# Journal of Medicinal Chemistry

Article

Subscriber access provided by The Library | University of Bath

# Design, Synthesis, and Biological Evaluation of Pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-diones as Potent and Selective Epidermal Growth Factor Receptor (EGFR) Inhibitors against L858R/T790M Resistance Mutation

Yongjia Hao, Jiankun Lyu, Rong Qu, Yi Tong, Deheng Sun, Fang Feng, Linjiang Tong, Tingyuan Yang, Zhenjiang Zhao, Lili Zhu, Jian Ding, Yufang Xu, Hua Xie, and Honglin Li *J. Med. Chem.*, Just Accepted Manuscript • DOI: 10.1021/acs.jmedchem.8b00346 • Publication Date (Web): 15 Jun 2018 Downloaded from http://pubs.acs.org on June 16, 2018

### **Just Accepted**

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

# Design, Synthesis, and Biological Evaluation of Pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-diones as Potent and Selective Epidermal Growth Factor Receptor (EGFR) Inhibitors against L858R/T790M Resistance Mutation

Yongjia Hao<sup>1,2,‡</sup>, Jiankun Lyu<sup>1,‡</sup>, Rong Qu<sup>3,‡</sup>, Yi Tong<sup>1</sup>, Deheng Sun<sup>1</sup>, Fang Feng<sup>3</sup>, Linjiang Tong<sup>3</sup>, Tingyuan Yang<sup>1</sup>, Zhenjiang Zhao<sup>1</sup>, Lili Zhu<sup>1</sup>, Jian Ding<sup>3</sup>, Yufang Xu<sup>1,\*</sup>, Hua Xie<sup>3,\*</sup>, Honglin Li<sup>1,\*</sup>

1 Shanghai Key Laboratory of New Drug Design, State Key Laboratory of Bioreactor Engineering, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China.

2 School of Pharmacy, Guizhou University of Chinese Medicine, Guiyang 550025, China.

3 Division of Anti-tumor Pharmacology, State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China.

<sup>‡</sup>Authors contributed equally to this work

\* To whom correspondence should be addressed. Email: yfxu@ecust.edu.cn, hxie@jding.dhs.org, hlli@ecust.edu.cn

#### ABSTRACT

First-generation epidermal growth factor receptor (EGFR) inhibitors, gefitinib and erlotinib have achieved initially marked clinical efficacy for non-small cell lung cancer (NSCLC) patients with EGFR activating mutations. However, their clinical benefit was limited by the emergence of acquired resistance mutations. In most cases (approximately 60%), the resistance was caused by the secondary EGFR T790M gatekeeper mutation. Thus, it is still desirable to develop novel third-generation EGFR inhibitors to overcome T790M mutation while sparing wild-type (WT) EGFR. Herein, a series of pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-dione derivatives were designed and synthesized. Among which, the most potent compound **20g** not only demonstrated significant inhibitory activity and selectivity for EGFR<sup>L858R/T790M</sup> and H1975 cells *in vitro*, but also displayed outstanding antitumor efficiency in H1975 xenograft mouse model. The encouraging mutant-selective results at both *in vitro* and *in vivo* levels suggested that **20g** might be used as a promising lead compound for further structural optimization as potent and selective EGFR<sup>L858R/T790M</sup> inhibitors.

#### INTRODUCTION

Lung cancer, especially non-small cell lung cancer (NSCLC) is one of the most prevalent malignant tumors, and also a major health problem worldwide, whose five-year survival rate is less than 15% after diagnosis.<sup>1</sup> It is well-known that the epidermal growth factor receptor (EGFR, also known as ErbB1, HER1)-mediated signaling pathways are essential for various biological processes and cancers development and progression, such as cell proliferation, adhesion, migration, differentiation and survival.<sup>2-4</sup> Thus, the EGFR tyrosine kinase has been proven to be an attractive and clinically validated drug target in cancer therapy, in particular for NSCLC treatment. First-generation reversible and guinazoline-based EGFR tyrosine kinase inhibitors (TKIs), including gefitinib (1) and erlotinib (2), have received approval from US Food and Drug Administration (FDA) for the treatment of NSCLC in 2003 and 2004, respectively (Figure 1).<sup>5,6</sup> Despite the clinical efficacy of the two drugs have been well established for NSCLC patients with the most common EGFR activating mutations, such as L858R (activating mutation in exon 21) and del E746–A750 (deletions in exon 19).<sup>7-9</sup> Unfortunately, a secondary T790M point mutation (the substitution threonine790 with methionine residue) at the gatekeeper position of EGFR has caused approximately 60% NSCLC patients who initially responded to the treatment of first-generation EGFR inhibitors to develop acquired resistance after a median of 10-14 months.<sup>10</sup> Occurrence of the T790M acquired resistance mutation not only seriously affected the improvement of survival rates, but also impaired the quality of life of NSCLC patients.

In order to prevent the acquired resistance caused by the T790M mutation, several secondgeneration irreversible EGFR inhibitors have been developed.<sup>11-14</sup> Afatinib (**3**) is a typical example, which has been approved by FDA as a new first-line treatment for late stage

(metastatic) NSCLC patients with EGFR exon 19 deletions or exon 21 L858R substitution mutations in 2013 (Figure 1).<sup>15</sup> Although afatinib was highly active *in vivo* in xenograft models driven by EGFR<sup>L858R/T790M</sup>, it didn't increase objective response rate in clinical trial of NSCLC patients who have been developed resistance to gefitinib or erlotinib on account of the narrow therapeutic window towards wild-type (WT) EGFR.<sup>16</sup> Therefore, there is still an unmet clinical need to develop third-generation EGFR TKIs, namely, mutant-selective EGFR-TKIs. These inhibitors have potent selectivity for EGFR T790M while sparing wild-type EGFR, so can improve the efficacy and reduce the toxicity associated with the inhibition of WT EGFR protein. WZ4002  $(4)^{17}$  with an aniline pyrimidine core was the first such type of mutant-selective EGFR-TKIs, nevertheless, which did not progress into clinical development. Simultaneously, another two mutant-selective small molecule EGFR inhibitors have been publicly reported, that are CO-1686 (5, Rociletinib)<sup>18,19</sup> and AZD9291 (6, Osimertinib)<sup>20,21</sup>. These two agents demonstrated good potency and prominent selectivity for EGFR L858R/T790M over WT EGFR (Figure 1). Furthermore, according to its significant clinical benefits, FDA has accelerated approved compound 6 to treat advanced NSCLC patients with EGFR<sup>T790M</sup> acquired resistance mutation, who have progressed on or after prior treatment with other EGFR-targeted therapy.<sup>22</sup>

#### Figure 1

Recently, we reported a study of developing a series of mutant-selective EGFR inhibitors with a C4-alkyl-1,4-dihydro-2*H*-pyrimido[4,5-*d*][1,3]oxazin-2-ones scaffold.<sup>23</sup> By a 3D-comformer based scaffold hopping strategy, we designed a new scaffold of pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-dione as a starting point to develop a series of novel EGFR<sup>L858R/T790M</sup> selective Page 5 of 52

inhibitors. At the end of a 3-round structural optimization, our candidate compound **20g** displays a favorable selectivity against double-mutant EGFR at both enzymatic and cellular levels. Compound **20g** also shows a good selectivity profile in *in vivo* study. Our study provides a new scaffold of mutant-selective EGFR inhibitors by an efficient *in-silico* scheme.

#### Figure 2

#### CHEMISTRY

As shown in Scheme 1, compounds **14a-e** were synthesized starting from 1,3-benzendiamine, followed by treatment with di-*tert*-butyl dicarbonate to give Boc-protected amine (**8**). Then, compound (**8**) was reacted with ethyl 2,4-dichloropyrimidine-5-carboxylate to yield ethyl 4-((3-((*tert*-butoxycarbonyl)amino)phenyl)amino)-2-chloropyrimidine-5-carboxylate (**9**) in a good yield. Hydrolysis of the carboxylic ethyl ester group by 1 M NaOH to afford carboxylic acid (**10**), and the key intermediates (**11**) were generated with arylamines by replacement of the chloride in the pyrimidine. The amide derivatives (**12**) were prepared from condensation carboxylic acid derivatives (**11**) with methylamine hydrochloride. Direct cyclization with 1,1'-carbonyldiimidazole to obtain the ring-closure intermediates (**13**). Finally, **13** were deprotected with trifluoroacetic acid in dichloromethane and reacted with acryloyl chloride to produce the target compounds **14a-e**.

#### Scheme 1

To compare the influence of activity, we synthesized a series of final compounds with different substituted alkyl groups at 3-position of pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-dione moiety (Scheme 2). Commercially available 2,4-dichloro-5-pyrimidinecarbonyl chloride was reacted with ammonium hydroxide to give 2,4-dichloropyrimidine-5-carboxamide (**15**). Intermediates (**17**) were generated with Boc-protected amine and appropriate arylamines by replacement of the chloride at 4, 2-position of pyrimidine, respectively. Then, the ring-closure compounds (**18**) were obtained based on the same procedure as described in Scheme 1, which further were alkylated with appropriate alkyl halides to afford the N3-alkylated compounds (**19**).<sup>24</sup> At last, the final compounds **20a**–**f** were synthesized using the same synthetic methods as described above. And compounds **20g-1** were prepared by a method similar to that for compound **20a**.

#### Scheme 2

#### **RESULTS AND DISCUSSION**

**Scaffold Hopping and Binding Mode Analysis.** Through a structure-based design strategy targeting a hydrophobic sub-pocket induced by a gatekeeper mutant T790M, compound 7 (Figure 2A) was previously identified as a nanomolar irreversible EGFR inhibitor with a double-mutant selective profile (IC<sub>50</sub> against EGFR<sup>L858R/T790M</sup> kinase over EGFR<sup>WT</sup> kinase: 4.7 nM vs 474 nM, 101-fold selectivity).<sup>25</sup> After superimposing the docked pose of compound 7 with another double-mutant-selective EGFR inhibitor **5** (Figure 2B), the binding mode of both double-mutant inhibitors share some similar characteristics: 1) The aminopyrimidine moiety of both inhibitors binds to Met793 residue at the hinge region through two hydrogen bonding

interactions. 2) The hydrophilic tail on the left-hand side faces toward solvent-exposure area. 3) The warhead, meta-acrylamide group, forms a covalent bond with Cys797. Besides, there are two sub-pockets induced by gatekeeper mutant T790M (Figure 2C). Based on the overlaid docked poses of compounds **5** and **7**, sub-pocket 1 (S1) is occupied by the fluorine group in the C5-CF<sub>3</sub> substituent of compound **5** or the methyl group at N5-position of compound **7**, respectively. In order to further improve the selectivity profile, by occupying both mutant-induced sub-pockets (S1 and S2), compound **14a** was designed with a novel scaffold: pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-dione. According to the docked pose of compound **5** and **7**. And hydrophobic groups with different lengths and volumes at the R<sup>3</sup> position could be introduced to explore the S2 pocket and to strengthen the nonpolar contacts with the gatekeeper mutant Met790.

