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Stereospecific fluorination of 1,3,5-tri-*O*-benzoyl-α-D-ribofuranose-2-sulfonate esters: preparation of a versatile intermediate for synthesis of 2'-[¹⁸F]-fluoro-arabinonucleosides

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Abstract

A detailed investigation on fluorination of 1,3,5-tri-*O*-benzoyl- α -D-ribofuranose-2-sulphonate esters is reported. Various combinations of sulfonate esters, fluorinating agents and solvents were evaluated in this study. Organic ammonium fluoride, in particular *n*-Bu₄NF, was found to be better fluorinating agent than inorganic fluoride, and 1,3,5-tri-*O*-benzoyl- α -D-ribofuranose-2-trifluoromethylsulphonate ester appeared to be the best substrate. The developed method is suitable for stereospecific (*arabino*) incorporation of radiofluorine (¹⁸F) into the sugar moiety. © 2000 Elsevier Science S.A. All rights reserved.

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1. Introduction

Pyrimidine-1-(2'-deoxy-2'-fluoro-β-D-arabinofuranosyl)nucleosides and their radiolabeled derivatives are potentially important compounds [1–5]. 2'-Fluoro-5-[¹¹*C*-methyl]-1-β-D-arabinofuranosyluracil (¹¹C-FMAU) **1a** (Fig. 1) is being studied as a marker for cell proliferation by positron emission tomography (PET) [3,4]. In addition, radiolabeled 2'fluoro-5-iodo-1-β-D-arabino-furanosyluracil (FIAU) **1b** is considered to be a useful agent for imaging the expression of transfected genes in vivo [5]. ¹¹C-FMAU **1a** was first synthesized in our laboratory [3], in quantities suitable for animal and patient studies. However, the short half-life of C-11 ($t_{1/2}$ =20 min) limits the clinical application of that compound. Therefore, labeling with the longer-lived PET isotope fluorine-18 ($t_{1/2}$ =110 min) in the glycone would be more advantageous.

The direct, stereospecific (*arabino*) introduction of fluorine by an $S_N 2$ substitution at the 2'-position of the furanosyl moiety in a pyrimidine nucleoside has not been possible [1,6-8]. Alternatively, the incorporation of fluorine in the *arabino* configuration at C-2 of the sugar followed by coupling with the pyrimidine base has been successful [1,9-12]. The common methods for the stereospecific (*arabino*) incorporation of fluorine involve use of KHF₂ or Et₃N·3HF as fluorinating agent in excess (6 eq) [11,13], and require long reaction times (6 h). These methods are not, however, satisfactory for incorporation of radiofluorine.



Fig. 1. Structure of FMAU (1a) and FIAU (1b).

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Fig. 2. General methodology of fluorination reactions.

Earlier, we developed a method suitable for stereospecific (*arabino*) incorporation of radioactive fluorine in the 2-position of the protected sugar [14,15]. The chemical yield was variable, but could be as high as 30%. However, the radiochemical yield was quite low (2–6%). In our continuing effort towards the synthesis of ¹⁸F-FMAU, we have investigated reactions of 1,3,5-tri-*O*-benzoyl- α -D-ribofuranose-2-sulphonate esters **2a–c** with several fluorinating agents, under a variety of experimental conditions (Fig. 2). Here, we report a procedure for stereospecific (*arabino*) incorporation of radiofluorine into the protected sugar, much superior to the currently available method in terms of simplicity and radiochemical yield.

2. Experimental

2.1. Reagents and instrumentation

All reagents and solvents were purchased from Aldrich Chemical Co. (Milwaukee, WI), and used without further purification. 2-*O*-(Fluorosulfonyl)-1,3,5-tri-*O*-benzoyl- α -Dribofuranose **2a**, 2-*O*-(methylsulfonyl)-1,3,5-tri-*O*-benzoyl- α -D-ribofuranose **2b** and 2-*O*-(trifluoromethylsulfonyl)-1,3,5-tri-*O*-benzoyl- α -D-ribofuranose **2c** were prepared following literature methods [11,13].

