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Note

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Synthesis and structural analyses of 3-acetamido-1,4-di-*O*-acetyl-2,3,5-trideoxy-5-*C*- (isopropylphosphinyl)-D-*erythro*-pentopyranoses

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Abstract

¹H NMR spectroscopy of phosphorus containing hetero sugars (phospha sugars), revealed the α and β configurations and chair conformations for 3-acetamido-1,4-di-*O*-acetyl-2,3,5-trideoxy-5-*C*-(isopropylphosphinyl)- α - and β -D-*erythro*-pentopyranoses. The conformation of the title compounds was determined by ¹H NMR as ¹C₄ in CDCl₃ and the conformation was in accord with that in solid state determined by X-ray crystallographic analysis. © 2002 Elsevier Science Ltd. All rights reserved.

Keywords: Phospha sugar; ¹C₄ conformation; NMR; X-ray

In order to prepare potentially bioactive hetero sugars, the present phospha sugar synthesis was performed and the conformations of 3-acetamido-1,4-di-O-acetyl-2,3,5-trideoxy-phospha sugar derivatives elucidated. Yamamoto et al. reported the syntheses of phospha sugars¹ and their NMR analysis which established the configuration at the phosphorus in the pyranoid rings and the conformation of the six membered heterocycles. The X-ray analyses of phospha sugar analogs had been reported by Luger et al. and ourselves.^{2,3} In the present paper the phospha sugar 16 was synthesized from D-xylose as a starting material via the process shown in Scheme 1. Methyl 3,5-O-isopropylidene-D-xylofuranoside (1) was prepared according to the reported procedure,⁴ 2-O-methylthiocarbonylation of methylcompound (1) with an excess amount of carbon disulfide and methyl iodide in DMSO under alkaline solution was carried out to afford 2α and 2β , easily separated by column chromatography on silica gel. Treatment of 2β with acidic methanol gave compound

3. 5-O-Triphenylmethylation of 3 with 1 equiv. of triphenylmethyl chloride in pyridine afforded 4. 3-O-Mesylation of 4 with methanesulfonyl chloride in pyridine afforded 5 in good yield. 2-O-Deoxygenation of compound 5 in accordance with the reported procedure^{5,6} gave compound 6 in quantitative yield. Conversion of 6 into 8 was readily achieved by heating with sodium azide and successive treatment with acidic methanol. Compounds 8α and 8β were separated by column chromatography on silica gel. 5-O-Tosylation of the separated 8β with 1 equiv. of tosyl chloride in pyridine afforded 9 in quantitative yield. 3-Acetoamido-2-deoxy derivative 11 was prepared by reduction of 9 with n-Bu₃SnH, followed by acetylation with acetic anhydride. On heating compound 11 with sodium iodide in acetone in a sealed tube methyl 3-acetoamido-2,3,5-trideoxy-5-iodo-β-D-erythro-pentofuranoside 12 was obtained in quantitative yield. Conversion of 12 into the phosphinate 13 was readily performed by heating with an excess amount of diethyl isopropylphosphonite at 150 °C. Treatment of compound 13 with sodium dihydrobis(2-methoxyethoxy)aluminate (SDMA) gave compound 14 which was subsequently followed by ring-enlargement

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caused by the opening of the furanoid ring (15) and the successive closing to prepare a pyranoid ring under acidic conditions. The known two step procedure^{3,7} gave trideoxy-ribopyranose type phospha sugars 16a–d. Diastereomers 16a, 16b, 16c, and 16d were separated, respectively, by using flash chromatography on silica gel (eluent; ethyl acetate/methanol = 4/1, v/v) from the mixture of products. The overall chemical yields of diastereomers 16a, 16b, 16c, and 16d were 7, 5, 5 and 11%, respectively, from compound 13.

