

## SYNTHESIS OF NONREDUCING-SUGAR SUBUNIT ANALOGS OF BACTERIAL LIPID A CARRYING AN AMIDE-BOUND (3R)-3-ACYLOXY-TETRADECANOYL GROUP

MAKOTO KISO, SHINJI TANAKA, MINORU FUJITA, YUSHUN FUJISHIMA, YUJI OGAWA, AND AKIRA HASEGAWA

*Department of Agricultural Chemistry, Gifu University, Gifu 501-11 (Japan)*

(Received July 21st, 1986; accepted for publication in revised form, October 8th, 1986)

### ABSTRACT

Two types of optically active, 4-*O*-phosphono-D-glucosamine derivatives related to the nonreducing-sugar subunit of bacterial lipid A, one being 2-[(3R)-3-acyloxytetradecanamido]-2-deoxy-4-*O*-phosphono-3-*O*-tetradecanoyl-D-glucose (GLA-27 type; GLA-57 and GLA-58), and the other 2-[(3R)-3-acyloxytetradecanamido]-2-deoxy-3-*O*-[(3R)-3-hydroxytetradecanoyl]-4-*O*-phosphono-D-glucose (GLA-59 type; GLA-61 and GLA-62), have been synthesized. The amino group of benzyl 2-amino-2-deoxy-4,6-*O*-isopropylidene- $\beta$ -D-glucopyranoside was first acylated with the (3R)-3-dodecanoyloxytetradecanoyl or (3R)-3-hexadecanoyloxytetradecanoyl group, and then the remaining hydroxyl group was esterified with the tetradecanoyl or (3R)-3-(benzyloxymethoxy)tetradecanoyl group, respectively. The resulting protected intermediates were each converted, by the sequence of *O*-deisopropylideneation, 6-*O*-tritylation, and 4-*O*-phosphorylation, into the desired compounds.

### INTRODUCTION

Lipid A has been proved<sup>1</sup> to be the active center responsible for most of the endotoxic activity of bacterial lipopolysaccharide (LPS).

Since discovery that a 4-*O*-phosphono-D-glucosamine derivative named<sup>2</sup> GLA-27, which has a structure analogous to that of the nonreducing-sugar subunit of bacterial lipid A, can express some distinct biological activities, such as *Limulus* amoebocyte-lysate gelation, interferon- and tumor necrosis factor (TNF)-induction, and mitogenic and polyclonal B cell activation activities, a variety of analogs have been synthesized<sup>3</sup> and their biological activities tested<sup>4</sup>. 2-Deoxy-3-*O*-(3-hydroxytetradecanoyl)-4-*O*-phosphono-2-(3-tetradecanoyloxytetradecanamido)-D-glucose (GLA-59)<sup>3c</sup> has also been found<sup>5</sup> as an immunological active molecule similar to GLA-27. It is of interest that the two compounds have a common, amide-bound tetradecanoyloxytetradecanoyl group at N-2 of the D-glucosamine moiety.

It has been revealed that the amide-bound acyloxyacyl group in lipid A varies

with the bacterial species<sup>6</sup>. The lipid A of *Salmonella minnesota*<sup>7</sup> has both a dodecanoyloxy-, and a hexadecanoyloxy-tetradecanoyl group, whereas that of the *Proteus mirabilis* Re mutant<sup>8</sup> has the tetradecanoyloxy- and hexadecanoyloxy-tetradecanoyl groups. In the *Escherichia coli* lipid A, however, only the dodecanoyloxy-tetradecanoyl group has been characterized<sup>9</sup>. In addition, a D-glucosamine-derived phospholipid named<sup>10</sup> lipid Y, found in certain *E. coli* mutants, also carries the amide-bound hexadecanoyloxytetradecanoyl group.

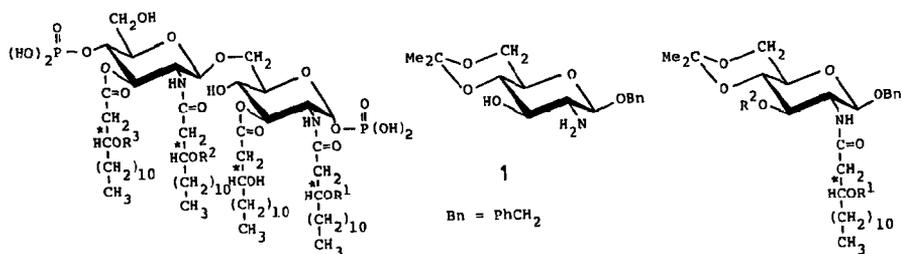
We now describe the synthesis of some nonreducing-sugar subunit analogs of bacterial lipid A which carry the amide-bound dodecanoyloxy- and hexadecanoyloxy-tetradecanoyl group at N-2 of the D-glucosamine backbone.

## RESULTS AND DISCUSSION

(3*R*)-3-Dodecanoyloxytetradecanoic acid (**2**) and (3*R*)-3-hexadecanoyloxy-tetradecanoic acid (**3**) were prepared *via* the phenacyl ester of (3*R*)-3-hydroxy-tetradecanoic acid as previously described<sup>3c</sup>. These (3*R*)-3-acyloxytetradecanoic acids were each treated<sup>2b</sup> with benzyl 2-amino-2-deoxy-4,6-*O*-isopropylidene- $\beta$ -D-glucopyranoside (**1**), in the presence of 3-(3-dimethylamino)propyl-1-ethylcarbodiimide hydrochloride (WSC) in dichloromethane, to give the corresponding benzyl 2-(acyloxytetradecanamido)-2-deoxy-4,6-*O*-isopropylidene- $\beta$ -D-glucopyranoside (**5** or **6**, respectively). Esterification of the remaining 3-hydroxyl group with tetradecanoic and (3*R*)-3-(benzyloxymethoxy)tetradecanoic acid in the presence of WSC (or dicyclohexylcarbodiimide, DCC) and 4-(dimethylamino)pyridine (DMAP), gave **7**, **8**, **9**, and **10**, from which **11**, **14**, **17**, and **20** were obtained by hydrolytic removal of the isopropylidene group. After 6-*O*-tritylation, the diphenylphosphono group was introduced at O-4 in the usual manner<sup>2b,3c</sup>, and the product was purified by chromatography.

