3-BROMO-2-BROMOMETHYLPROPYL GLYCOSIDES IN THE PREPARA-TION OF DOUBLE-CHAIN BIS-SULFIDE NEO-GLYCOLIPIDS*

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ABSTRACT

Boron trifluoride etherate-induced glycosidation of 3-bromo-2-bromomethylpropan-1-ol with sugar acetates gave the title glycosides of the following sugars of the D series: Glcp, Galp, GlcpA, GlcNPhthp, Xylp, β -Galp-(1->4)-Glcp, and α -Galp-(1->4)-Galp. Treatment of the fully acetylated glycosides with alkanethiols and cesium carbonate in N,N-dimethylformamide followed by deacetylation gave the corresponding bis-sulfide glycolipids.

INTRODUCTION

Glycolipids are localised in the outer half of cell-surface membrane bilayers where, *inter alia*, they exert different biological receptor functions¹⁻³. Although unnatural synthetic glycolipids (neo-glycolipids) have been prepared⁴, apparently there has been no report on the synthesis of neo-glycolipids that mimic the structure and amphiphilic properties of the naturally occurring compounds. We now describe the synthesis of some such neo-glycolipids related to 1-3.

Neo-glycolipids are useful in biological receptor studies for coating of thinlayer plates, microtiter wells and cells, and for forming such aggregates as micelles and liposomes for agglutination studies². The aglycon portion of glycolipids greatly influences the type of aggregates that are formed in aqueous solution; single-chain glycolipids give rise to spherical micelles, whereas double-chain glycolipids form double-layer liposomes⁵. In addition, neo-glycolipids would generally show increased stability against enzymic breakdown. Finally, it is desirable to be able to transform (or synthesise) oligosaccharides that are present uniquely on glycoproteins into the corresponding neo-glycolipids for use in receptor studies and coating experiments.

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1 neo-glycolipid



2 glycosphingolipid



The stepwise synthesis^{6a} of neo-glycolipids allows variation of, for example, the unsaturation and length of the hydrocarbon chains, and glycoside synthesis steps need to be performed only once. Also, 2-bromoethyl glycosides can be used to alkylate alkanethiols, thereby, *inter alia*, giving access to single-chain glycolipids with various chain-lengths⁶.

RESULTS AND DISCUSSION

We have synthesised glycosphingolipid- and glycoglycerolipid-like doublechain neo-glycolipids by alkylation of thiols with 3-bromo-2-bromomethylpropyl glycosides [or dibromoisobutyl (DIB) glycosides]. Boron trifluoride etheratemediated glycosidation⁷ of 3-bromo-2-bromomethylpropan-1-ol (dibromoisobutyl alcohol, DIBol; **26**) with sugar acetates gave the DIB glycosides **4–10**. The method works best with primary alcohols and sugar 1,2-*trans* acetates⁸ which can be prepared in near-quantitative yields and with a *trans/cis* ratio of >20:1 by treatment of acetylated 2-trimethylsilylethyl glycosides with boron trifluoride etherate in the presence of 1 equiv. of acetic anhydride⁹.

Treatment of the DIB glycosides 4-10 with alkanethiols of various chainlengths in N,N-dimethylformamide-cesium carbonate⁶ gave the acetylated compounds 11-19. Deacetylation in methanolic sodium methoxide-dichloromethane gave the neo-glycolipids 20-24. The utilisation of these glycolipids will be reported elsewhere.

Surprisingly, DIBol (26) appears to be a novel compound, and was prepared

R'O CH2OR

r'o



HO





òΡ

25

27

Br

Br

Br

Br

8,15,22

ноос

H₂C

R'O'

R'O



10, 17, 24

Br

26

Br

28

Br



6,13





CH2OR



CH2OR

by borane reduction of the known¹⁰ acid **25**. The hydroxyl group of DIBol could be protected by acetylation (\rightarrow **27**) or tetrahydropyranylation (\rightarrow **28**) to give useful synthons.

