THE USE OF 2,6-ANHYDRO-2-THIO SUGAR FOR A HIGHLY STEREOCONTROLLED GLYCOSYLATION: A NOVEL STRATEGY FOR SYNTHESIS OF 2,6-DIDEOXY-α-GLYCOSIDES

Kazunobu Toshima, Satsuki Mukaiyama, Takashi Ishiyama, and Kuniaki Tatsuta* Department of Applied Chemistry, Keio University 3-14-1 Hiyoshi, Kohoku-ku, Yokohama 223, JAPAN

<u>Summary</u>: A novel and efficient synthesis of 2,6-dideoxy- α -glycosides was developed by use of phenyl 2,6-anhydro-1,2-dithio-D-altropyranosides as glycosyl donors in a highly stereocontrolled glycosylation.

Highly stereocontrolled synthesis of 2,6-dideoxy glycosides is of considerable interest from points of view of carbohydrate chemistry and natural products synthesis. Many biologically important organic compounds contain 2,6-dideoxy sugars¹) as significant functions in appearance of biological activity and, recently, a novel 2,6-dideoxy sugar was found in new attractive antitumor antibiotics, esperamicin²) and calichemicin³). In this article, we would like to report the novel synthesis of 2,6-dideoxy- α -glycosides having 1,3-diaxial groups by use of phenyl 2,6-anhydro-1,2-dithio-glycosides (1 and 2) as glycosyl donors to illustrate a highly stereocontrolled glycosylation⁴).

We designed the activated glycosyl donors 1 and 2 which had a very rigid structure of the 2,6-anhydro-2-thio bridge and could be good precursors of 2,6-dideoxy glycosides. Phenyl thioglycosides $1^{5, 6}$ and $2^{5, 6}$ were synthesized from $3^{7, 8}$, which was readily prepared from methyl α -D-glucopyranoside, in good overall yields as shown in Scheme I. The anomeric mixture of 1 and the β -anomer of 2^{9} were used for the following glycosylations according to Nicolaou's glycosylation method^{5, 10}).



Scheme I. Preparation of phenyl 2,6-anhydro-1,2-dithio-glycosides 1 and 2. (a) Ac₂O, cat. 4-DMAP, Py, 26°C, 15min; (b) 2.2 equiv BnBr, 2.6 equiv NaH, DMF, 26°C, 75min; (c) 5 equiv Me₃SiSPh, 1.2 equiv TMSOTf, CH₂Cl₂, O°C, 30min. ⁵⁾

First, we examined the glycosylation of 1 by using cyclohexylmethanol (6) as the glycosyl acceptor in several solvents. The results (entries 1~6) in Table 1 showed some excellent feature of

the present glycosylation. These reactions proceeded very rapidly (within 30 min) even at low temperature (-25°C) to give the glycoside 7⁶) in excellent yields (>90%) and the selectivities of the glycosylations were quite α -specific in all cases. Remarkably, the stereoselectivity of the glycosylation was completely independent on both solvent effect and the stereochemistry of anomeric center of phenyl thioglycoside 1. Further, we examined the glycosylation of 1 with several alcohols. The results (entries 7~10) in Table 1 showed some additional efficiencies of our method. Even hindered 2,4-dimethyl-3-pentanol (8) and *t*-butanol (9) were smoothly glycosylated with high stereocontrol in excellent yields. Our next attention was turned to the effect of protecting groups of 1 in selectivity of glycosylation. Results of the glycosylation of 2 having the bulky protecting groups in CH₂Cl₂ at two different temperatures are summarized as entries 11~14. Only in the glycosylation of a hindered secondary alcohol 8 at higher temperature, surprisingly, the selectivity was dramatically changed and the β -glycoside was obtained in excellent yield (entry 13), although the reaction at lower temperature gave exclusively the α -glycoside (entry 14).



To accomplish the synthesis of 2,6-dideoxy glycosides, the 2,6-anhydro-2-thio system in glycosides was converted into the desired 2,6-dideoxy system which was often found in natural products. For example, di-O-benzyl glycoside $15\alpha^{60}$ was desulfurized with de-O-benzylation by hydrogenolysis in the presence of Raney-Ni (W4) at 40°C to afford the 2,6-dideoxy- α -glycoside 16^{60} in 86% yield. On the other hand, di-O-acetyl glycoside 12^{60} was deacetylated to 17^{60} , followed by desulfurization to give 16 in 76% overall yield (Sheme II).