Proton and ¹⁹F NMR spectra were recorded on a Brucker 360 or 500 MHz spectrometer in chloroform-D₃ using tetramethylsilane as an internal reference, and trichlorofluoromethane as an external reference, respectively. Mass spectra were obtained on a Finnigan 400 mass spectrometer at the University of Minnesota using ammonia chemical ionization technique. High performance liquid chromatography (HPLC) was performed on a Waters Associates system using a 510 pump, UV detector (Isco) operated at 254 nm, and a radioactivity detector with single-channel analyzer (Technical Associates, Canoga Park, CA) using semi-preparative or analytical C₁₈ reverse phase columns (Alltech). A MeCN/ H₂O solvent system (70% MeCN) was used.

2.2. Fluorinating reagents

Tetraethylammonium fluoride (Et_4NF), tetrapropylammonium fluoride (*n*- Pr_4NF) and tetrabutylammonium

fluoride (*n*-Bu₄NF) were prepared from their hydroxycompounds by neutralization with aqueous hydrofluoric acid (48% HF) to pH 7.00. The aqueous solution was dried by azeotropic evaporation of water with acetonitrile before the fluorination reaction. Triethylamine tri-(hydrogen fluoride) (Et₃N·3HF) was prepared by adding cold, anhydrous HF into Et₃N at low temperature (dry ice acetone).

2.3. 2-Deoxy-2-fluoro-1,3,5-tri-O-benzoyl-α-Darabinofuranose 3

All fluorination reactions were performed in a similar manner; a representative procedure is described here. Each precursor (fluorosulfonate, methylsulfonate or trifluoromethylsulfonate, 10-12 mg) was dissolved in either EtOAc or MeCN (0.5 ml), and the solution was added to the dry fluorination reagent under argon in a V-vial. The vial was heated in a heating block at 71-72°C for 20 min, when TLC (12% acetone in hexane) showed no starting material remained. The reaction mixture was cooled to room temperature, solvent evaporated, and purified by HPLC (C18 semi-prep. column, 70% MeCN/H₂O, flow 4.5 ml/min). The appropriate fraction was collected and evaporated to dryness to provide the pure product. ¹H NMR: δ =8.05–8.15 (m, 6H, aromatic), 7.55-7.64 (m, 3H, aromatic), 7.39-7.48 (m, 6H, aromatic), 6.74 (d, 1H, C₁H, J=8.6 Hz), 5.61 (dd, 1H, C₃H, J = 19.4 and 2.9 Hz), 5.37 (d, 1H, C₂H, J = 48.6 Hz), 4.68– 4.79 (m, 3H, C₄H and C₅H). ¹⁹F NMR: $\delta = -190.8$. MS: 465 (M+1, 10), 391 (35), 343 (75), 242 (100).

2.4. 1,3,5-Tri-O-benzoyl-α-D-arabinofuranose 4

This compound was essentially the major product in most of the fluorination reactions. It was isolated during HPLC purification of the crude material in 44–60% yield, and characterized by ¹H NMR spectroscopy and mass spectrometry. ¹H NMR: δ =8.04–8.14 (m, 6H, aromatic), 7.60–7.66 (m, 3H, aromatic), 7.41–7.49 (m, 6H, aromatic), 6.75 (s, 1H, C₁H), 5.82 (s, 1H, C₂H), 5.68 (d, 1H, C₃H, *J*=3.5 Hz), 4.81– 4.84 (m, 1H, C₄H), 4.71–4.75 (m, 2H, C₅H). MS: 461 (M–1, 5), 445 (100), 242 (85), 201 (70).