EXPERIMENTAL

General. — Optical rotations were measured on solutions in $CDCl_3$ with a Perkin–Elmer 141 polarimeter. N.m.r. spectra were recorded for solutions in $CDCl_3$ (internal Me₄Si) or $CDCl_3$ –CD₃OD–D₂O (CMD, 75:45:10) with Varian XL-300 and Nicolet WB 360 spectrometers. Solvents were removed at <0.1 Torr.

3-Bromo-2-bromomethylpropan-1-ol (DIBol; **26**). — To a stirred solution of 3-bromo-2-bromomethylpropanoic acid¹⁰ (**25**; 15.3 g, 62.3 mmol) in dichloromethane (400 mL) at 0° under nitrogen was added diborane (M BH₃ in tetrahydrofuran; 187 mL) dropwise during ~10 min. After 1 h, the mixture was stirred overnight at room temperature, M hydrochloric acid (200 mL) was then added dropwise. the mixture was stirred for ~30 min, the aqueous phase was extracted with dichloromethane (3 × 50 mL), and the combined extracts were dried (Na₂SO₄) and concentrated. The resulting oil that was eluted from a column (200 × 45 mm) of silica gel with CH₂Cl₂ gave DIBol (**26**; 13.8 g, 96%), b.p. ~45°/0.1 Torr), $n_D^{2,3}$ 1.5439; ν_{max} 3340 cm⁻¹. N.m.r. data (CDCl₃) ¹H, δ 3.79 (d, 2 H, J 5.7 Hz, CH₂O), 3.58 and 3.57 (2 ABq, each 2 H, J_{AB} 10.0 and J 5.5 Hz, CH₂Br), 2.27 [septet, 1 H, CH(CH₂)₃]; ¹³C, δ 62.4 (CH₂OH), 44.4 (CH), 32.8 (CH₂Br).

Anal. Calc. for C₄H₈Br₂O: C, 20.7; H, 3.5. Found: C, 21.0; H, 3.7.

3-Bromo-2-bromomethylpropyl glycosides (4–10). — To a solution of the acetylated sugar (1.5 mmol) and DIBol (26, 2 mmol) in dry dichloromethane (10 mL) at room temperature was added boron trifluoride etherate (10 mmol). The reaction was monitored by t.l.c. (SiO₂; ethyl acetate-hexane). The sugar acetate was normally consumed within 1–4 h. The mixture was washed with water and saturated aqueous sodium hydrogencarbonate, dried (Na₂SO₄), and concentrated. Column chromatography (SiO₂; ethyl acetate-heptane) gave the following glycosides.

3-Bromo-2-bromomethylpropyl 2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranoside (4, 54% from 1,2,3,4,6-penta-*O*-acetyl-β-D-glucopyranose), $[\alpha]_{D}^{23} - 5^{\circ}$ (c 0.6). ¹H-N.m.r. data (CDCl₃): δ 5.22 (t, 1 H, $J_{2,3} = J_{3,4} = 9.7$ Hz, H-3), 5.1 (t, 1 H, $J_{4,5}$ 9.4 Hz, H-4), 4.99 (t, 1 H, H-2), 4.51 (d, 1 H, $J_{1,2}$ 7.9 Hz, H-1), 4.27 and 4.15 (ABq with further coupling, each 1 H, J_{AB} 12.6, $J_{5,6}$ 4.0 Hz, H-6,6'), 3.71 (m, 1 H, H-5), 2.34 [m, 1 H, CH(CH₂)₃].

Anal. Calc. for C₁₈H₂₆Br₂O₁₀: C, 38.5; H, 4.7. Found: C, 38.4; H, 4.7.

3-Bromo-2-bromomethylpropyl 2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside (5, 50% from 1,2,3,4,6-penta-*O*-acetyl-β-D-galactopyranose), $[\alpha]_{15}^{23}$ +1° (*c* 0.7). ¹H-N.m.r. data (CDCl₃): δ 5.40 (d, 1 H, $J_{3,4}$ 3.2 Hz, H-4), 5.19 (dd, 1 H, $J_{2,3}$ 10.4 Hz, H-2), 5.03 (dd, 1 H, H-3), 4.47 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 4.19 and 4.13 (ABq with further coupling, each 1 H, J_{AB} 11.2, $J_{5,6} = J_{5,6'} = 6.5$ Hz, H-6,6'), 3.92 (t, 1 H, $J_{4,5}$ 0.4 Hz, H-5), 2.35 [septet, 1 H, J 5.8 Hz, $CH(CH_2)_3$].

