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Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 16 (2006) 2773-2776

## Synthesis and antiproliferative activity of two new tiazofurin analogues with 2'-amido functionalities

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> Received 17 January 2006; revised 1 February 2006; accepted 1 February 2006 Available online 21 February 2006

Abstract—Two novel tiazofurin analogues 2 and 3 were synthesized starting from D-glucose. The key step of the synthesis was the efficient one-step hydrogen sulfide-mediated conversion of 2-azido-3-*O*-acyl-ribofuranosyl cyanides to the corresponding 2-amido thiocarboxamides. Compounds 2 and 3 were evaluated for their in vitro antiproliferative activity against certain human tumour cell lines. Remarkably, compound 2 was found to be 570-fold more potent than tiazofurin against MCF-7 cells, while compound 3 showed the most powerful cytotoxicity against HT-29 cancer cells, being almost 100-fold more active than tiazofurin. © 2006 Elsevier Ltd. All rights reserved.

Tiazofurin (1, Fig. 1) is a synthetic<sup>1</sup> C-nucleoside with a potent antitumour activity.<sup>2</sup> It is a prodrug as once within the cell it is converted to thiazole adenine dinucleotide (TAD) which blocks the key step in the de novo synthesis of GTP.<sup>3</sup> In phase I/II clinical trials, it induced complete haematological remissions in patients with endstage acute nonlymphocytic leukaemia, or in myeloblastic crisis of chronic myeloid leukaemia.<sup>4</sup> Despite the remarkable efficacy of tiazofurin, lack of specificity and a significant neurotoxicity remains a problem in its clinical use.<sup>2</sup> In the search for new antineoplastic agents of improved therapeutic effects, many tiazofurin derivatives were synthesized, including a number of those with a modified sugar segment.<sup>5</sup> However, none of these compounds showed favourable biological properties. We have recently reported on the synthesis of several tiazofurin analogues with modified sugar moieties that showed increased antitumour activities with respect to the lead compound  $1.^{6,7}$  The sub-micromolar activity of 2'-benzamido tiazofurin derivative7 directed our further work in this field to the synthesis and biological evaluation of related molecules bearing different amide functions at C-2'. Herein, we describe the synthesis of two new tiazofurin analogues 2 and 3 with hexan- and



Figure 1. Tiazofurin (1) and analogues 2 and 3.

dodecanamido functions at C-2', along with their effects on the proliferation of some malignant cell lines.

Our strategy for the synthesis of both targets 2 and 3 assumes previous transformation of D-glucose into the ribofuranosyl thioamides 10 and 16 (Scheme 1), followed by their subsequent cyclocondensation with ethyl bromopyruvate to form the thiazole ring. It was further assumed that the key intermediates 10 and 16 could be accessible from the 2-azido-2-deoxy-D-ribofuranosyl cyanides 9 and 15, through a one-step H<sub>2</sub>S-mediated cascade similar to that recently observed in our laboratory.<sup>7</sup> As the previous conversion included a single aromatic ester derivative, further evaluation of this method in the saturated ester series represents an important task of the present work.

The 2,5-anhydro-D-glucose derivative 4, readily available from D-glucose,<sup>8</sup> was used as a convenient starting material for the synthesis of both targets 2 and 3. Com-

*Keywords*: 2,5-Anhydro sugars; *C*-Nucleosides; Antitumour agents; De novo synthesis; Tiazofurin analogues; Thiazoles.

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<sup>0960-894</sup>X/\$ - see front matter @ 2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2006.02.001



Scheme 1. Reagents and conditions: (a)  $C_5H_{11}COCl$ , Py,  $CH_2Cl_2$ , 0 °C  $\rightarrow$  rt, 24 h, 87% of 5, 22 h, 99% of 11; (b) KOBz, DMF, 100 °C, 12 h, 86% of 6, 8 h, 77% of 12; (c) NaN<sub>3</sub>, DMSO, 110 °C, 24 h, 47% of 7, 70 h, 44% of 13; (d) i—4:1 TFA/6 M HCl, 4 °C, 5 days for 7, 8 days for 13, ii—NH<sub>2</sub>OH·HCl, NaOAc, EtOH, rt, 4 h; (e) MsCl, Py, -15 °C  $\rightarrow$  rt, 2 h, 46% of 9, 36% of 15; (f) H<sub>2</sub>S, Py, rt, 12 h, 99% of 10, or H<sub>2</sub>S, DMAP, EtOH, rt, 8 h, 92% of 16.

