| 1  | The influences of different substituents on spectral properties of  |
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| 2  | Rhodamine B based chemosensors for mercury ion and application in   |
| 3  | EC109 cells   |
| 4  |   |
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Abstract: Six rhodamine-based "Turn-on" fluorescence chemosenors $(L_{1-6})$  with 14 different substituents for Hg<sup>2+</sup> were readily synthesized and investigated, which 15 displayed high selectivity and chelation enhanced ratiometric fluorescence change and 16 colorimetric change with Hg<sup>2+</sup> among the metal ions examined. Based on UV spectral 17 data and fluorescence spectral data, the effect of different substituents on spectral 18 properties of the probes were presented and discussed. The detection limit of  $Hg^{2+}$  to 19 probe  $L_1$  was as low as 50 nM, which attribute to its electron donating group. 20 Theoretical calculation also support the process of reaction. Confocal laser scanning 21 microscopy experiments showed that probe could be used to detect  $\mathrm{Hg}^{2+}$  in living 22 23 cells.

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Keywords: Rhodamine B; Fluorescent chemosesor ; Sensing mercury ion; Livingcells.

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## 28 1.Introduction

Various transition-metal ions are essential to the life of creatures. Mercury is a 29 dangerous and hazardous toxic which has posed a great threat to our environment<sup>1-4</sup>. 30 Mercury is heavy metal, which can be dispersed in the atmosphere, the ocean, and 31 the soil. Different speciation mercury can penetrate our environment by various ways, 32 such as methyl mercury produced by aquatic microbes which bioaccumulates 33 through the food chain and oxidation of mercury vapor in atmosphere to 34 water-soluble Hg<sup>2+</sup> ions<sup>5-7</sup>. After long-term exposure in the environment, mercury ion 35 accumulate in organisms which can cause nausea, vomiting, abdominal pain, renal 36 dysfunction and other diseases. 37

At present, UV spectrophotometry, electrochemical methods, ionization coupled plasma mass spectrometry, ionization coupled plasma atomic emission spectrometry, atomic absorption spectroscopy and other analytical methods can be used to identify and detect metal ions<sup>8-12</sup>. However, these methods have some disadvantages in the application, such as high detection costs, narrow range of applications, and can't achieve real-time detection. In contrast, the fluorescent molecular probe has no

damage to the sample, and gives its priority to easy operability and high sensitivity, so fluorescence analytical methods are effective and efficient ways to detect ions<sup>13-15</sup>. Compared with other fluorescent dyes, rhodamine dyes have some excellent photophysical properties, such as high fluorescence quantum yield, pH insensitive, high molar absorptivity and high extinction coefficient, particularly in its nucleotide and nucleic acid conjugates<sup>16</sup>. Some rhodamine-based chemosensors for Hg<sup>2+</sup> ions have been reported<sup>17-22</sup>, but dual colorimetric and fluorescent chemosensors for mercury ion were still rare<sup>23</sup> and some of them were not efficient enough to be selective toward mercury ion.

Here, we report six rhodamine-based caprolactam derivatives as a sensor for Hg<sup>2+</sup>, when binding phenomena could be probed through binding-induced changes in an electronic spectral pattern. The difference between these probes are that we selected different substitutional group such as methyl group, nitro group, aldehyde group, halogen group, phenyl group. These rhodamine-based derivatives that behaves as a fluorescent Hg<sup>2+</sup> sensor with high emission sensitivity and selectivity, which exhibited sensitive detection toward Hg<sup>2+</sup> via significant fluorescence enhancement in solution, and in the meantime they showed a significant color change from colorless to rose red. We have designed some similar structures to compounds  $L_{1-6}$  and conduct a series of experiments to explore their properties 62 which have the correlations of nature of substituent group. After comparison of 63 optical properties, probe L<sub>1</sub> showed a good performance in the recognition toward 64 Hg<sup>2+</sup>, which indicated that the different electronic distribution among the senors' 65 structures make an influence on their properties. For this reason, we selected  $L_1$  as a 66 typical example to expound in the following discussion. 67

### 68 2. Experimental

2.1 Apparatus and Materials 69

Fluorescence spectra measurement were performed on Horiba Jobin Yvon Inc. 70 Fluorolog 3-TSCPC. <sup>1</sup>H NMR spectrum was run on a Bruker 300 MHz spectrometer 71 using TMS as the internal standard. Mass spectrum was recorded with a VG ZAB-HS 72 double focusing mass spectrometer. Absorption spectra were measured on a UV-2201 73

double-beam UV/VIS spectrometer. Melting point was determined using an SGW X-4
digital melting point apparatus.