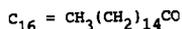
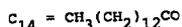
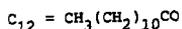
Hydrolytic removal of the trityl group gave the protected synthetic intermediates **13**, **16**, **19**, and **22**, from which **23**, **24**, **25**, and **26** were obtained in nearly quantitative yield by hydrogenolytic removal of the benzyl group in the presence of palladium catalyst. The pair **23** and **24**, or **25** and **26**, showed <sup>1</sup>H-n.m.r. spectra quite similar to that of the corresponding precursor of GLA-27<sup>2b</sup> or GLA-59<sup>3c</sup>, respectively, except for minor differences in the number of methylene protons and the chemical shift of the hydroxyl protons. In all these compounds, the  $\alpha$ -D-pyranose form preponderates in the equilibrium mixture in chloroform-*d*, as previously described for a series of homologous compounds. Finally, the phenyl groups were cleaved by hydrogenolysis in the presence of platinum catalyst, to afford the desired GLA-57(*R*), GLA-58(*R*), GLA-61(*R,R*) and GLA-62(*R,R*) as colorless powders which gave a positive test with the specific spray-reagent<sup>11</sup> for the phosphate group. It may be noted that, like GLA-27(*R*), these compounds are essentially insoluble in most of the organic solvents.

The structures of GLA-57(*R*) and GLA-61(*R,R*) are closely related to those of the nonreducing-sugar subunit of the lipid A of *S. minnesota* and *E. coli*, whereas



Bacterial lipid A

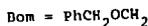
	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>
<i>S. minnesota</i>	C <sub>16</sub>	C <sub>12</sub>	C <sub>14</sub>
<i>P. mirabilis</i>	C <sub>16</sub>	C <sub>14</sub>	C <sub>14</sub>
<i>E. coli</i>	H	C <sub>12</sub>	C <sub>14</sub>



\* (R) -configuration

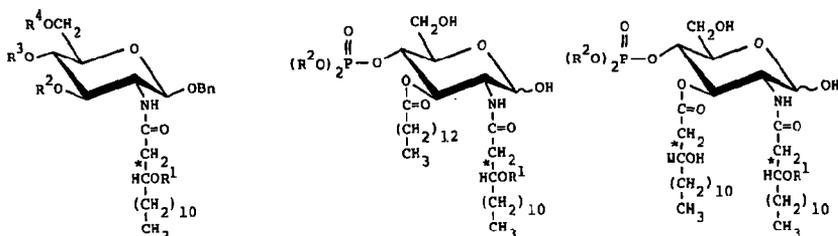


- 2 R = C<sub>12</sub>  
3 R = C<sub>16</sub>  
4 R = Bom



	R <sup>1</sup>	R <sup>2</sup>
5	C <sub>12</sub>	H
6	C <sub>16</sub>	H
7	C <sub>12</sub>	C <sub>14</sub>
8	C <sub>16</sub>	C <sub>14</sub>
9	C <sub>12</sub>	C <sub>14</sub> -OBom
10	C <sub>16</sub>	C <sub>14</sub> -OBom

C<sub>14</sub>-OBom = CH<sub>3</sub>(CH<sub>2</sub>)<sub>10</sub><sup>\*</sup>CHCH<sub>2</sub>CO  
OBom



	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>		R <sup>1</sup>	R <sup>2</sup>		R <sup>1</sup>	R <sup>2</sup>
11	C <sub>12</sub>	C <sub>14</sub>	H	H	23	C <sub>12</sub>	Ph	25	C <sub>12</sub>	Ph
12	C <sub>12</sub>	C <sub>14</sub>	H	Tr	24	C <sub>16</sub>	Ph	26	C <sub>16</sub>	Ph
13	C <sub>12</sub>	C <sub>14</sub>	(PhO) <sub>2</sub> PO	H	GLA-57 (R)	C <sub>12</sub>	H	GLA-61 (R,R)	C <sub>12</sub>	H
14	C <sub>16</sub>	C <sub>14</sub>	H	H	GLA-58 (R)	C <sub>16</sub>	H	GLA-62 (R,R)	C <sub>16</sub>	H
15	C <sub>16</sub>	C <sub>14</sub>	H	Tr	[GLA-27 (R)	C <sub>14</sub>	H	[GLA-59 (R,R)	C <sub>14</sub>	H
16	C <sub>16</sub>	C <sub>14</sub>	(PhO) <sub>2</sub> PO	H						
17	C <sub>12</sub>	C <sub>14</sub> -OBom	H	H						
18	C <sub>12</sub>	C <sub>14</sub> -OBom	H	Tr						
19	C <sub>12</sub>	C <sub>14</sub> -OBom	(PhO) <sub>2</sub> PO	H						
20	C <sub>16</sub>	C <sub>14</sub> -OBom	H	H						
21	C <sub>16</sub>	C <sub>14</sub> -OBom	H	Tr						
22	C <sub>16</sub>	C <sub>14</sub> -OBom	(PhO) <sub>2</sub> PO	H						

Tr = Ph<sub>3</sub>C

GLA-58(*R*) and GLA-62(*R,R*) have partial structures similar to lipid Y. Furthermore, the importance of the chain length of the C<sub>14</sub> *O*-acyl group will be clarified by comparing the biological activities of these compounds with those of GLA-27(*R*) and GLA-59(*R,R*), which carry the amide-bound tetradecanoyloxytetradecanoyl (C<sub>14</sub>-*O*-C<sub>14</sub>) group at N-2 of the D-glucosamine backbone.

## EXPERIMENTAL

*General methods.* — The instrumental and chromatographic procedures employed were those previously given<sup>2b</sup>. <sup>1</sup>H-N.m.r. spectra were recorded at 270 MHz. For details of the reaction procedures employed, see also, ref. 3c.