Structure-activity Relationship. 14a inhibited the enzymatic activity against EGFR<sup>L858R/T790M</sup> and EGFR<sup>WT</sup> with IC<sub>50</sub> values of 46 and 180 nM, respectively, and showed nearly 4-fold selectivity for EGFR<sup>L858R/T790M</sup> in comparison with EGFR<sup>WT</sup> (Table 1). Its *in vitro* antiproliferation activity were tested by using the sulforhodamine B (SRB) colorimetric assay on H1975 cell lines (expressing EGFR-L858R/T790M) and A431 cell lines (harboring overexpressed EGFR<sup>WT</sup>). However, it exhibited weak antiproliferative effects on both the H1975 and A431 cell lines (> 10  $\mu$ M). Thus, we decided to improve the physical properties of 14a by exploring the  $R^2$  position. With a *N*.*N*-dimethylpiperidin-4-amine substituent at the  $R^2$  position (compound 14d), the predicted cell permeability score was improved from 64.1 (14a) to 80.3 (14d) (the higher value, the better cell permeability, predicted by Qikprop in Maestro software package<sup>26</sup>). As we expected, the enzymatic activity of 14d against EGFR kinases and the

selectivity between EGFR<sup>L858R/T790M</sup> and EGFR<sup>WT</sup> remain the same level as **14a** (Table 1). Besides, compound **14d** displayed potent inhibitory activity against H1975 cell line (230 nM), and showed selectivity against the H1975 over the A431 cell lines (approximately 17-fold). As a contrast, we also synthesized compound **14e** with no modification on R<sup>1</sup> and R<sup>2</sup>. As expected, **14e** lost the selectivity between EGFR<sup>L858R/T790M</sup> and EGFR<sup>WT</sup>, and showed weak antiproliferative effects on both the H1975 and A431 cell lines (> 9  $\mu$ M). The results once again verify the importance of the hydrophilic tail on the left-hand side. Since compound **14d** showed better inhibitory activity at both enzymatic and cellular levels, and acceptable aqueous solubility (531  $\mu$ g/mL), it was selected as a new starting point for the next-round structure modifications.

#### Table 1

Based on the strategy we described in Figure 2D, probing the S2 pocket is one of the most important structure-based optimizations to improve the selectivity against double mutants over wild type. Considering the feasibility of synthesis, hydrophobic groups with varied length and volume were introduced at  $R^3$  position (Table 2). Thus, compounds **20a-f** were synthesized (Scheme 2). When varying the substituted group at  $R^3$  position from methyl, ethyl, propyl to isopropyl (compounds **14d**, **20a-c**), the enzymatic inhibition against double-mutant EGFR also boosted correspondingly with the increase of group length and volume. For compounds **20b** and **20c**, their values of IC<sub>50</sub> against double-mutant EGFR reach the subnanomolar range (0.4 and 0.8 nM, respectively, Table 2). Based on the docking simulation (Figure 2E), the hydrophobic groups at  $R^3$  position is able to occupy the T790M mutant induced S2 pocket as well as strengthen van der Waals (VDWs) contacts with gatekeeper Met790. Compared with the positive

control compound 5, all three compounds (20a-c) showed a double-mutant preferred selectivity (14-fold vs. 44, 33 and 25-fold), but the longer and bulkier substituents at  $R^3$  position (compounds **20d-f**) keep potent inhibitory activity against double-mutant EGFR, these structural modifications also increase the inhibition against wild-type EGFR, resulting in complete loss of double-mutant selective profile (0.6, 0.7, 2.7-fold, respectively). There are many structures that show the flexibility of S2 pocket to accommodate a wide range of hydrophobic moieties in both EGFR double-mutant and wild-type structures.<sup>27,28</sup> According to our docking simulations, compound **20f** is able to be docked into both double-mutant and wild-type EGFR structures with a wide open S2 pocket (Figure 2F and Figure S1). Based on the docked pose of **20f** with doublemutant EGFR, we believe that the energy gain from occupying at S2 pocket with bulkier group compensates the steric strain (1.9 Å: the distance in Figure 2F shows unfavorable VDW interactions) between the carbonyl group at C4-position and wild type gatekeeper Thr790. Thus, compounds with longer and bulkier substituents at R<sup>3</sup> position regain inhibitory activity against wild-type EGFR. At the second-round optimization, compounds 20a-c also showed a better selectivity against H1975 cell lines over A431 cell lines than that of compounds **20d-f** (12, 11 and 4-fold, respectively). This trend is in consistent with that at the enzymatic level.

At the third round, we moved forward with the most selective one at cellular level (**20c**) for further structural optimizations. Previously, some literatures have also reported to modify the left-hand side chain of the compounds to discovery active candidates.<sup>29-31</sup> Inspired by that, we synthesized a set of compounds (compounds **20g-1**) that possess different left-hand side chains on the basis of **20c** (Table 2). The data revealed that compounds **20h**, **20j-1** displayed varying degrees selectivity losses in EGFR enzymatic inhibitory activity compared with **20c**. Encouragingly, compounds **20g** and **20i** demonstrated more potent selectivity for double-mutant

EGFR enzyme (263-fold and 34-fold, respectively) and increase of aqueous solubility (262  $\mu$ g/mL and 280  $\mu$ g/mL, respectively) than **20c**. Especially **20g**, which showed 263-fold selectivity between the EGFR<sup>L858R/T790M</sup> and the EGFR<sup>WT</sup>, and showed more potent inhibitory activity and selectivity than the positive control compound **5**. In consistent with their enzyme inhibitory activity, these compounds also showed different results in antiproliferation assays. Compounds **20g-i** were roughly equipotent to **20c** for the H1975 cell lines, but resulted in varying degrees drops of selectivity for A431 cells, especially **20h** and **20i**. And **20j-i** exhibited diminished antiproliferation activity for H1975 cells and increased antiproliferation efficiency for A431 cells as compared to **20c** (Table 2).

#### Table 2

**Inhibitory Effects of EGFR and Downstream Signaling Transduction.** The selective inhibition of the representative compound **20g** for the phosphorylation of EGFR and the downstream signaling transduction was further confirmed by using the Western blot analysis (Figure 3). In H1975 cells, **20g** inhibited the phosphorylation of EGFR<sup>L858R/T790M</sup> at the concentration as low as 1 nM, and potently inhibited EGFR phosphorylation and downstream molecules AKT phosphorylation at 100 nM, which even showed more potent inhibitory activity relative to compound **6** (Figure 3A). In A431cells, **20g** moderately inhibited the phosphorylation of EGFR<sup>WT</sup> even at 100 nM (Figure 3B). The results clearly showed that compound **20g** more potently inhibited the phosphorylation of EGFR and downstream signaling transduction (p-AKT) in cellular level, further suggesting **20g** was a potent and selective EGFR<sup>L858R/T790M</sup> inhibitor.

#### Figure 3

**Target Duration of Action of Compound 20g in Vitro**. In order to get an insight into the duration of target inhibition *in vitro*, we performed washout experiment. Compound **20g** was washed out after incubation for 2 h in H1975 cells. Then phosphorylation of target at different time points was detected by Western blot. As shown in Figure 4A, inhibition of target EGFR<sup>L858R/T790M</sup> activation can last for 24 h at least. And inhibition of the signaling protein ERK activation can also last for more than 12 h. Furthermore, the target duration was similar to that of control compound **6** (Figure 4B).

#### Figure 4

**DMPK Properties of Compound 20g.** In the permeability properties assay, compound **20g** showed modest permeability in the MDCK II cells ( $P_{app}$ ,  $_{A\to B}$  of  $1.5 \times 10^{-6}$  cm/s, and  $P_{app}$  ratio of 9.1 at 2 µM). We then examined the *in vitro* metabolic stability of compound **20g** with human and mouse liver microsomes. At the assay concentration of 0.1 µM, the results demonstrated that **20g** exhibited acceptable microsomal stability with half-life ( $T_{1/2}$ ) of 85.4 min (human liver microsomes) and 46.1 min (mouse liver microsomes), and MF% of 44.3 and 49.4, respectively (Table S1). In addition, **20g** showed weak cytochrome P450 (CYP450) inhibition against five major human CYP450 enzymes (1A2, 2C9, 2C19, and 2D6: IC<sub>50</sub> > 50 µM; 3A4: IC<sub>50</sub> = 23.9 µM). We also examined the preliminary *in vivo* pharmacokinetic properties of compound **20g** in mice following intravenous (IV) and oral administration. As shown in Table 3, **20g** demonstrated a reasonable area under the curve (AUC<sub>0-∞</sub>) value of 658 ng·h/mL, moderate half-life ( $T_{1/2}$ =1.1 h) and oral bioavailability (F = 35%) during the oral dose of 10 mg/kg. The results indicate that it is meaningful to further evaluate the preliminary *in vivo* antitumor efficacy studies of compound

20g.

#### Table 3

In Vivo Antitumor Efficiency Study. On the basis of its outstanding *in vitro* inhibitory activity, we selected compound **20g** to further evaluate its preliminary *in vivo* antitumor efficacy in H1975 and A431 xenograft mouse models (Figure 5). In this study, 4-6 weeks-old mice (n = 6/group) in treatment group were dosed orally once daily with **20g** at 50 mg/kg for 14 days. As shown in Figure 5A, compared with the vehicle group, **20g** significantly inhibited the tumor growth (TGI = 73%, p<0.001) during the 14 day period in H1975 xenograft mouse mode. However, **20g** exhibited much less potent tumor growth inhibition (TGI = 37%, p<0.05) in A431 xenograft mouse side effects were observed (Figure 5C and 5D), indicating that compound **20g** was well-tolerated at this dose.

#### Figure 5

#### CONCLUSIONS

Through a 3D conformer based scaffold hopping protocol based on previously reported mutantselective EGFR inhibitors, we designed and synthesized a series of pyrimido[4,5-*d*]pyrimidine-2,4(1*H*,3*H*)-dione derivatives as novel potent and selective EGFR<sup>L858R/T790M</sup> inhibitors. For three compounds (**20b**, **20c** and **20g**) in this series, our design strategy to occupy S2 pocket boosts the inhibitions against double-mutant EGFR and brings the IC<sub>50</sub> to a subnanomolar level. After three

round SAR explorations, one of the most promising compounds **20g** shows a favorable selectivity at both *in vitro* and *in vivo* levels, indicating that compound **20g** might be used as a promising drug candidate to overcome EGFR<sup>L858R/T790M</sup> drug-resistance mutation. Further evaluation for the candidate compound is still ongoing, and will be reported in due course.

#### **EXPERIMENTAL SECTION**

**Molecular Modeling.** The X-ray structures (PDB code: 3IKA and 4G5J) were downloaded from the Protein Data Bank (PDB, <u>http://www.pdb.org</u>). The protocol of comparative docking study was reported in our previous publications<sup>23</sup>: First, compounds **5** and **7** was docked into the EGFR<sup>T790M</sup> (PDB code: 3IKA) as reversible inhibitors using Glide<sup>23</sup> (version 5.5) in extra precision (XP) mode with default settings; Then, the covalent bond between the Cys797 and the electrophilic group was formed by the build panel in Maestro<sup>26</sup>; Finally, to fix the bond lengths and eliminate steric clashed, the inhibitor-kinase covalent complex was minimized in water solvent using MacroModel application with a flexible ligand and a restrained protein.

The scaffold hopping was conducted by an in-house software: SHAFTS<sup>32</sup>. The query molecule is the docked pose of compound **7** obtained from the above docking study. The aligned pose of compound **20c** (similarity score is 1.2875) was generated by a fast 3D-similarity calculation in SHAFTS<sup>33</sup>. Then, formed the covalent bond and minimized complexes with MacroModel application with the same settings as described above.

For getting docked poses of compound **20f** with wild-type and double-mutant EGFRs. Two structures (PDB code: 2RGP and 3W2S) were prepared in Maestro and aligned to the structure with PDB code 3IKA. Compound **20f** (similarity score is 1.4038) was aligned to the docked pose

of compound 7 by SHAFTS. Then manually formed the covalent bond and minimized in MacroModel.