Scheme II. Synthesis of 2,6-dideoxy-α-glycoside. (a) H₂, Raney-Ni (W4), EtOH-dioxane (3:1), 40°C, 0.5h; (b) NaOMe, MeOH, 26°C, 0.5h.

In conclusion, we developed a novel strategy for the highly stereocontrolled synthesis of 2, 6-dideoxy- α -glycosides by use of 2,6-anhydro-2-thio sugar as an efficient glycosyl donor.

	s	ξ	R'OH	- K	1s	Ŕ	Is In o	R'
R	RO	SPh	NBS, MS 4A	RÓ		RÓ R' R		
1: R=Ac 2: R=Bn					α		β	
Entry	Sugar	Alcohol	Solvent	Temp.	Time	Product ⁶⁾	Yield (%) ^{b)}	α/β ^{c)}
1	1	6	CH ₂ Cl ₂	-25°C	15min	7	94	α
2	1	6	(CH ₂ Cl) ₂	-25°C	15min	7	92	α
3	1	6	Et ₂ O	-25°C	30min	7	96	α
4	1	6	THF	-25°C	15min	7	98	α
5	1	6	CH₃CN	-25°C	15min	7	97	α
6	1	6	PhMe	-25°C	30min	7	92	α
7	1	C ₈ H ₁₇ OH	CH₃CN	-40°C	15min	10	91	α
8	1	<i>c</i> -C ₆ H ₁₁ OH	CH₃CN	-40°C	10min	11	92	α
9	1	8	CH ₃ CN	-40°C	10min	12	96	97/3
10	1	9	CH ₃ CN	-40°C	10min	13	92	83/17
11	2	6	CH ₂ Cl ₂	-10ºC	10min	14	90	α
12	2	6	CH ₂ Cl ₂	-40°C	10min	14	99	α
13	2	8	CH ₂ Cl ₂	-10°C	10min	15 β	90	β
14	2	8	CH ₂ Cl ₂	-40°C	10min	15α	92	α

Table 1. Glycosylations of 1 or 2 with several alcohois.^{a)}

.

a) All reactions were carried out by use of 2.0 equiv. of alcohol and 1.1 equiv. of NBS to the glycosyl donor.

b) Isolated yields after purification by column chromatography.

c) a: \$ Ratios were determined by ¹H-NMR spectroscopy¹¹ and /or isolation of pure isomers.

Mechanistic studies of this reaction and application of this method to the synthesis of other types of 2, 6-dideoxy glycosides and the relating natural products are now in progress.

<u>Acknowledgement:</u> We are grateful to the Institute of Microbial Chemistry for the generous support of our program. Financial support by the Ministry of Education, Science and Culture (Grant-in-Aid Scientific Research) is gratefully acknowledged.

References and Notes:

- 1) B. W. Bycraft, "Dictionary of Antibiotics and Related Substances", Chapman and Hall Ltd., 1988.
- 2) J. Golik, G. Dubay, G. Groenewold, H. Kawaguchi, M. Konishi, B. Krishnan, H. Ohkuma, and K. Saitoh, J. Am. Chem. Soc., 109, 3462 (1987).
- 3) M. Lee, T. Dunne, M. Siegel, C. Chang, G. Morton, and D. Border, J. Am. Chem. Soc., 109, 3464 and 3466 (1987).
- 4) For recent reviews of glycosylation, see: (a) K. Krohn, Nachr. Chem. Tech. Lab., 35, 930 (1987). (b) R. R. Schmidt, Angew. Chem. Int. Ed. Engl., 25, 212 (1986). (c) H. Paulsen, Angew. Chem. Int. Eng., 21, 155 (1982). (d) S. Hanessian, D. M. Dixit, and T. J. Liak, Pure Appl. Chem., 53, 129 (1981). (e) H. Tsutsumi and Y. Ishida, J. Syn. Org. Chem. Jpn., 38, 473 (1980). (f) A. F. Bochkov and G. E. Zaikov, "Chemistry of the O-glycosidic Bond: Formation and Cleavage", Pergamon Press: Oxford, 1979. (g) P. Sinaÿ, Pure Appl. Chem, 50, 1437 (1978). (h) K. Igarashi, Adv. Carbohydr. Chem. Biochem., 34, 243 (1977). (i) S. Hanessian and J. Banoub, Adv. Chem. Ser., 39, 36 (1976). (j) G. Wulff and G. Rohle, Angew. Chem., 86, 173 (1974).
- 5) K. C. Nicolaou, S. P. Seitz, and D. P. Papahatjis, J. Am. Chem. Soc., 105, 2430 (1983).
- 6) All new compounds were purified by silica-gel column chromatography and were fully characterized by spectroscopic means. Significant 1 H-NMR(270MHz, CDCl₃) spectra [δ (TMS), J(Hz)] are the following. 1: 2.09 (4/23H, s, OAc(β)), 2.11 (4/23H, s, OAc(β)), 2.17 $(19/23H, s, OAc(\alpha))$, 2.20 $(19/23H, s, OAc(\alpha))$, 2.80 $(4/23H, dd, J=12.0 \text{ and } 3.0, H-6(\beta))$, 2.83 $(19/23H, dd, J=12.0 and 2.8, H-6(\alpha)), 3.25 (19/23H, dd, J=12.0 and 3.0, H-6'(\alpha)), 3.25-3.3$ (4/23H, m, H-2(β)), 3.34 (19/23H, d, J=3.7, H-2(α)), 3.68 (4/23H, dd, J=12.0 and 2.6, H-6'(β)), 4.33 (4/23H, dd, J=3.0 and 2.6, H-5(β)), 4.41 (19/23H, ddd, J=3.0, 2.8 and 0.4, H-5(α)), 5.09 (19/23H, dd, J=8.2 and 0.4, H-4(α)), 5.14 (4/23H, d, J=8.2, H-4(β)), 5.52 (19/23H, ddd, J=8.2, 3.7 and 1.0, H-3(a)), 5.58 (4/23H, dd, J=8.2 and 4.0, H-3(B)), 5.70 (19/23H, br s, H-1(a)), 5.82 (4/23H, d, J=3.4, H-1(β)); 2: 2.57 (1H, dd, J=11.8 and 2.8, H-6), 3.17 (1H, dd, J=3.6 and 4.1, H-2), 3.59 (1H, dd, J=11.8 and 2.4, H-6'), 3.84 (1H, d, J=7.9, H-4), 4.21 (1H, dd, J=7.9 and 4.1, H-3), 4.36 (1H, dd, J=2.8 and 2.4, H-5), 4.57 and 4.70 (each 1H, ABq, J=12.0), 4.68 and 4.72 (each 1H, ABq, J=12.2), 6.04 (1H, d, J=3.6, H-1), 7.2-7.6 (15H, m); 7: 0.9-1.1 (2H, m), 1.1-1.35 (3H, m), 1.5-1.9 (6H, m), 2.03 (3H, s, OAc), 2.12 (3H, s, OAc), 2.70 (1H, dd, J=12.0 and 2.0, H-6), 3.06 (1H, dd, J=2.6 and 1.2, H-2), 3.08 (1H, dd, J=12.0 and 4.0, H-6'), 3.21 (1H, dd, J=8.6 and 6.0), 3.63 (1H, dd, J=8.6 and 6.0), 4.38 (1H, ddd, J=4.0, 2.0 and 1.6, H-5), 5.02 (1H, dd, J=8.2 and 1.6, H-4), 5.21 (1H, dd, J=1.2 and 1.2, H-1), 5.30 (1H, ddd, J=8.2, 2.6 and 1.2, H-3); 16: 1.31 (1H, d, J=6.2, H-6), 2.48 (1H, br d, J=10.4, OH), 3.62 (1H, d, J=10.2, OH), 4.92 (1H, dd, J=3.6 and 1.4, H-1).
- 7) J. Kocourek, Carbohydr. Res., 3, 502 (1967).
- 8) A. B. Foster, J. M. Duxbury, T. D. Inch, and J. M. Webber, Chem. Commun., 1967, 881.
- 9) Only β -isomer was obtained.
- 10) In original paper, the reaction by using NBS as an activator in the presence of MS 4A was carried out in CH₂Cl₂ or CH₃CN at 25°C and selectivity of the glycosylation was dependent on solvent with low stereocontrol. See ref. 5.
- 11) ¹H-NMR spectroscopies of all α -isomers showed W-coupling between H-1 and H-3.