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Fe phthalocyanines and porphyrins immobilized by π -stacking interactions on various types of carbon nanotubes such as SWNTs, DWNTs, oxidized and non-oxidized MWNTs, was improved by the presence of the carbon nanotubes. However, as discussed by Rigsby *et al.* the nature and structure of the adsorbed catalyst are crucial parameters as well [14]. Thus, the formulation itself of a catalyst ink containing the same iron porphyrin, that is either deposited directly on glassy carbon (GC) or mixed with Vulcan and then deposited on GC electrode, has an effect at least as important as changing the structure of the molecular catalyst itself. In the same line, and more recently, we have reported on the synergistic effect on ORR of strapped porphyrins polymerized around carbon nanotubes [15]. It was shown that the presence of a potential proton relay in the hybrids materials in comparison with those lacking such a group did not generate any significant improvement. However, it is worth to mention that the spatial arrangement of the molecular catalyst, namely the iron porphyrin, properly functionalized with peripheral groups for the polymerization was expected to be somehow controlled by the covalent linkages between the porphyrin units. In the same line, it has been evidenced that a concomitant tuning of the ligand modification with the method of immobilization of the catalyst as well as the supporting material are three influential parameters [16–18].

Thus, in the present work, we now report on the ORR activity of MWNTs functionalized only by physisorption with specific iron (III) strapped porphyrins in a pH range from 13 to 6. These porphyrins contain a bridge bearing one or two carboxylic functions between the phenyl groups in 5 and 15 *meso* positions (Scheme 1). The overhanging bridge should prevent the aggregation of the porphyrins compared to the previous studies while preserving one side available for the interaction with the nanotubes by π -stacking. The goal of this study is first

to measure the ORR properties of strapped porphyrins bearing either one or two polar overhanging carboxylic acid(s) and second to evaluate the influence of the communication between the nanotube and the catalytic centers as well as the effects of the non-aggregation of the porphyrins adsorbed on the nanotube as depicted in Fig. 1.

RESULTS AND DISCUSSION

Strapped-porphyrin derivatives **1Fe** and **2Fe** bearing one and two carboxylic acid functions respectively were prepared according to a synthetic pathway previously reported in the case of a single strap porphyrin bearing 3,5-dihydroxyphenyl in the 10,20 *meso* positions [19]. Chiefly, the synthesis of **2**, summarized in Scheme 1, began by condensation of 5-(4-methoxyaryl)dipyrromethane **3** with 2-nitrobenzaldehyde in acidic conditions to obtain porphyrin **4** whose nitro groups were reduced by tin chloride in acidic medium leading to a mixture of the two atropisomers of the resulting aminophenyl porphyrin. Then, the atropisomer $\alpha\alpha$ **5** was separated by silica gel flash chromatography and acylated with 3-chloromethyl benzoyl chloride in the presence of triethylamine to obtain porphyrin **6**. Condensation of diethyl malonate in basic conditions allowed the formation of the strap in the 5,15 *meso* positions of the resulting porphyrin **7** and finally treatment by BBr_3 of the latter supplied free base porphyrin **2**. Actually, both porphyrins **1Fe** and **2Fe** were obtained by refluxing free-base porphyrin **2** in THF in the presence of iron(II) bromide and 2,6-lutidine in a glove box. During this process, the decarboxylation reaction leading partially to **1Fe** was observed and the two complexes were obtained in roughly equal proportions as indicated by TLC analysis and were separated by silica gel chromatography out of the glovebox after oxidation

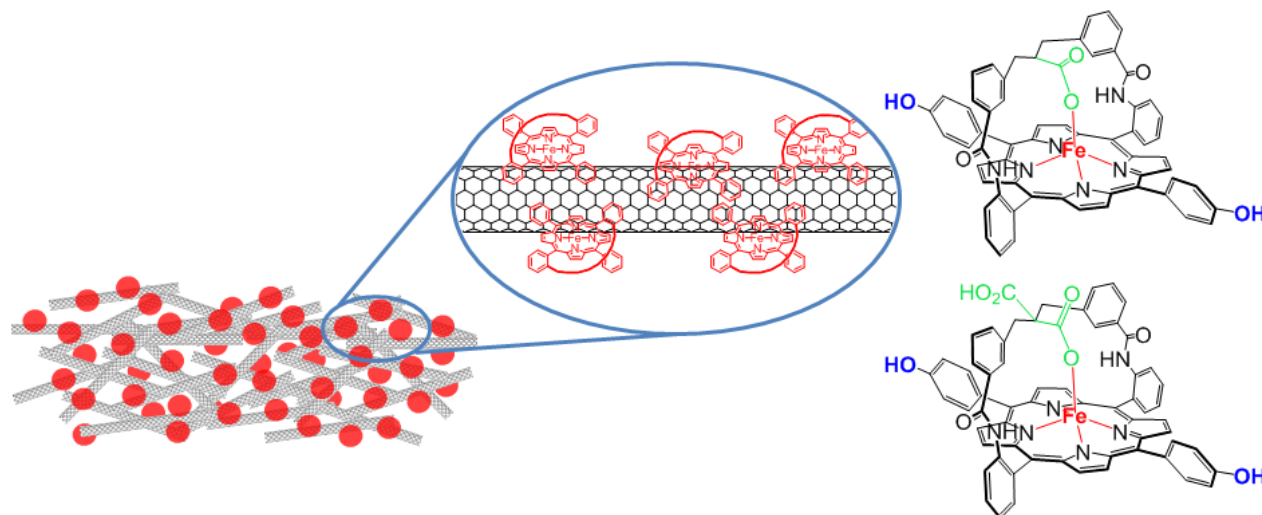
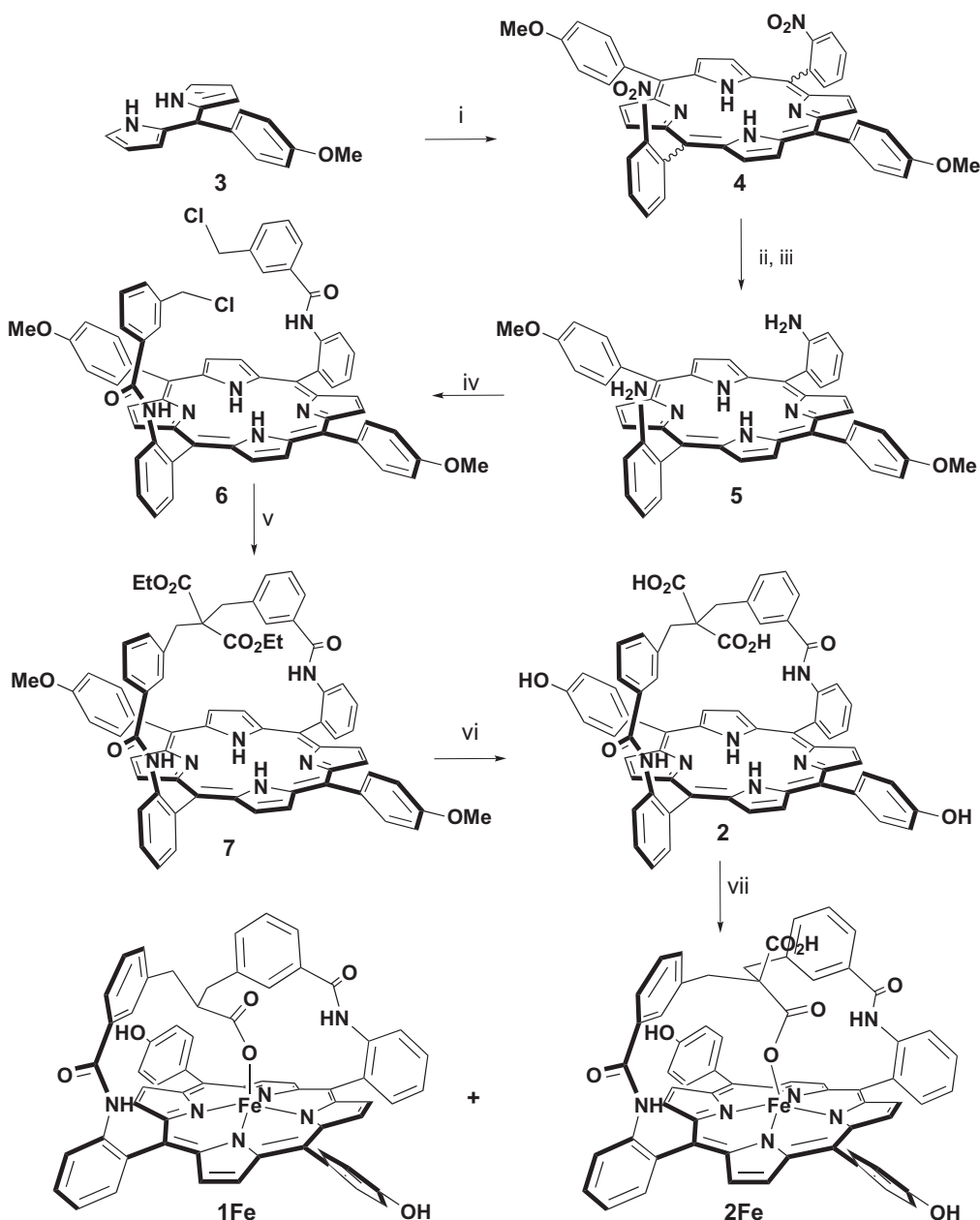