**Reagents and General Methods.** All chemical reagents and solvents used in experiments were purchased from commercial suppliers and used without further purification. Silica gel (300–400 mesh) used for column chromatography and thin-layer chromatography (TLC) used to monitor progress of the reaction were purchased from Qingdao Haiyang Chemical Co.,Ltd. <sup>1</sup>H and <sup>13</sup>C NMR spectra were performed on the Bruker spectrometer (AV-400) at 400 MHz and 100 MHz, respectively. The low-resolution mass spectra (LC-MS) were measured using Agilent 6120 LC-MS. The high-resolution mass spectra (HRMS, Waters LCT Premier XE TOF) were measured at the Institute of Fine Chemistry of ECUST. Melting points were determined with the digital melting point apparatus (WRS-1B). The purity of final compounds were analyzed by HPLC (Hewlett-Packard 1100), with the purity of all compounds being over than 95%. The HPLC instrument was equipped with a photodiode array detector using a Zorbax RP-18 column (5  $\mu$ m, 4.6 mm×250 mm, reverse phase column). The mobile phases A and B were acetonitrile and the solution of 10 mM ammonium acetate in purified water (pH = 6.0), respectively.

*tert*-Butyl(3-Aminophenyl)carbamate (8). To a mixture of 1,3-phenylenediamine (10.800 g, 100 mmol), Et<sub>3</sub>N (10.100 g, 100 mmol) in methanol (150 mL) was added Boc<sub>2</sub>O (21.800 g, 100 mmol) at 0 °C. The reaction solution was stirred for 24 h at room temperature. Then, the solution was concentrated with a rotary evaporator. The crude product was purified by silica gel chromatography (petroleum ether/ethylacetate = 4:1, v/v) to obtain the title compound 2 as white powder (13.310 g, 64%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.03 (t, *J* = 8.0 Hz, 1H), 6.96 (s, 1H), 6.55 (dd, *J* = 8.0 Hz, *J* = 1.2 Hz, 1H), 6.43 (s, 1H), 6.36 (dd, *J* = 8.0 Hz, *J* = 1.6 Hz, 1H), 3.54 (s, 2H), 1.51 (s, 9H). LC-MS: m/z: 209.1 (M+H)<sup>+</sup>.

#### Ethyl4-((3-((tert-butoxycarbonyl)amino)phenyl)amino)-2-chloropyrimidine-5-

**carboxylate (9).** A mixture of ethyl-2,4-dichloropyrimidine-5-carboxylate (2.210 g, 10 mmol), compound **8** (2.080 g, 10 mmol) and DIPEA (1.935 g, 15 mmol) in CH<sub>3</sub>CN (40 mL) was heated with reflux for 2 h. After cooling to room temperature, the white solid was collected by filtration and washed with CH<sub>3</sub>CN to give the product **9** (3.371 g, 86%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.23 (s, 1H), 9.50 (s, 1H), 8.80 (s, 1H), 7.70 (s, 1H), 7.35 (d, *J* = 7.6 Hz, 1H), 7.29 (t, *J* = 8.0 Hz, 1H), 7.24 (d, *J* = 8.4 Hz, 1H), 4.39 (q, *J* = 7.2 Hz, 2H), 1.49 (s, 9H), 1.36 (t, *J* = 7.2 Hz, 3H). LC-MS: m/z: 393.1 (M+H)<sup>+</sup>.

4-((3-((*tert*-Butoxycarbonyl)Amino)phenyl)amino)-2-chloropyrimidine-5-carboxylic acid (10). A mixture of compound 9 (3.136 g, 8 mmol) in THF and 1 M NaOH (8 mL) was heated at 50 °C for 3 h. After cooling to room temperature, the solution was acidified with 1 M HCl to give a white solid, which was collected by filtration and washed with THF and water to give the title compound 10 (2.796 g, 96%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.84 (s, 1H), 9.49 (s, 1H), 8.76 (s, 1H), 7.67 (s, 1H), 7.40 (d, *J* = 8.0 Hz, 1H), 7.28 (t, *J* = 8.0 Hz, 1H), 7.22 (d, *J* = 8.4 Hz, 1H), 1.49 (s, 9H). LC-MS: m/z: 365.1 (M+H)<sup>+</sup>.

#### 4-((3-((tert-Butoxycarbonyl)Amino)phenyl)amino)-2-(phenylamino)pyrimidine-5-

**carboxylic acid (11).** To a solution of compound **10** (2.184 g, 6 mmol) in CH<sub>3</sub>CN (40 mL) was added aniline (0.558 g, 6 mmol), the mixture was heated with reflux for 12 h. After cooling to room temperature, the residue was collected by filtration and washed with CH<sub>3</sub>CN to give the product **11** as a white solid (2.249 g, 89%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.44 (s, 1H), 9.82 (s, 1H), 9.36 (s, 1H), 8.70 (s, 1H), 7.69 (d, *J* = 8.0 Hz, 2H), 7.63 (s, 1H), 7.43 (s, 1H), 7.26-7.21 (m, 4H), 6.98 (t, *J* = 6.8 Hz, 1H), 1.47 (s, 9H). HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>22</sub>H<sub>24</sub>N<sub>5</sub>O<sub>4</sub> 422.1828, found, 422.1823.

*tert*-Butyl(3-((5-(Methylcarbamoyl)-2-(phenylamino)pyrimidin-4-yl)amino)phenyl)carba mate (12). To a solution of 11 (1.263 g, 3 mmol), DIPEA (0.774 g, 6 mmol) in dry DMF (15 mL) was added HATU (1.368 g, 3.6 mmol). After stirring for 1 h at room temperature, methylamine hydrochloride (0.306 g, 4.5 mmol) was added and stirred overnight. The reaction mixture was poured into ice water, the resulting solid was collected by filtration, washed with water and recrystallized from EtOH to give the product 12 (0.612 g, 47%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  11.33 (s, 1H), 9.59 (s, 1H), 9.31 (s, 1H), 8.64 (s, 1H), 8.50 (d, *J* = 4.8 Hz, 1H), 7.69 (d, *J* = 7.6 Hz, 2H), 7.62 (s, 1H), 7.50-7.48 (m, 1H), 7.23 (t, *J* = 7.6 Hz, 2H), 7.19 (d, *J* = 8.0 Hz, 1H), 7.15 (d, *J* = 8.0 Hz, 1H), 6.96 (t, *J* = 7.2 Hz, 1H), 2.79 (d, *J* = 4.4 Hz, 3H), 1.47 (s, 9H). LC-MS: m/z: 435.2 (M+H)<sup>+</sup>.

#### tert-Butyl(3-(3-Methyl-2,4-dioxo-7-(phenylamino)-3,4-dihydropyrimido[4,5-d]pyrimidin-

**1(2***H***)-yl)phenyl)carbamate (13).** To a solution of **12** (0.434 g, 1 mmol), K<sub>2</sub>CO<sub>3</sub> (0.276 g, 2 mmol) in dry THF (10 mL) was added 1,1'-carbonyldiimidazole (0.324 g, 2 mmol). The reaction mixture was heated with reflux overnight. After cooling to room temperature, the reaction mixture was poured into water, and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×20 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (petroleum ether/ethylacetate = 1:2, v/v) to give the product **13** as a white solid (0.193 g, 42%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.62 (s, 1H), 8.92 (s, 1H), 7.66 (s, 1H), 7.52 (d, *J* = 8.0 Hz, 1H), 7.45 (t, *J* = 8.0 Hz, 1H), 7.36-7.32 (m, 2H), 7.03 (d, *J* = 8.0 Hz, 3H), 6.91 (d, *J* = 7.2 Hz, 1H), 3.30 (s, 3H), 1.45 (s, 9H). HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>24</sub>H<sub>25</sub>N<sub>6</sub>O<sub>4</sub> 461.1937, found, 461.1939.

*N*-(3-(3-Methyl-2,4-dioxo-7-(phenylamino)-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)yl)phenyl)acrylamide (14e). To a solution of 13 (0.190 g, 0.41 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4 mL) was

added trifluoroacetic acid (1 mL). The mixture was stirred for 4 h at room temperature. The reaction solution was neutralized with saturated aqueous NaHCO<sub>3</sub>, and extracted with CH<sub>2</sub>Cl<sub>2</sub> ( $3 \times 15$  mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo to obtain a white solid (0.136 g, 90%), which was used in the next reaction without further purification. To a solution of 1-(3-aminophenyl)-3-methyl-7-(phenylamino)pyrimido[4,5-d] pyrimidine-2,4(1H,3H)-dione (0.136 g, 0.38 mmol) in NMP (1.5 mL) was added acryloyl chloride (37 µL,

2,4(1*H*,3*H*)-dione (0.136 g, 0.38 mmol) in NMP (1.5 mL) was added acryloyl chloride (37  $\mu$ L, 0.46 mmol) dissolved in CH<sub>3</sub>CN (0.5 mL) at 0 °C. The reaction solution was stirred overnight at room temperature. The mixture was poured into water, and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×15 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (petroleum ether/ethylacetate = 1:1.5, v/v) to give the product **14e** as a white solid (0.083 g, 53%). mp 290.3-291.2 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.38 (s, 1H), 8.93 (s, 1H), 7.82 (d, *J* = 3.2 Hz, 1H), 7.80 (s, 1H), 7.53 (t, *J* = 8.0 Hz, 1H), 7.33-7.29 (m, 2H), 7.15 (d, *J* = 7.6 Hz, 1H), 7.02-6.96 (m, 2H), 6.89 (t, *J* = 6.0 Hz, 1H), 6.44 (dd, *J* = 17.2 Hz, *J* = 10.4 Hz, 1H), 6.25 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 5.76 (dd, *J* = 10.4 Hz, *J* = 2.0 Hz, 1H), 3.30 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.24, 160.11, 158.97, 157.87, 150.86, 139.90, 138.91, 136.19, 131.61, 129.45, 128.11, 127.22, 124.30, 122.53, 120.07, 119.26, 119.07, 27.57. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>N<sub>6</sub>O<sub>3</sub> 415.1519, found, 415.1512. HPLC purity: 97.66%, retention time = 10.68 min.

The following compounds **14a-d** were prepared by a method similar to that for the synthesis of compound **14e**.

*N*-(3-(7-((4-(4-Acetylpiperazin-1-yl)-2-methoxyphenyl)amino)-3-methyl-2,4-dioxo-3,4dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (14a). Yellow solid (yield 49%). mp 228.5-229.8 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.34 (s, 1H), 8.83 (s, 1H), 8.59 (s, 1H), 7.86 (s, 1H), 7.69 (s, 1H),7.48 (t, *J* = 8.4 Hz, 1H), 7.18 (d, *J* = 8.8 Hz, 1H), 7.09 (d, *J* = 8.0 Hz, 1H), 6.55 (s, 1H), 6.45 (dd, *J* = 17.2 Hz, *J* = 10.4 Hz, 1H), 6.26 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 5.99 (s, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.2 Hz, 1H), 3.76 (s, 3H), 3.58-3.54 (m, 4H), 3.28 (s, 3H), 3.05-2.99 (m, 4H), 2.05 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  168.19, 163.21, 160.10, 157.83, 139.73, 135.99, 131.67, 129.33, 127.18, 49.15, 48.76, 45.42, 40.60, 27.54, 21.16. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>31</sub>N<sub>8</sub>O<sub>5</sub> 571.2417, found, 571.2417. HPLC purity: 97.63%, retention time = 9.50 min.