(3*R*)-3-Dodecanoyloxytetradecanoic acid (**2**) and (3*R*)-3-hexadecanoyloxytetradecanoic acid (**3**). — To a cooled, stirred solution of (3*R*)-3-hydroxytetradecanoic acid phenacyl ester<sup>3c</sup> (2 g) in pyridine (21 mL) were added a catalytic amount of DMAP and dodecanoyl or hexadecanoyl chloride (1.2 mol equiv.) in dry dichloromethane; the mixture was stirred overnight at room temperature. Methanol was added, and the mixture was processed as described<sup>3c</sup> for the preparation of (3*R*)-3-tetradecanoyloxytetradecanoic acid, to give the corresponding phenacyl ester of (3*R*)-3-dodecanoyloxytetradecanoic acid (2.87 g, 95.3%); m.p. 35.5°, [ $\alpha$ ]<sub>D</sub> +1.3° (c 0.7, chloroform);  $\nu_{\max}^{\text{film}}$  1740, 1730, 1710 (C=O), 780–670 (Ph), and complete loss of the peak at 3600–3200 cm<sup>-1</sup> (OH); and of (3*R*)-3-hexadecanoyloxytetradecanoic acid (3.3 g, 99.4%); m.p. 38.5–40°, [ $\alpha$ ]<sub>D</sub> +1° (c 0.9, chloroform).

Finally, the phenacyl esters of the (3*R*)-3-acyloxytetradecanoic acids (2 g) were each treated with zinc dust (3 g) in acetic acid (10 mL) at 50°. The resulting, free acids were each purified by chromatography on a column of silica gel (Wakogel C-200) with dichloromethane, to give **2** or **3** as a syrup in nearly quantitative yield.

Compound **2** had [ $\alpha$ ]<sub>D</sub> -1° (c 0.9, chloroform); i.r. data  $\nu_{\max}^{\text{film}}$  3700–2500 (CO<sub>2</sub>H), 1750, 1720 (C=O), and complete loss of the peaks at 780–670 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>26</sub>H<sub>50</sub>O<sub>4</sub> (426.66): C, 73.19; H, 11.81. Found: C, 73.42; H, 11.67.

Compound **3** had [ $\alpha$ ]<sub>D</sub> -1° (c 1, chloroform); i.r. data similar to those of **2**.

*Anal.* Calc. for C<sub>30</sub>H<sub>58</sub>O<sub>4</sub> (482.76): C, 74.63; H, 12.11. Found: C, 74.78; H, 12.23.

*Benzyl 2-deoxy-2-[(3R)-3-dodecanoyloxytetradecanamido]-4,6-O-isopropylidene-β-D-glucopyranoside (5) and benzyl 2-deoxy-2-[(3R)-3-hexadecanoyloxytetradecanamido]-4,6-O-isopropylidene-β-D-glucopyranoside (6).* — To a solution of **1** (0.65 g) in dry dichloromethane (5 mL) were added **2** (0.9 g) and WSC (0.62 g); the mixture was stirred at room temperature. After completion of the reaction (t.l.c., 2:1 ethyl acetate–hexane), the mixture was evaporated to a residue that was chromatographed on a column of silica gel (Wakogel C-200) with 400:1 dichloromethane–methanol, to afford **5** (1.36 g; 90%), which was lyophilized from 1,4-dioxane solution; m.p. 63–64°, [ $\alpha$ ]<sub>D</sub> -47° (c 1, chloroform);  $\nu_{\max}^{\text{film}}$  3500, 3375,

3300 (OH, NH), 1730 (ester), 1660, 1550, 1530 (amide), 860 (CMe<sub>2</sub>), and 740–680 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>42</sub>H<sub>71</sub>NO<sub>8</sub> (718.00): C, 70.25; H, 9.97; N, 1.95. Found: C, 70.39; H, 10.14; N, 1.81.

Compound **6** (1.44 g; 93.5%) was obtained by treatment of **1** (0.64 g) with **3** (1.2 g) as described for **5**; m.p. 71–72°, [ $\alpha$ ]<sub>D</sub> -48° (c 0.9, chloroform);  $\nu_{\text{max}}^{\text{film}}$  3600–3200 (OH, NH), 1730 (ester), 1660, 1530 (amide), 850 (CMe<sub>2</sub>), and 760–680 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>46</sub>H<sub>79</sub>NO<sub>8</sub> (774.10): C, 71.37; H, 10.29; N, 1.81. Found: C, 71.18; H, 10.35; N, 1.77.

*Benzyl 2-deoxy-2-[(3R)-3-dodecanoyloxytetradecanamido]-4,6-O-isopropylidene-3-O-tetradecanoyl- $\beta$ -D-glucopyranoside (7) and benzyl 2-deoxy-2-[(3R)-3-hexadecanoyloxytetradecanamido]-4,6-O-isopropylidene-3-O-tetradecanoyl- $\beta$ -D-glucopyranoside (8).* — To a solution of **5** (0.55 g) in dry dichloromethane (6 mL) were added tetradecanoic acid (0.21 g), WSC (0.294 g), and a catalytic amount of DMAP; the mixture was stirred at room temperature. After completion of the reaction (t.l.c., 80:1 dichloromethane–methanol), the mixture was processed as described for **5**. The product was purified by chromatography on a column of silica gel (Wakogel C-200) with 500:1 dichloromethane–methanol, to give **7** (0.69 g; 97%), which was lyophilized from 1,4-dioxane solution; m.p. 86–88°, [ $\alpha$ ]<sub>D</sub> -34° (c 0.9, chloroform);  $\nu_{\text{max}}^{\text{film}}$  3350 (NH), 1740 (ester), 1660, 1550, 1530 (amide), 860 (CMe<sub>2</sub>), and complete loss of the peak at 3500 cm<sup>-1</sup> (OH).

*Anal.* Calc. for C<sub>36</sub>H<sub>67</sub>NO<sub>9</sub> (928.35): C, 72.45; H, 10.53; N, 1.51. Found: C, 72.31; H, 10.40; N, 1.49.