The precise structures of these compounds **16a**, **16b**, **16c**, and **16d** were determined on the basis of the 500 MHz ¹H NMR spectra and by X-ray crystallography (Fig. 1). The assignments of all signals were readily made by employing first-order analysis with the aid of a decoupling technique. The analyzed results are summarized in Table 1. The conformations of these phospha sugar derivatives (in CDCl₃ solution) was derived by the careful analysis of the magnitudes of $J_{2R,P}$ (20.5 Hz for 16a vs. 2.0–9.8 Hz for **16b–d**), $J_{2S,P}$ (7.3 Hz for

16a vs. 20.5–21.7 Hz for **16b–d**), and $J_{4,P}$ (9.2 Hz for 16a vs. 21.4-22.0 Hz for 16b-d) with respect to the corresponding vicinal dihedral angles. As a result, compounds 16a and 16b-d were shown to exist predominantly in the ${}^{4}C_{1}$ and ${}^{1}C_{4}$ conformations, respectively. The orientation of the P=O group could be established by the δ values of H-4 for **16a**, and HR-2 and HS-2 for **16b**-d. An appreciable downfield shift (0.05–0.31 ppm) is observed for 16a compared with 16b-d, the observation shows the axial orientation of the ring P=O group for compound 16a. The equatorial P=O orientation was assigned to the result of upfield shift for products **16a**–**d**. The small magnitudes of $J_{5R,P}$ support the anti orientation of HR-5-C5-P=O for 16c and 16d. The equatorial P=O group orientation was assigned to the rest of the compounds, products 16c and 16d. The appreciable downfield shift observed for HR-2 of 16c and 16d could be explained by its axial orientation. The anomeric orientation at C-1 was derived from the magnitudes of $J_{1,2}$ and $J_{1,P}$. The moderate magnitudes of



Scheme 1. Reagents: (i) CS_2/MeI ; (ii) $MeOH/H_2SO_4$; (iii) TrCl/py; (iv) MsCl/py; (v) n- Bu_3SnH ; (vi) NaN_3 ; (viii) TsCl/py; (ix) n- Bu_3SnH ; (x) Ac_2O/py ; (xi) NaI; (xii) $P(OEt)_2i$ -Pr; (xiii) SDMA; (xiv) HCl; (xv) Ac_2O/py .





Chemica	l shift (ð	i), ppm																					
Compd	H-1	HR-2	HS-2	Н-3 а	H-4	HR-5	HS-5	Ac0-1,3,4 ^a	NH-3	HC-P	CH ₃ -C-P	³¹ P	1										
16a	2.45	2.17	2.44	4.64	5.38	2.20	2.15	2.16, 2.01, 1.97	6.03	1.94	1.23, 1.23	46.9											
16b	5.50	2.09	2.25	4.51	5.33	2.30	2.44	2.19, 2.10, 1.96	5.86	2.01	1.28, 1.21	39.2											
16c	5.28	2.57	2.07	4.30	5.07	1.77	2.60	2.15, 2.15, 1.98	5.99	1.95	1.20, 1.18	40.2											
16d	5.43	2.63	2.13	4.43	5.16	1.84	2.72	2.16, 2.16, 1.99	5.70	1.80	1.19, 1.13	41.0											
Couplin	g constar	it (Hz)																					
	$J_{1,2\mathrm{R}}$	$J_{1,2\mathrm{S}}$	$J_{\rm 1,P}$	$J_{1,5\rm R}$	$J_{1,5S}$	$J_{2R,2S}$	$J_{2R,3}$	$J_{2R,P}$	$J_{2S,3}$	$J_{2{ m S},{ m P}}$	$J_{2\mathrm{S},4}$	$J_{3,4}$	$J_{3,\mathrm{NH}}$	$J_{3,5R}$	J _{4,5R} J	4,5S	I _{4,P} J	5R,5S J	SR,P	5S,P ²	${}^{2}J_{\mathrm{H,P}}{}^{\mathrm{b}}$	${}^{3}J_{\rm H,H}{}^{\rm b}$	${}^{3}J_{\mathrm{H,P}}{}^{\mathrm{b}}$
16a	6.1	3.4	9.2	1.2	0	15.3	5.0	20.5	3.7	7.3	0	3.4	8.9	1.0	3.7 1	1.0	9.2 1	4.7 1	7.4	8.2	5.2	7.3	16.8
16b	2.8	6.1	5.0	0	1.8	14.7	11.0	6.7	3.1	21.7	1.2	3.1	9.1	0	3.4	6.1	21.7 1	6.2 1	8.9	9.5	3.1	7.3	16.2
16c	11.6	5.2	5.2	0	0	12.8	11.6	9.8	3.4	20.5	1.1	2.4	9.0	0	3.2	5.2	21.4 1	5.9	4.9 1	5.8]	10.7	7.3	16.2
16d	2.2	4.8	9.2	0	2.4	14.4	12.5	2.0	3.4	21.3	1.2	2.4	8.9	0	3.1	5.2	22.0 1	5.6	4.9 1	4.6	7.0	7.3	16.2
^a The ^b Couj Hz and	assignme pling con ${}^{3}J_{H,P} = 1$	ent of activity of	etyl grou risoprop or 16c an	ps may $\frac{1}{\text{yl group}}$ nd $^2 J_{\text{H,P}}$	have to on phose $= 7.0 \text{ H}$	be interc sphorus <i>i</i> z, ³ J _{H,H}	hanged. ttom are = 7.3 Hz	${}^{2}J_{\rm H,P} = 5.2 \text{ Hz}, {}^{3}J_{\rm H,F}$ and ${}^{3}J_{\rm H,P} = 16.2 \text{ Hz}$	r = 7.3 Hz for 16d ,	: and ³ J _H respective	_{1,P} = 16.8 Hz fr ely.	or 16a , ² ,	$J_{\rm H,P} = 3.$	1 Hz, ³ J	, _{Н,Н} = 7.	3 Hz ar	d ³ J _{H,P}	= 16.2 H	[z for 16	b, ² J _{H,I}	. = 10.7	Hz, ³ J _H	. _H = 7.3