#### N-(3-(7-((2-Methoxy-4-morpholinophenyl)amino)-3-methyl-2,4-dioxo-3,4-

**dihydropyrimido**[4,5-*d*]**pyrimidin-1**(2*H*)-**y**]**pheny**]**)acrylamide** (14b). Yellow solid (yield 51%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.36 (s, 1H), 8.80 (s, 1H), 8.44 (s, 1H), 8.03 (s, 1H), 7.86 (d, *J* = 8.0 Hz, 1H), 7.69 (t, *J* = 2.0 Hz, 1H), 7.52 (t, *J* = 8.4 Hz, 1H), 7.32 (d, *J* = 8.8 Hz, 1H), 7.09 (d, *J* = 7.6 Hz, 1H), 6.57 (d, *J* = 2.0 Hz, 1H), 6.45 (dd, *J* = 16.8 Hz, *J* = 10.0 Hz, 1H), 6.27 (dd, *J* = 17.2 Hz, *J* = 2.0 Hz, 1H), 5.77 (dd, *J* = 10.4 Hz, *J* = 2.0 Hz, 1H), 3.77 (s, 3H), 3.58-3.54 (m, 4H), 3.06-2.99 (m, 4H), 2.05 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.23, 160.09, 158.87, 157.82, 150.86, 149.81, 148.06, 139.74, 135.98, 131.65, 129.34, 127.19, 124.18, 121.08, 120.00, 119.12, 105.61, 99.65, 99.15, 66.03, 55.64, 48.78, 27.54. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>27</sub>H<sub>28</sub>N<sub>7</sub>O<sub>5</sub> 528.1995, found, 528.1993. HPLC purity: 96.86%, retention time = 10.73 min.

# *N*-(3-(7-((2-Methoxy-4-thiomorpholinophenyl)amino)-3-methyl-2,4-dioxo-3,4dihydropyrimido[4,5-d]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (14c). Yellow solid (yield

52%). mp 246.0-247.1 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.35 (s, 1H), 8.84 (s, 1H), 8.60 (s, 1H), 8.85 (s, 1H), 7.70 (s, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.17 (d, *J* = 8.4 Hz, 1H), 7.09 (d, *J* = 7.6 Hz, 1H), 6.45 (dd, *J* = 17.2 Hz, *J* = 10.0 Hz, 1H), 6.27 (dd, *J* = 16.8 Hz, *J* = 2.0 Hz, 1H), 5.97-5.95 (m, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 3.75 (s, 3H), 3.42-3.39 (m, 4H), 3.28 (s, 3H), 2.66-2.63 (m, 4H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.24, 160.08, 158.90, 157.81, 150.86, 149.99, 139.76, 135.99, 131.63, 129.33, 127.22, 124.19, 121.32, 120.01, 119.10, 106.85, 100.43, 99.63, 55.65, 51.46, 27.54, 25.62. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>27</sub>H<sub>28</sub>N<sub>7</sub>O<sub>4</sub>S 546.1923, found, 546.1924. HPLC purity: 95.21%, retention time = 12.49 min.

*N*-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-3-methyl-2,4dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (14d). Yellow solid (yield 47%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.36 (s, 1H), 8.83 (s, 1H), 8.54 (s, 1H), 7.88-7.86 (m, 1H), 7.70 (s, 1H),7.47 (t, *J* = 8.0 Hz, 1H), 7.16 (d, *J* = 8.4 Hz, 1H), 7.08 (d, *J* = 7.6 Hz, 1H), 6.50-6.43 (m, 2H), 6.26 (dd, *J* = 16.8 Hz, *J* = 1.6 Hz, 1H), 5.98-5.96 (m, 1H), 5.76 (dd, *J* = 10.0 Hz, *J* = 1.2 Hz, 1H), 3.75 (s, 3H), 3.59-3.58 (m, 2H), 3.28 (s, 3H), 2.60-2.55 (m, 2H), 2.22 (s, 6H), 1.81 (d, *J* = 7.6 Hz, 2H), 1.47-1.40 (m, 2H). HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>35</sub>N<sub>8</sub>O<sub>4</sub> 571.2781, found, 571.2780. HPLC purity: 97.96%, retention time = 11.35 min.

2,4-Dichloropyrimidine-5-carboxamide (15). To a solution of 2,4-dichloropyrimidine-5carbonyl chloride (4.220 g, 20 mmol) in dichloromethane (15 mL) was added ammonium hydroxide (1.400 g, 40 mmol) at 0 °C. The reaction mixture was stirred for 2 h at 0 °C, and the mixture was poured into ice water. The resulting solid was collected by filtration, washed with water to give the product **15** as a white solid (3.510 g, 92%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$ 8.91 (s, 1H), 8.20 (s, 1H), 8.08 (s, 1H).

*tert*-Butyl(3-((5-Carbamoyl-2-chloropyrimidin-4-yl)amino)phenyl)carbamate (16). A mixture of compound 15 (3.438 g, 18 mmol), DIPEA (3.483 g, 27 mmol), compound 8 (3.744 g, 18 mmol) in isopropanol (50 mL) was heated at 40 °C for 5 h. After cooling to room temperature, the white solid was collected by filtration and washed with isopropanol to give the product 16 (5.480 g, 84%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  11.51 (s, 1H), 9.49 (s, 1H), 8.78 (s, 1H), 8.47 (s, 1H), 8.00 (s, 1H), 7.64 (s, 1H), 7.41 (d, *J* = 8.0 Hz, 1H), 7.27 (t, *J* = 8.0 Hz, 1H), 7.20 (d, *J* = 8.0 Hz, 1H), 1.48 (s, 9H). LC-MS: m/z: 364.1 (M+H)<sup>+</sup>.

#### tert-Butyl(3-((5-Carbamoyl-2-((4-(4-(dimethylamino)piperidin-1-yl)-2-methoxyphenyl)

amino)pyrimidin-4-yl)amino)phenyl)carbamate (17). To a mixture of compound 16 (5.082 g, 14 mmol), trifluoroacetic acid (1.550 mL, 21 mmol) in isopropanol (100 mL) was added 1-(4amino-3-methoxyphenyl)-*N*,*N*-dimethylpiperidin-4-amine (4.183 g, 16.8 mmol). The reaction solution was heated with reflux overnight. After cooling to room temperature, the solution was concentrated with a rotary evaporator. And neutralized with saturated aqueous NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×50 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (dichloromethane/methanol = 15:1, v/v) to give the product 11 (5.450 g, 67%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  11.46 (s, 1H), 9.34 (s, 1H), 8.65 (s, 1H), 8.24 (s, 1H), 8.08 (s, 1H), 7.52 (d, *J* = 8.8 Hz, 1H), 7.41 (d, *J* = 7.6 Hz, 1H), 7.37 (s, 1H), 7.12-7.07 (m, 2H), 6.62 (d, *J* = 1.6 Hz, 1H), 6.45 (d, *J* = 8.4 Hz, 1H), 3.77 (s, 3H), 3.70 (d, *J* = 12.0 Hz, 2H), 2.66 (t, *J* = 11.2 Hz, 2H), 2.20 (s, 6H), 1.84 (d, *J* = 12.0 Hz, 2H), 1.54-1.49 (m, 2H), 1.47 (s, 9H). LC-MS: m/z: 577.4 (M+H)<sup>+</sup>.

*tert*-Butyl(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-2,4dioxo-3,4-dihydropyrimido[4,5-d]pyrimidin-1(2H)-yl)phenyl)carbamate (18). A mixture of

compound **17** (5.184 g, 9 mmol), K<sub>2</sub>CO<sub>3</sub> (1.863 g, 13.5 mmol), 1,1'-carbonyldiimidazole (2.916 g, 18 mmol) in dry THF (40 mL) was heated with reflux overnight. After cooling to room temperature, the solution was added ice water and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×30 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (dichloromethane/methanol = 12:1, v/v) to give the product **18** (3.289 g, 60%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  11.55 (s, 1H), 9.59 (s, 1H), 8.77 (s, 1H), 8.54 (s, 1H), 7.53 (s, 2H), 7.38 (t, *J* = 8.0 Hz, 1H), 7.17 (d, *J* = 6.8 Hz, 1H), 6.96 (d, *J* = 7.2 Hz, 1H), 6.50 (s, 1H), 6.02-5.99 (m, 1H), 3.76 (s, 3H), 3.61-3.59 (m, 2H), 2.62-2.57 (m, 2H), 2.19 (s, 6H), 1.82 (d, *J* = 12.0 Hz, 2H), 1.51-1.48 (m, 2H), 1.46 (s, 9H). LC-MS: m/z: 603.3 (M+H)<sup>+</sup>. *tert*-Butyl(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-3-ethyl-2,4-dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)carbamate (19). To a

2,4-dioxo-3,4-dinydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yi)pinenyi)carbamate (19). To a mixture of compound **18** (0.602 g, 1 mmol), Cs<sub>2</sub>CO<sub>3</sub> (0.489 g, 1.5 mmol) in DMF (5 mL) was added iodoethane (95 µL, 1.2 mmol). The mixture was stirred at room temperature overnight. The solution was added ice water and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×25 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (dichloromethane/methanol = 13:1, v/v) to give the title compound **19** (0.378 g, 60%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.60 (s, 1H), 8.83 (s, 1H), 8.58 (s, 1H), 7.57 (s, 1H), 7.53-7.51 (m, 1H), 7.39 (t, *J* = 7.6 Hz, 1H), 7.18 (d, *J* = 7.6 Hz, 1H), 6.97 (d, *J* = 7.2 Hz, 1H), 6.51 (s, 1H), 6.03-6.01 (m, 1H), 3.93 (q, *J* = 6.4 Hz, 2H), 3.76 (s, 3H), 3.63-3.60 (m, 2H), 2.63-2.58 (m, 2H), 2.24 (s, 6H), 1.83 (d, *J* = 12.0 Hz, 2H), 1.52-1.47 (m, 2H), 1.46 (s, 9H), 1.17 (t, *J* = 6.8 Hz, 3H). LC-MS: m/z: 631.3 (M+H)<sup>+</sup>.

*N*-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-3-ethyl-2,4dioxo-3,4-dihydropyrimido[4,5-d]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20a). To a solution of compound 19 (0.348 g, 0.55 mmol) in  $CH_2Cl_2$  (5 mL) was added trifluoroacetic acid (1.5 mL). The mixture was stirred for 5 h at room temperature. The reaction solution was neutralized with saturated aqueous NaHCO<sub>3</sub>, and extracted with  $CH_2Cl_2$  (3×30 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo to obtain a yellow solid (0.241 g, 82%), which was used in the next reaction without further purification.

To solution 1-(3-aminophenyl)-7-((4-(4-(dimethylamino)piperidin-1-yl)-2а of methoxyphenyl)amino)-3-ethylpyrimido[4,5-d]pyrimidine-2,4(1H,3H)-dione (0.220)0.4 g, mmol) in NMP (1.5 mL) was added acryloyl chloride (42  $\mu$ L, 0.5 mmol) dissolved in CH<sub>3</sub>CN (0.5 mL) at 0 °C. The reaction solution was stirred overnight at room temperature. The mixture was poured into water, and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×20 mL). The organic layer was washed with saturated aqueous NaCl, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel chromatography (dichloromethane/methanol = 12:1, v/v) to give the title product **20a** as a yellow solid (0.116 g, 49%). mp 212.1-213.6 °C. <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ 10.64 (s, 1H), 10.56 (s, 1H), 8.84 (s, 1H), 8.62 (s, 1H), 7.92-7.90 (m, 1H), 7.71 (s, 1H), 7.48 (t, J = 8.0 Hz, 1H), 7.17 (d, J = 6.8 Hz, 1H), 7.10 (d, J = 7.2 Hz, 1H), 6.55-6.49 (m, 2H), 6.28 (dd, J = 16.8 Hz, J = 1.6 Hz, 1H), 6.00 (s, 1H), 5.78 (dd, J = 11.2 Hz, J = 1.2 Hz, 1H), 3.94 (q, J = 6.4 Hz, 2H), 3.76 (s, 3H), 3.72-3.68 (m, 2H), 2.73 (s, 6H), 2.60 (t, J = 8.0 Hz, 1H), 2.63-2.58 (m, 2H), 2.09 (d, J = 11.2 Hz, 2H), 1.72-1.68 (m, 2H), 1.18 (t, J = 6.4 Hz, 3H). HRMS(ESI) (m/z):  $(M+H)^+$  calcd for  $C_{31}H_{37}N_8O_4$  585.2938, found, 585.2928. HPLC purity: 96.26%, retention time = 9.95 min.