Anal. Calc. for C₁₈H₂₆Br₂O₁₀: C, 38.5; H, 4.7. Found: C, 39.3; H, 4.4.

Methyl (3-bromo-2-bromomethylpropyl 2,3,4-tri-O-acetyl- β -D-glucopyranosid)uronate (6, 26% from methyl 1,2,3,4-tetra-O-acetyl- β -D-glucopyranuronate), $[\alpha]_{D}^{23}$ +3° (c 1.1). ¹H-N.m.r. data (CDCl₃): δ 5.33–5.16 (m, 2 H, H-3,4), 5.01 (m, 1 H, H-2), 4.55 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 4.04 (d, 1 H, $J_{4,5}$ 9.4 Hz, H-5), 3.77 (s, 3 H, OMe), 2.34 [septet, 1 H, J 6.1 Hz, CH(CH₂)₃].

Anal. Calc. for C₁₇H₂₄Br₂O₁₀: C, 37.2; H, 4.6. Found: C, 37.5; H, 4.6.

3-Bromo-2-bromomethylpropyl 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (7, 52% from 1,3,4,6-tetra-O-acetyl-2-deoxy-2-phthalimidoα,β-D-glucopyranose^{6d,11}, αβ-ratio 1:1), $[\alpha]_D^{23} + 20^\circ$ (c 1). ¹H-N.m.r. data (CDCl₃): δ 5.82 (t, 1 H, $J_{3,4}$ 10.1 Hz, H-3), 5.36 (d, 1 H, $J_{1,2}$ 8.3 Hz, H-1), 5.17 (t, 1 H, $J_{4,5}$ 10.1 Hz, H-4), 4.32 (dd, 1 H, $J_{2,3}$ 10.4 Hz, H-2), 4.33 and 4.19 (ABq with further coupling, each 1 H, J_{AB} 12.2, $J_{5,6}$ 5.0, $J_{5,6'}$ 2.2 Hz, H-6,6'), 3.89 (m, 1 H, H-5), 2.24 [m, 1 H, J 5.8 Hz, CH(CH₂)₃].

Anal. Calc. for C₂₄H₂₇Br₂NO₁₀: C, 44.4; H, 4.2. Found: C, 44.9; H, 4.3.

3-Bromo-2-bromomethylpropyl 2,3,4-tri-*O*-acetyl-β-D-xylopyranoside (**8**, 50% from 1,2,3,4-tetra-*O*-acetyl-α,β-D-xylopyranose; αβ-ratio, 1:1), $[\alpha]_D^{23} - 25^\circ$ (c 0.9). ¹H-N.m.r. data (CDCl₃): δ 5.18 (t, 1 H, $J_{2,3} = J_{3,4} = 8.3$ Hz, H-3), 4.98–4.89 (m, 2 H, H-2,4), 4.49 (d, 1 H, $J_{1,2}$ 6.7 Hz, H-1), 4.14 and 3.39 (ABq with further coupling, J_{AB} 11.5, $J_{4,5}$ 5.0, $J_{4,5'}$ 9.0 Hz, H-5,5'), 2.34 [septet, J 5.6 Hz, CH(CH₂)₃].

Anal. Calc. for C₁₅H₂₂Br₂O₈: C, 36.8; H, 4.5. Found: C, 37.3; H, 4.7.

3-Bromo-2-bromomethylpropyl 2,3,6-tri-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl)-β-D-glucopyranose], $[\alpha]_{D}^{23}$ -6° (*c* 0.7). ¹H-N.m.r. data (CDCl₃): δ 5.35 (d, 1 H, $J_{3',4'}$ 2.9 Hz, H-4'), 5.20 (t, 1 H, $J_{2,3}$ 9.0 Hz, H-2), 5.11 (dd, 1 H, $J_{1',2'}$ 7.9, $J_{2',3'}$ 10.1 Hz, H-2'), 4.95 (dd, 1 H, H-3'), 4.89 (t, 1 H, $J_{3,4}$ 9.0 Hz, H-3), 4.50, 4.47 (2 d, each 1 H, J 7.9 Hz, H-1,1'), 2.32 [septet, 1 H, J 5.8 Hz, CH(CH₂)₃].