All the materials for synthesis were purchased from Sinopharm Chemical Reagent Co.,Ltd (Shanghai, China) and used without further purification. Metal solutions were prepared from their analytical grade nitrate salts. The metal ions were prepared as 0.2 mM in water solution.

80 2.2 Syntheses

As shown in Scheme 1, the compounds  $L_1 \sim L_6$  were prepared by reacting RBH with aromatic aldehyde. RBH was synthesized according to literature.

83 2.2.1 Synthesis of RBH

As for the synthesis of RBH, sereval different procedures have been reported<sup>24-26</sup>. In this work, RBH was synthesized by a modified method according to Xiang<sup>27</sup>. m/z: 457.2265 ( $[M+H]^+$ ); M<sup>+</sup> calculated 456.2265. IR(KBr,cm<sup>-1</sup>): v = 3428, 2987, 2926, 1689, 1614, 1514, 1380, 1218, 1117, 818, 767. 1H NMR (300 MHz, CDCl<sub>3</sub>,  $\delta$  ppm),  $\delta$ (ppm):1.17 (t, 12H), 3.34 (q, 8H), 3.62 (bs, 2H), 6.29 (dd, 2H), 6.43(d, 2H), 6.45 (d, 2H), 7.11 (dd, 1H), 7.42 (d, 1H), 7.44(d, 1H), 7.93 (dd, 1H). (Fig.S1~S3)

90 2.2.2 Synthesis of  $L_1$ 

RBH (0.5 mmoL, 0.24 g) was dissolved in 30 mL ethanol, and then p-tolualdehyde
(0.5 mmoL, 0.12 g) was slowly added. The mixture was stirred and refluxed for 12 h
at 80 °C. After distillation in vacuum, the residue was recrystallized with methanol
and water to give the final product L1 in yield of 79.8%. m/z: 559.3020([M+H]<sup>+</sup>);
M<sup>+</sup>calculated 558.3020. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, δ ppm): 8.64 (s, 1H),7.97 (m,
1H), 7.49 (d, 2H), 7.45 (m, 2H),7.26 (d, 1H), 7.09 (m, 2H), 6.54 (d, 2H), 6.43 (s, 2H),
6.26 (d, 2H), 3.33 (q, 8H), 2.29 (s, 3H), 1.15(t, 12H). (Fig.S4~S6)

98 2.2.3 Synthesis of  $L_{2\sim6}$ 

Compound L<sub>2~6</sub> was prepared using a general procedure which is essentially similar
to that used for L<sub>1</sub>. (Fig.S7~S21)

### 101 **3. Results and Discussion**

102 The structure of compounds  $L_{1\sim6}$  were characterized by <sup>1</sup>H NMR, HR-MS. The 103 results were in good agreement with the structure showed in Scheme 1. Fluorescence and UV-vis studies were performed using a 20  $\mu$ M solution of L<sub>1~6</sub> in a CH<sub>3</sub>CN/H<sub>2</sub>O

105 (1:1, v/v) solution with appropriate amounts of metal ions.

106 3.1 Comparison of six rhodamine B based senors

Under the same conditions, we explore the sensitivity measurement of ion probes. 107 As shown in Fig.1, rhodamine B derivatives with methyl exhibits excellent sensitivity 108 towards mercury ion. Derivatives with halogen group also show good sensitivity 109 owing to its electron donating groups. Based on these phenomena, it suggested that 110 chelation-enhanced fluorescence appears when compounds with electron donating 111 groups are complexed with Hg<sup>2+</sup>. In order to further explore the affinity capacity 112 between probes and metal ions, association constants were calculated to verify the 113 experiment we have conducted. It is indicated in the Scheme 2, the numerical size of 114 association constants minished in order, which in accordance with the results of 115 fluorescence intensity when different substituents with rhodamine B binds to mercury. 116 After comparison between these probes, we can come to conclude that methyl 117 group, halogen group and phenyl group belong to electron donating group while nitro 118 group, aldehyde group belong to electron-attracting group. When interacting with 119 120 mercury icon, electron donating group provide electron to coordination site which 121 make it easier to combine. On the contrary, electron-attracting group make it more difficult for probe to interact with mercury icon. So, we prefer to select electron 122 withdrawing substituents to detect metal ions which reveals high sensitivity. To 123 investigate photoelectric properties of these probes further, we selected  $L_1$  as an 124 typical example to expound in the discussion. 125