pound 4 readily reacted with hexanovl chloride under the standard acylation conditions to afford the corresponding 4-O-hexanoyl derivative 5 in 87% yield. Reaction of 5 with potassium benzoate in DMF gave the corresponding 6-O-benzoyl derivative 6 (86%), which was subsequently treated with sodium azide in dimethylsulfoxide to afford the 3-azido derivative 7 in 47% yield. Hydrolytic removal of the dioxolane protective group in 7 was achieved in a mixture of trifluoroacetic acid and 6 M hydrochloric acid at +4 °C. The resulting unstable aldehyde (not shown in the scheme) was not purified, but was rather immediately treated with hydroxylamine hydrochloride to yield the corresponding oxime 8 (68%) as a mixture of the corresponding *E*- and *Z*-isomers. The mixture was not separated (except for the characterisation purpose), but was rather further treated with mesyl chloride to give a 68% yield of the corresponding nitrile 9. Moreover, 9 was treated with hydrogen sulfide in pyridine,<sup>7</sup> to provide an almost quantitative yield of the desired thioamide  $10^{,9}$  a key intermediate in the synthesis of analogue 2.

After the synthesis of thioamide 10, we were also interested in preparation of its six-carbon homologue 16, a key intermediate in the synthesis of dodecan-amido derivative 3. By applying essentially the same multistep sequence as that used for the preparation of 9, compound 4 was converted to the nitrile 15. The conversion of 13 to 15 was carried out without purification of both aldehydo and oximino intermediates to afford the nitrile 15 in 36% overall yield (three steps). Although the nitrile 15 can be converted to the thioamide 16 under the reaction conditions similar to those already used for the conversion of 9 to 10 (H<sub>2</sub>S, Py, rt), at this point we wanted to explore alternative reagent system for the preparation of 16 (H<sub>2</sub>S/DMAP in EtOH), which was also successfully applied in the aromatic ester series.<sup>7</sup> Accordingly, the nitrile 15 was treated with hydrogen sulfide gas and 4dimethylaminopyridine in ethanol, whereby the desired thioamide  $16^{10}$  was obtained in 92% yield.

The last H<sub>2</sub>S-mediated conversion of 2-azido-3-*O*-acyl-2-deoxy-D-ribofuranosyl cyanides **9** and **15** into the corresponding thioamides **10** and **16** deserves some additional comments. This cascade process is presumably comprised of an initial addition of hydrogen sulfide to the nitrile group,<sup>11</sup> followed by the azide reduction and spontaneous *O*,*N*-acyl rearrangement. This sequence already gave good results in the aromatic ester series,<sup>7</sup> but the present work reveals that this method is even more efficient when applied to the saturated ester derivatives. The transformation is likely to be a general method that provides an access to a variety of 2-amidoribofuranosyl-thiocarboxamides, which may be further converted to an array of tiazofurin analogues with different amido functions at the C-2' position.

With the requisite intermediates 10 and 16 in hands, we next focused on their transformation to the target *C*-nucleosides 2 and 3, by using a modified Hantzsch thiazole synthesis, <sup>12</sup> as outlined in Scheme 2. Treatment of both 10 and 16 with ethyl bromopyruvate in refluxing ethanol gave the required thiazoles 17 and 18 in 54% and 56% yield, respectively. Final exposure of 17 and 18 to methanolic ammonia provided the tiazofurin analogues  $2^{13}$  and  $3^{14}$  in 66% and 80% yield, respectively.

