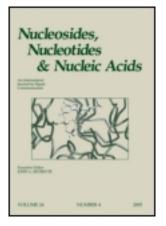
This article was downloaded by: [University of Arizona] On: 23 November 2012, At: 05:40 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/lncn20

Fluorinated Peptide Nucleic Acid

Marcel Hollenstein ^{a b}, Daniel Gautschi ^a & Christian J. Leumann ^a

^a Department of Chemistry and Biochemistry, University of Bern, Bern, Switzerland

^b Department of Chemistry and Biochemistry, University of Bern, Freiestrasse 3, CH-3012, Bern, Switzerland

Version of record first published: 31 Aug 2006.

To cite this article: Marcel Hollenstein, Daniel Gautschi & Christian J. Leumann (2003): Fluorinated Peptide Nucleic Acid, Nucleosides, Nucleotides and Nucleic Acids, 22:5-8, 1191-1194

To link to this article: <u>http://dx.doi.org/10.1081/NCN-120022833</u>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-conditions</u>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

NUCLEOSIDES, NUCLEOTIDES & NUCLEIC ACIDS Vol. 22, Nos. 5–8, pp. 1191–1194, 2003

Fluorinated Peptide Nucleic Acid

Marcel Hollenstein,* Daniel Gautschi, and Christian J. Leumann

Department of Chemistry and Biochemistry, University of Bern, Bern, Switzerland

ABSTRACT

The fluorinated olefinic peptide nucleic acid analogue (F-OPA) monomer containing the base thymine was synthesised in 13 steps. PNAs containing this unit were prepared and their pairing properties assessed by means of UV-melting experiments.

Polyamide or peptide nucleic acids 1, first described in 1991, are DNA analogues entirely based on an achiral polyamide backbone.^[1] The PNAs undergo sequencespecific and efficient Watson-Crick base pairing with complementary DNA and RNA.^[2,3] One structural feature of PNA is the central amide linker connecting the base to the backbone. The carbonyl oxygens of this unit, uniformly point towards the carboxy termini in PNA/DNA,^[4,5] PNA/RNA^[6] and PNA/PNA^[7] complexes, whereas both rotameric forms co-exist in the free monomer. In order to elucidate this structural ambiguity, the olefinic peptide nucleic acids (OPAs) have been synthesised and studied (Fig. 1).^[3] Fully modified OPA oligoamides resulted in a marked decrease in affinity towards complementary DNA, compared to PNA. In order to investigate the effect of the dipole moment of the linker carboxy group while

1191

DOI: 10.1081/NCN-120022833 Copyright © 2003 by Marcel Dekker, Inc.

Downloaded by [University of Arizona] at 05:40 23 November 2012

1525-7770 (Print); 1532-2335 (Online) www.dekker.com

^{*}Correspondence: Marcel Hollenstein, Department of Chemistry and Biochemistry, University of Bern, Freiestrasse 3, CH-3012 Bern, Switzerland; E-mail: hollenstein@ioc. unibe.ch.

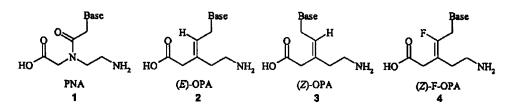
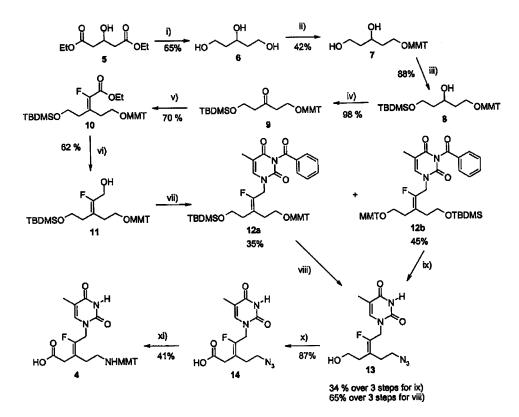


Figure 1. Chemical structure of the monomeric units of the different peptide nucleic acids.

maintaining the geometry of the C=C double bond, (Z) -t-F-OPA 4 was synthesised and incorporated into PNA.

The synthesis of the monomeric unit **4** containing the base thymine is outlined in Sch. 1.



Scheme 1. i) LiAlH₄, THF, RT, 3 h. ii) MMTrCl, Pyridine, RT, overnight. iii) TBDMSCl, Pyridine, RT, overnight. iv) IBX, THF/DMSO 1/1, RT, 6 h. v) 1) n-BuLi, $(EtO)_2P(O)-CHFCO_2Et$, THF, $-78^{\circ}C$, 2 h. 2) Ketone, $-78^{\circ}C$ to RT, 4 h. vi) LiAlH₄, Et₂O, RT, 2 h. vii) TBz, PPh₃, DIAD, THF, RT, overnight. viii) 1) BCl₃, CH₂Cl₂, $-40^{\circ}C$, 30 min. 2) LiN₃, PPh₃, CBr₄, DMF, RT, overnight. 3) TBAF, THF, RT, overnight. ix) 1) TBAF, THF, RT, 5 h. 2) LiN₃, PPh₃, CBr₄, DMF, RT, overnight. 3) BCl₃, CH₂Cl₂, $-40^{\circ}C$, 30 min. x) 1) Dess-Martin, CH₂Cl₂, RT. 2) NaClO₂, NaH₂PO₄, 2-methyl-2-butene, t-BuOH. xi) 1) Lindlar catalyst, H₂, RT. 2) MMTrCl, Pyridine, RT.