Anal. Calc. for C₃₀H₄₂Br₂O₁₈: C, 42.4; H, 5.0. Found: C, 42.4; H, 4.9.

3-Bromo-2-bromomethylpropyl 2,3,6-tri-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl-α-D-galactopyranosyl)-β-D-galactopyranoside [**10**, 43% from 1,2,3,6-tetra-*O*-acetyl-α-D-galactopyranosyl)-α-D-galactopyranose¹²], $[\alpha]_D^{2^3}$ +69° (*c* 1.5). ¹H-N.m.r. data (CDCl₃): δ 5.58 (dd, 1 H, $J_{4',5'}$ 1.0 Hz, H-4'), 5.39 (dd, 1 H, $J_{2',3'}$ 10.8 Hz, H-2'), 5.20 (dd, 1 H, $J_{3',4'}$ 3.6 Hz, H-3'), 5.17 (dd, 1 H, $J_{2,3}$ 10.8 Hz, H-2), 5.01 (d, 1 H, $J_{1',2'}$ 3.2 Hz, H-1'), 4.82 (dd, 1 H, $J_{3,4}$ 2.9 Hz, H-3), 4.47 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 2.37 [septet, 1 H, J 5.8 Hz, CH(CH₂)₃].

Anal. Calc. for C₃₀H₄₂Br₂O₁₈: C, 42.4; H, 5.0. Found: C, 43.1; H, 5.0.

Acetylated bis-sulfide glycolipids (11-19). — A mixture of the 3-bromo-2bromomethylpropyl glycoside (4-10, 0.37 mmol), an alkanethiol (1 mmol), cesium carbonate (0.6 mmol), and N,N-dimethylformamide (2 mL) was stirred at room temperature for 24-48 h. The reaction was monitored by t.l.c. $(SiO_2; \text{ ethyl acetate})$ heptane). When the starting glycoside had been consumed, dichloromethane (30 mL) was added and the solution was washed with water (15 mL). The rate of phase separation could be increased by the addition of a small amount of aqueous sodium chloride. The organic phase was dried (Na₂SO₄) and concentrated, and the residue was subjected to chromatography on a column (300 \times 15 mm) of silica gel with ethyl acetate-heptane, to give the following bis-sulfide glycolipids.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranoside (**11**, 70% from **4** and hexadecanethiol), $[\alpha]_D^{23} - 2^\circ$ (*c* 1.1). ¹H-N.m.r. data (CDCl₃): δ 5.20 (t, 1 H, $J_{2,3}$ 9.3 Hz, H-3), 5.06 (t, 1 H, $J_{3,4} = J_{4,5} = 9.5$ Hz, H-4), 4.98 (dd, 1 H, H-2), 4.48 (d, 1 H, $J_{1,2}$ 7.9 Hz, H-1), 2.26 and 2.11 (ABq with further coupling, each 1 H, J_{AB} 12.4, $J_{5,6}$ 4.8, $J_{5,6'}$ 2.5 Hz, H-6,6'), 2.6-2.4 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₅₀H₉₂O₁₀S₂: C, 65.5; H, 10.1. Found: C, 65.7; H, 10.2.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside (**12**, 79% from **5** and hexadecanethiol), $[\alpha]_D^{2^3} + 1^\circ$ (*c* 1.6). ¹H-N.m.r. data (CDCl₃): δ 5.37 (dd, 1 H, $J_{4,5}$ 0.8 Hz, H-4), 5.17 (dd, 1 H, $J_{2,3}$ 10.3 Hz, H-2), 4.99 (dd, 1 H, $J_{3,4}$ 3.4 Hz, H-3), 4.44 (d, 1 H, $J_{1,2}$ 7.8 Hz, H-1), 2.7–2.4 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₅₀H₉₂O₁₀S₂: C, 65.5; H, 10.1. Found: C, 65.3; H, 10.2.