The newly synthesized tiazofurin analogues 2 and 3 were evaluated for their antiproliferative activity against human myelogenous leukaemia K562, promyelocytic leukaemia HL-60, colon adenocarcinoma HT-29, estrogen receptor positive breast adenocarcinoma MCF7, cervix carcinoma HeLa and normal foetal lung fibroblasts MRC-5. In vitro cytotoxicity was evaluated after 24-h



Scheme 2. Reagents and conditions: (a) BrCH<sub>2</sub>COCO<sub>2</sub>Et, EtOH, 80 °C, 50 min, 54% of 17, 56% of 18; (b) NH<sub>3</sub>, MeOH, rt, 7 days, 66% of 2, 80% of 3.

cell treatment by using the MTT assay.<sup>15</sup> The results, including the data for the reference compound tiazofurin (1), are presented in Table 1.

Compound 2 was inactive against the K562, HL-60 and HT-29 malignant cell lines, but showed a powerful cytotoxic activity against MCF-7 cells being 570-fold more potent than the reference compound 1. This analogue was also active against HeLa cells, but it was almost 2-fold less potent with respect to the parent molecule 1. However, compound 3 exhibited a significant cytotoxicity against HL-60, HT-29 and HeLa cell lines, but devoid of any activity towards the MCF-7 neoplastic cells. This analogue showed the most pronounced cytotoxicity against the HT-29 cells, being almost 100-fold more potent than tiazofurin. Compound 3 was approximately 2and 1.5-fold more active than tiazofurin against HL-60 and HeLa cells, respectively. It also showed a moderate antiproliferative activity against K562 cells, but it was more than 6-fold less active than the reference compound 1. Remarkably, both analogues were devoid of any cytotoxicity against the normal foetal lung fibroblasts MRC-5. These results do suggest that 2 and 3 are more selective than tiazofurin, but such a generalization should be supported by additional in vitro experiments with a larger number of cell lines.

Due to such a powerful cytotoxicity against certain malignant cell lines, and their nontoxicity towards the normal MRC-5 cells, the analogues 2 and 3 may serve as important leads in the synthesis of more potent and selective anticancer agents. However, the difference in their cytotoxic activity is large and suggests that compounds 2 and 3 are not acting at the same biological target (IMPDH) as tiazofurin.<sup>2,3</sup> In order to explain the biological activity, structure–activity relationship should be performed that would include the synthesis and a more detailed biological evaluation of a series of 2'-amido tiazofurin analogues bearing different R groups in

**Table 1.** In vitro cytotoxicity of 1, 2 and 3

Compound	$IC_{50}^{a}$ ( $\mu$ M)					
	K562	HL 60	HT-29	MCF-7	HeLa	MRC-5
1	5.29	9.32	1.01	7.98	4.76	0.85
2	>100	>100	>100	0.014	8.75	>100
3	35.62	4.82	0.011	>100	3.22	>100

<sup>a</sup>  $IC_{50}$  is the concentration of compound required to inhibit the cell growth by 50% compared to an untreated control.

their amide moiety. Such a study is currently underway in our laboratory the results will be reported elsewhere.

In conclusion, two novel tiazofurin derivatives bearing a hexan- (2), or a dodecanamido function (3) in the C-2' position, were synthesized starting from D-glucose. The key intermediates for this approach, that is, 2-amido-D-ribofuranosyl thiocarboxamides 10 and 16, were easily accessible from the 2-azido-2-deoxy-D-ribofuranosyl cyanides 9 and 15, through the one-step H<sub>2</sub>S-mediated cascade, comprised of an initial addition of hydrogen sulfide to the nitrile group, followed by the azide reduction and spontaneous O,N-acyl rearrangement. The sequence represents a general and particularly versatile approach to a variety of 2-amido-p-ribofuranosyl thiocarboxamides, which can be easily converted to the corresponding ribofuranosyl thiazole-4-carboxamides, as exemplified by the conversion of 10 and 16 to 2 and 3, respectively. Analogues 2 and 3 have shown a potent cytotoxic activity against some human leukaemia and solid tumour cell lines, but did not exhibit any significant cytotoxicity towards normal foetal lung MRC-5 cells. These results, along with our previous findings,<sup>7</sup> confirmed that the introduction of 2'-nitrogen functionalities into the tiazofurin sugar moiety may provide analogues of improved antiproliferative effects against some neoplastic cells, and therefore, it may be of use in the search for new anticancer agents derived from the lead compound 1.