2.5. 2-Acetyl-1,3,5-tri-O-benzoyl- α -D-arabinofuranose 5

The precursor (**2a**, 7.7 mg) was dissolved in EtOAc (0.5 ml) in a V-vial under argon. Acetic acid (1.2 eq) and *n*-Bu₄NF (1.2 eq) were added and the reaction mixture was heated in a heating block at 71–72°C for 95 min. The reaction mixture was cooled to room temperature, solvent evaporated, and purified by HPLC. The desired product was isolated in 75% yield. ¹H NMR: δ =8.07–8.12 (m, 6H, aromatic), 7.55–7.64 (m, 3H, aromatic), 7.41–7.47 (m, 6H, aromatic), 6.58 (s, 1H, C₁H), 5.58 (s, 1H, C₂H), 5.52 (d, 1H, C₃H, *J*=3.4 Hz), 4.71–4.77 (m, 3H, C₄H and C₅H), 2.09 (s, 3H, acetate). MS: 522, M+NH₄ (42), 383 (100), 343

(28), 104 (65). Exact mass calculated for $C_{28}H_{28}NO_9$ (M+NH₄) 522.1766, found 522.1766.

2.6. 2-Deoxy-2-[^{18}F]-fluoro-1,3,5-tri-O-benzoyl- α -D-arabinofuranose **3**

[¹⁸F]-Fluoride was reacted with *n*-Bu₄NHCO₃ and evaporated to dryness azeotropicaly with MeCN (1.2 ml). To the dry residue, *n*-Bu₄N¹⁸F, a solution of **2a** or **2c** (5–7 mg) in ethyl acetate (0.5 ml) was added, and the reaction mixture was heated at 71–72°C for 20 min. The crude reaction mixture was passed through a sep-pack cartridge (silica gel), and eluted with EtOAc. After evaporation of the solvent, the residue was dissolved in the HPLC solvent (70% MeCN/H₂O), and chromatographed using a C₁₈ semi-prep column. The appropriate fraction containing the radioactive species was isolated and evaporated to dryness. The radiochemical yields were 32–41% for **2c** and 3–6% for **2a**, respectively, both in >99% radiochemical purity.

3. Results and discussion

The precursors, fluorosulfonate 2a, mesylate 2b and triflate 2c were prepared following literature methods in high yields [11,13]. Like other investigators, we were not successful in preparing the tosylate 2d by the reaction of tosyl chloride on 1,3,5-tri-*O*-benzoyl ribofuranose [11]. We carried out extensive studies on the preparation of

Table 1

Fluorination reactions of substrates with various fluorinating agents and solvents at 71-72°C

2-deoxy-2-fluoro-1,3,5-tri-O-benzoyl- α -D-arabinofuranose **3** by fluorination of the fluorosulfonate **2a**. Most of these experiments were performed in EtOAc, which has been reported to be the best solvent for this reaction [13]. In addition, MeCN and DMF were also used in some experiments for comparison. Fig. 2 represents the general methodology of such fluorination reactions.

Table 1 represents the results of fluorination with various substrates, fluorinating agents, solvents and time at 71-72°C. We have investigated each of three substrates against various fluorinating agents. When reacted with n-Bu₄NF, the triflate 2c and fluorosulfonate 2a produced 3 in 30-40 and 12-15% yield, respectively. In contrast, the mesylate 2b did not produce any product. n-Bu₄NF in either EtOAc or MeCN led to successful fluorination of 2a and 2c, although MeCN was optimal for isolation of the pure product. Fluorinations with either tetra-alkyl ammonium fluorides resulted in relatively higher yield (12-16%) in EtOAc. In MeCN, the yields were slightly lower (10%) in the reactions of Et₄NF and *n*-Pr₄NF, but this difference was not significant in the case of *n*-Bu₄NF. The small difference of yields (Table 1) in the reactions of tetra-alkyl ammonium fluorides with these solvents, may be due to the relative solubility of the reagents, and thus the availability of the fluoride ion in solution. It has been suggested, that the solvent plays an important role in reactions of the fluoride ion [16]. Our results also suggest that choice of solvent is important, with EtOAc appearing to be a better solvent for organic ammonium fluorides in these fluorination reactions in terms of yields. EtOAc was also