Compound **8** (0.74 g, 97%) was obtained by treatment of **6** (0.6 g) with tetradecanoic acid (0.212 g) in the presence of WSC (0.297 g) and DMAP (10 mg) as described for **7**; m.p. 88–89°, [ $\alpha$ ]<sub>D</sub> -32° (c 1, chloroform);  $\nu_{\text{max}}^{\text{film}}$  3400 (NH), 1740 (ester), 1680, 1520 (amide), 860 (CMe<sub>2</sub>), and complete loss of the peak at 3500 cm<sup>-1</sup> (OH).

*Anal.* Calc. for C<sub>60</sub>H<sub>105</sub>NO<sub>9</sub> (984.45): C, 73.20; H, 10.75; N, 1.42. Found: C, 73.46; H, 10.78; N, 1.33.

*Benzyl 3-O-[(3R)-3-(benzyloxymethoxy)tetradecanoyl]-2-deoxy-2-[(3R)-3-dodecanoyloxytetradecanamido]-4,6-O-isopropylidene- $\beta$ -D-glucopyranoside (9).* — Compound **5** (0.55 g) in dichloromethane (6 mL) was treated with **4** (ref. 3c) (0.334 g) in the presence of WSC (0.294 g) and DMAP (9 mg), and the mixture was processed as described for **7** and **8**. The product was purified by chromatography on a column of silica gel (Wakogel C-200) with 500:1 dichloromethane–methanol, to give **9** (0.8 g, 98.2%); m.p. 54–56°, [ $\alpha$ ]<sub>D</sub> -22° (c 1, chloroform);  $\nu_{\text{max}}^{\text{film}}$  3320 (NH), 1740, 1730 (ester), 1660, 1550, 1530 (amide), 860 (CMe<sub>2</sub>), and 760–680 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>64</sub>H<sub>105</sub>NO<sub>11</sub> (1064.49): C, 72.21; H, 9.94; N, 1.32. Found: C, 72.03; H, 9.81; N, 1.29.

*Benzyl 3-O-[(3R)-3-(benzyloxymethoxy)tetradecanoyl]-2-deoxy-2-[(3R)-3-hexadecanoyloxytetradecanamido]-4,6-O-isopropylidene- $\beta$ -D-glucopyranoside (10).*

— Compound **10** (0.79 g, 97.7%) was obtained by treatment of **6** (0.56 g) with **4** (0.314 g) in the presence of WSC (0.277 g) and DMAP (9 mg) as described for **7**, **8**, and **9**; m.p. 61–63°,  $[\alpha]_D -21^\circ$  (c 1, chloroform);  $\nu_{\max}^{\text{film}}$  3340 (NH), 1740 (ester), 1660, 1540 (amide), 860 (CMe<sub>2</sub>), and 760–690 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>68</sub>H<sub>113</sub>NO<sub>11</sub> (1120.58): C, 72.88; H, 10.16; N, 1.25. Found: C, 73.14; H, 9.99; N, 1.17.

*Benzyl 2-deoxy-4-O-(diphenylphosphono)-2-[(3R)-3-dodecanoyloxytetradecanamido]-4,6-O-isopropylidene-3-O-tetradecanoyl-β-D-glucopyranoside (13).* — *O*-Deisopropylideneation of **7** (0.65 g) was performed by treatment with 95% acetic acid (16 mL) as described previously<sup>2b</sup>, to give **11** in 82% yield; m.p. 144–146°,  $[\alpha]_D -24^\circ$  (c 0.8, chloroform).

A solution of **11** (0.48 g) in dry pyridine (7 mL) was stirred at 90°, and then trityl chloride (0.3 g) was added; stirring was continued for 6 h at 90°, the mixture was cooled, methanol was added, and the solvents were evaporated. After extractive processing, the product was purified by chromatography on a column of silica gel (Wakogel C-200) with 500:1 dichloromethane–methanol, to afford **12** (0.61 g; quantitative); m.p. 100–101°,  $[\alpha]_D -27^\circ$  (c 0.9, chloroform); <sup>1</sup>H-n.m.r. data (CDCl<sub>3</sub>): δ 0.88 (~t, 9 H, CH<sub>3</sub>), 1.0–1.4, 1.4–1.7 (m, 54 H + 6 H, -CH<sub>2</sub>-), 2.15–2.5 (m, 6 H, -COCH<sub>2</sub>-), 2.63 (d, 1 H, *J* 3.7 Hz, OH), 3.3–3.5 (m, 3 H, H-5,6,6'), 3.75 (m, 1 H, H-4), 4.05 (~q, 1 H, H-2), 4.5 (d, 1 H, *J*<sub>1,2</sub> 8.4 Hz, H-1), 4.63, 4.88 (2 d, 2 H, *J*<sub>gem</sub> 12.1 Hz, CH<sub>2</sub>Ph), 4.93 (dd, 1 H, *J*<sub>2,3</sub> 9, *J*<sub>3,4</sub> 10 Hz, H-3), 5.04 (m, 1 H, H-3 of C<sub>14</sub>-O-C<sub>12</sub>), 5.81 (d, 1 H, *J* 9.2 Hz, NH), and 7.1–7.6 (m, 20 H, Ph-H).

The introduction of the diphenylphosphono group at O-4 of **12** (0.58 g) was accomplished with diphenyl phosphorochloridate (0.217 g) and DMAP (94 mg) in 2:1 dichloromethane–pyridine as described in ref. 2b. The product was purified by chromatography on a column of silica gel (Wakogel C-300) with 5:1 hexane–ethyl acetate, to give benzyl 2-deoxy-4-*O*-(diphenylphosphono)-2-[(3R)-3-dodecanoyloxytetradecanamido]-3-*O*-tetradecanoyl-6-*O*-trityl-β-D-glucopyranoside (0.52 g; 74.4%), which was then treated with tetrafluoroboric acid<sup>12</sup> in acetone, to afford **13** in almost quantitative yield; m.p. 85.5–86°,  $[\alpha]_D -32^\circ$  (c 0.9, chloroform);  $\nu_{\max}^{\text{film}}$  3500, 3300 (OH, NH), 3150–3000 (Ph), 1740 (ester), 1650, 1560 (amide), 960 (P–O–Ph), and 780–680 cm<sup>-1</sup> (Ph).