Table 1. Mass spectrometry data and Tm values [°C] (UV-melting curves, 260 nm) of PNA sequences containing (*E*)-t-OPA, (*Z*)-t-OPA or (*Z*)-t-F-OPA units with parallel and antiparallel DNA (c=4 μ M in 100 mM NaCl, 10 mM Na₂HPO₄, pH 7.0). Lowercase letters: PNA units; t^{*Z*} = (*Z*)-t-OPA, t^{*E*} = (*E*)-t-OPA and t^{*F*} = (*Z*)-t-F-OPA.

		m/z calcd	m/z found (ESI ⁺ -TOF)	$T_{\rm m}$ (antiparallel DNA) ^a	$T_{\rm m}$ (parallel DNA) ^b
15	Lys-ttttaatata-Gly-NH ₂	2900.9	2900.10	33.2	< 0
	Lys-ttttaat ^E ata-Gly-NH ₂		2883.30	36.7	n.d. ^c
17	Lys-ttttaat ^z ata-Gly-NH ₂	2883.9	2883.32	28.0	< 0
18	Lys-ttttaat ^F ata-Gly-NH ₂	2901.9	2901.26	35.6	n.d.
19	Lys-tttt ^E aatata-Gly-NH ₂	2883.9	2883.17	30.0	11.0, 34.0
20	Lys-ttt ^E taatata-Gly-NH ₂	2883.9	2883.19	28.1	n.d.

^ad(AAAATTATAT).

^bd(TATATTAAAA).

^cNot determined.

Downloaded by [University of Arizona] at 05:40 23 November 2012

In order to study the pairing properties, oligomers **15–20** were prepared and the stability of the duplexes formed with anti-parallel and parallel DNA was assessed by means of UV-melting curves (Table 1). Introduction of the modified units leads to a marked difference in $T_{\rm m}$ as a function of the position of the modification in the sequence. Indeed, positioning of a (*E*)-t-OPA unit between 2 purine bases leads to a stabilisation of the duplex ($\Delta T_{\rm m} = +3.5^{\circ}$ C), while introduction of this unit between 2 pyrimidine units leads to a marked destabilisation ($\Delta T_{\rm m} = -5.1^{\circ}$ C). Positioning between one pyrimidine and one purine base leads, as expected, to an intermediate value ($\Delta T_{\rm m} = -3.2^{\circ}$ C). The (*Z*)-t-F-OPA modification leads to a stabilisation comparable to the one observed for (*E*)-t-OPA ($\Delta T_{\rm m} = +2.4^{\circ}$ C), whereas a substantial decrease of duplex stability is observed for the (*Z*)-t-OPA unit ($\Delta T_{\rm m} = -5.2^{\circ}$ C).

The introduction of the fluorine atom at that location could alter the electrostatic properties and result in a reduced stacking ability. This could account for the lower T_m value obtained for oligomer **18** compared to the one for oligomer **16**. However, the effect on the dipole moment on the whole oligomer is yet unknown, and only a fully modified (Z)-t-F-OPA strand could provide with an answer.

REFERENCES

- Nielsen, P.E.; Egholm, M.; Berg, R.H.; Burchardt, O. Science 1991, 254, 1497– 1500.
- Egholm, M.; Burchardt, O.; Christensen, L.; Behrens, C.; Freier, S.M.; Driver, D.A.; Berg, R.H.; Kim, S.K.; Norden, B.; Nielsen, P.E. Nature 1993, 365, 566–568.
- 3. Schütz, R.; Cantin, M.; Roberts, C.; Greiner, B.; Uhlmann, E.; Leumann, C. Angew. Chem. Int. Ed. **2000**, *39*, 1250–1253.
- 4. Betts, L.; Josey, J.A.; Veal, J.M.; Jordan, S.R. Science 1995, 270, 1838–1841.

1193

- Leijon, M.; Gräslund, A.; Nielsen, P.E.; Burchardt, O.; Nordén, B.; Kristensen, S.M.; Eriksson, M. Biochemistry 1994, 33, 9820–9825.
- 6. Brown, S.C.; Thomson, S.A.; Veal, J.M.; Davis, D.G. Science 1994, 265, 777–780.
- 7. Rasmussen, H.; Kastrup, J.S.; Nielsen, J.N.; Nielsen, J.M.; Nielsen, P.E. Nature Struct. Biol. **1997**, *4*, 98–101.