126 3.2 Metal ion selectivity and competition experiments

As shown in Fig.2, UV–vis spectrum of compound  $L_1$  (20µM) exhibited only very weak bands over 450 nm, which could be attributed to the presence of a trace amount of the ring-opened form of compounds. On addition of 10 equiv. Hg<sup>2+</sup> into solution,  $L_1$  immediately resulted in a significant enhancement of absorbance at about 556 nm simultaneously the color changed into rose red. Other metal ions such as Zn<sup>2+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>, Pb<sup>2+</sup>, Ba<sup>2+</sup>, Ni<sup>2+</sup>, Mn<sup>2+</sup>, K<sup>+</sup>, Li<sup>+</sup>, Ag<sup>+</sup>, Co<sup>2+</sup>, except for Fe<sup>3+</sup>, did not show any significant color and spectral change under any significant color and

spectral change under identical conditions. These phenomena suggest that these 134 compounds can serve as a "naked-eye" chemosensor for Hg<sup>2+</sup>. To further investigate 135 the interaction of  $Hg^{2+}$  and the probe, the  $Hg^{2+}$  binding stoichiometry of the probe can 136 be determined from titration and the Job plot<sup>28, 29</sup>. As shown in Fig.3, it is obvious that 137 the fluorescence intensity reached a maximum when the ratio was 0.5, which 138 suggesting that a 1:1 stoichiometry of the Hg<sup>2+</sup> to the probes respectively in the 139 complex. Furthermore, we also conducted the competitive experiments which 140 indicates the background metal ions showed very low interference with the detection 141 of  $Hg^{2+}$  in the water solutions (Fig.4.). 142

143 3.3 Emission spectra and detection limit of sensor

A fluorescence titration experiment was carried out in order to further explore the 144 interaction of probes and Hg<sup>2+</sup>. On addition and gradual increase concentration of 145 mercury ions to the  $L_1$  in CH<sub>3</sub>CN/H<sub>2</sub>O(V:V=1: 1), the fluorescence intensity gradually 146 increased and the solutions turned colorless to rose red. Limit detection of metal ions 147 plays an important role in evaluating fluorescence chemosensor. As for probe  $L_1$ , the 148 linear response for the fluorescence intensity response of compound  $L_1$  was between 0 149 and 140  $\mu$ M (Fig. 5), and the detection limit of Hg<sup>2+</sup> was measured to be 0.05  $\mu$ M. 150 The association constant K of the complex  $L_1$ -Hg<sup>2+</sup> was then calculated to be 151  $3.24 \times 10^3$  M<sup>-1</sup> with a linear relationship (Fig. 6) by Benesi–Hildebrand method, Eq. 152  $(1)^{30, 31}$ 153

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$$\frac{1}{F - F_0} = \frac{1}{K(F_{Max} - F_{Min})[Hg^{2+}]} + \frac{1}{F_{Max} - F_{Min}}$$
 (1)

As shown in the Fig.7, probe  $L_1$  show a weak fluorescence in the absence of metal ions. On the addition of 10 eqiv. metal ions, the fluorescence intensity has changed. It is obvious that when 10 eqiv.  $Hg^{2+}$  was introduced into a solution of  $L_1$  in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1, v/v), the fluorescence intensity increased sharply. Under this conditions, Fe<sup>3+</sup> induce a mild fluorescence enhancement while Zn<sup>2+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>, Pb<sup>2+</sup>, Ba<sup>2+</sup>, Ni<sup>2+</sup>, Mn<sup>2+</sup>, K<sup>+</sup>, Li<sup>+</sup>, Ag<sup>+</sup>, Al<sup>3+</sup>, Co<sup>2+</sup> and Na<sup>+</sup> did not show remarkable changes in fluorescence intensity and the color.