Fig. 1. Schematic representation of the nanotube/porphyrins hybrids



Scheme 1. Synthesis of iron porphyrins **1Fe** and **2Fe** bearing one and two overhanging carboxylic acid groups respectively, for MWNT coating. (i) 2-nitrobenzaldehyde, CH₂Cl₂, BF₃·Et₂O, 2 h, then DDQ, 20%; (ii) SnCl₂, HCl, 80%; (iii) silica gel chromatography, CH₂Cl₂, 66%; (iv) 3-(chloromethyl)benzoyl chloride (3 eq), NEt₃, THF, 85%; (v) CH₂(CO₂Et)₂ (10 equiv.), THF, EtONa, room temp., 12 h, 80%; (vi) BBr₃, CH₂Cl₂, room temp., 12 h, 80%; (vii) FeBr₂, 2,6-lutidine, THF, reflux overnight, silica gel column chromatography after air oxidation and HCl (1M) washing, **1Fe** (41%), **2Fe** (37%)

and identified by MALDI-TOF mass spectrometry (see experimental part).

Among the two final iron porphyrins, we were able to obtain single crystals of **1Fe** and to solve its X-ray structure (Fig. 2 and Table 1). The latter indicates that the iron cation is square-pyramidal five-coordinate with the intramolecular carboxylate bound on it with a Fe–O1 bond length of 1.997 Å. This intramolecular binding implies a significant distortion of the strap which retains a “W-shape” as observed on the apical view (Fig. 2,

right). Although a similar coordination polyhedron has already been reported for the bis-carboxylato analogous complex of **1Fe** [15], in the present case, the strap is more distorted with the carbon atom C2 diving toward the macrocycle plane (3.935 Å from iron against 4.301 Å for the non-decarboxylated complex) and with a distance of the iron atom to the mean porphyrin plane of 0.449 Å, against 0.512 Å for the non-decarboxylated complex. The very same distorted conformation was also found in an analogous zinc complex in which the distance of

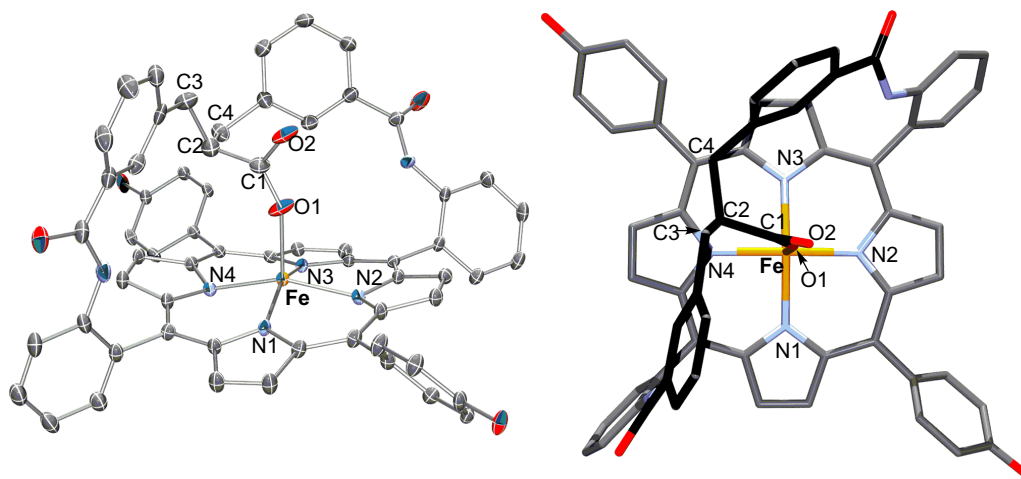


Fig. 2. X-ray structure of **1Fe**. Top: ORTEP lateral view (30% thermal ellipsoids) and bottom: apical rods view with the carbon atoms of the strap colored in black. Selected distances (Å): N1-Fe 2.066, N2-Fe 2.049, N3-Fe 2.061, N4-Fe 2.056, O1-Fe 1.997, C1-O1 1.232, C1-O2 1.230, (Fe, 24MP) 0.449

the metal to the mean porphyrin plane was only 0.386 Å [20], clearly indicating that this type of strap remains quite flexible. The macrocycle is strongly saddle-shaped with almost no ruffling, as indicated by the angle with the mean plane of the two pairs of diametrically opposed pyrrole rings (N1 and N3: -13.02° and -12.68° ; N2 and N4: 11.14° and 14.06°).