Substrate	Fluorinating agent	Solvent	Time	Yield (%)
2a	(Et) ₄ NF (1.2 eq)	EtOAc	20 min	13
2a	$(Et)_4 NF (1.2 eq)$	EtOAc/THF	20 min	9.0
2a	$(Et)_4NF$ (1.2 eq)	MeCN	20 min	10.5
2a	$(nPr)_4NF$ (1.2 eq)	EtOAc	20 min	12.5
2a	$(n Pr)_4 NF (1.2 eq)$	MeCN	20 min	10.5
2a	$(nBu)_4NF$ (1.2 eq)	EtOAc	20 min	15.7
2a	$(nBu)_4NF$ (1.2 eq)	MeCN	20 min	14.5
2a	CsF (10.0 eq)	EtOAc	45 min	5.0
2a	CsF (10.0 eq)	EtOAc	96 min	10.5
2a	CsF (3.4 eq)	MeCN	45 min	12.3
2a	CsF (3.4 eq)	MeCN	75 min	17.5
2a	CsF (3.3 eq)	DMF	75 min	5.3
2a	CsF (8.7 eq)	DMF	45 min	14.0
2a	$(Et)_3N\cdot 3HF/(Et)_3N$ (6 eq)	EtOAc	2.5 h	56.0
2a	$(Et)_3N\cdot 3HF/(Et)_3N$ (6 eq)	EtOAc	4 h	60.5
2a	$(Et)_3N\cdot 3HF/(Et)_3N$ (6 eq)	MeCN	4 h	35.5
2a	(nBu) ₄ NF+Et ₃ N·3HF	EtOAc	30 min	30
2a	$(nBu)_4N(F-18)F+Et_3N\cdot 3HF$	EtOAc	30 min	6 ^a
2b	$(nBu)_4NF$ (1.2 eq)	EtOAc	20 min	no product
2c	$(nBu)_4NF$ (1.2 eq)	EtOAc	20 min	31
2c	$(nBu)_4NF$ (1.2 eq)	EtOAc	20 min	34
2c	$(nBu)_4NF$ (1.2 eq)	EtOAc	20 min	30
2c	$(nBu)_4 N^{18} F$ (trace)	EtOAc	20 min	32 ^a
2c	$(nBu)_4 N^{18} F$ (trace)	EtOAc	20 min	41 ^a
2c	$(nBu)_4 N^{18}F$ (trace)	EtOAc	20 min	35 ^a

^a Radiochemical yield.

found to be much better solvent in the reactions with the other organic fluoride $Et_3N.3HF/Et_3N$, but the reaction time was much longer. Cesium fluoride (CsF), in the presence of a crown ether, produced higher yields in MeCN than in EtOAc. A highly polar aprotic solvent, DMF, however, did not produce a high yield of product with CsF unless the amount of CsF was increased to more than two-fold compared to that used in MeCN.

All reactions using the fluorosulfonate ester 2a produced low yield of the desired product 3 within 20-30 min except in one instance when two reagents were used. Combination of n-Bu₄NF and Et₃N·3HF (3:2) appeared to produce the higher yield within shorter reaction time (30 min). The chemical yield could be improved from 15 to 30% by manipulation of the reagent composition, with the radiochemical yield improved proportionally from 3 to 6%. This persistent low radiochemical yield was due to an undesirable competitive exchange, between radioactive fluoride and the covalently bound fluorine on the leaving group. The nucleophilic radiofluorine (fluoride) is consumed by displacing the isotopic fluorine on the sulfonate ester leaving group (Fig. 3) and thus becomes unavailable for the desired substitution reaction. The net result of this facile exchange is a significant reduction in radiochemical yield. Although, 2a is known to be stable [13], the sulfur fluorine bond have been reported to be very labile [17]. Therefore, it is likely that the radioactive fluoride in solution is equilibrated fast with the non-radioactive fluoride before the apparently slower displacement of the sulfonate ester is completed.