Methyl (3-hexadecylthio-2-hexadecylthiomethylpropyl 2,3,4-tri-O-acetyl-β-D-glucopyranosid)uronate (13, 68% from 6 and hexadecanethiol), $[\alpha]_D^{23} - 2^\circ$ (c 0.9). ¹H-N.m.r. data (CDCl₃): δ 5.25 (t, 1 H, $J_{3,4}$ 9.0 Hz, H-3), 5.20 (t, 1 H, $J_{4,5}$ 9.4 Hz, H-4), 5.01 (dd, 1 H, $J_{2,3}$ 9.0 Hz, H-2), 4.54 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 4.03 (d, 1 H, H-5), 3.76 (s, 3 H, OMe), 2.60–2.45 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₄₉H₉₀O₁₀S₂: C, 64.7; H, 10.2. Found: C, 64.9; H, 10.2.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 3,4,6-tri-*O*-acetyl-2-deoxy-2phthalimido-β-D-glucopyranoside (14, 81% from 7 and hexadecanethiol), $[\alpha]_D^{23}$ +12° (c 1.1). ¹H-N.m.r. data (CDCl₃): δ 5.80 (dd, 1 H, $J_{2,3}$ 10.7 Hz. H-3), 5.32 (d, 1 H, $J_{1,2}$ 8.5 Hz, H-1), 5.16 (t, 1 H, $J_{3,4} = J_{4,5} = 9.2$ Hz, H-4), 2.5–2.2 (m, 8 H, 4 CH₂S).

Anal. Calc. for $C_{55}H_{93}NO_{10}S_2$: C, 67.0; H, 9.3; N, 1.4. Found: C, 67.0; H, 9.5; N, 1.4.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 2,3,4-tri-*O*-acetyl-β-D-xylopyranoside (**15**, 61% from **8** and hexadecanethiol), $[\alpha]_D^{23} - 18^\circ$ (c 1.1). ¹H-N.m.r. data (CDCl₃): δ 5.15 (t, 1 H, $J_{2,3} = J_{3,4} = 8.5$ Hz, H-3), 4.98-4.85 (m, 2 H, H-2,4), 4.45 (d, 1 H, $J_{1,2}$ 6.7 Hz, H-1), 4.10 and 3.34 (ABq with further coupling, each 1 H, J_{AB} 12.0, $J_{4,5}$ 5.0, $J_{4,5'}$ 8.8 Hz, H-5,5'), 2.7-2.4 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₄₇H₈₈O₈S₂: C, 66.8; H, 10.5. Found: C, 66.7; H, 10.6.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 2,3,4-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- β -D-glucopyranoside (16, 88% from 9 and hexadecanethiol), $[\alpha]_D^{23} - 4^\circ$ (c 0.8). ¹H-N.m.r. data (CDCl₃): δ 5.34 (d, 1 H, $J_{4',5'}$ 2.5 Hz, H-4'), 5.19 (t, 1 H, $J_{2,3}$ 9.0 Hz, H-2), 5.10 (dd, 1 H, $J_{2',3'}$ 10.4 Hz, H-2'), 4.95 (dd, 1 H, $J_{3',4'}$ 3.6 Hz, H-3'), 4.89 (dd, 1 H, $J_{3,4}$ 7.9 Hz, H-3), 4.47 and 4.45 (2 d, each 1 H, J 7.6 and 7.9 Hz, H-1,1'), 2.6–2.45 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₆₂H₁₀₈O₁₈S₂: C, 61.8; H, 9.0. Found: C, 62.0; H, 9.3.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 2,3,6-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl-α-D-galactopyranosyl)-β-D-galactopyranoside (17, 51% from 10 and hexadecanethiol), $[\alpha]_D^{23}$ +52° (c 0.6). ¹H-N.m.r. data (CDCl₃): δ 5.57 (dd, 1 H, $J_{4',5'}$ 0.8 Hz, H-4'), 5.38 (dd, 1 H, $J_{2',3'}$ 11.0 Hz, H-2'), 5.18 (dd, 1 H, $J_{3',4'}$ 3.7 Hz, H-3'), 5.16 (dd, 1 H, $J_{2,3}$ 11.0 Hz, H-2), 4.99 (d, 1 H, $J_{1',2'}$ 3.3 Hz, H-1'), 4.79 (dd, 1 H, $J_{3,4}$ 2.8 Hz, H-3), 4.44 (d, 1 H, $J_{1,2}$ 7.7 Hz, H-1), 2.7–2.45 (m, 8 H, 4 CH₂S).