The following compounds **20b-1** were prepared by a synthesis route similar to that for compound **20a**.

#### N-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-2,4-dioxo-3-

propyl-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20b). Yellow solid (yield 39%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.54 (s, 1H), 8.84 (s, 1H), 8.63 (s, 1H), 7.92 (s, 1H), 7.69 (s, 1H), 7.49 (t, *J* = 8.0 Hz, 1H), 7.19 (d, *J* = 6.8 Hz, 1H), 7.10 (d, *J* = 7.8 Hz, 1H), 6.54-6.48 (m, 2H), 6.28 (dd, *J* = 16.8 Hz, *J* = 1.2 Hz, 1H), 6.00 (s, 1H), 5.78 (dd, *J* = 10.4 Hz, *J* = 1.6 Hz, 1H), 3.86 (t, *J* = 6.8 Hz, 2H), 3.76 (s, 3H), 3.72-3.69 (m, 2H), 3.26-3.21 (m, 1H), 2.72 (s, 6H), 2.62-2.57 (m, 2H), 2.07 (d, *J* = 11.2 Hz, 2H), 1.70-1.67 (m, 2H), 1.65-1.60 (m, 2H), 0.90 (t, *J* = 7.2 Hz, 3H). HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>32</sub>H<sub>39</sub>N<sub>8</sub>O<sub>4</sub> 599.3094, found, 599.3099. HPLC purity: 97.85%, retention time = 10.96 min.

*N*-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-3-isopropyl-2,4dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20c). Yellow solid (yield 41%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.55 (s, 1H), 8.83 (s, 1H), 8.60 (s, 1H), 7.91 (s, 1H), 7.69 (s, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.19 (d, *J* = 5.6 Hz, 1H), 7.10 (d, *J* = 7.6 Hz, 1H), 6.55-6.48 (m, 2H), 6.28 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 6.00 (s, 1H), 5.78 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 5.17-5.10 (m, 1H), 3.76 (s, 3H), 3.72-3.68 (m, 2H), 3.26-3.20 (m, 1H), 2.72 (s, 6H), 2.63-2.59 (m, 2H), 2.07 (d, *J* = 10.8 Hz, 2H), 1.70-1.67 (m, 2H), 1.44 (d, *J* = 6.8 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.29, 160.19, 157.83, 139.83, 135.83, 135.95, 131.81, 129.22, 127.00, 124.26, 120.04, 119.06, 106.58, 99.91, 62.21, 55.67, 47.77, 45.00, 25.55, 19.25. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>32</sub>H<sub>39</sub>N<sub>8</sub>O<sub>4</sub> 599.3094, found, 599.3079. HPLC purity: 98.38%, retention time = 11.12 min. *N*-(3-(3-Benzyl-7-((4-(4-(dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-2,4dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20d). Yellow solid (yield 39%). mp 285.4-286.6 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.54 (s, 1H), 8.87 (s, 1H), 8.69 (s, 1H), 7.91 (s, 1H), 7.69 (s, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.38 (d, *J* = 7.2 Hz, 2H), 7.33 (t, *J* = 7.2 Hz, 2H), 7.26 (t, *J* = 7.2 Hz, 1H), 7.19 (d, *J* = 8.0 Hz, 1H), 7.11 (d, *J* = 6.8 Hz, 1H), 6.54-6.47 (m, 2H), 6.27 (dd, *J* = 16.8 Hz, *J* = 1.6 Hz, 1H), 6.01-5.99 (m, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 5.10 (s, 2H), 3.76 (s, 3H), 3.72-3.69 (m, 2H), 3.27-3.21 (m, 1H), 2.73 (s, 6H), 2.62-2.57 (m, 2H), 2.08 (d, *J* = 11.2 Hz, 2H), 1.71-1.68 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO*d*<sub>6</sub>)  $\delta$  163.31, 159.94, 139.88, 136.98, 135.84, 131.82, 129.27, 128.30, 127.63, 127.14, 127.01, 62.24, 59.74, 55.68, 47.72, 43.83, 25.47, 20.74. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>36</sub>H<sub>39</sub>N<sub>8</sub>O<sub>4</sub> 647.3094, found, 647.3088. HPLC purity: 95.30%, retention time = 12.72 min.

*N*-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-2,4-dioxo-3phenethyl-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20e). Yellow solid (yield 40%). mp 196.5-197.7 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.58 (s, 1H), 8.85 (s, 1H), 8.67 (s, 1H), 7.91 (s, 1H), 7.72 (s, 1H), 7.49 (t, *J* = 8.0 Hz, 1H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.26-7.23 (m, 3H), 7.19 (d, *J* = 12.8 Hz, 1H), 7.07 (d, *J* = 6.8 Hz, 1H), 6.56-6.49 (m, 2H), 6.28 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 6.01-5.99 (m, 1H), 5.78 (dd, *J* = 10.4 Hz, *J* = 1.6 Hz, 1H), 4.10-4.08 (m, 2H), 3.77 (s, 3H), 3.72-3.69 (m, 2H), 3.24-3.19 (m, 1H), 2.90 (t, *J* = 7.6 Hz, 2H), 2.71 (s, 6H), 2.62-2.57 (m, 2H), 2.07 (d, *J* = 11.2 Hz, 2H), 1.70-1.68 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.33, 159.68, 139.90, 138.51, 135.82, 131.83, 129.26, 128.59, 128.50, 127.02, 126.39, 62.23, 55.70, 47.74, 41.92, 33.28, 25.52. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>37</sub>H<sub>41</sub>N<sub>8</sub>O<sub>4</sub> 661.3251, found, 661.3251. HPLC purity: 99.09%, retention time = 13.71 min.

*N*-(3-(7-((4-(4-(Dimethylamino)piperidin-1-yl)-2-methoxyphenyl)amino)-2,4-dioxo-3-(3phenylpropyl)-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20f). Yellow solid (yield 38%). mp 252.1-253.8 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.72 (s, 1H), 8.81 (s, 1H), 8.60 (s, 1H), 7.92 (s, 1H), 7.78 (s, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.28-7.19 (m, 5H), 7.14 (t, *J* = 7.2 Hz, 1H), 7.07 (d, *J* = 7.2 Hz, 1H), 6.61-6.52 (m, 2H), 6.29 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 6.02 (s, 1H), 5.78 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 3.94 (t, *J* = 7.2 Hz, 2H), 3.76 (s, 3H), 3.22-3.17 (m, 1H), 2.69 (s, 6H), 2.68-2.63 (m, 4H), 2.08 (d, *J* = 10.4 Hz, 2H), 1.94 (t, *J* = 6.8 Hz, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.33, 159.89, 157.82, 141.32, 139.88, 135.89, 131.84, 129.23, 128.21, 128.12, 126.99, 125.68, 62.20, 47.76, 40.52, 32.51, 28.57, 25.57. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>38</sub>H<sub>43</sub>N<sub>8</sub>O<sub>4</sub> 675.3407, found, 675.3403. HPLC purity: 95.85%, retention time = 14.42 min.

*N*-(3-(3-Isopropyl-7-((2-methoxy-4-(4-methylpiperazin-1-yl)phenyl)amino)-2,4-dioxo-3,4dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20g). Yellow solid (yield 39%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.64 (s, 1H), 8.83 (s, 1H), 8.63 (s, 1H), 7.85 (d, *J* = 7.2 Hz, 1H), 7.73 (s, 1H), 7.47 (t, *J* = 8.0 Hz, 1H), 7.20 (d, *J* = 8.4 Hz, 1H), 7.09 (d, *J* = 7.6 Hz, 1H), 6.56-6.51 (m, 2H), 6.27 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 6.03-6.01 (m, 1H), 5.77 (dd, *J* = 10.4 Hz, *J* = 1.2 Hz, 1H), 5.17-5.10 (m, 1H), 3.76 (s, 3H), 3.29 (t, *J* = 4.0 Hz, 4H), 3.20 (t, *J* = 4.0 Hz, 4H), 2.75 (s, 3H), 1.44 (d, *J* = 6.4 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.33, 160.20, 157.83, 139.79, 135.94, 131.79, 129.21, 127.05, 120.09, 100.00, 55.74, 52.25, 46.05, 45.02, 42.29, 19.24. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>35</sub>N<sub>8</sub>O<sub>4</sub> 571.2781, found, 571.2775. HPLC purity: 98.71%, retention time = 11.25 min.

*N*-(3-(3-Isopropyl-7-((3-methoxy-4-(4-methylpiperazin-1-yl)phenyl)amino)-2,4-dioxo-3,4dihydropyrimido[4,5-d]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20h). Yellow solid (yield 47%). mp 221.3-222.5 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.51 (s, 1H), 10.18 (s, 1H), 8.88 (s, 1H), 7.89 (s, 1H), 7.72 (s, 1H), 7.50 (t, *J* = 8.0 Hz, 1H), 7.13 (d, *J* = 8.0 Hz, 1H), 6.91 (s, 1H), 6.49 (dd, *J* = 16.8 Hz, *J* = 10.0 Hz, 1H), 6.43-6.41 (m, 1H), 6.27 (dd, *J* = 16.8 Hz, *J* = 1.6 Hz, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 5.18-5.11 (m, 1H), 3.56 (s, 3H), 3.04-3.01 (m, 8H), 2.63 (s, 3H), 1.44 (d, *J* = 6.8 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.29, 160.17, 151.55, 150.38, 139.87, 136.09, 134.47, 131.73, 129.33, 127.13, 124.33, 120.10, 119.23, 55.29, 53.27, 47.95, 45.06, 43.21, 19.23. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>35</sub>N<sub>8</sub>O<sub>4</sub> 571.2781, found, 571.2773. HPLC purity: 97.35%, retention time = 10.11 min.

*N*-(3-(3-Isopropyl-7-((3-methyl-4-(4-methylpiperazin-1-yl)phenyl)amino)-2,4-dioxo-3,4dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20i). Yellow solid (yield 49%). mp 295.5-296.9 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.54 (s, 1H), 10.22 (s, 1H), 8.88 (s, 1H), 7.90 (d, *J* = 7.2 Hz, 1H), 7.74 (s, 1H), 7.51 (t, *J* = 8.0 Hz, 1H), 7.18 (s, 1H), 7.13 (d, *J* = 8.0 Hz, 1H), 7.08 (d, *J* = 6.4 Hz, 1H), 6.69 (d, *J* = 7.6 Hz, 1H), 6.50 (dd, *J* = 17.2 Hz, *J* = 10.4 Hz, 1H), 6.26 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 5.76 (dd, *J* = 10.4 Hz, *J* = 1.6 Hz, 1H), 5.18-5.11 (m, 1H), 3.12 (t, *J* = 4.0 Hz, 4H), 2.92 (t, *J* = 4.0 Hz, 4H), 2.70 (s, 3H), 1.97 (s, 3H), 1.44 (d, *J* = 7.2 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.29, 160.18, 159.05, 157.91, 150.40, 139.92, 136.16, 134.60, 132.03, 131.74, 129.37, 127.10, 124.26, 121.38, 119.95, 119.35, 118.62, 117.58, 53.35, 48.94, 45.04, 42.84, 19.23, 17.44. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>35</sub>N<sub>8</sub>O<sub>3</sub> 555.2832, found, 555.2831. HPLC purity: 99.05%, retention time = 10.93 min.