## Acknowledgments

Financial support from the Ministry of Science and Environment Protection of the Republic of Serbia (Project No. 142005) is gratefully acknowledged. The authors thank Mr. D. Djoković (Faculty of Chemistry, University of Belgrade, S&M) for recording the mass spectra.

## **References and notes**

- For recent syntheses of tiazofurin, see: (a) Brown, R. S.; Dowden, J.; Moreau, C.; Potter, B. V. L. Tetrahedron Lett. 2002, 43, 6561; (b) Ramasamy, K. S.; Lau, J. Y. N. Nucleosides Nucleotides Nucleic Acids 2001, 20, 1329; (c) Ramasamy, K. S.; Banderu, R.; Averett, D. J. Org. Chem. 2000, 65, 5849; (d) Ramasamy, K. S.; Averett, D. Nucleosides Nucleotides 1999, 18, 2425.
- 2. Grifantini, M. Curr. Opin. Invest. Drugs 2000, 1, 257.

- 3. (a) Pankiewicz, K. W.; Patterson, S. E.; Black, P. L.; Jayaram, H. N.; Risal, D.; Goldstein, B. M.; Stuyver, L. J.; Schinazi, R. F. Curr. Med. Chem. 2004, 11, 887; (b) Szekeres, T.; Fitzer, M.; Pillwein, K.; Felzmann, T.; Chiba, P. Life Sci. 1992, 51, 1309.
- 4. Malek, K.; Boosalis, M.; Waraska, K.; Mitchell, B. S.; Wright, D. G. Leuk. Res. 2004, 28, 1125, and references therein.
- 5. For recent syntheses of tiazofurin analogues with modified sugar moieties, see: (a) Chun, M. W.; Kim, M. J.; Shin, J. H.; Jeong, L. S. Nucleosides Nucleotides Nucleic Acids 2005, 24, 975; (b) Nair, V.; Wenzel, T. ARKIVOC 2004, 14, 128, <a href="http://arkat-usa.alfahosting.net/ark/journal/">http://arkat-usa.alfahosting.net/ark/journal/</a> 2004/I14 General/1184/04-1184B.pdf>; (c) Liang, C. W.; Kim, M. J.; Jeong, L. S.; Chun, M. W. Nucleosides Nucleotides Nucleic Acids 2003, 22, 2039; (d) Cappellacci, L.; Barboni, G.; Franchetti, P.; Martini, C.; Jayaram, H. N.; Grifantini, M. Nucleosides Nucleotides Nucleic Acids 2003, 22, 869; (e) Zhang, H. Y.; Yu, H. W.; Ma, L. T.; Min, J. M.; Zhang, L. H. Tetrahedron: Asymmetry 1998, 9, 141
- 6. Popsavin, M.; Torović, Lj.; Kojić, V.; Bogdanović, G.; Spaić, S.; Popsavin, V. Bioorg. Med. Chem. Lett. 2003, 13, 3167.