Table 1 $^1{\rm H}$ NMR (500 MHz) parameters and $^{31}{\rm P}$ chemical shifts for 16a, 16b, 16c, and 16d (Scheme 2)



Scheme 2.

Table 2 ¹³C NMR (67.80 MHz) chemical shifts for **16a**, **16b**, **16c**, and **16d**

Chemical	shift δ ppm,	$(J_{\rm P-C}, {\rm Hz})$							
Compd.	C-1	C-2	C-3	C-4	C-5	$P-CH(CH_3)_2$	$P-CH(CH_3)_2$	<i>C=</i> 0	<i>C</i> H ₃ –C=O
16a	65.81 (73.1)	46.32 (2.4)	29.62	68.61 (0.0)	24.87 (68.3)	25.10 (54.8)	14.57 (4.2)	167.92, 169.27, 169.60	20.86 20.96 23.45
16b	64.33 (70.8)	45.44 (6.1)	30.32	69.99 (4.8)	26.45 (71.1)	25.63 (56.3)	14.77 (7.3)	169.99, 169.78, 169.52	23.11 20.95 20.85
16c	65.07 (70.7)	47.78 (0.0)	29.27	69.32 (6.1)	27.18 (67.1)	24.77 (57.3)	15.15 (0.0)	169.24, 169.61, 170.71	20.79 21.15 23.17
16d	65.09 (75.7)	44.86 (6.1)	28.56	69.78 (7.3)	24.57 (70.8)	25.36 (58.6)	14.32 (0.0)	170.78, 169.25, 168.98	23.29 21.28 20.99

 $J_{1,P}$ of 16a and 16d suggest the syn connection of H–C-1–P=O for these compounds. The large magnitude of $J_{1,2R}$ (11.6 Hz) indicated axial H-1 orientation for 16c, whereas the smaller $J_{1,2R}$ values (2.2–2.8 Hz) suggested the equatorial H-1 orientation for 16b and 16d. Small coupling constants $J_{1.5S}$ and $J_{2S,4} = 1.1 - 2.4$ Hz were shown in the compounds 16b-d. Obviously compounds 16b-d retain the conformation of ${}^{1}C_{4}$ from the above conformation analyses, and the long range couplings $(J_{1.58} = 1.8 \text{ Hz for } 16b, J_{1.58} = 2.4 \text{ Hz for } 16d,$ and $J_{28,4} = 1.1 - 1.2$ Hz for **16b**-d) of compounds **16a**-d support. As for the more precise conformation of 16b**d**, it is noted that the magnitudes of $J_{2R,P}$ and $J_{2S,P}$ tend to become appreciably closer to intermediate values, which suggest an equilibrium mixture of ${}^{4}C_{1}$ and ${}^{1}C_{4}$ conformers at room temperature (ca. 20% of the ${}^{4}C_{1}$ conformation was contained in the compounds 16b-d from the observed coupling constant values). A part of the ¹H NMR spectra of compounds 16a-d were assigned with difficulty, because the methylene protons on C-2 and C-5 overlapped with the acetyl group, and it was reexamined more precisely by using a simulation

program,⁸ giving a good fit with the measured spectrum. The chemical shift values of ¹³C NMR are shown in Table 2.