*Anal.* Calc. for C<sub>65</sub>H<sub>102</sub>NO<sub>12</sub>P (1120.46); C, 69.67; H, 9.18; N, 1.25. Found: C, 69.83; H, 9.20; N, 1.14.

*Benzyl 2-deoxy-4-O-(diphenylphosphono)-2-[(3R)-3-hexadecanoyloxytetradecanamido]-4,6-O-isopropylidene-3-O-tetradecanoyl-β-D-glucopyranoside (16).* — A mixture of **8** (0.67 g) and 95% acetic acid (16 mL) was processed as described for **11**, to give **14** in almost quantitative yield; m.p. 135–137°,  $[\alpha]_D -22^\circ$  (c 0.8, chloroform);  $\nu_{\max}^{\text{film}}$  3500, 3300 (OH, NH), 1740, 1705 (ester), 1660, 1570 (amide), 750–680 (Ph), and complete loss of the peak at 860 cm<sup>-1</sup> (CMe<sub>2</sub>).

Tritylation of the primary hydroxyl group of **14** (0.56 g) was conducted with trityl chloride (0.3 g) in pyridine (8 mL) as described for **12**, to afford **15** (0.7 g;

99.4%); m.p. 74–76°,  $[\alpha]_D -25^\circ$  (c 1, chloroform);  $^1\text{H-n.m.r. data}$  ( $\text{CDCl}_3$ ):  $\delta$  0.88 (t, 9 H,  $\text{CH}_3$ ), 1.0–1.7 (m, 68 H,  $-\text{CH}_2-$ ), 2.15–2.5 (m, 6 H,  $-\text{COCH}_2-$ ), 2.61 (d, 1 H,  $J_{4,\text{OH}}$  3.7 Hz, OH), 3.3–3.5 (m, 3 H, H-5,6,6'), 3.75 (m, 1 H, H-4), 4.05 (~q, 1 H,  $J \sim 9$  Hz, H-2), 4.50 (d, 1 H,  $J_{1,2}$  8.4 Hz, H-1), 4.63, 4.88 (2 d, 2 H,  $\text{CH}_2\text{Ph}$ ), 4.93 (~t, 1 H, H-3), 5.04 (m, 1 H, H-3 of  $\text{C}_{14}\text{-O-C}_{16}$ ), 5.75 (d, 1 H,  $J$  9.2 Hz, NH), and 7.1–7.55 (m, 20 H, Ph-H).

The introduction of the diphenylphosphono group at O-4 of **15** (0.7 g) was performed by the procedure described for **13**, to give benzyl 2-deoxy-4-*O*-(diphenylphosphono)-2-[(3*R*)-3-hexadecanoyloxytetradecanamido]-3-*O*-tetradecanoyl-6-*O*-trityl- $\beta$ -D-glucopyranoside (0.76 g; 91%), which was then treated with aqueous acetic acid. The resulting **16** (nearly quantitative) was purified by chromatography on a column of silica gel (Wakogel C-300) with 5:2 hexane–ethyl acetate; m.p. 83–85°,  $[\alpha]_D -31^\circ$  (c 1, chloroform); i.r. data similar to those of **13**.

*Anal. Calc.* for  $\text{C}_{69}\text{H}_{110}\text{NO}_{12}\text{P}$  (1176.56): C, 70.43; H, 9.42; N, 1.19. Found: C, 70.62; H, 9.27; N, 1.04.

*Benzyl 3-O-[(3R)-3-(benzyloxymethoxy)tetradecanoyl]-2-deoxy-4-O-(diphenylphosphono)-2-[(3R)-3-dodecanoyloxytetradecanamido]- $\beta$ -D-glucopyranoside (19).* — Compound **9** (0.77 g) was treated with aqueous acetic acid as described for **11** and **14**, to give **17** (0.7 g, 94.5%); m.p. 96–98°,  $[\alpha]_D -38^\circ$  (c 0.9, chloroform). The 6-*O*-trityl derivative **18** (0.75 g; 91%) was obtained from **17** (0.67 g) in the usual way; m.p. 84–86°,  $[\alpha]_D -33^\circ$  (c 1, chloroform);  $^1\text{H-n.m.r. data}$  ( $\text{CDCl}_3$ ):  $\delta$  0.88 (~t, 9 H,  $\text{CH}_3$ ), 1.0–1.7 (m, 58 H,  $-\text{CH}_2-$ ), 2.15–2.65 (m, 4 H, H-4,5,6,6'), 3.9–4.1 (m, 2 H, H-2, and H-3 of  $\text{C}_{14}\text{-OBom}$ ), 4.44 (d, 1 H,  $J_{1,2}$  8.4 Hz, H-1), 4.45–4.95 (6 d, 6 H,  $-\text{OCH}_2\text{O-}$  and  $\text{CH}_2\text{Ph}$ ), 4.85 (dd, 1 H, H-3), 5.05 (m, 1 H, H-3 of  $\text{C}_{14}\text{-O-C}_{12}$ ), 5.71 (d, 1 H, NH), and 7.1–7.55 (m, 25 H, Ph-H).

4-*O*-Phosphorylation of **18** (0.72 g), followed by detritylation, afforded compound **19** (62% in 2 steps); m.p. 68–71°,  $[\alpha]_D -16^\circ$  (c 1, chloroform);  $\nu_{\text{max}}^{\text{film}}$  3500, 3280 (OH, NH), 3150–3000 (Ph), 1740 (ester), 1650, 1560 (amide), 960 (P–O–Ph), and 780–680  $\text{cm}^{-1}$  (Ph).

*Anal. Calc.* for  $\text{C}_{73}\text{H}_{110}\text{NO}_{14}\text{P}$  (1256.60): C, 69.77; H, 8.82; N, 1.11. Found: C, 70.05; H, 8.73; N, 1.00.