162 3.4 Time-dependence and Fluorescent lifetime

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Time-dependence for binding of the probe  $L_1$  with  $Hg^{2+}$  is given in Fig.8. 163 Following the addition of 10 equiv.  $Hg^{2+}$  ion to 20.0  $\mu$ M probe L<sub>1</sub>, the fuorescence 164 intensity of probe  $L_1$  was turn on rapidlly and reached a stable value within 2 min. 165 The fluorescence lifetime was measured at an excitation 460 nm of the NanoLED 166 source. The decays of probes were found to be monoexponential. The lifetime decays 167 in the absence of  $Hg^{2+}$  and in the presence of  $Hg^{2+}$  are shown in Fig.9. The average 168 lifetime of L<sub>1</sub> was 1.87 ns and 5.00 ns (XSQ = 1.06) while the lifetime of L<sub>1</sub> + Hg<sup>2+</sup> 169 was 1.91 ns (XSQ = 1.14). Double exponential fitting equation was used to describe 170 the fluorescence lifetime of probe itself, because xanthene was a chromophore which 171 172 formed a conjugated system. The upper part of structure formed a plane and orthogonalized with the ring of xanthene which also can be seen as a chromophore. 173 After adding metal ions, the upper part including carbanyl group and phenyl group do 174 not conjugate with xanthene, and the stereo-hindrance effect of carbanyl group limits 175 the rotation of the benzene ring. So the major chromophore is still xanthene which 176 used single exponential fitting equation. 177

178 3.5 Application

179 3.5.1 Test papers

To demonstrate the practical application of our sensor, we prepared the test papers of sensor  $L_1$ . As depicted in Fig.10, the color of the test paper changed from colorless to purple and deepened gradually with the increasing of  $Hg^{2+}$  concentration. These paper-made test kits may be used as a simple tool for detecting  $Hg^{2+}$  in environmental samples.

185 3.5.2 Fluorescence imaging for EC109 cells

Owing to its favourable spectroscopic properties of the response to  $Hg^{2+}$ ,  $L_1$  should be suited for fluorescence imaging in living cells. Laser scanning confocal microscopy was used to investigate this proposition. As determined by laser scanningconfocal microscope, EC109 cells with a 10µM solution of probe  $L_1$  in CH<sub>3</sub>CN-H<sub>2</sub>O (1:1, v/v) 30 min at 25 °C led to no intracellular fluorescence(Fig.11b.). After washed with PBS three times, the staining cells were then supplemented with 10 µM of  $Hg^{2+}$  under the same condition for 30 min, a significant increase of 193 fluorescence from the intracellular area was observed (Fig.11c).

### 194 3.6 Mechanism

To investigate the interaction of probes and metal ions, probe  $L_1$  was selected as an 195 example to explain mechanism. IR spectra of  $L_1$  and  $L_1$ +Hg<sup>2+</sup> were taken in KBr disks 196 (Fig.S22). It can be seen that the peak at 1698 cm<sup>-1</sup> which relates to the amide 197 carbonyl absorption disappeared upon with the addition of Hg<sup>2+</sup>. To further 198 investigate the mechanism of the reaction, we selected  $L_1$  as an typical example to 199 explain the phenomena. In addition, EDTA-adding experiment has been conducted to 200 confirm the reversibility of reaction and the result was shown in the Fig.12, when the 201 30 eqiv. EDTA was added into  $L_1 + Hg^{2+}$  in CH<sub>3</sub>CN/H<sub>2</sub>O solution, the fluorescence 202 intensity was decreased rapidly(green line). Then after adding the 30 eqiv. Hg<sup>2+</sup>, the 203 fluorescence intensity was recovered (red line). This phenomenon indicates that the 204 reaction between the compound and the metal ion are reversible. 205

As shown in the Fig.13, it indicates that these sereval chemosenors exhibit 206 reversible and sensitive detection to Hg<sup>2+</sup>. While hydrolytic action of compounds 207 might be happen after a long-term interaction with metal ions. Theoretical calculation 208 has been conducted to verify the stability of compound  $L_7$ , we assume the structure  $L_7$ 209 was existed after interacting with Hg<sup>2+</sup>. Frequency calculation successfully provide 210 the evidence for the existence of the stable structure. To investigate the possible 211 mechanism of the absorption and fluorescent emission of  $L_1$ , theoretical calculation of 212  $L_1$  which based on the optimized structures of the ground S0 state and the first 213 excited S1 state were performed in Fig.14. After adding Hg<sup>2+</sup>, the energy gap between 214 the HOMO and LUMO is greatly decreased<sup>32</sup>. 215