Rod-shaped crystals of 16c and 16d were grown from ethyl acetate-hexane. Precise lattice constants and three dimensional intensity data were obtained by a RIGAKU AFC7R four-circle diffractometer with Nifiltered CuKa radiation. Phase determination was made by a direct method (SIR92).⁹ the molecular structures for compounds 16c and 16d are shown in Fig. 2. A water molecule is present in crystal 16c ($16c/H_2O = 1/1$) but is not shown in Fig. 2. A summary of crystal data for compounds 16c and 16d is shown in Table 3. In 16c, the substituent groups at C-1, C-3, and P are orientated equatorially, while those are at C-4 and P=O are axial. However, in molecular structure of 16d, the substituents at C-3 and P are equatorial, while those at C-1, C-4, and P=O are axial. The acetoxy groups on C-1, C-3, and C-4 have usual syn arrangement between the C=O bond and the adjacent C-H bond. The Cremer-Pople puckering parameters¹⁰ are Q = 0.600 Å, $\theta = 168.5^{\circ}$, $\psi = 252.5^{\circ}$ for **16c** and Q = 0.5602 Å, $\theta = 167.4^{\circ}$, $\psi =$

262.2° for **16d**, respectively, and the six membered rings of the compounds **16c** and **16d** were distorted slightly from the ${}^{1}C_{4}$ conformation. The X-ray analysis of crystals **16c** and **16d**, the structures were first solved at 223 K, but they could not be resolved at 295 K because carbon and phosphorus atoms of six membered ring are disordered at 295 K. In the crystals **16c** and **16d** may exist in the same way of equilibrium as the obtained results by ¹H NMR, where the compounds exist in an equilibrium mixture of ${}^{1}C_{4}$ and ${}^{4}C_{1}$ in CDCl₃ solution. This is the first observed ${}^{1}C_{4}$ conformation of phospha sugar by X-ray crystal structure analysis.

1. Experimental

1.1. General procedures

Synthetic procedures for compound **5** from **1** as starting material were in with a previous paper.¹ ¹H NMR spectra were measured in CDCl₃ (TMS as the internal standard) on a Brucker 500 (500 MHz) and ¹³C NMR were measured in CDCl₃ (CDCl₃ as the internal standard) on a JASCO EX270 (67.80 MHz). Melting points were measured with a micro melting point apparatus (Yanagimoto Co., Ltd., Japan) and are uncorrected. Column chromatography was performed by Merck Lobar silica gel. The reaction were monitored by TLC on Kieselgel 60 F254 (Merck) with detection by H₂SO₄. Optical rotations were determined with a digital polar-

imeter DIP-4 (JASCO Ltd.). Mass spectra were recorded on Shimadzu GCMS-AP5050 gas-chromatograph-mass spectrometer using electron impact 70 eV. X-Ray single crystallographic measurements were conducted on a RIGAKU AFC7R using cut crystals of size ($0.2 \times 0.2 \times 0.6$ mm). Phase determination was made by a direct method (SIR 92) and expanded using Fourier techniques. After convergence of all parameters, final *R* values were 5.6% for 16c and 6.4% for 16d, respectively.