*Benzyl 3-O-[(3R)-3-(benzyloxymethoxy)tetradecanoyl]-2-deoxy-4-O-(diphenylphosphono)-2-[(3R)-3-hexadecanoyloxytetradecanamido]- $\beta$ -D-glucopyranoside (22).* — Compound **20** (0.66 g, 91.3%) was obtained from **10** (0.75 g) as described for **17**; m.p. 97–97.5°,  $[\alpha]_D -38^\circ$  (c 0.8, chloroform); i.r. data similar to those of **17**.

The 6-*O*-tritylation of **20** (0.57 g) gave **21** (0.69 g, 99%); m.p. 79–81.5°,  $[\alpha]_D -30^\circ$  (c 1, chloroform);  $^1\text{H-n.m.r. data}$  ( $\text{CDCl}_3$ ):  $\delta$  0.88 (t, 9 H,  $\text{CH}_3$ ), 1.0–1.7 (m, 66 H,  $-\text{CH}_2-$ ), 2.1–2.7 (m, 6 H,  $-\text{COCH}_2-$ ), 3.08 (~s, 1 H, OH), 3.2–3.5 (m, 4 H, H-4,5,6,6'), 3.9–4.1 (m, 2 H, H-2, and H-3 of  $\text{C}_{14}\text{-O-C}_{16}$ ), 5.73 (d, 1 H,  $J$  8.8, NH), and 7.1–7.55 (m, 25 H, Ph-H).

Compound **21** (0.68 g) was converted, by the sequence described for **13**, **16**, and **19**, into **22** (66% in 2 steps); m.p. 68–69°,  $[\alpha]_D -15^\circ$  (c 1, chloroform); i.r. data similar to those of **19**.

*Anal.* Calc. for  $C_{77}H_{118}NO_{14}P$  (1312.70): C, 70.45; H, 9.06; N, 1.07. Found: C, 70.73; H, 9.18; N, 0.96.

2-Deoxy-2-[(3R)-3-dodecanoyloxytetradecanamido]-4-O-phosphono-3-O-tetradecanoyl-D-glucose [GLA-57(R)] and 2-deoxy-2-[(3R)-3-hexadecanoyloxytetradecanamido]-4-O-phosphono-3-O-tetradecanoyl-D-glucose [GLA-58(R)]. — Compound **13** (0.23 g) was dissolved in methanol–ethanol–toluene (10 mL), and hydrogenolyzed in the presence of 10% palladium–carbon catalyst (0.1 g). The catalyst was filtered off, and washed with the same solvent. The filtrate and washings were combined, and evaporated to a residue which was chromatographed on a column of silica gel (Wakogel C-300) with 2:1 ethyl acetate–hexane, to give **23** in almost quantitative yield; m.p. 95–96°,  $[\alpha]_D +4.3^\circ$  (c 1.7, chloroform);  $^1H$ -n.m.r. data for the  $\alpha$  anomer ( $CDCl_3$ ):  $\delta$  0.88 (t, 9 H,  $CH_3$ ), 1.0–1.7 (m, 60 H,  $-CH_2-$ ), 3.28 (~t, 1 H, OH-6), 3.5–3.7 (m, 2 H, H-6,6'), 3.78 (d, 1 H,  $J_{1,OH} \sim 3$  Hz, OH-1), 4.00 (~d, 1 H,  $J_{4,5}$  9.9 Hz, H-5), 4.24 (m, 1 H, H-2), 4.76 (~q, 1 H,  $J_{3,4} = J_{4,5} = J_{4,P} = 9$ –10 Hz, H-4), 5.09 (m, 1 H, H-3 of  $C_{14}-O-C_{12}$ ), 5.28 (~t, 1 H,  $J$  3–4 Hz, H-1), 5.48 (~t, 1 H,  $J_{2,3}$  10.6,  $J_{3,4}$  9.5 Hz, H-3), 6.07 (d, 1 H,  $J$  9.2 Hz, NH), and 7.1–7.45 (m, 10 H, Ph-H).

Compound **24** was obtained from **16** (0.21 g) as described for **23**; m.p. 92.5–93.5°,  $[\alpha]_D +4.2^\circ$  (c 1.6, chloroform);  $^1H$ -n.m.r. data for the  $\alpha$  anomer ( $CDCl_3$ ):  $\delta$  0.88 (t, 9 H,  $CH_3$ ), 1.0–1.7 (m, 68 H,  $-CH_2-$ ), 2.0–2.5 (m, 6 H,  $-COCH_2-$ ), 3.29 (~t, 1 H, OH-6), 3.5–3.7 (m, 2 H, H-6,6'), 3.86 (~s, 1 H, OH-1), 4.01 (~d, 1 H,  $J_{4,5}$  9.9 Hz, H-5), 4.23 (m, 1 H, H-2), 4.76 (~q, 1 H,  $J_{3,4} = J_{4,5} = J_{4,P} = \sim 9.5$  Hz, H-4), 5.09 (m, 1 H, H-3 of  $C_{14}-O-C_{16}$ ), 5.27 (t, 1 H,  $J_{1,2} = J_{1,OH} = 3$ –4 Hz, H-1), 5.48 (~t, 1 H, H-3), 6.08 (d, 1 H,  $J$  9.2 Hz, NH), and 7.1–7.45 (m, 10 H, Ph-H).

The phenyl groups of **23** (0.16 g) and **24** (0.15 g) were removed by hydrogenolysis in the presence of pre-reduced, Adams' platinum catalyst (50 mg) in ethanol–methanol as described<sup>2b</sup> for the preparation of GLA-27, and the products were lyophilized from suspensions in 1,4-dioxane, to give colorless, fine powders. Because, like GLA-27(R), both compounds were essentially insoluble in the usual organic solvents, their  $[\alpha]_D$  values could not be measured accurately.

GLA-57(R) had m.p. 162–163.5°;  $\nu_{max}^{KBr}$  3700–2500 (OH, NH), 1740, 1720 (ester), 1640, 1560 (amide), and complete loss of the peaks at 960 (P–O–Ph) and 780–680  $cm^{-1}$  (Ph).