In order to prepare the hybrid materials, MWNT (Nanocyl NC3100TM) were purified by treatment with nitric acid (35%) at 100°C for 5 h (see experimental section). After treatment the nanotubes were diluted with iced-water, filtered through $0.45\ \mu\text{m}$ PTFE membrane and extensively washed with water, methanol and then dried under vacuum. The catalysts were prepared by mixing MWNT with porphyrins **1Fe** or **2Fe** (in a 1:1 ratio in weight) in THF under bath sonication. After mixing, THF was evaporated and the catalyst inks were prepared by dispersing, using bath sonication, the different MWNT/porphyrin hybrids (3 mg) in 750 μL of ethanol and 75 μL of Nafion solution (5% in alcohol). Similarly, the inks of the reference compounds MWNT, **1Fe** and **2Fe** were prepared by mixing the nanotubes or the porphyrins in 750 μL of ethanol and 75 μL of Nafion solution (5% in alcohol). The catalysts were drop-casted on the Glassy Carbon disk and tested in a series of Rotating Ring Disk Electrode experiments at pH 13 (NaOH, 0.1 M), pH 10, 8 and 6 (phosphate buffers).

We first compared the electrocatalytic properties of the different components: MWNT, iron porphyrins **1Fe** and **2Fe** and the 1:1 mixture MWNT-**1Fe** and MWNT-**2Fe** at pH 13 (Fig. 3) and pH 10 (Fig. S48). All the curves correspond to the average (reduction and reoxidation) of the cyclic voltammetry curves.

Figures 3a–3b present the comparison between the ORR activity of MWNT-**1Fe**, MWNT-**2Fe** and those of the iron porphyrins deposited directly on the Glassy

Carbon (GC) electrode. The curves show that the catalyst inks made by mixing the nanotubes with the porphyrins exhibit higher current density and lower overpotential (of about 0.1 V) than porphyrins alone. This result is not surprising since similar observations were made by Rigsby *et al.* in the case of iron porphyrins deposited on different carbon supports [14]. In addition, it is interesting to notice that the voltammetry curves of MWNT-**1Fe** and MWNT-**2Fe** exhibit two waves at around -0.4 and -0.7 V, explained by the own activity of the nanotubes (see below). Figure 3c shows the activity of MWNT and Fig. 3d compares the voltammetry curves (0 and 2000 rpm) of MWNT, **1Fe** and MWNT-**1Fe**. First, we observe that, at pH 13, MWNT reduced oxygen with lower overpotential and higher current density than **1Fe**. Second, the reduction starts roughly at the same potential ($-0.2\text{ V vs. Ag/AgCl}$) for MWNT and MWNT-**1Fe**. Multi-walled carbon nanotubes are active materials for oxygen reduction in alkaline media [21, 22] and it is likely that the nanotubes are more competitive than the nanotube/porphyrin hybrids at pH 13. It can explain the two waves observed on the reduction curves of the nanotube/porphyrin hybrids. We believe that the reduction initiated on the nanotubes is then performed by the porphyrin at lower potential. Nevertheless, when the plateau is reached at $-0.8\text{ V vs. Ag/AgCl}$ the current density is larger of ca. 2 mA/cm^2 for MWNT-**1Fe** compared to MWNT alone.

Figure S48 shows the same comparison between the ORR activity of MWNT, MWNT-**1Fe**, MWNT-**2Fe** and those of **1Fe** and **2Fe** deposited on the GC electrode, but at pH = 10. Once again, the curves clearly demonstrate that the catalyst inks made by mixing the nanotubes with the porphyrins exhibit higher current density than porphyrins alone. The reduction of oxygen starts with an overpotential of almost 0.4 V for **1Fe** and **2Fe** compared to the same porphyrin deposited on the nanotubes.

Table 1. X-ray structural data of **1Fe**

Empirical formula	C ₆₂ H ₄₁ FeN ₆ O ₆
Formula weight	1021.86 g/mol
Temperature	150 (2) K
Wavelength	0.71073 Å
Crystal system, space group	triclinic <i>P</i> -1
Unit cell dimensions	<i>a</i> = 14.499 (2) Å, α = 90.838 (5)° <i>b</i> = 14.729 (2) Å, β = 109.567 (6)° <i>c</i> = 17.848 (3) Å, γ = 117.362 (5)
Volume	3124.0 (8) Å ³
Z, Calculated density	2, 1.086 g·cm ⁻³
Absorption coefficient	0.291 mm ⁻¹
<i>F</i> (000)	1058
Crystal size	0.320 × 0.260 × 0.110 mm
Crystal color	black
Theta range for data collection	3.026 to 27.484°
<i>h</i> _min, <i>h</i> _max	-18, 18
<i>k</i> _min, <i>k</i> _max	-19, 19
<i>l</i> _min, <i>l</i> _max	-23, 23
Reflections collected/unique	57073/13966 [<i>R</i> (int) ^a = 0.1159]
Reflections [<i>I</i> > 2σ]	9825
Completeness to theta_max	0.975
Absorption correction type	multi-scan
Max. and min. transmission	0.968, 0.776
Refinement method	Full-matrix least-squares on <i>F</i> ²
Data/restraints/parameters	13966/0/672
^b <i>S</i> (Goodness-of-fit)	1.036
Final <i>R</i> indices [<i>I</i> > 2σ]	<i>R</i> ₁ ^c = 0.0888, <i>wR</i> ₂ ^d = 0.2340
<i>R</i> indices (all data)	<i>R</i> ₁ ^c = 0.1213, <i>wR</i> ₂ ^d = 0.2579
Largest diff. peak and hole	1.852 and -1.601 e ⁻ ·Å ⁻³

$$^a R_{\text{int}} = \sum |F_o^2 - \langle F_o^2 \rangle| / \sum [F_o^2]$$

$$^b S = \{ \sum [w(F_o^2 - F_c^2)^2] / (n - p) \}^{1/2}$$

$$^c R_1 = \sum ||F_o| - |F_c|| / \sum |F_o|$$

$$^d wR_2 = \{ \sum [w(F_o^2 - F_c^2)^2] / \sum [w(F_o^2)^2] \}^{1/2}$$

$$w = 1 / [\sigma(F_o^2) + aP^2 + bP] \text{ where } P = [2F_c^2 + \max(F_o^2, 0)]/3$$