1.2. Methyl 2-deoxy-3-*O*-methanesulfonyl-5-*O*triphenylmethyl-β-D-*threo*-pentofuranoside (6)

To a dried toluene (80 mL) solution of compound 5 (4.17 g) were added *n*-Bu₃SnH (2.83 g) and a catalytic amount of AIBN. The reaction mixture was heated for 2 h under a nitrogen atmosphere. Toluene was removed under reduced pressure. The residue was dissolved in acetonitrile, and washed several times with n-hexane to remove organic tin compound. The hexane layer was further extracted with acetonitrile. The acetonitrile extract was collected and concentrated under reduced pressure. The crude product was purified by column chromatography on silica gel to afford syrupy product 6 in quantitative yield. ¹H NMR (CDCl₃): δ , 2.3–2.4 (m, 2H, H-2 and 2'), 2.78 (s, 3H, OSO₂CH₃), 3.2-3.6 (m, 2H, H-5 and 5'), 3.40 (s, 3H, OCH₃), 4.2-4.3 (m, 1H, H-4), 5.08 (dd, J = 4.3 and 2.5 Hz, 1H, H-1), 5.2-5.3 (m, 1H, H-3), 7.2-7.5 (m, 15H, aroma).



Fig. 2. Molecular structure for compounds 16c (left) and 16d (right).

Table 3 Crystal and structure refinements for **16c** and **16d**

	16c	16d
Molecular formula	$C_{14}H_{24}NPO_6 \cdot H_2O$	$C_{14}H_{24}NPO_6$
Molecular weight	351.34	333.32
Temperature (K)	223	223
Crystal system	orthorhombic	monoclinic
Space group	$P2_12_12_1 (\# 19)$	P2 ₁ (#4)
Unit cell dimensions (Å)	••• • • •	• • •
a	14.877(3)	9.7660(8)
b	23.416(3)	8.588(1)
С	5.253(2)	10.1716(5)
β (°)	90.296(5)	
Volume (Å ³)	1830.1(6)	853.1(1)
Z (molecules/cell)	4	2
D_{calcd} (g cm ⁻³)	1.275	1.235
Absorption coefficient (mm^{-1})		
F(000)	712.00	340.00
Crystal size (mm)	$0.20 \times 0.20 \times 0.30$	$0.2 \times 0.2 \times 0.6$
Reflections collected	1359	1457
Independent reflections	1346 ($R_{\rm int} = 0.302$)	1633 ($R_{\rm int} = 0.059$)
Refinement method	Full-matrix least-squares on F^2	Full-matrix least-squares on F^2
Goodness-of-fit indicator	3.44	2.77
Final R indices $[I > 2\sigma(I)]$		
R_1	0.056	0.064
wR	0.059	0.180

1.3. Methyl 3-azido-2,3-dideoxy-5-O-(triphenylmethyl)- α - and -D-*erythro*-pentofuranoside (7)

To a solution of compound 6 (3.39 g) in DMF (40 mL)was added sodium azide (7.5 g), and the mixture heated for 2 h at 110 °C. The reaction mixture was filtered, and the filtrate was concentrated under reduced pressure. The syrupy residue was dissolved in chloroform and washed with water several times. The water layer was further extracted with chloroform several times. The organic layer were collected, dried with anhydrous sodium sulfate, and evaporated. The residue was purified by column chromatography on silica gel (eluent; ethyl acetate/hexane = 1/2, v/v) to obtain syrupy product 7 (2.74 g) in 93% yield. ¹H NMR (CDCl₃): δ , 1.9-2.2 (m, 2H, H-2 and 2'), 2.86 and 2.91 (2s, 2H, H-5 and 5'), 3.28 (s, 3H, OCH₃), 3.9-4.3 (m, 2H, H-3 and 4), 5.03 (dd, J = 4.8 and 2.0 Hz, 1H, H-1), 7.2–7.6 (m, 15H, aroma).

1.4. Methyl 3-azido-2,3-dideoxy- α - and β -D-*erythro*-pentofuranoside (8)

To a solution of 7 (2.0 g) in dried methanol (10 mL) was added HCl in methanol solution (0.5 wt%, 0.4 mL), and the mixture was stirred for 1 h at 40-50 °C. The reaction mixture was neutralized with lead carbonate and left overnight in a refrigerator. The precipitate was