Anal. Calc. for C₆₂H₁₀₈O₁₈S₂: C, 61.8; H, 9.0. Found: C, 60.7; H, 9.0.

3-Octadecylthio-2-octadecylthiomethylpropyl2,3,6-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- β -D-glucopyranoside (18, 67% from 9 and octadecanethiol), $[\alpha]_D^{23} - 3^\circ$ (c 0.8). ¹H-N.m.r. data (CDCl₃): the signals for the sugar moiety were practically identical with those of the spectra of 16 and 19.

Anal. Calc. for C₆₆H₁₁₆O₁₈S₂: C, 62.8; H, 9.3. Found: C, 62.1; H, 9.1.

3-Octylthio-2-octylthiomethylpropyl 2,3,6-tri-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- β -D-glucopyranoside (**19**, 73% from **9** and octane-thiol), $[\alpha]_D^{23} - 5^\circ$ (c 0.8). ¹H-N.m.r. data (CDCl₃): the signals for the sugar moiety were practically identical with those of the spectra of **16** and **18**.

Anal. Calc. for C₄₆H₇₆O₁₈S₂: C, 56.3; H, 7.8. Found: C, 57.1; H, 7.5.

Bis-sulfide glycolipids (20-24). — To a solution of each acetylated glycolipid (11, 12, 15-17; 0.2 mmol) in dichloromethane (15 mL) was added methanolic sodium methoxide (10 mL; from \sim 1 mg of sodium). The reaction was monitored by t.l.c. (chloroform-methanol-water, 65:35:10). One drop of acetic acid was added, the mixture was concentrated, filtered through silica gel (solvent as in t.l.c. above), and concentrated, and the residue was suspended in water (10 mL) and freeze-dried to give a quantitative yield of the glycolipid. Carbon analyses were outside normally accepted limits, probably due to remaining traces of water that could not be removed by freeze-drying. The following compounds were prepared in this way.

3-Hexadecylthio-2-hexadecylthiomethylpropyl β -D-glucopyranoside (20, from 11), $[\alpha]_D^{23} - 7^\circ$ (c 0.9, CMD). ¹H-N.m.r. data (CMD, 50°): δ 4.29 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 2.70 [d, 4 H, J 6.4 Hz, CH(CH₂S)₂], 2.53 (t, 4 H, J 7.3 Hz, SCH₂CH₂).

Anal. Calc. for C₄₂H₈₄O₆S₂: C, 67.3; H, 11.3. Found: C, 65.9; H, 11.3.

3-Hexadecylthio-2-hexadecylthiomethylpropyl β -D-galactopyranoside (21, from 12), $[\alpha]_D^{23} -3^\circ$ (c 0.5, CMD). ¹H-N.m.r. data (CMD, 20°): δ 4.24 (virtual coupling¹³, $J_{1,2}$ 7.6 Hz, H-1), 2.71 [d, 4 H, J 6.7 Hz, CH(CH₂S)₂], 2.53 (t, 4 H, J 7.2 Hz, SCH₂CH₂).

Anal. Calc. for C₄₂H₈₄O₆S₂: C, 67.3; H, 11.3. Found: C, 65.6; H, 11.1.

3-Hexadecylthio-2-hexadecylthiomethylpropyl β -D-xylopyranoside (22, from

15), $[\alpha]_{D}^{23} - 6^{\circ} (c \ 0.5, \text{ CMD})$. ¹H-N.m.r. data (CMD, 50°): $\delta 4.25 (d, 1 \text{ H}, J 7.1 \text{ Hz}, H-1)$, 2.69 [d, 4 H, J 6.4 Hz, CH(CH₂S)₂], 2.53 (t, 4 H, J 7.5 Hz, SCH₂CH₂).