### 216 **4. Conclusions**

In summary, we reported the design and synthesis of six new rhodamine-based derivatives  $L_{1-6}$  used as sensitive and selective chemosenor, which could specifically recognize Hg<sup>2+</sup> ion in solutions by naked eyes. These six probe display 1:1 complex formation with Hg<sup>2+</sup> which could be monitored by the spectral changes as well as color changes. Based on the experiment above, after a series comparison of these probes with different substituent group, L<sub>1</sub> shows better optical properties than others

owing to its electron donating group and excellent properties. Confocal laser scanning 223 microscopy experiments showed that  $L_1$  could be used to detect  $Hg^{2+}$  in living cells. 224 Futhermore, these six new probes show a great potential for biotic environmental 225 226 detection. Supplementary Information (SI) 227 The structure of compounds RBH,  $L_{1-6}$  were characterized by <sup>1</sup>H NMR, HR-MS, IR in 228 Supplementary Information is 229 Supplementary Information. available www.ias.ac.in/chemsci. 230 231 232 Acknowledgements This work was supported by the Fundamental Research Funds for the Central 233 Universities (KYLX15\_0125) and National Major Scientific Instruments and 234 Equipment Development Projects (2014YQ060773) and A Project Funded by the 235 Priority Academic Program Development of Jiangsu Higher Education Institutions 236 (1107047002) and Policy Guidance Program (Research Cooperation)--Prospective 237 Joint Research Project (BY2016076-02). 238 239 References 240 Boening, D. W. Chemosphere 2000, 40, 1335 241 1. 2. Nolan, E. M.; Lippard, S. J.Chem. Rev. 2008,108, 3443 242 Nolan, E. M.; Lippard, S. J. J.Am.Soc. 2003, 125, 14270 3. 243 Wang, C. Inorg.Chem. 2013, 52, 13432 244 4. 5. Clarkson, T. W.; Magos, L.; Myers, G. J. New. Engl. J. Med. 2003, 349, 1731 245 246 6. Llobet, J.J.Agr. Food. Chem. 2003 51 838 247 7. Suresh, M. Org. Lett. 2008, 10, 3013 Chen, Y.; Han, K. Y.; Liu, Y. Bioorgan.Med.Chem.2007, 15, 4537 248 8. 9. Roy, P. Inorg.Chem. 2007, 46, 6405 249 10. Royzen, M. J. Am. Soc .2006, 128, 3854 250 11. Banerjee, A. Analyst 2012,137, 2166

at

12. Ding, P. New. J. Chem.2015, 39, 342 252

- 253 13. Lee, H. Y. Sensor. Actuat. B-Che.2013, 182, 530
- 254 14. Sen, S. Analyst. 2012, 137, 3975
- 255 15. Patil, R. Dal. Trans. 2014, 43, 2895
- 16. Haugland, R. P The handbook: a guide to fluorescent probes and labeling
  technologies 2005,p.52
- 258 17. Huang, J.; Xu, Y. J. Org.Chem. 2009, 74, 2167
- 259 18. Suresh, M. Org. Lett. 2009, 11, 2740
- 260 19. Zhou, Y. Org. Lett. 2009,11,4442
- 261 20. Du, J. Org. Lett. 2010, 12 476
- 262 21. McClure, D. S. J. Chem. Phys. 1952, 20, 682
- 263 22. Suresh, M.; Ghosh, A. Chem. Commun. 2008, 33, 3906
- 264 23. Renzoni, A.; Zino, F.; Franchi, E. Environ. Res. 1998, 77, 68
- 265 24. Dujols, V. F.; Ford, A. W.; J. Am. Soc. 1997,119, 7386
- 266 25. Yang, X.; Guo, R.; Zhao, Y, B.; Talanta. 2002, 57, 883
- 267 26. Geng, T.; R, Huang.; Wu D. RSC Adv. 2014, 4, 46332
- 268 27. Xiang, Y. Anal. Chim. Acta. 2007, 581, 132
- 269 28. Huang, C. Y. Method. Enzymol. 1981, 87, 509
- 270 29. Renny, J. S. Angew. Chem. Int. Edit. 2013, 52, 11998
- 271 30. Liu, Y, J. Helv. Chim. Acta. 2004, 87, 3119
- 272 31. Yan, L. Spectrochim. Acta. A. 2016, 155, 116
- 273 32. Liu, X. J. Tetrahedro. 2015, 71, 8194
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280 Fig.1. Fluorescence intensity (at 580 nm) of  $L_{1-6}$  (20µM) in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1, v/v) with the

281 presence of Hg<sup>2+</sup> (100  $\mu$ M) ( $\lambda$ ex = 500 nm)

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| Coordination constant                    |                 |        |        |          |        |        |  |  |
|--|-----------------|--------|--------|----------|--------|--------|--|--|
| -R                                       | CH <sub>3</sub> | Br     | F      | $C_6H_6$ | $NO_2$ | СНО    |  |  |
| Coordination<br>constant/M <sup>-1</sup> | 3.24E3          | 2.67E3 | 2.38E3 | 2.27E3   | 3.70E2 | 2.56E2 |  |  |

284 285 Scheme 2. The coordination constant of six probes





10 equiv. of various species.