MWNT-1Fe and **MWNT-2Fe** exhibit very similar properties, it appears that the presence of one or two carboxylate functions does not influence the properties under alkaline conditions. At pH 10, **MWNT** become less competitive for oxygen reduction than **MWNT-1Fe** or **MWNT-2Fe** and only one reduction wave is observed.

The comparison of the ORR activity (RDE and RRDE) of **MWNT**, **1Fe**, **2Fe**, **MWNT-1Fe** and **MWNT-2Fe** for pH 8 and pH 6 are given in Fig. S46 and Fig. S47, respectively. The catalytic activities of **MWNT-1Fe** and **MWNT-2Fe** (and of the reference compounds) exhibit the same trend with respect to those observed at pH 13 and 10. We also performed tests at pH 6 and pH 8 with

a porphyrin that does not exhibit proton relay and similar results to those obtained for **MWNT-1Fe** were obtained [15]. The presence of the proton in close proximity to the iron center does not seem to be mandatory to improve the reaction in our case. Indeed the presence of Nafion in the mixture probably ensures the availability of protons close to the reaction center.

It is worth mentioning that at pH 6 and lower, the ORR activity of the porphyrins decreases rapidly and almost completely vanishes after the 3 cycles requiring the deposition of a new catalyst ink for each rotation cycle. This loss of ORR activity can be due to demetalation of the porphyrin during the electrochemical processes [23, 24], this effect being well documented in the case of iron phthalocyanine [25–27] but still subject to debate in the case of porphyrins.

Figure 4 shows the RRDE curves registered at a rotation rate of 400 rpm for **MWNT**, **MWNT-1Fe** and **1Fe** at pH 13 and 10. The numbers of electrons *n* involved in the reduction, summarized in Table 2 were determined following the equation $n = 4I_d / (I_d + I_r/N_c)$ as recommended by Qiao *et al.* [28], from the disk and ring currents and with a collection coefficient *N_c* = 0.2 determined using the one-electron Fe(CN)₆³⁻/Fe(CN)₆⁴⁻ redox couple. From the curves, it is observed that the reduction of O₂ is accompanied by the production of hydrogen peroxide both for **MWNT** and **1Fe**. Conversely, for **MWNT-1Fe**, almost no production of H₂O₂ is detected at the plateau. At pH 13 a bump in the ring current between -0.20 and -0.70 V reflecting the production of H₂O₂ is observed; however, this phenomenon is attributed to the initial reduction of oxygen by the nanotubes and appears before the plateau at disk is reached. However, at both pH, the evaluated value of *n* remains close to the ideal value of 4.

CONCLUSION

Herein we formulated a series of catalyst inks for oxygen reduction reaction containing strapped porphyrins and MWNTs. The combination of the nanotubes with the iron porphyrins systematically gives better catalytic properties than those of two components taken separately. While carbon nanotubes are known to be slightly active in oxygen reduction, they produce significant amounts of hydrogen peroxide. When porphyrins are simply mixed in Nafion and deposited on the glassy carbon electrode, a low efficiency is generally observed. We attribute this behavior to the lack of electrons available for the reduction. Indeed in the catalyst ink, the porphyrins are embedded in Nafion and only the porphyrins close to the glassy carbon disk can benefit from efficient electron transfers from