*Anal.* Calc. for  $C_{46}H_{88}NO_{12}P$  (878.15): C, 62.91; H, 10.10; N, 1.60. Found: C, 63.25; H, 9.86; N, 1.39.

GLA-58(R) had m.p. 158–161°;  $\nu_{max}^{KBr}$  3700–2500 (OH, NH), 1740 (ester), 1640, 1560 (amide), and complete loss of the peaks at 960 (P–O–Ph) and 780–680  $cm^{-1}$  (Ph).

*Anal.* Calc. for  $C_{50}H_{96}NO_{12}P$  (934.26): C, 64.28; H, 10.36; N, 1.50. Found: C, 64.03; H, 10.21; N, 1.36.

2-Deoxy-2-[(3R)-3-dodecanoyloxytetradecanamido]-3-O-[(3R)-3-hydroxytetradecanoyl]-4-O-phosphono-D-glucose [GLA-61(R,R)] and 2-deoxy-2-[(3R)-3-hexadecanoyloxytetradecanamido]-3-O-[(3R)-3-hydroxytetradecanoyl]-4-O-phos-

*phono-D-glucose* [GLA-62(R,R)]. — Compound **19** (0.2 g) was hydrogenolyzed in methanol (10 mL) in the presence of 10% palladium-carbon catalyst (0.1 g), and the mixture was processed as described for GLA-57(R) and GLA-58(R), to give **25**; m.p. 88.5–90.5°,  $[\alpha]_D -1.5^\circ$  (c 1.5, chloroform);  $^1\text{H-n.m.r.}$  data for the  $\alpha$  anomer ( $\text{CDCl}_3$ ):  $\delta$  0.88 (t, 9 H,  $\text{CH}_3$ ), 1.0–1.8 (m, 58 H,  $-\text{CH}_2-$ ), 2.1–2.5 (m, 6 H,  $-\text{COCH}_2-$ ), 3.26 (~d, 1 H, OH-3 of the 3-hydroxytetradecanoyl group), 3.38 (broad t, 1 H, OH-6), 3.5–3.7 (broad s, 2 H, H-6,6'), 3.90 (m, 1 H, H-3 of the 3-hydroxytetradecanoyl group), 4.04 (~d, 1 H, H-5), 4.29 (m, 1 H, H-2), 4.38 (d, 1 H, OH-1), 4.77 (~q,  $J_{3,4} = J_{4,5} = J_{4,P} = \sim 9.5$  Hz, H-4), 5.08 (m, 1 H, H-3 of  $\text{C}_{14}\text{-O-C}_{12}$ ), 5.24 (t, 1 H,  $J_{1,\text{OH}} = J_{1,2} = 3\text{--}4$  Hz, H-1), 5.51 (~t, 1 H,  $J_{3,4} \sim 10$  Hz, H-3), 6.30 (d, 1 H,  $J$  9.2 Hz, NH) and 7.1–7.4 (m, 10 H, Ph-H).

Compound **26** was obtained from **22** as just described; m.p. 87–90°,  $[\alpha]_D -1.2^\circ$  (c 1.3, chloroform);  $^1\text{H-n.m.r.}$  data for the  $\alpha$  anomer ( $\text{CDCl}_3$ ):  $\delta$  0.88 (t, 9 H,  $\text{CH}_3$ ), 1.0–1.7 (m, 66 H,  $-\text{CH}_2-$ ), 2.1–2.5 (m, 6 H,  $-\text{COCH}_2-$ ), 3.23 (~d, 1 H, OH-3 of the 3-hydroxytetradecanoyl group), 4.0–4.1 (m, 2 H, H-5 and OH-1), 4.30 (m, 1 H, H-2), 4.78 (~q,  $J$  9–10 Hz, H-4), 5.08 (m, 1 H, H-3 of  $\text{C}_{14}\text{-O-C}_{16}$ ), 5.25 (t, 1 H,  $J$  3–4 Hz, H-1), 5.51 (~t, 1 H, H-3), 6.27 (d, 1 H,  $J$  9.2 Hz, NH), and 7.1–7.4 (m, 10 H, Ph-H).

Hydrogenolytic removal of the phenyl groups from **25** and **26** was performed in ethanol solution, and the reaction mixture was processed as described for GLA-57(R) and GLA-58(R), to give, almost quantitatively, GLA-61(R,R) and GLA-62(R,R) as colorless, fine powders.

GLA-61(R,R) had m.p. 177.5–180°,  $[\alpha]_D +11^\circ$  (c 0.1, 1:1 dichloromethane-ethanol);  $\nu_{\text{max}}^{\text{KBr}}$  3700–2600 (OH, NH), 1740 (ester), 1640, 1560 (amide), and complete loss of the peaks at 960 (P–O–Ph) and 780–680  $\text{cm}^{-1}$  (Ph).

*Anal. Calc.* for  $\text{C}_{46}\text{H}_{88}\text{NO}_{13}\text{P}$  (894.15): C, 61.79; H, 9.92; N, 1.57. Found: C, 61.47; H, 10.18; N, 1.69.

GLA-62(R,R) had m.p. 175–178°,  $[\alpha]_D +10^\circ$  (c 0.14, 1:1 dichloromethane-ethanol); i.r. data (KBr), same as those for GLA-61(R,R).

*Anal. Calc.* for  $\text{C}_{50}\text{H}_{96}\text{NO}_{13}\text{P}$  (950.26): C, 63.19; H, 10.18; N, 1.47. Found: C, 63.53; H, 9.99; N, 1.42.