Anal. Calc. for C₄₁H₈₂O₅S₂: C, 68.5; H, 11.5. Found: C, 66.3; H, 10.4.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 4-O- β -D-galactopyranosyl- β -D-glucopyranoside (23, from 16), $[\alpha]_{D}^{23}$ -3.5° (c 1.6, CMD). ¹H-N.m.r. data (CMD, 40°): δ 4.31 (d, 2 H, J 7.8 Hz, H-1,1'), 2.71 [d, 4 H, J 6.6 Hz, CH(CH₂S)₂], 2.53 (t, 4 H, J 7.3 Hz, SCH₂CH₂).

Anal. Calc. for C₄₈H₉₄O₁₁S₂: C, 63.3; H, 10.4. Found: C, 62.6; H, 10.6.

3-Hexadecylthio-2-hexadecylthiomethylpropyl 4-*O*-α-D-galactopyranosyl-β-D-galactopyranoside (**24**, from **17**), $[\alpha]_D^{23}$ +28° (*c* 0.6, CMD). ¹H-N.m.r. data (CMD, 50°): δ 5.01 (m, 1 H, H-4'), 4.27 (d, 1 H, J_{1,2} 7.2 Hz, H-1), 2.72 [d, 4 H, J 6.3 Hz, CH(CH₂S)₂], 2.53 (t, 4 H, J 7.4 Hz, SCH₂CH₂).

Anal. Calc. for C₄₈H₉₄O₁₁S₂: C, 63.3; H, 10.4. Found: C, 61.6; H, 10.6.

3-Bromo-2-bromomethylpropyl acetate (27). — A mixture of 3-bromo-2bromomethylpropan-1-ol (26; 512 mg, 2.21 mmol), pyridine (10 mL), and acetic anhydride (10 mL) was stirred at room temperature for 17 h and then co-concentrated with toluene, ethyl acetate (20 mL) was added, and the solution was washed with water (2 × 10 mL). The aqueous phase was extracted with ethyl acetate (10 mL), and the combined organic phases were dried (Na₂SO₄) and concentrated. The residue was eluted from silica gel with heptane–ethyl acetate (4:1) to give 27 (483 mg, 81%); ν_{max} 1752, 1230, and 1050 cm⁻¹. ¹H-N.m.r. data (CDCl₃): δ 4.18 (d, 2 H, J 6.4 Hz, AcOCH₂), 3.58 and 3.53 (dABq, 4 H, J_{AB} 10.6, J 5.3, J 6.2 Hz, CH₂Br), 2.41 (septet, 1 H, CH), 2.09 (s, 3 H, Me).

Anal. Calc. for C₆H₁₀Br₂O₂: C, 26.3; H, 3.7. Found: C, 26.8; H, 4.0.

3-Bromo-2-bromomethyl-1-(tetrahydropyran-2-yloxy)propane (28). - To a solution of 3-bromo-2-bromomethylpropan-1-ol (26; 1.0 g, 4.3 mmol) and dihydropyran (1.81 g, 21.6 mmol) in dry dichloromethane (20 mL) at 0° was added a solution of toluene-p-sulfonic acid (10 mg) in dichloromethane (2 mL). After 4 h, the mixture was left at room temperature for 5.5 h and then cooled to 0°, and more (10 mg, 2 mL) of the toluene-p-sulfonic acid solution was added. After 7 h, toluene (30 mL) and ether (20 mL) were added, and the mixture was washed with saturated aqueous sodium hydrogencarbonate (50 mL) and saturated aqueous sodium chloride (50 mL). The aqueous phases were extracted with toluene (50 mL), and the combined organic phases were dried (Na₂SO₄) and concentrated. The residue was distilled to give 28 (1.07 g, 79%), b.p. 85-105% 0.08 mmHg. Chromatography on a column of silica gel, using ethyl acetate-heptane (1:10), gave purc 28 (0.88 g, 65%), n_D^{23} 1.5120; ν_{max} 1130, 1060 cm⁻¹. Mass spectrum: m/z 85 (C₅H₉O, 100%), 133, 135 (C₃H₂OBr; 7 and 9%). ¹H-N.m.r. data (CDCl₃): δ 4.62 (t. 1 H, J 3 Hz, OCHO), 3.75-3.90 (m, 2 H), 3.40-3.70 (m, 6 H), 2.35 (septet, 1 H, J ~5 Hz, BrCH₂CH).

