (d, J = 6.0 Hz, 1H), 4.68 (d, J = 1.2 Hz, 1H), 4.60 (t, J = 6.0 Hz, 1H), 3.84 (d, J = 6.6 Hz, 1H), 2.02 (m, 1H), 1.77 (m, 1H), 1.84 (dd, J = 9.0, 4.2 Hz, 1H), 0.83 (m, 1H); ¹³C NMR (150.9 MHz, MeOD): δ 153.5, 141.5, 141.1, 140.0, 128.1, 98.8, 77.9, 72.9, 65.5, 24.7, 19.6, 8.43; HRMS Calcd for C₁₂H₁₅N₄O₂ [M+H]* 247.1195, found 247.1188.
- Compound 3, white solid; ¹H NMR (600 MHz, MeOD): δ 8.11 (s, 1H), 7.78 (s, 1H), 6.36 (m, 1H), 6.29 (m, 1H), 6.13 (m, 1H), 4.71 (m, 1H), 4.27 (m, 1H); ¹³C

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NMR (150.9 MHz, MeOD): δ 153.2, 143.6, 142.1, 138.4, 137.2, 133.6, 129.7, 90.9, 79.8, 74.8, 67.2; HRMS Calcd for C₁₁H₁₂N₄O₂Br [M+H]⁺ 311.0144, found 311.0132. Anal. Calcd for C₁₁H₁₁N₄O₂Br: C, 42.46; H, 3.56; N, 18.01. found: C, 42.63; H, 3.63; N, 17.74.

- Compound 4, white solid; ¹H NMR (600 MHz, MeOD): δ 8.32 (s, 1H), 7.77 (s, 1H), 4.64 (d, J = 1.2 Hz, 1H), 4.58 (t, J = 5.4 Hz, 1H), 3.94 (d, J = 6.0 Hz, 1H), 2.0 (m, 1H), 1.76 (m, 1H), 1.40 (dd, J = 9.0, 4.2 Hz, 1H), 0.79 (m, 1H); ¹³C NMR (150.9 MHz, MeOD): δ 151.7, 141.9, 140.6, 135.0, 128.3, 89.3, 75.9, 71.4, 63.6, 22.9, 19.2, 6.69; HMSS Calcd for C₁₂H₁₄N₄O₂Br [M+H]* 325.0300, found 325.0313.
- 23. Crystallographic data (excluding structure factors) for 3 and 4 has been deposited with the Cambridge Crystallographic Data Centre with the following deposition number CCDC 1544960 and CCDC 1544961 respectively. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK, fax: +44 (0)1223 336033 or e-mail: deposit@ccdc.cam. ac.uk.
- 24. For the antiviral and hydrolase assay details: Chen Q, Liu C, Komazin G, Bowlin SW, Schneller SW. *Bioorg Med Chem*. 2014;22:6961.
- 25. No significant activities were found for 1-4 towards Adenovirus, Chikungunya virus, Dengue virus, Poliovirus, Punta Toro virus, Rift Valley fever virus, Tacaribe virus, Usutu virus, Venezuelan equine encephalitis virus, West Nile virus, Yellow fever virus, Japanese encephalitis virus, Zika virus and measles virus.
- 26. De Clercq E, Cools M, Balzarini J, et al. Antimicrob Agents Chemother. 1989;33:1291.
- 27. Compound **3** towards Dengue virus (Vero IC₅₀=7.2 μg/mL, CC₅₀=28 μg/mL); Rift Valley fever virus (Vero IC₅₀=31 μg/mL, CC₅₀=39 μg/mL); Venezuelan equine encephalitis virus (Vero IC₅₀=27 μg/mL, CC₅₀=27 μg/mL); West Nile virus (Vero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Yellow fever virus (Vero IC₅₀=32 μg/mL, CC₅₀=32 μg/mL); Japanese encepahalitis virus (Vero IC₅₀=32 μg/mL, CC₅₀=32 μg/mL); Japanese encepahalitis virus (Vero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Mest Nile virus (Vero IC₅₀=32 μg/mL); Vero IC₅₀=32 μg/mL); Japanese encepahalitis virus (Vero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Cero IC₅₀=32 μg/mL); Vero IC₅₀=32 μg/mL); Cero IC₅₀ Ag/mC); Cero IC₅₀=30 µg/mL); Cero IC₅₀ Ag/mC); Cero IC₅₀ Ag/mC); Cero IC₅₀ Ag/mC