## REFERENCES

- (a) C. GALANOS, O. LÜDERITZ, E. T. RIETSCHEL, AND O. WESTPHAL, in T. W. GOODWIN (Ed.), *Biochemistry of Lipids II, Int. Rev. Biochem.*, 14 (1977) 239–335; (b) O. LÜDERITZ, C. GALANOS, V. LEHMANN, H. MAYER, E. T. RIETSCHEL, AND J. WECKESSER, *Naturwissenschaften*, 65 (1978) 578–585; (c) C. GALANOS, O. LÜDERITZ, E. T. RIETSCHEL, O. WESTPHAL, H. BRADE, L. BRADE, M. FREUDENBERG, U. SCHADE, M. IMOTO, H. YOSHIMURA, S. KUSUMOTO, AND T. SHIBA, *Eur. J. Biochem.*, 148 (1985) 1–5; (d) S. KOTANI, H. TAKADA, M. TSUJIMOTO, T. OGAWA, I. TAKAHASHI, T. IEDA, K. OTSUKA, H. SHIMAUCHI, N. KASAI, J. MASHIMO, S. NAGAO, A. TANAKA, S. TANAKA, K. HARADA, K. NAGAKI, H. KITAMURA, T. SHIBA, S. KUSUMOTO, M. IMOTO, AND H. YOSHIMURA, *Infect. Immun.*, 49 (1985) 225–237; (e) J. Y. HOMMA, M. MATSUURA, S. KANEGASAKI, Y. KAWAKUBO, Y. KOJIMA, N. SHIBUKAWA, Y. KUMAZAWA, A. YAMAMOTO, K. TANAMOTO, T. YASUDA, M. IMOTO, H. YOSHIMURA, S. KUSUMOTO, AND T. SHIBA, *J. Biochem. (Tokyo)*, 98 (1985) 395–406.

- 2 (a) M. KISO, H. ISHIDA, AND A. HASEGAWA, *Agric. Biol. Chem.*, 48 (1984) 251–252; (b) M. KISO, S. TANAKA, M. TANAHASHI, Y. FUJISHIMA, Y. OGAWA, AND A. HASEGAWA, *Carbohydr. Res.*, 148 (1986) 221–234.
- 3 (a) M. KISO, H. ISHIDA, T. KITO, S. TANAKA, AND A. HASEGAWA, *Gifu Daigaku Nogakubu Kenkyu Hokoku*, 49 (1984) 171–181; (b) M. KISO, Y. OGAWA, S. TANAKA, H. ISHIDA, AND A. HASEGAWA, *J. Carbohydr. Chem.*, 5 (1986) 621–630; (c) M. KISO, S. TANAKA, M. FUJITA, Y. FUJISHIMA, Y. OGAWA, H. ISHIDA, AND A. HASEGAWA, *Carbohydr. Res.*, 162 (1987) 127–140.
- 4 (a) M. MATSUURA, Y. KOJIMA, J. Y. HOMMA, Y. KUBOTA, A. YAMAMOTO, M. KISO, AND A. HASEGAWA, *FEBS Lett.*, 167 (1984) 226–230; (b) Y. KUMAZAWA, M. MATSUURA, J. Y. HOMMA, Y. NAKATSURU, M. KISO, AND A. HASEGAWA, *Eur. J. Immunol.*, 15 (1985) 199–201; (c) M. MATSUURA, A. YAMAMOTO, Y. KOJIMA, J. Y. HOMMA, M. KISO, AND A. HASEGAWA, *J. Biochem. (Tokyo)*, 98 (1985) 1229–1237; (d) T. S. TAKI, M. NAKANO, M. KISO, AND A. HASEGAWA, *Microbiol. Immunol.*, 29 (1985) 1111–1120; (e) M. MATSUURA, Y. KOJIMA, J. Y. HOMMA, Y. KUMAZAWA, A. YAMAMOTO, M. KISO, AND A. HASEGAWA, *J. Biochem. (Tokyo)*, 99 (1986) 1377–1384; (f) Y. KUMAZAWA, M. MATSUURA, Y. MARUYAMA, J. Y. HOMMA, M. KISO, AND A. HASEGAWA, *Eur. J. Immunol.*, 16 (1986) 1099–1103.
- 5 Y. KUMAZAWA, M. NAKATSUKA, A. YAMAMOTO, S. IKEDA, H. TAKIMOTO, C. NISHIMURA, J. Y. HOMMA, M. KISO, AND A. HASEGAWA *Proc. Jpn. Soc. Immunol.*, 19 (1986) 580.
- 6 (a) H.-W. WOLLENWEBER, U. SEYDEL, B. LINDNER, O. LÜDERITZ, AND E. T. RIETSCHEL, *Eur. J. Biochem.*, 145 (1984) 265–272; (b) E. T. RIETSCHEL, H.-W. WOLLENWEBER, R. RUSSA, H. BRADE, AND U. ZÄHRINGER, *Rev. Infect. Dis.*, 6 (1984) 432–438.
- 7 (a) E. T. RIETSCHEL, Z. SIDORCZYK, U. ZÄHRINGER, H.-W. WOLLENWEBER, AND O. LUDERITZ, in L. ANDERSON AND F. M. UNGER (Eds.), *Bacterial Lipopolysaccharides: Structure, Synthesis, and Biological Activities*, *ACS Symp. Ser.*, 231 (1983) 195–218; (b) N. QURESHI, P. MASCAGNI, E. RIBI, AND K. TAKAYAMA, *J. Biol. Chem.*, 260 (1985) 5271–5278.
- 8 Z. SIDORCZYK, U. ZÄHRINGER, AND E. T. RIETSCHEL, *Eur. J. Biochem.*, 137 (1983) 15–22.
- 9 M. IMOTO, S. KUSUMOTO, T. SHIBA, E. T. RIETSCHEL, C. GALANOS, AND O. LUDERITZ, *Tetrahedron Lett.*, (1985) 907–908.
- 10 K. TAKAYAMA, N. QURESHI, P. MASCAGNI, L. ANDERSON, AND C. R. H. RAETZ, *J. Biol. Chem.*, 258 (1983) 14,245–14,252.
- 11 J. C. DITTMER AND R. L. LESTER, *J. Lipid Res.*, 5 (1964) 126–127.
- 12 R. ALBERT, K. DAX, R. PLESCHKO, AND A. E. STÜTZ, *Carbohydr. Res.*, 137 (1985) 282–290.