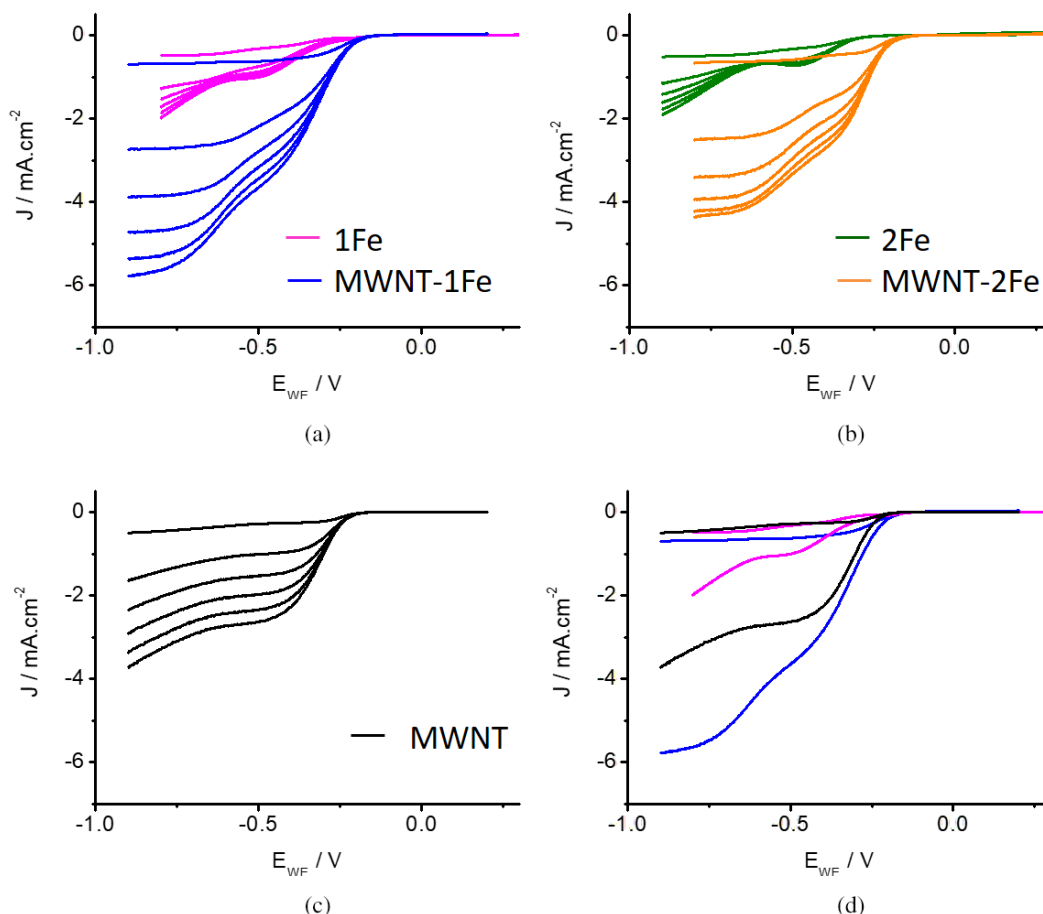


Fig. 3. Polarization curves at different rotation rates (0, 400, 800, 1200, 1600 and 2000 rpm) recorded for ORR in O_2 -saturated 0.1 M NaOH solution (pH 13) (scan rate = 5 mV/s, room temperature) on GC with predeposited (a) **1Fe** (pink) and **MWNT-1Fe** (blue); (b) **2Fe** (green) and (c) **MWNT-2Fe** (orange); **MWNT** (black). (d) Comparison between **MWNT**, **MWNT-1Fe** and **1Fe** at 0 and 2000 rpm

the electrodes. Conversely, carbon nanotubes create a percolating pathway for the charges in the catalyst film that ensure electron availabilities for the reduction of oxygen. In this case almost no hydrogen peroxide is produced and the reduction of oxygen is mostly performed via a 4 electrons pathway. However, under acidic conditions, at pH 6 and lower, the catalytic activity of these inks decreased rapidly.

EXPERIMENTAL

Materials and methods

Mass spectra: ESI: *Micromass MS/MS ZABSpec TOFF* spectrometer. MALDI-TOF: *Microflex-LT Bruker Daltonics* were performed at the C.R.M.P.O. (University of Rennes 1) using α -cyano-4-hydroxycinnamic acid as the matrix. 1H - and ^{13}C -NMR spectra were recorded either on *BrukerAvance500* or *BrukerAvance400* spectrometers equipped with a BBFO probe. Spectra were referenced with residual solvent protons. UV/vis spectra

were recorded on an Uvikon XL spectrometer. Chemicals were purchased from Aldrich and were used as received. Solvents were purchased from Aldrich or VWR and were used as received. THF (K/benzophenone, N_2) was distilled before use. MWNT commercial grade NC3100 (>95%) were purchased from Nanocyl.

Synthesis of porphyrin ligands

5-(4-Methoxyphenyl)-dipyrromethane 3. In a two neck round bottom flask equipped with a stir bar and a gas inlet, 4-methoxy benzaldehyde (92.8 mmol, 11.3 mL) and pyrrole (161.2 mL, 25 equiv.) were mixed. The reaction mixture was degassed for 15 min in argon under dark at room temperature, and then TFA (707 μ L, 0.1 equiv.) was added. The solution was stirred for further half an hour. The reaction was monitored by TLC, after that the reaction mixture was quenched by Et_3N . The excess pyrrole was recovered under reduced pressure. The resulting solid was dissolved in CH_2Cl_2 and directly loaded on a silica gel chromatography column. The desired compound eluted with 70% CH_2Cl_2 -cyclohexane

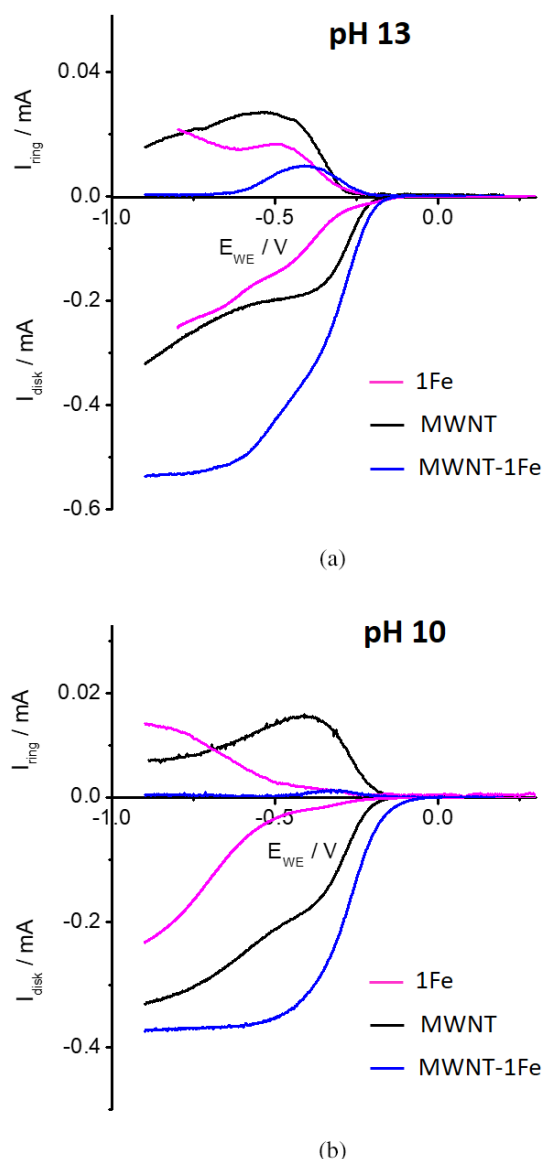


Fig. 4. RRDE measurements of oxygen reduction (negative current) and hydrogen peroxide oxidation (positive current) for MWNT (black), 1Fe (pink) and MWNT-1Fe (blue) at (a) pH 13 and (b) pH 10 in O_2 -saturated solutions. The ring electrode was polarized at 0.260 V vs. Ag/AgCl. Rotation rate: 400 rpm; scan rate: 5 mV s^{-1}

was obtained in 50% yield (11.65 g, 46.17 mmol). 1H NMR ($CDCl_3$, 298 K, 500.13 MHz): δ 7.90 (2H, pyr_{NH}), 7.18 (2H, d, $^3J = 8.61$ Hz, aro_2), 6.91 (2H, d, $^3J = 8.61$ Hz, aro_3), 6.71 (2H, m, pyr_4), 6.22 (2H, m, pyr_3), 5.97 (2H, m, pyr_2), 5.44 (1H, CH_α), 3.84 (3H, s, OMe). ^{13}C NMR ($CDCl_3$, 298 K, 500.13 MHz): δ 158.5, 134.3, 132.9, 129.4, 117.2, 114, 108.3, 107.2, 55.3, 43.2. ESI-HRMS: calcd $m/z = 275.1154$ [$M-H+Na$] $^+$ for $C_{16}H_{16}N_2NaO$, found 275.1159.