*N*-(3-(7-((4-((2-(Dimethylamino)ethyl)(methyl)amino)-2-methoxyphenyl)amino)-3isopropyl-2,4-dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20j). Yellow solid (yield 37%). mp 150.7-152.4 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.94 (s, 1H), 8.45 (s, 1H), 7.95 (s, 1H), 7.83 (s, 1H), 7.68 (d, *J* = 8.0 Hz, 1H), 7.46 (t, *J* = 8.0 Hz, 1H), 7.35 (d, *J* = 8.8 Hz, 1H), 7.04 (d, J = 7.6 Hz, 1H), 6.38 (dd, J = 16.8 Hz, J = 1.6 Hz, 1H), 6.27-6.17 (m, 2H), 5.83 (dd, J = 9.6 Hz, J = 1.6 Hz, 1H), 5.69 (d, J = 10.4 Hz, 1H), 5.36-5.29 (m, 1H), 3.81 (s, 3H), 3.39 (t, J = 7.6 Hz, 2H), 2.88 (s, 3H), 2.45 (t, J = 7.6 Hz, 2H), 2.32 (s, 6H), 1.55 (d, J = 6.8 Hz, 6H). HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>37</sub>N<sub>8</sub>O<sub>4</sub> 573.2938, found, 573.2932. HPLC purity: 95.07%, retention time = 10.56 min.

*N*-(3-(7-((4-(2-(Dimethylamino)ethoxy)-2-methoxyphenyl)amino)-3-isopropyl-2,4-dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (20k). Yellow solid (yield 41%). mp 195.4-196.7 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.39 (s, 1H), 8.84 (s, 1H), 8.64 (s, 1H), 7.80 (d, *J* = 4.4 Hz, 1H), 7.71 (s, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.23 (d, *J* = 5.6 Hz, 1H), 7.10 (d, *J* = 7.6 Hz, 1H), 6.52-6.42 (m, 2H), 6.27 (dd, *J* = 17.2 Hz, *J* = 1.6 Hz, 1H), 6.01-5.99 (m, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.2 Hz, 1H), 5.15-5.11 (m, 1H), 3.95 (t, *J* = 5.2 Hz, 2H), 3.75 (s, 3H), 2.60 (t, *J* = 5.2 Hz, 2H), 2.22 (s, 6H), 1.44 (d, *J* = 6.8 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.19, 160.20, 159.16, 157.83, 155.23, 150.33, 139.68, 136.00, 131.65, 129.27, 127.16, 124.32, 121.37, 120.08, 119.10, 103.84, 100.07, 98.76, 65.73, 57.64, 55.73, 45.47, 44.99, 19.23. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>29</sub>H<sub>34</sub>N<sub>7</sub>O<sub>5</sub> 560.2621, found, 560.2625. HPLC purity: 98.57%, retention time = 8.78 min.

*N*-(3-(3-Isopropyl-7-((2-methoxy-4-(4-(4-methylpiperazin-1-yl)piperidin-1-yl)phenyl) amino)-2,4-dioxo-3,4-dihydropyrimido[4,5-*d*]pyrimidin-1(2*H*)-yl)phenyl)acrylamide (201). Yellow solid (yield 43%). mp >300 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.57 (s, 1H), 8.82 (s, 1H), 8.57 (s, 1H), 7.91 (s, 1H), 7.70 (s, 1H), 7.47 (t, *J* = 8.0 Hz, 1H), 7.18 (d, *J* = 6.8 Hz, 1H), 7.09 (d, *J* = 7.6 Hz, 1H), 6.56-6.49 (m, 2H), 6.27 (dd, *J* = 16.8 Hz, *J* = 1.6 Hz, 1H), 5.99-5.97 (m, 1H), 5.77 (dd, *J* = 10.0 Hz, *J* = 1.6 Hz, 1H), 5.17-5.10 (m, 1H), 3.76 (s, 3H), 3.64-3.61 (m, 2H), 2.96-2.75 (m, 6H), 2.60-2.56 (m, 6H), 1.87-1.85 (m, 1H), 1.54-1.50 (m, 2H), 1.44 (d, *J* = 6.8 Hz, 6H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  163.31, 160.19, 159.07, 157.81, 150.34, 139.83, 135.92, 131.79, 129.22, 126.99, 119.99, 99.79, 60.77, 55.61, 52.86, 48.21, 46.52, 45.02, 43.23, 27.01, 19.24. HRMS(ESI) (m/z): (M+H)<sup>+</sup> calcd for C<sub>35</sub>H<sub>44</sub>N<sub>9</sub>O<sub>4</sub> 654.3516, found, 654.3519. HPLC purity: 99.38%, retention time = 11.76 min.

In Vitro Enzymatic Inhibition and Cell Antiproliferation Activity Assay. The enzymatic inhibition and cell antiproliferation activity assays were evaluated by using the well-established ELISA-based assay and sulforhodamine B (SRB) colorimetric assay, respectively. The detailed experiments were performed according to the published literature,<sup>25</sup> and the detailed characterizations are provided in the Supporting Information.

Western Blot Analysis. H1975 and A431 cells were collected and suspended in lysis buffer (Tris-HCl, pH 6.8, 100 mmol/L; DTT, 200 mmol/L; 0.2% bromophenol blue; 4% SDS; 20% glycerol). Equivalent amounts of proteins were loaded and separated by SDS-PAGE (8%), then transfered to nitrocellulose membranes. Western blot analysis was subsequently performed with standard procedures. The antibodies were used for immune detection of proteins, including p-EGFR (Y1068; #3777S), EGFR (#4267S), p-AKT (Ser473), AKT, and GAPDH (Cell Signaling Technologies, Cambridge, MA).

Human and Mouse Liver Microsomal Stability Assays. The incubation is performed as follows: microsomes in 0.1 M Tris/HCl buffer of pH 7.4 (0.33 mg/mL microsomal protein), co-factor MgCl<sub>2</sub> (5 mM), tested compound (final concentration of 0.1  $\mu$ M, co-solvent (0.01% DMSO) and 0.005% Bovin serum albumin) and NADPH (1 mM) at 37 °C for 60 min. The reaction can be started by the addition of liver microsomes, or the tested compound or NADPH. Aliquots were sampled at 0, 7, 17, 30 and 60 min incubation and enzymatic reaction is stopped

by protein precipitation in methanol. After centrifugation, samples are then analysed by LC-MS/MS (Waters UPLC I-Class system; Xevo TQ-S mass spectrometer).

Measurements of Aqueous Solubility, CYP Inhibition Assays, and Measurement of Permeability. Determination of the aqueous solubility was performed according to the published literature.<sup>34</sup> The CYP inhibition assays and measurement of permeability of compound **20g** were conducted by WuXi AppTec, and the detailed characterizations are provided in the Supporting Information.

In Vivo Pharmacokinetics Study in Mice. The *in vivo* pharmacokinetic study in mice was conducted by Shanghai Medicilon, Inc. Briefly, the dosing solution (1 mg/mL) was prepared by dissolving the appropriate amount of the compound in 0.5% CMC-Na aqueous solution for oral administration, and the 0.2 mg/mL dosing solution was prepared by dissolving the appropriate amount of the compound in mixed solvent (0.15 mL DMSO+1.200 mL PEG400+1.650 mL Saline) for intravenous administration. Male ICR mice were separately administration. At time points 0.083, 0.25, 0.5, 1, 2, 4, 8, and 24 h after dosing, the blood sample was collected from each animal and stored in 0–4 °C, then separated by centrifugation (8000 r/min for 6 min). All samples were analyzed using LC–MS/MS (SIMADZU LC system; Applied Biosystems mass spectrometer), and the acquired data were analyzed by using the WinNonlin (v5.2).

In Vivo Antitumor Efficiency Study. BALB/c mice (4-6 weeks-old) were purchased from Shanghai Lab. Animal Research Center (Shanghai, China). Briefly, H1975 cell lines and A431 cell lines were cultured in RPMI-1640 (GE Healthcare) supplemented with 10% (v/v) fetal bovine serum (Gibico). Then a total volume of 0.1 mL/mouse (approximately  $2 \times 10^6$  cells) were

implanted subcutaneously into the right flank of the animals. When the size of the tumor reached about 200–400 mm<sup>3</sup>, the mice were randomly divided into vehicle group and treated group (six mice /group). For antitumor efficacy studies, mice were dosed once daily by oral administration with vehicle or inhibitor with the indicated dose for 14 consecutive days. The average tumor volume was measured every 2 or 3 days using vernier calipers, followed by calculated with the formula  $V = (L \times W^2)/2$ , where L stands for length, and W stands for width. All the animal experiments in this article, including animal handling, care and treatment were performed in compliance with Agreement of the ethics committee on laboratory animal care and the Guidelines for the Care and Use of Laboratory Animals in Shanghai, China.

#### **ASSOCIATED CONTENT**

#### **Supporting Information**

The docked pose of **20f**, and the metabolic stability of **20g** in human and mouse liver microsomes are in the supporting material. Also include availability of molecular formula strings (CSV). This material is available free of charge via the Internet at http://pubs.acs.org.

#### **AUTHOR INFORMATION**

#### **Corresponding Authors**

\*E-mail: yfxu@ecust.edu.cn (Y.X.). Phone: +86-21-64251399.

\*E-mail: hxie@jding.dhs.org (H.X.). Phone: +86-21-50805897.

\*E-mail: hlli@ecust.edu.cn (H.L.). Phone/Fax: +86-21-64250213.

#### 

Author Contributions

<sup>‡</sup>These authors contributed equally.

#### Notes

The authors declare no competing financial interest.

#### ACKNOWLEDGMENTS

The research is supported in part by the National Key Research and Development Program (Grant 2016YFA0502304), the Shanghai Committee of Science and Technology (Grants 14431902100), Special Program for Applied Research on Super Computation of the NSFC-Guangdong Joint Fund (the second phase) under Grant No.U1501501. Honglin Li is also sponsored by National Program for Special Supports of Eminent Professionals and National Program for Support of Top-notch Young Professionals.

#### **ABBREVIATIONS USED**

EGFR, epidermal growth factor receptor; NSCLC, non-small cell lung cancer; WT, wild-type; TKIs, tyrosine kinase inhibitors; AKT, protein kinase B; ERK, extracellular signal-regulated kinases; TGI, tumor growth inhibition; DIPEA, *N*,*N*-diisopropylethylamine; CH<sub>3</sub>CN, acetonitrile; HATU, 1-[Bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxid hexaflu orophosphate; THF, tetrahydrofuran; DMF, *N*,*N*-dimethylformamide; NMP, 1-Methyl-2-pyrrolidinone

#### REFERENCES

(1) Siegel, R.; Ward, E.; Brawley, O.; Jemal, A. Cancer statistics, 2011. *Ca-Cancer J. Clin.* 2011, *61*, 212-236.

(2) Yarden, Y.; Sliwkowski, M. X. Untangling the ErbB signalling network. *Nat. Rev. Mol. Cell Biol.* 2001, *2*, 127-137.