- 7. Popsavin, M.; Torović, Lj.; Kojić, V.; Bogdanović, G.; Popsavin, V. Tetrahedron. Lett. 2004, 45, 7125.
- 8. Popsavin, M.; Popsavin, V.; Vukojević, N.; Csanádi, J.;
- Miljković, D. *Carbohydr. Res.* **1994**, *260*, 145. Selected data for **10** (syrup):  $[\alpha]_D^{25}$  +15.36 (*c* 1.34, CHCl<sub>3</sub>). <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>+D<sub>2</sub>O):  $\delta$  0.84 (t, 3H, J = 7.1 Hz, Me), 1.27 (m, 4H, 2× CH<sub>2</sub>), 1.60 (m, 2H, CH<sub>2</sub>), 2.24 (m, 2H, CH<sub>2</sub>CO), 4.05 (ddd, 1H, J<sub>2,3</sub> = 9.1,  $J_{3,4} = 5.2, J_{3,\text{NH}} = 5.4 \text{ Hz}, \text{ H-3}$ , 4.29 (br s, 1H, exchangeable with D<sub>2</sub>O, OH), 4.36 (m, 1H, H-5), 4.45 (dd, 1H,  $J_{5,6a} = 3.1, J_{6a, 6b} = 12.1 \text{ Hz}, \text{H-}6a), 4.50-4.62 \text{ (m, 2H, H-}4$ and H-6b), 4.68 (d, 1H,  $J_{2,3} = 9.1$  Hz, H-2), 6.86 (d, 1H, exchangeable with D<sub>2</sub>O,  $J_{3,NH} = 5.4$  Hz, CONH), 7.33– 8.10 (m, 5H, Ph), 8.24 and 8.47 (2× br s, 1H each, CSNH<sub>2</sub>); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$  13.81 (Me), 22.22, 25.05 and 31.25 (3× CH<sub>2</sub>), 36.48 (CH<sub>2</sub>CO), 58.95 (C-3), 64.74 (C-6), 71.79 (C-4), 83.99 (C-5), 84.56 (C-2), 128.53, 129.15, 129.62 and 133.47 (Ph), 166.92 (PhC=O), 174.74 (C<sub>5</sub>H<sub>11</sub>C=O), 203.46 (C=S). CI MS: m/z 395  $(M^{+}+H).$
- 10. Selected data for **16**: mp 110.5–111 °C (from CH<sub>2</sub>Cl<sub>2</sub>– hexane),  $[\alpha]_D^{25}$  +15.36 (*c* 1.34, CHCl<sub>3</sub>). <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>):  $\delta$  0.87 (t, 3H, *J* = 6.7 Hz, Me), 1.24 (m, 16H, 8× CH<sub>2</sub>), 1.60 (m, 2H, CH<sub>2</sub>), 2.24 (dd, 2H, J = 7.3, J = 8.2 Hz, CH<sub>2</sub>CO), 4.06 (ddd, 1H,  $J_{2.3} = 9.0$ ,  $J_{3,4} = 5.2, J_{3,\text{NH}} = 5.5 \text{ Hz}, \text{ H-3}$ , 4.13 (br s, 1H, exchangeable with D<sub>2</sub>O, OH), 4.36 (m, 1H, H-5), 4.41-4.61 (m, 3H,  $2 \times$  H-6 and H-4), 4.65 (d, 1H,  $J_{2,3}$  = 9.0 Hz, H-2), 6.85 (br