5,15-Bis-(2-nitrophenyl)-10,20-bis-(4-methoxyphenyl)-porphyrin 4. Samples of 4-methoxyphenyldipyromethane **3** (7.9 mmol, 2 g) and 2-nitrobenzaldehyde (1.19 g,

Table 2. Number of electrons involved in the reduction of O_2 at -0.75V a vs. Ag/AgCl

	1Fe		MWNT-1Fe
pH 10	3.22	3.54	3.98
pH 13	2.81	2.82	3.97

a Potential chosen on the plateau for MWNT-1Fe.

1 equiv.) were dissolved in freshly prepared distilled CH_2Cl_2 (600 mL) in a 1 L round-bottomed flask containing molecular sieves, degassed with a stream of Ar for 15 min. Then $BF_3 \cdot Et_2O$ (110 μ L, 0.1 equiv.) was added slowly over 30 s. The reaction was stirred at room temperature and monitored by TLC and MALDI-TOF. After 2 h, DDQ (2.7 g, 1.5 equiv.) was added, and the reaction mixture was stirred at room temperature for a further 1 h. The complete reaction mixture was quenched by Et_3N and evaporated under reduced pressure to give a black solid which was dissolved in CH_2Cl_2 . The mixture of two atropisomers *i.e.* $\alpha\beta$ and $\alpha\alpha$ -bis-2-nitrophenylporphyrin were purified by silica gel column chromatography using CH_2Cl_2 as eluent. The atropisomers could not be separated by the usual method of column chromatography on silica gel due to the same polarity. Overall yield: 20 % (600 mg, 0.78 mmol).

α -5,15-Bis-(2-aminophenyl)-10,20-bis-(4-methoxyphenyl)-porphyrin 5. $\alpha\beta$ and $\alpha\alpha$ atropisomers of the dinitroporphyrin **4** (5.2 mmol, 4 g) were dissolved in the mixture of CH_2Cl_2 -MeOH (100 mL), taken in a 2 L conical flask along with a reducing agent $SnCl_2 \cdot 2H_2O$ (11.8 g, 10 equiv.) and concentrated HCl (200 mL) was added slowly to the mixture. The resulting green solution was stirred for 2 days at RT. After completion of the reaction (monitored by MALDI-TOF), it was quenched by aqueous KOH solution at 0 $^\circ$ C under ice. The resulting violet solution was washed several times with water and $CHCl_3$. The organic layers were collected and dried over $MgSO_4$. Yield: 80% (2.95 g, 4.19 mmol).

Steric decompression of two atropisomers. In a 1 L two necks round bottom flask equipped with a stir bar and a condenser, 200 g of silica (60 μ m) were added to toluene (400 mL). The reaction mixture was heated to 80 $^\circ$ C and degassed with argon during 45 min. Then 2.9 g of the $\alpha\beta$ and $\alpha\alpha$ atropisomers of the bis-2-aminophenylporphyrin obtained previously were dissolved in toluene and added to the silica gel mixture in toluene. After evaporation of the solvent, the compound was dissolved in minimum amount of CH_2Cl_2 and purified by column chromatography. The two atropisomers were separated on a silica gel chromatography eluted with CH_2Cl_2 /MeOH ($\alpha\beta$ 0.2%, $\alpha\alpha$ 0.5%). Yield: **5** $\alpha\alpha$ (66%, 1.9 g, 2.70 mmol), $\alpha\beta$ (34%, 0.9 g, 1.28 mmol). It is worth to note that the atropisomer $\alpha\alpha$ **5** remains contaminated with by-products resulting from scrambling reactions not separable by silica gel flash chromatography and has not been fully characterized.

α -5,15-Bis-(2-[[3-chloromethyl]benzoylamido]-phenyl)-10,20-bis-(4-methoxyphenyl)-porphyrin 6. A 500 mL two neck round bottom flask equipped with a stirrer and cooled in an ice bath was charged with compound **5** (0.99 mmol, 700 mg), dry CH_2Cl_2 (300 mL) and NEt_3 (350 μL , 2.5 equiv.). 3-(chloromethyl)benzoyl chloride (420 μL , 3 equiv.) was then added dropwise under argon atmosphere. The reaction mixture was allowed to stir for three hours. Then the reaction was quenched by water and the organic layer was separated. The solvent was removed under vacuum. The resulting solid was dissolved in CH_2Cl_2 and directly loaded on a silica gel chromatography column. The expected compound eluted with 0.2% $\text{CH}_2\text{Cl}_2/\text{MeOH}$, was obtained in 90% yield (902 mg, 0.89 mmol). ^1H NMR (CDCl_3 , 298 K, 500.13 MHz): δ 8.93 (4H, d, $^3J = 4.81$ Hz, βpyr), 8.87 (4H, d, $^3J = 4.81$ Hz, βpyr), 8.93 (2H, d, $^3J = 8.06$ Hz, aro_2), 8.13 (2H, d, $^3J = 7.49$ Hz, aro_5), 8.11 (2H, d, $^3J = 7.60$ Hz, aro_9), 8.02 (2H, d, $^3J = 7.60$ Hz, aro_9), 7.91 (2H, t, $^3J = 8.06$ Hz, aro_3), 7.63 (2H, s, NHCO), 7.59 (2H, t, $^3J = 7.49$ Hz, aro_4), 7.28 (4H, bs, aro_8 , aro_8), 6.73 (2H, d, $^3J = 7.68$ Hz, aro_4), 6.55 (2H, d, $^3J = 7.90$ Hz, aro_6), 6.49 (2H, t, $^3J = 7.68$, aro_5), 6.33 (2H, s, aro_2), 4.09 (6H, s, OMe), 3.30 (4H, s, CH_2bz), -2.61 (2H, s, NH_{int}). ^{13}C NMR (CDCl_3 , 298 K, 500.13 MHz): δ 164.7, 159.7, 138.7, 137.1, 135.7, 135.6, 134.9, 134.7, 133.6, 131.7, 130.8, 129.9, 128.4, 126.2, 126.1, 123.2, 120.9, 120.7, 113.7, 112.5, 112.4, 55.6, 44.3. ESI-HRMS: calcd $m/z = 1009.3030$ $[\text{M} + \text{H}]^+$ for $\text{C}_{62}\text{H}_{47}\text{N}_6\text{O}_4^{35}\text{Cl}_2$, found 1009.3031, calcd $m/z = 973.3263$ $[\text{M} - \text{HCl} + \text{H}]^+$ for $\text{C}_{62}\text{H}_{46}\text{N}_6\text{O}_4^{35}\text{Cl}$, found 973.3268. UV-vis (DMF): λ/nm (10^{-3} ϵ , $\text{dm}^3 \text{mol}^{-1} \text{cm}^{-1}$): 426 (364), 521 (14), 558 (8), 599 (4), 652 (3.4).