(3) Oda, K.; Matsuoka, Y.; Funahashi, A.; Kitano, H. A comprehensive pathway map of epidermal growth factor receptor signaling. *Mol. Syst. Biol.* **2005**, *1*, 1-17.

(4) Peters, S.; Zimmermann, S.; Adjei, A. A. Oral epidermal growth factor receptor tyrosine kinase inhibitors for the treatment of non-small cell lung cancer: comparative pharmacokinetics and drug–drug interactions. *Cancer Treat. Rev.* **2014**, *40*, 917-926.

(5) Cohen, M. H.; Williams, G. A.; Sridhara, R.; Chen, G.; McGuinn, W. D.; Morse, D.; Abraham, S.; Rahman, A.; Liang, C.; Lostritto, R.; Baird, A.; Pazdur, R. United states food and drug administration drug approval summary: gefitinib (ZD1839; Iressa) tablets. *Clin. Cancer Res.* **2004**, *10*, 1212-1218.

(6) Cohen, M. H.; Johnson, J. R.; Chen, Y.-F.; Sridhara, R.; Pazdur, R. FDA drug approval summary: erlotinib (Tarceva®) tablets. *Oncologist.* **2005**, *10*, 461-466.

(7) Cataldo, V. D.; Gibbons, D. L.; Pérez-Soler, R.; Quintás-Cardama, A. Treatment of nonsmall-cell lung cancer with erlotinib or gefitinib. *N. Engl. J. Med.* **2011**, *364*, 947-955.

(8) Mitsudomi, T.; Morita, S.; Yatabe, Y.; Negoro, S.; Okamoto, I.; Tsurutani, J.; Seto, T.; Satouchi, M.; Tada, H.; Hirashima, T.; Asami, K.; Katakami, N.; Takada, M.; Yoshioka, H.; Shibata, K.; Kudoh, S.; Shimizu, E.; Saito, H.; Toyooka, S.; Nakagawa, K.; Fukuoka, M. Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung cancer harbouring mutations of the epidermal growth factor receptor (WJTOG3405): an open label, randomised

phase 3 trial. Lancet Oncol. 2010, 11, 121-128.

(9) Rosell, R.; Carcereny, E.; Gervais, R.; Vergnenegre, A.; Massuti, B.; Felip, E.; Palmero, R.; Garcia-Gomez, R.; Pallares, C.; Sanchez, J. M.; Porta, R.; Cobo, M.; Garrido, P.; Longo, F.; Moran, T.; Insa, A.; De Marinis, F.; Corre, R.; Bover, I.; Illiano, A.; Dansin, E.; de Castro, J.; Milella, M.; Reguart, N.; Altavilla, G.; Jimenez, U.; Provencio, M.; Moreno, M. A.; Terrasa, J.; Muñoz-Langa, J.; Valdivia, J.; Isla, D.; Domine, M.; Molinier, O.; Mazieres, J.; Baize, N.; Garcia-Campelo, R.; Robinet, G.; Rodriguez-Abreu, D.; Lopez-Vivanco, G.; Gebbia, V.; Ferrera-Delgado, L.; Bombaron, P.; Bernabe, R.; Bearz, A.; Artal, A.; Cortesi, E.; Rolfo, C.; Sanchez-Ronco, M.; Drozdowskyj, A.; Queralt, C.; de Aguirre, I.; Ramirez, J. L.; Sanchez, J. J.; Molina, M. A.; Taron, M.; Paz-Ares, L. Erlotinib versus standard chemotherapy as first-line treatment for european patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): a multicentre, open-label, randomised phase 3 trial. *Lancet Oncol.* 2012, *13*, 239-246.

(10) Yu, H. A.; Arcila, M. E.; Rekhtman, N.; Sima, C. S.; Zakowski, M. F.; Pao, W.; Kris, M. G.; Miller, V. A.; Ladanyi, M.; Riely, G. J. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clin. Cancer Res.* **2013**, *19*, 2240-2247.

(11) Smaill, J. B.; Rewcastle, G. W.; Loo, J. A.; Greis, K. D.; Chan, O. H.; Reyner, E. L.; Lipka,
E.; Showalter, H. D. H.; Vincent, P. W.; Elliott, W. L.; Denny, W. A. Tyrosine kinase inhibitors.
irreversible inhibitors of the epidermal growth factor receptor: 4-(phenylamino)quinazoline- and
4-(phenylamino)pyrido[3,2-*d*]pyrimidine-6-acrylamides bearing additional solubilizing
functions. *J. Med. Chem.* 2000, *43*, 1380-1397.

(12) Tsou, H.-R.; Overbeek-Klumpers, E. G.; Hallett, W. A.; Reich, M. F.; Floyd, M. B.;

Johnson, B. D.; Michalak, R. S.; Nilakantan, R.; Discafani, C.; Golas, J.; Rabindran, S. K.; Shen, R.; Shi, X.; Wang, Y.-F.; Upeslacis, J.; Wissner, A. Optimization of 6,7-disubstituted-4-(arylamino)quinoline-3-carbonitriles as orally active, irreversible inhibitors of human epidermal growth factor receptor-2 kinase activity. *J. Med. Chem.* **2005**, *48*, 1107-1131.

(13) Engelman, J. A.; Zejnullahu, K.; Gale, C.-M.; Lifshits, E.; Gonzales, A. J.; Shimamura, T.;
Zhao, F.; Vincent, P. W.; Naumov, G. N.; Bradner, J. E.; Althaus, I. W.; Gandhi, L.; Shapiro, G.
I.; Nelson, J. M.; Heymach, J. V.; Meyerson, M.; Wong, K.-K.; Jänne, P. A. PF00299804, an
irreversible pan-ERBB inhibitor, is effective in lung cancer models with EGFR and ERBB2
mutations that are resistant to gefitinib. *Cancer Res.* 2007, *67*, 11924-11932.

(14) Li, D.; Ambrogio, L.; Shimamura, T.; Kubo, S.; Takahashi, M.; Chirieac, L. R.; Padera, R.
F.; Shapiro, G. I.; Baum, A.; Himmelsbach, F.; Rettig, W. J.; Meyerson, M.; Solca, F.; Greulich,
H.; Wong, K. K. BIBW2992, an irreversible EGFR/HER2 inhibitor highly effective in preclinical lung cancer models. *Oncogene* 2008, *27*, 4702-4711.

(15) Dungo, R.; Keating, G. Afatinib: first global approval. Drugs 2013, 73, 1503-1515.

(16) Miller, V. A.; Hirsh, V.; Cadranel, J.; Chen, Y.-M.; Park, K.; Kim, S.-W.; Zhou, C.; Su, W.-C.; Wang, M.; Sun, Y.; Heo, D. S.; Crino, L.; Tan, E.-H.; Chao, T.-Y.; Shahidi, M.; Cong, X. J.; Lorence, R. M.; Yang, J. C.-H. Afatinib versus placebo for patients with advanced, metastatic non-small-cell lung cancer after failure of erlotinib, gefitinib, or both, and one or two lines of chemotherapy (LUX-lung 1): a phase 2b/3 randomised trial. *Lancet Oncology*. **2012**, *13*, 528-538.

(17) Zhou, W.; Ercan, D.; Chen, L.; Yun, C.; Li, D.; Capelletti, M.; Cortot, A. B.; Chirieac, L.; Iacob, R. E.; Padera, R.; Engen, J. R.; Wong, K.; Eck, M. J.; Gray, N. S.; Jänne, P. A. Novel mutant-selective EGFR kinase inhibitors against EGFR T790M. *Nature*. **2009**, *462*, 1070-1074.

(18) Walter, A. O.; Sjin, R. T. T.; Haringsma, H. J.; Ohashi, K.; Sun, J.; Lee, K.; Dubrovskiy, A.; Labenski, M.; Zhu, Z.; Wang, Z.; Sheets, M.; St Martin, T.; Karp, R.; van Kalken, D.; Chaturvedi, P.; Niu, D.; Nacht, M.; Petter, R. C.; Westlin, W.; Lin, K.; Jaw-Tsai, S.; Raponi, M.; Van Dyke, T.; Etter, J.; Weaver, Z.; Pao, W.; Singh, J.; Simmons, A. D.; Harding, T. C.; Allen, A. Discovery of a mutant-selective covalent inhibitor of EGFR that overcomes T790M-mediated resistance in NSCLC. *Cancer Discovery.* **2013**, *3*, 1404-1415.

(19) Tjin Tham Sjin, R.; Lee, K.; Walter, A. O.; Dubrovskiy, A.; Sheets, M.; Martin, T. S.;
Labenski, M. T.; Zhu, Z.; Tester, R.; Karp, R.; Medikonda, A.; Chaturvedi, P.; Ren, Y.;
Haringsma, H.; Etter, J.; Raponi, M.; Simmons, A. D.; Harding, T. C.; Niu, D.; Nacht, M.;
Westlin, W. F.; Petter, R. C.; Allen, A.; Singh, J. In vitro and in vivo characterization of
irreversible mutant-selective EGFR inhibitors that are wild-type sparing. *Mol. Cancer Ther.*2014, *13*, 1468-1479.

(20) Ward, R. A.; Anderton, M. J.; Ashton, S.; Bethel, P. A.; Box, M.; Butterworth, S.;
Colclough, N.; Chorley, C. G.; Chuaqui, C.; Cross, D. A. E.; Dakin, L. A.; Debreczeni, J. É.;
Eberlein, C.; Finlay, M. R. V.; Hill, G. B.; Grist, M.; Klinowska, T. C. M.; Lane, C.; Martin, S.;
Orme, J. P.; Smith, P.; Wang, F.; Waring, M. J. Structure- and reactivity-based development of
covalent inhibitors of the activating and gatekeeper mutant forms of the epidermal growth factor
receptor (EGFR). *J. Med. Chem.* 2013, *56*, 7025-7048.

(21) Cross, D. A. E.; Ashton, S. E.; Ghiorghiu, S.; Eberlein, C.; Nebhan, C. A.; Spitzler, P. J.;
Orme, J. P.; Finlay, M. R. V.; Ward, R. A.; Mellor, M. J.; Hughes, G.; Rahi, A.; Jacobs, V. N.;
Brewer, M. R.; Ichihara, E.; Sun, J.; Jin, H.; Ballard, P.; Al-Kadhimi, K.; Rowlinson, R.;
Klinowska, T.; Richmond, G. H. P.; Cantarini, M.; Kim, D.-W.; Ranson, M. R.; Pao, W.
AZD9291, an irreversible EGFR TKI, overcomes T790M-mediated resistance to EGFR

inhibitors in lung cancer. Cancer Discovery. 2014, 4, 1046-1061.

(22) FDA approves new pill to treat certain patients with non-small cell lung cancer. November 13, 2015. http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm 472525.htm.

(23) Hao Y.; Lyu J.; Qu R.; Sun D.; Zhao Z.; Chen, Z.; Ding, J.; Xie, H.; Xu, Y.; Li, H. Structureguided design of C4-alkyl-1,4-dihydro-2hpyrimido[4,5-*d*][1,3]oxazin-2-ones as potent and mutant-selective epidermal growth factor receptor (EGFR) L858R/T790M inhibitors. *Scientific Reports.* **2017**. *7*, 3830.

(24) Goto, S.; Tsuboi, H.; Kanoda, M.; Mukai, K.; Kagara, K. The process development of a novel aldose reductase inhibitor, FK366. part 1. Improvement of discovery process and new syntheses of 1-substituted quinazolinediones. *Org Process Res Dev.* **2003**, *7*, 700-706.