s, 1H, after treatment with D<sub>2</sub>O changed to d,  $J_{3,\text{NH}} = 5.5 \text{ Hz}$ , CONH), 7.36–8.04 (m, 5H, Ph), 8.21 and 8.47 (2× br s, 1H each, CSNH<sub>2</sub>); <sup>13</sup>C NMR (62.9 MHz, CDCl<sub>3</sub>):  $\delta$  14.00 (Me), 22.56, 25.43, 29.20, 29.22, 29.41, 29.50, 29.52, 29.5 and 31.78 (9× CH<sub>2</sub>), 36.59 (CH<sub>2</sub>CO), 58.99 (C-3), 64.76 (C-6), 71.90 (C-4), 84.05 (C-5), 84.63 (C-2), 128.50, 129.17, 129.61 and 133.43 (Ph), 166.87 (PhC=O), 174.74 (C<sub>11</sub>H<sub>23</sub>C=O), 203.65 (C=S). CI MS: m/z 479 (M<sup>+</sup>+H).

- 11. After shorter reaction time, the main reaction product was accompanied with the thiocarboxamide bearing an unchanged azido group at C-2. This proves that addition of hydrogen sulfide to the nitrile group precedes reduction of the azido function.
- 12. Aguilar, E.; Meyers, A. I. Tetrahedron Lett. 1994, 35, 2773.
- 13. Selected data for 2: mp 155–156 °C (from MeOH–<sup>1</sup>Pr<sub>2</sub>O),  $[\alpha]_D^{25}$  -16.44 (c 1.11, MeOH). <sup>1</sup>H NMR (250 MHz, methanol- $d_4$ ):  $\delta$  0.85 (t, 3H, J = 7.1 Hz, Me), 1.16–1.36 (m, 4H, 2× CH<sub>2</sub>), 1.56 (m, 2H, CH<sub>2</sub>), 2.24 (t, 2H, J = 7.7 Hz, CH<sub>2</sub>CO), 3.74 (d, 2H,  $J_{4',5'} = 4.6$  Hz, 2× H-5'), 4.13 (m, 1H, H-4'), 4.26 (dd, 1H,  $J_{2',3'} = 5.4$ ,  $J_{3',4'} = 2.4$  Hz, H-3'), 4.53 (dd, 1H,  $J_{1',2'} = 8.7$ ,  $J_{2',3'} = 5.4$  Hz, H-2'), 5.10 (d, 1H,  $J_{1',2'} = 8.7$  Hz, H-1'), 7.56 and 7.88 (2× br s, 0.4H each, CONH<sub>2</sub>), 8.12 (d, 0.3H, J = 8.9 Hz, NH), 8.21 (s, 1H, H-5); <sup>13</sup>C NMR (62.9 MHz, methanol- $d_4$ ):  $\delta$  14.30 (Me), 23.35, 26.59 and 32.30 (3× CH<sub>2</sub>), 36.89 (CH<sub>2</sub>CO), 60.02 (C-2'), 63.44 (C-5'), 72.70 (C-3'), 80.61 (C-1'), 88.79 (C-4'), 126.32 (C-5), 150.33 (C-4), 165.67 (C-2), 172.23 and 176.78 (2× C=O); CI MS: m/z 358 ( $M^+$ +H); Anal. Calcd for  $C_{15}H_{23}N_3O_5S$ : C, 50.40; H, 6.49, N, 11.76; S, 8.97. Found: C, 50.12; H, 6.21, N, 11.96; S, 8.66.
- 14. Selected data for 3: mp 133.5-134.5 °C (from MeOH-<sup>i</sup>Pr<sub>2</sub>O),  $[\alpha]_{D}^{25}$  -17.03 (c 0.88, MeOH). <sup>1</sup>H NMR (250 MHz, methanol- $d_4$ ):  $\delta$  0.87 (t, 3H, J = 7.0 Hz, Me), 1.18–1.62 (m, 18H, 9× CH<sub>2</sub>), 2.25 (t, 2H, J = 7.3 Hz, CH<sub>2</sub>CO), 3.74 (d, 2H,  $J_{4',5'} = 4.5$  Hz, 2× H-5'), 4.12 (m, 1H, H-4'), 4.26 (dd, 1H,  $J_{2',3'} = 5.1$ ,  $J_{3',4'} = 2.2$  Hz, H-3'), 4.52 (dd, 1H,  $J_{1',2'} = 8.7$ ,  $J_{2',3'} = 5.1$  Hz, H-2'), 5.09 (d, 1H,  $J_{1',2'} = 8.7$  Hz, H-1'), 8.21 (s, 1H, H-5); <sup>13</sup>C NMR (62.9 MHz, methanol- $d_4$ ):  $\delta$  14.41 (Me), 23.61, 26.86, 30.10, 30.32, 30.46, 30.59 and 32.93 (9× CH<sub>2</sub>), 36.94 (CH<sub>2</sub>CO), 59.92 (C-2'), 63.42 (C-5'), 72.65 (C-3'), 80.66 (C-1'), 88.74 (C-4'), 126.27 (C-5), 150.36 (C-4), 165.59 (C-2), 172.17 and 176.69 (2× C=O); CI MS: m/z 442 (M<sup>+</sup>+H); Anal. Calcd for C<sub>21</sub>H<sub>35</sub>N<sub>3</sub>O<sub>5</sub>S: C, 57.12; H, 7.99, N, 9.52; S, 7.26. Found: C, 56.92; H, 7.77, N, 9.86; S, 6.90.
- 15. Scudiero, D. A.; Shoemaker, R. H.; Paull, K. D.; Monks, A.; Tierney, S.; Nofziger, T. H.; Currens, M. J.; Seniff, D.; Boyd, M. R. Cancer. Res. 1988, 48, 4827.