α -5,15-Bis-({2,2-(3,3-[2,2-(diethoxycarbonyl)propane-1,3-diyl]-dibenzoyl-amido]-diphenyl)-10,20-bis-(4-methoxyphenyl)-porphyrin 7. Sodium metal (182 mg, 10 equiv.) was added to the absolute alcohol (20 mL) in a small round bottom flask and stirred for few minutes until the complete consumption of Na. Diethyl malonate (1.2 mL, 10 equiv.) was added to this solution at room temperature and stirred for half an hour. The resulting mixture was added to a solution of porphyrin **6** (0.79 mol, 800 mg, 1 equiv.) in CH_2Cl_2 (600 mL) and the solution was turned immediately from violet to green. After 2 h of stirring the reaction was quenched by H_2O , the organic layer was separated and removed under vacuum. The desired product was purified on a silica gel chromatography column eluted with 0.3% $\text{CH}_2\text{Cl}_2/\text{MeOH}$. The expected compound was obtained in 80% yield (702 mg, 0.64 mmol). ^1H NMR (CDCl_3 , 298 K, 500 MHz): δ 9.15 (2H, d, $^3J = 8.39$ Hz, aro_2), 8.93 (4H, d, $^3J = 4.58$ Hz, βpyr), 8.86 (4H, d, $^3J = 4.58$ Hz, βpyr), 8.22 (2H, s, NHCO), 8.19 (2H, bs, aro_9), 8.01 (2H, d, $^3J = 7.54$ Hz, aro_5), 7.98 (2H, bs, aro_9), 7.91 (2H, t, $^3J = 7.70$ Hz, aro_3), 7.70 (2H, d, 2H_b , $^3J = 7.98$ Hz, aro_6), 7.55 (2H, t, $^3J = 7.70$ Hz, aro_4), 7.31 (4H, bs, aro_8 , aro_8), 6.98 (2H, t, $^3J = 7.90$ Hz, aro_5), 6.74 (2H, d, $^3J = 7.90$ Hz, aro_4), 4.99 (2H, s, aro_2), 4.08 (6H, s, OCH_3), 1.70 (4H, s, CH_2bz),

1.23 (4H, bs, CH_2ester), -0.55 (6H, t, $^3J = 6.58$ Hz, CH_3ester), -2.40 (2H, s, NH_{int}). ^{13}C NMR (CDCl_3 , 298 K, 500 MHz): δ 167.5, 164.4, 159.6, 138.8, 135.9, 135.2, 134.9, 133.8, 133.7, 132.4, 131.2, 129.9, 128.2, 127.4, 125.9, 122.8, 120.7, 119.7, 113.7, 112.5, 59.9, 58.8, 55.5, 42.5, 11.9. ESI-HRMS: calcd $m/z = 1097.4232$ $[\text{M} + \text{H}]^+$ for $\text{C}_{69}\text{H}_{57}\text{N}_6\text{O}_8$, found 1097.4234, calcd $m/z = 1119.4051$ $[\text{M} + \text{Na}]^+$ for $\text{C}_{69}\text{H}_{56}\text{N}_6\text{O}_8\text{Na}$, found 1119.4047. UV-vis (DMF): λ/nm (10^{-3} ϵ , $\text{dm}^3 \text{mol}^{-1} \text{cm}^{-1}$): 425 (380), 520 (15.8), 558 (8.2), 596 (4.4), 653 (3.4).

α -5,15-Bis-({2,2-(3,3-[2,2-(dicarboxylicacid)propane-1,3-diyl]-dibenzoyl-amido]-diphenyl)-10,20-bis-(4-hydroxyphenyl)-porphyrin 2. Boron tribromide (2.5 mL, 50 equiv.) was added to compound **7** (0.54 mmol, 600 mg) was dissolved in DCM (100 mL). After 12 h of stirring at RT, the reaction was completed. The mixture was quenched by water. The precipitated compound was filtered and the green solid was washed with water at pH = 7. The product was purified by silica gel chromatography column and eluted with $\text{CHCl}_3/\text{MeOH}/\text{AcOH}$ (90/9/1). Yield: 80% (440 mg, 0.43 mmol). ^1H NMR ($\text{DMSO}-d_6$, 298 K, 500.13 MHz): δ 9.92 (2H, s, OH), 8.84 (4H, d, $^3J = 4.55$ Hz, βpyr), 8.77 (4H, d, $^3J = 4.55$ Hz, βpyr), 8.59 (2H, s, NHCO), 8.37 (4H, bs, aro_2 , aro_5), 8.12 (2H, bs, aro_9), 7.92 (2H, t, $^3J = 7.92$ Hz, aro_3), 7.84 (2H, d, $^3J = 6.25$ Hz, aro_9), 7.75 (2H, t, $^3J = 7.60$ Hz, aro_4), 7.24 (2H, d, $^3J = 8.04$ Hz, aro_6), 7.16 (4H, bs, aro_8 , aro_8), 6.90 (2H, t, $^3J = 7.49$ Hz, aro_5), 6.72 (2H, d, $^3J = 7.77$ Hz, aro_4), 4.57 (2H, s, aro_2), 1.19 (4H, s, CH_2bz), -2.70 (2H, bs, NH_{int}). ^{13}C NMR ($\text{DMSO}-d_6$, 298 K, 500 MHz): δ 171.5, 165.7, 157.8, 139.2, 136.2, 135.9, 135.7, 135.3, 134.9, 134.8, 132.1, 132.0, 129.7, 128.1, 126.8, 126.4, 124.6, 124.3, 120.7, 115.3, 114.3, 59.3, 40.5. ESI-HRMS: calcd $m/z = 1013.3293$ $[\text{M} + \text{H}]^+$ for $\text{C}_{63}\text{H}_{45}\text{N}_6\text{O}_8$, found 1013.3288, calcd $m/z = 1035.3112$ $[\text{M} - \text{H} + \text{Na}]^+$ for $\text{C}_{63}\text{H}_{43}\text{N}_6\text{NaO}_8$, found 1035.3102. UV-vis (DMF): λ/nm (10^{-3} ϵ , $\text{dm}^3 \text{mol}^{-1} \text{cm}^{-1}$): 428 (344), 522 (19), 560 (12), 599 (7.6), 655 (6).