Fig. 3. Exchange reaction between radiofluorine and isotopic stable fluorine.

All fluorinating agents, except $Et_3N \cdot 3HF/Et_3N$, produced a common by-product (45–60%), which was characterized as 1,3,5-tri-*O*-benzoyl-1- α -D-arabinofuranose **4**. This product was produced by the competitive nucleophilic substitution of the sulfonate esters with a hydroxide ion from either an excess of tetra-alkyl ammonium hydroxide or traces of water present in the reaction mixture. In the reactions with $Et_3N \cdot 3HF/Et_3N$, this by-product was not formed, since the overall reaction was under acidic conditions.

A second by-product was formed in small amounts when the reactions were performed in EtOAc. It was identified as 2-acetyl-1,3,5,-tri-O-benzoyl-1- α -D-arabinofuranose **5**. The formation of **5** may be explained by the nucleophilic substitution of the sulfonate ester by trace of acetate ion present in EtOAc. Fluoride ion is known to be a strong base, a potent hydrogen bond electron donor, and a weak nucleophile [18]. As an electron donor it forms strong hydrogen bonded complex with acetic acid producing acetate nucleophile, which competes with fluoride as a nucleophile and produces the acetoxy compound. In order to verify the formation of the compound **5**, separate experiments were performed with *n*-Bu₄NF (1.2 eq) in the presence of same equivalent acetic acid. The major product in these experiments was **5** in 70–75% yields.

Of the three substrates studied 2a, 2b, and 2c, only 2a and 2c produced the desired product. The failure of the mesylate 2b to yield 3 may be due to an electronic effect; i.e. the mesylate may not be a good leaving group in this system. When fluorination of the triflate 2c was performed using radioactive fluoride the radiochemical yield was 32-41%. In this reaction, the radioactive fluoride was consumed in the substitution reaction producing high yield of the desired compound. An HPLC chromatogram of radiolabeled fluorosugar 3 is represented in Fig. 4.

Earlier studies demonstrated that the reaction of sulfonate esters 2b and 2c with n-Bu₄NF resulted, either in an elimination product or in the recovery of the starting material [11]. These authors explained their results on the basis of an earlier suggestion indicating that nucleophilic displacement at C₂ creates an unfavorable alignment of dipoles in the transition state, particularly when the substituent at C1 is the α-D-anomer [19]. Our results, however, showed the absence of any elimination product or starting material, and instead two products, 2-fluoro-and 2-hydroxy-arabinose derivatives produced by nucleophilic substitution (Fig. 2). These results, therefore, suggest that the fluorosulfonate and trifluormethylsulfonate esters of 1,3,5-tri-O-benzoyl-2-ribose do undergo nucleophilic substitution with fluoride, acetate and hydroxide ions, while mesylate is inactive in this system. Furthermore, nucleophilic substitution of trifluoromethanesulfonate ester at the C₂ position of a xylo sugar has also been reported [20]. Since chemical yield in fluorination reaction of sulfonate esters with *n*-But₄NF in EtOAc and in MeCN are comparable, use of MeCN might be advantageous to avoid the formation of the acetate 5 which



Fig. 4. HPLC chromatogram of [F-18]-labeled-2-fluoro-1,3,5-tri-O-ben-zoyl-α-D-arabinofuranose.

is, rather, difficult to separate from the desired product during HPLC purification.

4. Conclusion

We have investigated, fluorination of 1,3,5-tri-*O*-benzoyl- α -D-ribofuranose-2-sulfonyl esters with several fluorides in various solvents, and developed a suitable method for incorporation of radiofluorine into the protected sugar. The 2-*O*-trifluoromethyl-sulfonyl-1,3,5-tri-*O*-benzoyl- α -D-ribofuranose was found to be the best substrate. Tetrabuty-lammonium fluoride in either EtOAc or MeCN led to successful fluorination, although MeCN is optimal. For radiolabeling with ¹⁸F, the triflate is the optimal substrate, which consistently produces the higher radiochemical yield in the series studied. The 2-fluoro-1,3,5-tri-*O*-benzoyl- α -D-arabinofuranose **3**, successfully labeled with ¹⁸F, will be a potential intermediate for synthesis of ¹⁸F labeled nucleoside analogues for PET imaging.