filtered and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (eluent; ethyl acetate/hexan = 1/2, v/v) to obtain syrupy product **8** α (0.44 g) in 51% yield and **8** β (0.26 g) in 30% yield. ¹H NMR (CDCl₃) for **8** α : δ , 1.90–2.60 (m, 3H, H-2, 2' and OH), 3.38 (s, 3H, OCH₃), 3.5–4.0 (m, 4H, H-3, 4, 5 and 5'), 5.07 (d, J = 5.3 Hz, 1H, H-1). ¹H NMR (CDCl₃) for **8** β : δ , 2.0–2.5 (m, 2H, H-2 and 2'), 2.9–3.1 (br s, 1H, OH), 3.38 (s, 3H, OCH₃), 3.6–3.8 (br m, 2H, H-5 and 5'), 4.0–4.3 (m, 2H, H-3 and 4), 5.09 (dd, J = 2.1 and 5.0 Hz, 1H, H-1). Anal. Calcd for C₆H₁₁N₃O₃: C, 41.61; H, 6.40; N, 24.27. Found: C, 41.65; H, 6.32; N, 24.25.

1.5. Methyl 3-azido-2,3-dideoxy-5-*O-p*-toluenesulfonylβ-D-*erythro*-pentofuranoside (9)

To a solution of **8** β (1.10 g) in pyridine (15 mL) was added *p*-toluenesulfonyl chloride (1.2 g), and the reaction and wash-up were the same as that for compound **8**. Compound **9** was purified by column chromatography on silica gel (ethyl acetate/hexane = 1/2, v/v) (1.85 g, 98%). ¹H NMR (CDCl₃): δ , 1.9–2.4 (m, 2H, H-2,2'), 2.45 (s, 3H, CH3 of tosyl), 3.27 (s, 3H, OCH₃), 3.9–4.2 (m, 4H, H-3,4,5, and 5'), 5.01 (dd, *J* = 1.9 and 4.7 Hz, 1H, H-1), 7.31 and 7.84 (2H each, 2 br d, *J* = 8.3 Hz, C₆H₄–S).

1.6. Methyl 3-acetamido-2,3-dideoxy-5-*O-p*-toluenesulfonyl-β-D-*erythro*-pentofuranoside (11)

To a solution of 9 (1.85 g) in dried toluene (100 mL) was added *n*-Bu₃SnH (2.17 g) under a nitrogen atmosphere at 120 °C for 15 min. Concentration of the reaction mixture afforded compound 10, which was dissolved in pyridine (10 mL). Acetic anhydride (0.75 mL) was added, the reaction mixture stirred for overnight at room temperature, and then the solvents were removed under reduced pressure. The crude products was dissolved in acetonitrile, and excess organo tin was removed by hexane extraction (several times). The syrupy crude product was dissolved in CHCl₃, and the organic layer was washed with water, dried over anhydrous sodium sulfate, and evaporated. The product was purified by column chromatography on silica gel (ethyl acetate/methanol = 10/1, v/v) and gave pure 11 in 89% yield. ¹H NMR (CDCl₃): δ , 1.9–2.3 (m, 2H, H-2,2'), 1.92 (s, 3H, N-Ac), 2.44 (s, 3H, CH₃ of tosyl), 3.16 (s, 3H, OCH₃), 3.9–4.6 (m, 4H, H-3, 4. 5, and 5'), 4.98 (d, J = 3.7 Hz, 1H, H-1), 3.89 (br d, J = 7.5 Hz, 1H, NH), 7.31 and 7.84 (2H each, 2br d, J = 8.3 Hz, C₆H₄–S). Anal. Calcd for C₁₅H₂₁NO₆S: C, 52.46; H, 6.16; N, 4.08. Found: C, 52.44; H, 6.19; N, 4.10.

1.7. Methyl 3-acetamido-2,3,5-trideoxy-5-iodo-β-D-ribofuranoside (12)

A mixture of compound **11** (1.97 g) and sodium iodide (1.8 g) in acetone (30 mL) in a sealed tube was heated for 4 h in a boiling water bath, and precipitate was filtered, the filtrate was concentrated. The residue was dissolved in chloroform and washed with water. The water layer was further extracted with chloroform. The organic layer was collected, dried over anhydrous sodium sulfate, and evaporated to afford crystalline 12 (1.43 g) in 96% yield. ¹H NMR (CDCl₃): δ , 1.9–2.6 (m, 2H, H-2 and 2'), 1.98 (s, 3H, N–Ac), 3.2–3.5 (m, 2H, H-5,5'), 3.39 (s, 3H, OMe), 3.9–4.7 (m, 2H, H-3,4), 5.08 (dd, J = 1.3 and 5.3 Hz, 1H, H-1), 6.10 (br s, 1H, NH).