Iron insertion. A free-base solution of porphyrin **2** in THF in the presence of an excess of iron bromide and 2,6-lutidine was heated at reflux overnight inside a glove box. During this process, the decarboxylation reaction leading to both **1Fe** and **2Fe** was observed and the two complexes were obtained in roughly equal proportions as indicated by TLC analysis. The resulting mixture was taken out of the glove box, washed with HCl (1M), and dried. There were easily separated by silica gel chromatography using a gradient of MeOH in CHCl_3 (from 0.2% to 2.2%) and identified by MALDI-TOF mass spectrometry (**1Fe**: 41%, calcd $m/z = 1022.25$ $[\text{M} + \text{H}]^+$ for $\text{C}_{62}\text{H}_{42}\text{FeN}_6\text{O}_6$ found 1022.30; **2Fe**: 37%, calcd $m/z = 1066.24$ $[\text{M} + \text{H}]^+$ for $\text{C}_{63}\text{H}_{42}\text{FeN}_6\text{O}_8$ found 1066.41).

MWNT. MWNTs (60 mg) were sonicated in nitric acid (35 vol%) (150 mL) with a sonic bath (Fisherbrand, 37 kHz, power 100% for 10 min and then 40% for 30 min) and then heated at 100 °C for 5 h. The suspension was then cooled and vacuum filtered through a 0.2 μm PTFE

membrane and washed with water. The nanotubes were redispersed in NaOH 2 M (100 mL) using the sonic bath (100% for 10 min) and then filtered through a PTFE membrane and washed with deionized water, and then HCl 1 M followed by deionized water until the filtrate was neutral.

Electrochemical experiments

Sample preparation. For the preparation of the sample containing nanotubes (**MWNT-1Fe**, **MWNT-2Fe**), a mixture of purified MWNT (9 mg) and porphyrin **1Fe** (9 mg) or porphyrin **2Fe** (9 mg) in dry THF (10 ml) was homogenized using a sonic bath (Fisherbrand, 37 kHz, power 100%) for 15 min. The THF was gently evaporated with a stream of N₂ and the mixtures were dried under vacuum. 3 mg of mixture were dispersed in 750 µl of ethanol and 75 µl of Nafion solution (5% in alcohol). The mixtures were homogenized using a sonic bath until they formed homogenous inks. For the **MWNT** ink, the same procedure was followed but without adding porphyrins. For the reference **1Fe** and **2Fe** inks, 3 mg of porphyrins were directly dispersed in 750 µl of ethanol and 75 µl of Nafion solution (5% in alcohol).

Electrode preparation. Before each measurement, the glassy carbon (GC) disk (5 mm, 0.196 cm²) used as rotating electrode was polished with aqueous dispersions of synthetic diamonds (1 µm), then rinsed and sonicated with water. 5 µl of the catalyst inks were deposited by drop-casting onto the GC disk, then dried in air. For pH 6, a new ink was deposited onto the GC disk for each rotation step.

Electrochemical measurements. The instrument used was a VSP bipotentiostat (Bio-Logic SAS). The electrochemical tests were carried out in 0.5 M H₂SO₄ solution in a three electrode glass cell, thermostated at 25 °C. A “CE to Ground” connection with a saturated KCl Ag/AgCl electrode as reference and a graphite plate as counter electrode was used. As working electrode, a Pine rotating ring disk electrode (RRDE) with catalyst-loaded GC disk (0.196 cm²) and Pt ring (0.110 cm²) was controlled by a speed control unit from Princeton Applied Research Model 636 Electrode Rotator. The voltammograms were recorded at 5 mV · s⁻¹ in stationary conditions (with various rotating rates: 0, 400, 800, 1200, 1600, and 2000 rpm) in O₂-saturated solutions. An average current was calculated from the forward and backward scans. All potentials reported in this paper refer to that of the Ag/AgCl electrode. H₂O₂ production was monitored in the RRDE configuration at 400 rpm with a CV at the GC disk (5 mV · s⁻¹). The collection coefficient of the RRDE (0.20) was measured using the one-electron Fe(CN)₆³⁻/Fe(CN)₆⁴⁻ redox couple, according to the manufacturer’s instructions.

X-ray crystallographic studies

CCDC 1916858, C₆₂H₄₁FeN₆O₆, *M* = 1021.86. D8 VENTURE Bruker AXS diffractometer equipped

with a (CMOS) PHOTON 100 detector founded by FEDER, Mo-*K*α radiation ($\lambda = 0.71073$ Å, multilayer monochromator), *T* = 150 (2) K; triclinic *P* -1 (I.T.#2), *a* = 14.499 (2), *b* = 14.729 (2), *c* = 17.848 (3) Å, $\alpha = 90.838$ (5), $\beta = 109.567$ (6), $\gamma = 117.362$ (5)°, *V* = 3124.0 (8) Å³. *Z* = 2, *d* = 1.086 g · cm⁻³, $\mu = 0.291$ mm⁻¹. The structure was solved by dual-space algorithm using the *SHELXT* program [29], and then refined with full-matrix least-squares methods based on *F*² (*SHELXL*) [30]. The contribution of the disordered solvents to the calculated structure factors was estimated following the *BYPASS* algorithm [31], implemented as the *SQUEEZE* option in *PLATON* [32]. A new data set, free of solvent contribution, was then used in the final refinement. All non-hydrogen atoms were refined with anisotropic atomic displacement parameters. H atoms were finally included in their calculated positions and treated as riding on their parent atom with constrained thermal parameters. A final refinement on *F*² with 13966 unique intensities and 672 parameters converged at $\omega R(F^2) = 0.2340$ (*R*(*F*) = 0.0888) for 9825 observed reflections with *I* > 2σ(*I*).

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Supporting information

Detailed spectroscopic data and detailed electrochemical measurements, Figures S1–S48 are given in the supplementary material. This material is available free of charge via the Internet at <http://www.worldscinet.com/jpp/jpp.shtml>. Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited at the Cambridge Crystallographic Data Center under deposition number 1916858. Copies can be obtained on request, free-of-charge, at www.ccdc.cam.ac.uk/conts/retrieving.html or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK (fax: +44 1223-336-033 or email: deposit@ccdc.cam.ac.uk).

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