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References

[1] U. Reichman, K.A. Watanabe, J.J. Fox, Carbohydr. Res. 42 (1975) 233.

- [2] J.J. Fox, K.A. Watanabe, T.C. Chou, R.F. Schinazi, K.E. Soike, I. Fourel, O. Hantz, C. Trepo. in: N. F. Taylor (Ed.), Fluorinated Carbodydrates, Am. Chem. Soc., 1988, p. 176.
- [3] P.S. Conti, M.M. Alauddin, J.D. Fissekis, B. Schmall, K.A. Watanabe, Nucl. Med. Biol. 22 (1995) 783.
- [4] P.S. Conti, J.H. Anderson, D.F. Wong, J.R. Bading, M.M. Alauddin, H.T. Ravert, W.B. Mathews, J.L. Musachio, R.F. Dannals, U. Scheffel, J. Nucl. Med. 37 (1996) 107P.
- [5] J.G. Tjuvajev, G. Stockhammar, R. Desai, H. Uehara, K.A. Watanabe, B. Gansbacher, R.G. Blasberg, Cancer Res. 55 (1995) 6126.
- [6] K.W. Pankiewicz, B. Nawrot, H. Gadler, R.W. Price, K.A. Watanabe, J. Med. Chem. 30 (1987) 2314.
- [7] K.W. Pankiewicz, K.A. Watanabe, Chem. Pharm. Bull. 35 (1987) 4494.
- [8] L.C. Cheng, T.L. Su, K.W. Pankiewicz, K.A. Watanabe, Nucleosides Nucleotides 8 (1989) 1179.
- [9] K.A. Watanabe, T.L. Su, R.S. Klein, K.C. Chu, A. Mastuda, M.W. Chun, C. Lopez, J.J. Fox, J. Med. Chem. 26 (1983) 152.
- [10] K.A. Watanabe, T.L. Su, R.S. Klein, U. Riechman, N. Greenberg, C. Lopez, J.J. Fox, J. Med. Chem. 27 (1984) 91.
- [11] C.H. Tann, P.R. Brodfuehrer, S.P. Brundidge, C. Sapino, H.G. Howell, J. Org. Chem. 50 (1985) 3644.
- [12] H.G. Howell, P.R. Brodfuehrer, S.P. Brundidge, D.A. Beningni, C. Sapino Jr., J. Org. Chem. 53 (1988) 85.
- [13] T.S. Chou, L.M. Becke, J.C. O'Toole, M.A. Carr, B.E. Parker, Tetrahedron Lett. 37 (1996) 17.
- [14] M.M. Alauddin, P.S. Conti, T. Mathew, H. Abrahamian, J.D. Fissekis, G.K.S. Prakash, K.A. Watanabe, J. Nucl. Med. 40 (1999) 311P.
- [15] M.M. Alauddin, P.S. Conti, T. Mathew, H. Abrahamian, G.K.S. Prakash, K.A. Watanabe, Anticancer Res. 18 (1998) 4991.
- [16] M.M. Alauddin, J.M. Miller, J.H. Clark, Can. J. Chem. 62 (1984) 263.
- [17] P.J. Durrant, B. Durrent, Introduction to Advanced Inorganic Chemistry, 2nd Edition, Wiley, New York, 1970, p. 857.
- [18] J.H. Clark, Chem. Rev. 80 (1980) 429.
- [19] A.C. Richardson, Carbohydr. Res. 10 (1969) 395.
- [20] T. Haradahira, A. Kato, M. Maeda, Y. Kanazawa, M. Yamada, Y. Tori, Y. Ichiya, K. Masuda, J. Label. Compd. Radiopharm. (1995) 552.