1.8. Methyl 3-acetamido-2,3,5-trideoxy-5-*C*-(ethoxyiso-propylphosphinyl)-β-D-*erythro*-pentofuranoside (13)

The mixture of **12** (1.09 g) and diethyl isopropylphosphonite (10 mL) was heated at 150 °C for 8 h. Excess amount of phosphonite was remove under reduced pressure, and crude product **13** was purified by column chromatography on silica gel (ethyl acetate/methanol = 10/1, v/v) to afford in quant. yield. ¹H NMR (CDCl₃): δ , 1.0–1.4 (m, 10H, P–CH(CH₃)₂ and P–OCCH₃), 1.6– 2.4 (m, 4H, H-2,2',5, and 5'). 1.97 (s, 3H, N–Ac), 3.33 (s, 3H, OCH₃), 3.8–4.6 (m, 4H, H-3,4,P–OCH₂), 4.97 (br d, J = 4.6 Hz, 1H, H-1), 7.9 (br d, J = 7.1 Hz, 1H, NH). Anal. Calcd for C₁₃H₂₆NO₅P: C, 50.81; H, 8.53; N, 4.56. Found: C, 50.83; H, 8.50; N, 4.55.

1.9. Preparation of phospha sugar derivatives (16)

To a solution 13 (776 mg) in dry THF (25 mL) SDMA (70% in toluene, 1.2 g) was added at 0 °C, and the mixture was stirred for 1 h. A small amount of water containing concd HCl was added. The precipitate was filtered off, the filtrate was evaporated in vacuo. The residue was purified by column chromatography on silica gel (ethylacetate/methanol = 3/1, v/v) to give syrupy compound 14 (305 mg). This syrup was immediately dissolved in water containing a small amount of THF and hydrochloric acid (0.2 mL) was added. The mixture was heated under nitrogen for 3.5 h at 110 °C (bath temp.). The mixture was cooled, diluted with water, and the acid neutralized with Amberlite IR-45A ion-exchenge resin, and filtered. The resin was washed with water and ethanol and filtered. The filtrates were combined, and evaporated in vacuo to give syrupy 15 (223 mg). This syrup was treated with acetic anhydride (1.0 mL) in dry pyridine (15 mL). Pyridine was evaporated in vacuo to give syrupy 16. This syrup was immediately purified by column chromatography on silica gel (ethylacetate/methanol = 2/1, v/v), and separated by column chromatography on silica gel (ethylacetate/methanol = 4/1-2/1, v/v) to give **16a-d**. Chemical vields of 16a, 16b, 16c and 16d were 59 mg (7% from 13), 46 mg (5%), 44 mg (5%), and 90 mg (11%), respectively. and optical rotations were $[\alpha]_{\rm D} - 21.7^{\circ}$ (c 1.15) for **16a**, -34.5° (c 1.45) for **16b**, -28.1° (c 1.78) for 16c, and -66.4° (c 1.28) for 16d in CHCl₃, respectively. Anal. Calcd for C₁₄H₂₄NO₆P (16a): C, 50.45; H, 7.26; N, 4.20. Found: C, 50.40; H, 7.29; N, 4.22. Anal. Calcd for C₁₄H₂₄NO₆P (16b): C, 50.45; H, 7.26; N, 4.20. Found: C, 50.44; H, 7.24; N, 4.25. Anal. Calcd for C₁₄H₂₄NO₆P (16c): C, 50.45; H, 7.26; N, 4.20. Found: C, 50.44; H, 7.26; N, 4.22. Anal. Calcd for C₁₄H₂₄NO₆P (16d): C, 50.45; H, 7.26; N, 4.20. Found: C, 50.48; H, 7.20; N, 4.21.

2. Supplementary material

Full crystallographic details excluding structure features have been deposited with the Cambridge Crystallographic Data Centre (CCDC). These data may be obtained, on request, from The Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (e-mail: deposit@ccdc.cam.ac.uk or www: http://www.ccdc.cam. ac.uk).

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