(25) Hao Y.; Wang X.; Zhang T.; Sun D.; Tong Y.; Xu Y.; Chen H.; Tong L.; Zhu L.; Zhao Z.; Chen, Z.; Ding, J.; Xie, H.; Xu, Y.; Li, H. Discovery and structural optimization of N5-substituted 6, 7-dioxo-6, 7-dihydropteridines as potent and selective epidermal growth factor receptor (EGFR) inhibitors against L858R/T790M resistance mutation. *J Med Chem.* **2016**. *59*, 7111-7124.

(26) Maestro, version 9.0; Schrödinger, LLC: New York, NY, 2009.

(27) Zhao, Z.; Wu, H.; Wang, L.; Liu, Y.; Knapp, S.; Liu, Q.; Gray, N. S. Exploration of type II binding mode: a privileged approach for kinase inhibitor focused drug discovery? *ACS Chem. Biol.* **2014**, *9*, 1230-1241.

(28) Roskoski, R. Classification of small molecule protein kinase inhibitors based upon the structures of their drug-enzyme complexes. *Pharmacol. Res.* **2016**, *103*, 26-48.

(29) Chang, S.; Zhang, L.; Xu, S.; Luo, J.; Lu, X.; Zhang, Z.; Xu, T.; Liu, Y.; Tu, Z.; Xu, Y.; Ren,X.; Geng, M.; Ding, J.; Pei, D.; Ding, K. Design, synthesis, and biological evaluation of novel

#### Journal of Medicinal Chemistry

→ methionine790 mutant. J. Med. Chem. 2012, 55, 2711-2723.

(30) Zhou, W.; Liu, X.; Tu, Z.; Zhang, L.; Ku, X.; Bai, F.; Zhao, Z.; Xu, Y.; Ding, K.; Li, H. Discovery of pteridin-7(8*H*)-one-based irreversible inhibitors targeting the epidermal growth factor receptor (EGFR) kinase T790M/L858R mutant. *J. Med. Chem.* **2013**, *56*, 7821-7837.

(31) Xu, S.; Xu, T.; Zhang, L.; Zhang, Z.; Luo, J.; Liu, Y.; Lu, X.; Tu, Z.; Ren, X.; Ding, K. Design, synthesis, and biological evaluation of 2-oxo-3,4-dihydropyrimido[4,5-*d*]pyrimidinyl derivatives as new irreversible epidermal growth factor receptor inhibitors with improved pharmacokinetic properties. *J. Med. Chem.* **2013**, *56*, 8803-8813.

(32) Liu, X.; Jiang, H.; Li, H. SHAFTS: a hybrid approach for 3D molecular similarity calculation. 1. Method and assessment of virtual screening. *J. Chem. Inf. Model.* **2011**, *51*, 2372-2385.

(33) Gong, J.; Cai, C.; Liu, X.; Ku, X.; Jiang, H.; Gao, D.; Li, H. ChemMapper: a versatile web server for exploring pharmacology and chemical structure association based on molecular 3D similarity method. *Bioinformatics*. **2013**, *29*, 1827-1829.

(34) Zhang, L.; Yuan, J.; Xu, Y.; Zhang, Y. H. P.; Qian, X. New artificial fluoro-cofactor of hydride transfer with novel fluorescence assay for redox biocatalysis. *Chem. Commun.* **2016**, *52*, 6471–6474.

Figure 1. Structures of representative first-, second- and third-generation EGFR inhibitors.

**Figure 2.** A) The 2D drawings of compound **5** and **7**; B) The overlaid docked poses of **5** (shown in orange sticks) and **7** (shown in blue sticks). The gatekeeper, hinge and covalent regions of both wild-type (PDB ID: 4G5J) and double-mutant (PDB ID: 3IKA) EGFR kinases are displayed in yellow sticks and cyan sticks, respectively; C) The sliced view of the predicted binding mode of **14a**. The docked pose of compound **14a** is in yellow sticks. The key residues Met793 and Met790 are shown in cyan sticks. The rest of the protein is demonstrated in a surface mode (PDB ID: 3IKA). D) The 2D interactions of the new scaffold to show strategies of structural modifications. E) The docked pose of compound **20c**. The protein (PDB ID: 3IKA) is shown in a blue cartoon and the key residues are shown in blue sticks. Key H-bond and VDW interactions are measured by distances. F) The docked pose of compound **20f**. The compound **20f** is in orange sticks. The mesh surface is shown to display the shape of the binding site and the S2 pocket. The key residues in the covalent, hinge and S2-pocket regions of both wild-type (PDB ID: 2RGP) and double-mutant (PDB ID: 3W2S) EGFR kinases are displayed in cyan sticks and green sticks, respectively

**Figure 3.** Inhibitory effects of compound **20g** for EGFR and downstream signaling transduction in H1975 cells (A) and A431 cells (B).

**Figure 4.** Target duration *in vitro*. NCI-H1975 cells were treated with **20g** (A) or compound **6** (B) for 2 hours, then washed with PBS for 3 times. Target inhibition at different time points was detected by Western blot.

**Figure 5.** Preliminary *in vivo* antitumor efficiency study of compound **20g** (50 mg/kg/day) against H1975 and A431 NSCLC xenograft mouse models. (A) H1975 xenograft mouse model (n = 6) tumor volumes and (C) body weights, (B) A431 xenograft mouse model (n = 6) tumor volumes and (D) body weights were recorded every 2-3 days. All values expressed as mean  $\pm$ SEM. (\*) p < 0.05 or (\*\*\*) p < 0.001 (Student's *t* test) *vs* the vehicle group.

## SCHEMES



## Scheme 1. Synthesis of Pyrimido[4,5-d]pyrimidine-2,4(1H,3H)-dione Derivatives 14a-e<sup>a</sup>

<sup>*a*</sup>Reagents and conditions: (a) (Boc)<sub>2</sub>O, Et<sub>3</sub>N, CH<sub>3</sub>OH, r.t., 24 h, 64%; (b) ethyl 2,4dichloropyrimidine-5-carboxylate, DIPEA, CH<sub>3</sub>CN, reflux, 2 h, 86%; (c) 1M NaOH, THF, 50 <sup>o</sup>C, 3 h, 96%; (d) arylamine, CH<sub>3</sub>CN, reflux, overnight, 42-89%; (e) methylamine hydrochloride, HATU, DIPEA, dry DMF, r.t., overnight, 31-47%; (f) 1,1'-carbonyldiimidazole, K<sub>2</sub>CO<sub>3</sub>, dry THF, reflux, overnight, 35-42%; (g) trifluoroacetic acid, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 4 h, 81-90%; (h) acrylyl chloride, NMP/CH<sub>3</sub>CN, 0 <sup>o</sup>C to r.t., overnight, 47-53%.





<sup>*a*</sup>Reagents and conditions: (a) ammonium hydroxide, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 2 h, 92%; (b) *tert*-butyl(3aminophenyl)carbamate, DIPEA, isopropanol, 40 °C, 5 h, 84%; (c) arylamine, trifluoroacetic acid, isopropanol, reflux, overnight, 67%; (d) 1,1'-carbonyldiimidazole, K<sub>2</sub>CO<sub>3</sub>, dry THF, reflux, overnight, 60%; (e) haloalkane, Cs<sub>2</sub>CO<sub>3</sub>, DMF, r.t., overnight, 55-61%; (f) trifluoroacetic acid, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 5 h, 78-82%; (g) acrylyl chloride, NMP/CH<sub>3</sub>CN, 0 °C to r.t., overnight, 37-49%.

## FIGURES

## Figure 1



## Figure 2



## Figure 3



### Figure 4



## Figure 5



## TABLES

## Table 1. In Vitro EGFR Inhibitory Activity and Antiproliferation Activity of Compounds

**14a-e**<sup>*a*</sup>



Compd.	$R^1$	R <sup>2</sup>	Enzyme inhibitory activity (IC <sub>50</sub> , nM)		Enzyme Selectivity	Cellular antiproliferative activity (IC <sub>50</sub> , nM)		Cellular Selectivity	Solubility	
			WT	L858R/ T790M	WT:DM	A431	H1975	A431: H1975	$(\mu g/mL) (a) PBS (pH 7.4)^c$	
14a	-OCH <sub>3</sub>		180±79	46±26	3.9	> 10000	> 10000	$\mathrm{ND}^b$	< 10	
14b	-OCH <sub>3</sub>		341±341	70±34	4.9	> 10000	> 10000	ND	61	
14c	-OCH <sub>3</sub>	N S	314±113	91±12	3.5	8373±1970	573±144	15	45	
14d	-OCH <sub>3</sub>		137±2.8	30±6.9	4.6	3892±3642	230±142	17	531	
14e	Н	Н	82±63	68±39	1.2	> 10000	9264±4791	ND	< 10	
5			272±127	20±2.9	14	641±282	31±34	21		
	<sup><i>a</i></sup> Kinase antiproli	activity iferation	assays we	ere examin he compou	ed by using nds were em	the ELISA	-based EGF	R-TK assay	r. The (SRB)	

colourimetric assay. Data are averages of at least three independent determinations and reported as the means  $\pm$  SDs (standard deviations). <sup>b</sup>Not determined. <sup>c</sup>Aqueous solubility of these derivatives was examined by using UV-visible spectrophotometer in PBS buffer (0.1 M, pH 7.4).

ACS Paragon Plus Environment



## Table 2. In Vitro EGFR Inhibitory Activity and Antiproliferation Activity of Compounds



**20a-l**<sup>*a*</sup>





14a-l

21 22 23 24 23 24 20 20 24	R <sup>3</sup>	$R^4$	R <sup>5</sup>	EGFR enzyme inhibitory activity (IC <sub>50</sub> , nM)		Enzyme Selectivity	Cellular antiproliferative activity (IC <sub>50</sub> , nM)		Cellular Selectivity	Solubility (µg/mL) @PBS	
24 25 26			-	WT	L858R/ T790M	WT:DM	A431	H1975	A431: H1975	$(pH 7.4)^{c}$	
27 28 29 30 31 32	22	2-OCH <sub>3</sub>	× N N	111±31	2.5±0.2	44	> 10000	175±122	$\mathrm{ND}^b$	229	
33 34 35 <b>20b</b> 36 37		2-OCH <sub>3</sub>	N N N	13±1.6	0.4±0.3	33	4130±2559	60±34	69	184	
38 39 40 41 <b>20c</b> 42 43	22.2	2-OCH <sub>3</sub>	N N N	20±14	0.8±0.3	25	> 10000	50±27	ND	176	
44 45 46 47 48 40	2	2-OCH <sub>3</sub>	N N N	1.2±0.3	1.9±0.2	0.6	3300±909	278±166	12	137	
50 51 52 <b>20</b> e 53 54	3	2-OCH <sub>3</sub>	N N N	0.7±0.4	1.0±0.2	0.7	2043±963	189±150	11	99	
55 56 57 58 59 60				F	\CS Paragon	Plus Environr	nent				



Table 3. Mouse Pharmacokinetics Parameters for Compo	ind 20g
--	---------

Dose (route)	<i>T</i> <sub>1/2</sub> (h)	T <sub>max</sub> (h)	C <sub>max</sub> (ng/mL)	AUC <sub>(0-t)</sub> (ng·h/mL)	$\begin{array}{c} AUC_{(0-\infty)}\\ (ng \cdot h/mL) \end{array}$	V <sub>z</sub> (mL/kg)	CL (mL/h/kg)	F (%)
1 mg/kg (IV)	0.9	0.1	311	181	186	4530	5370	-
10 mg/kg (PO)	1.1	2.0	271	650	658	-	-	35

# **Table of Contents Graphic**