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Fig.3. Job's plot of the complexation between  $L_1$  and  $Hg^{2+}$ , the total concentration of  $L_1$  and  $Hg^{2+}$ 

292 is 20.0 μM .

293



Fig.4. Fluorescence intensity (at 580 nm) of  $L_1$  upon the addition of 20  $\mu$ M Hg<sup>2+</sup> in the presence

296 of  $20\mu$ M background metal ions in CH<sub>3</sub>CN/H<sub>2</sub>O (1/1, v/v) ( $\lambda$ ex = 500 nm).





300  $\text{Hg}^{2+}$  concentration in CH<sub>3</sub>CN/H<sub>2</sub>O(1:1, v/v) solution ( $\lambda$ ex = 500 nm, slit = 3 nm)



303 Fig.5(b). Linear relationship of  $L_1$  (20  $\mu$ M) in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1,v/v) upon addition of different

amounts of  $Hg^{2+}$  ions.



307 Fig.6. Benesi–Hildebrand plot ( $\lambda ex = 500$  nm) of L<sub>1</sub>, assuming 1:1 stoichiometry for association

308 between  $L_1$  and  $Hg^{2+}$  in the solutions.



- 311 Fig.7. Fluorescence spectra of  $L_1$  (20  $\mu$ M) in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1,v/v) with the presence of 10 equiv.
- 312 of various metal ions ( $\lambda ex = 500 \text{ nm}$ ).



Fig. 8. Effects of time on the reaction of  $Hg^{2+}$  with  $L_1$ 



Fig.9. Fluorescence decay curves of  $L_1$  and  $L_1 + Hg^{2+}$  in solutions obtained at  $\lambda_{ex} = 500$  nm. 318



- Fig. 10. Photographs of the test kits with  $L_1$  for detecting  $Hg^{2+}$  in(CH<sub>3</sub>CN/H<sub>2</sub>O = 1:1, v/v) solution with different concentrations:(1) 0; (2)  $1.0 \times 10^{-5}$ M; (3)  $1.0 \times 10^{-4}$ M; (4)  $1.0 \times 10^{-3}$ M;(5) $1.0 \times 10^{-3}$ M;(5) $1.0 \times 10^{-3}$ M;(5) $1.0 \times 10^{-5}$ M; (3)  $1.0 \times 10^{-4}$ M; (4)  $1.0 \times 10^{-3}$ M;(5) $1.0 \times 10^{-5}$ M;(5) $1.0 \times 10^{-5}$ M;(5) $1.0 \times 10^{-5}$ M;(7)
- 323  $10^{-2}$ M;(6)1.0 ×  $10^{-1}$ M.



Fig.11. Laser confocal scanning microscopy experiments of EC109 cells: (a) Bright-field transmission image. (b) Cells with  $10\mu$ M solution of L<sub>1</sub> in CH<sub>3</sub>CN-H<sub>2</sub>O (1:1, v/v) for 30 min at

328 25 °C.(c)The staining EC109 cells exposed to  $L_1$  (10µM) for 30 min and then to a CH<sub>3</sub>CN-H<sub>2</sub>O

329 (1:1, v/v) solution of  $Hg^{2+}$  (10µM) for 30 min.

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332 Fig.12. Fluorescence intensity (at 580 nm) of  $L_1$  (20  $\mu$ M) to  $Hg^{2+}$  in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1,v/v)

solutions (1) blue line: 10  $\mu$ M L<sub>1</sub> only; (2) red line: 20  $\mu$ M L<sub>2</sub> with 10 equiv. Hg<sup>2+</sup>; (3) green line:

334  $20 \ \mu M \ L_1$  with 10 equiv. Hg<sup>2+</sup> and then addition of 30 equiv. EDTA; (4) black line:  $10 \ \mu M \ L_1$  with

335 10 equiv. Hg<sup>2+</sup> and 30 equiv. EDTA then addition of 10 equiv. Hg<sup>2+</sup> ( $\lambda$ ex = 500 nm).





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Fig.14. Theoretical molding of the absorption of  $L_1$  without and with Hg<sup>2+</sup> at the TDDFT level.