Anal. Calc. for C₉H₁₆Br₂O₂: C, 34.2; H, 5.1. Found: C, 34.7; H, 5.1.

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REFERENCES

- 1 S. HAKOMORI, Annu. Rev. Biochem., 50 (1981) 733-764.
- 2 K.-A. KARLSSON, in D. CHAPMAN (Ed.), Biological Membranes, Vol. 4, Academic Press, New York, 1982, pp. 1–74; K. BOCK, K.-A. KARLSSON, N. STRÖMBERG, AND S. TENEBERG, in A. WU (Ed.), Molecular Immunology of Complex Carbohydrates, Plenum Press, in press.
- 3 R. U. LEMIEUX, Proc. Int. Symp. Med. Chem., VIIIth, Uppsala, 1 (1984) 329-351.
- 4 R. GIGG, Chem. Phys. Lipids, 26 (1980) 287-404; M. M. PONPIPOM, R. L. BUGIANESI, AND T. Y. SHEN, Can. J. Chem., 58 (1980) 214-220; J. SLAMA AND R. R. RANDO, Carbohydr. Res., 88 (1981) 213-221; Biochemistry, 19 (1980) 4595-4600; R. R. RANDO, J. SLAMA, AND F. W. BAUGERTER, Proc. Natl. Acad. Sci. U.S.A., 77 (1980) 2510-2513; G. MAGNUSSON, in D. LARK (Ed.), Molecular Biology of Microbial Pathogenicity, Academic Press, New York, 1986, pp. 215-228.
- 5 I. N. ISRAELACHVILI, S. MARCELJA, AND R. G. HORN, Q. Rev. Biophys., 13 (1980) 121-200.
- 6 (a) J. DAHMÉN, T. FREJD, G. MAGNUSSON, AND G. NOORI, Carbohydr. Res., 111 (1982) C1-C4; (b)
 J. DAHMÉN, T. FREJD, G. GRÖNBERG, T. LAVE, G. MAGNUSSON, AND G. NOORI, *ibid.*, 116 (1983)
 303-307; (c) J. DAHMÉN, T. FREJD, G. GRÖNBERG, T. LAVE, G. MAGNUSSON, AND G. NOORI, *ibid.*, 118 (1983) 292-301; (d) J. DAHMÉN, T. FREJD, G. MAGNUSSON, G. NOORI, AND A.-S. CARLSTRÖM, *ibid.*, 125 (1984) 237-245; (e) *idem*, *ibid.*, 127 (1984) 15-25; (f) *idem*, *ibid.*, 127 (1984) 27-33; (g) *idem*, *ibid.*, 129 (1984) 63-71.
- 7 G. MAGNUSSON, G. NOORI, J. DAHMÉN, T. FREJD, AND T. LAVE, Acta Chem. Scand., Ser. B, 35 (1981) 213–216; J. DAHMÉN, T. FREJD, G. MAGNUSSON, AND G. NOORI, Carbohydr. Res., 114 (1983) 328–330.
- 8 H. PAULSEN AND M. PAAL, Carbohydr. Res., 135 (1984) 53-69.
- 9 K. JANSSON, T. FREJD, J. KIHLBERG, AND G. MAGNUSSON, Tetrahedron Lett., 27 (1986) 753-756.
- 10 A. F. FERRIS, J. Org. Chem., 20 (1955) 780-787.
- 11 R. U. LEMIEUX, T. TAKEDA, AND B. Y. CHUNG, ACS Symp. Ser., 39 (1976) 90-115.
- 12 J. DAHMÉN, T. FREID, T. LAVE, F. LINDH, G. MAGNUSSON, G. NOORI, AND K. PÅLSSON, Carbohydr. Res., 113 (1983) 219–224.
- 13 J. DAHMÉN, T. FREJD, G. GRÖNBERG, G. MAGNUSSON, AND G. NOORI, Carbohydr. Res., 125 (1984) 161–164.