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Synthesis, Characterization, and Antimicrobial Evaluation of Sulfonamides Containing N-Acyl Moieties Catalyzed by Bismuth(III) Salts Under Both Solvent and Solvent-Free Conditions

Hadi Adibi $^{\rm a}$, Ahmad Reza Massah $^{\rm b}$, Mohammad Bagher Majnooni $^{\rm a}$, Sherita Shahidi $^{\rm b}$, Maryam Afshar $^{\rm b}$, Ramin Abiri $^{\rm c}$ & Hamid Javaherian Naghash $^{\rm b}$

^a Department of Medicinal Chemistry , Faculty of Pharmacy, Kermanshah University of Medical Sciences , Kermanshah, Islamic Republic of Iran

^b Department of Chemistry, Islamic Azad University, Shahreza Branch, Shahreza, Islamic Republic of Iran

^c Department of Microbiology , Faculty of Medicine, Kermanshah University of Medical Sciences , Kermanshah, Islamic Republic of Iran

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SYNTHESIS, CHARACTERIZATION, AND ANTIMICROBIAL EVALUATION OF SULFONAMIDES CONTAINING *N*-ACYL MOIETIES CATALYZED BY BISMUTH(III) SALTS UNDER BOTH SOLVENT AND SOLVENT-FREE CONDITIONS

Hadi Adibi,¹ Ahmad Reza Massah,² Mohammad Bagher Majnooni,¹ Sherita Shahidi,² Maryam Afshar,² Ramin Abiri,³ and Hamid Javaherian Naghash²

¹Department of Medicinal Chemistry, Faculty of Pharmacy, Kermanshah University of Medical Sciences, Kermanshah, Islamic Republic of Iran ²Department of Chemistry, Islamic Azad University, Shahreza Branch, Shahreza, Islamic Republic of Iran

³Department of Microbiology, Faculty of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Islamic Republic of Iran

Efficient N-acylation of sulfonamides with both readily available carboxylic acid chlorides and anhydrides has been carried out with catalysis by bismuth(III) salts including BiCl₃ and Bi (OTf)₃. The reactions proceed rapidly in both heterogeneous and solvent-free conditions and afforded the corresponding N-acylsulfonamides in good to excellent yields. The mild reaction conditions and low toxicity of bismuth salts make this procedure attractive and in close agreement with the goals of green chemistry. Some of the synthesized compounds were evaluated in vitro as antimicrobial agents against representative strains of Gram-positive (Staphylococcus aureus ATCC 25922, clinical strains of Staphylococcus aureus VISA and Enterococcus spp.) and Gram-negative bacteria (Pseudomonas aeruginosa ATCC 27853, clinical strains of Klebsiella pneumonia and Escherichia coli) and as antifungal agents against Candida albicans (clinically isolated) by both disc diffusion and minimal inhibition concentration (MIC) methods. All these bacteria and fungi studied were screened against some antibiotics to compare with our chemicals' zone diameters.

Keywords: Antimicrobial activity; bismuth(iii) salts; N-acylation; N-acylsulfonamide; solvent-free

INTRODUCTION

Sulfonamides are among the most widely used antibacterial agents in the world, chiefly because of their low cost, low toxicity, and excellent activity against common bacterial diseases.^[1] The synergetic action of sulfonamides with trimethoprim has

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Address correspondence to Hadi Adibi, Department of Medicinal Chemistry, Faculty of Pharmacy, Kermanshah University of Medical Sciences, Kermanshah 67145-1673, Islamic Republic of Iran, and Ahmad Reza Massah, Department of Chemistry, Islamic Azad University, Shahreza Branch, Shahreza 86145-311, Islamic Republic of Iran. E-mail: hadibi@kums.ac.ir; massah@iaush.ac.ir

brought about enormous resurgence of sulfonamide usage everywhere over the past decade. *N*-Acylsulfonamides have received considerable attention because of their diverse biological activities as precursors of therapeutic agents for Alzheimer's disease,^[2] antibacterial inhibitors of tRNA synthetases,^[3] prostaglandin F1a sulfonamides for the potential treatments of osteoporosis,^[4] antagonists for angiotensin II,^[5] and leukotriene D4-receptors.^[6]

The most common methods for N-acylation of sulfonamides with different acylating agents have utilized suitable bases such as pyridine,^[7,8] 4-N.N-dimethyl pyridine,^[9,10] and alkali hydroxides.^[11–13] Formation of bis-acvlated by-products is the most important restriction of these methods. Recently, the first solvent-free procedure for the N-acylation of sulfonamides in the presence of potassium carbonates has been performed that overcomes the restriction of the other methods.^[14] On the other hand, there are a few reports mentioning this transformation under acidic conditions. Some of the methods reported include acylation of sulfonamides with concentrated H₂SO₄ in the carboxylic acid anhydride as solvent^[15] or in acetonitrile^[16] and also using Fe-exchanged montmorillonite K10.^[17] Very recently, we have demonstrated the applicability of protic solid acids such as silica chloride^[18] and silica phosphoric acid^[19] for N-acylation of sulfonamides. Unfortunately, many of these methods have drawbacks such as long reaction times, vigorous reaction conditions, use of expensive or unavailable reagents, poor yields of products, and tedious workups. Furthermore, most of these methods occur in solvent media, contrary to green chemistry. Clearly, there is a need for development of a new, inexpensive, benchtop acid that can promote this reaction in a catalytic way.

Bismuth compounds have recently received a lot of attention because of their low toxicity, low cost, and good stability.^[20–23] Bismuth(III) salts have been reported as catalyst for epoxide opening,^[24] Mannich-type reactions,^[25] Mukaiyama-aldol reactions,^[26] Claisen rearrangement,^[27] deprotection of 1,1-diacetates,^[28] and acetylation and benzoylation of alcohols and phenols.^[29] Bi(OTf)₃ and BiCl₃ are particularly attractive because they are commercially available or can be easily prepared from commercially available starting materials. In continuation of our studies in this regard,^[30] we decided to apply these reagents to the N-acylation of sulfonamides and evaluate their antimicrobial activity.

RESULTS AND DISCUSSION

Chemistry

An efficient method is reported here using $Bi(OTf)_3$ and $BiCl_3$ as catalysts for N-acylation of different sulfonamides with some carboxylic acid anhydride and chloride in solvent-free and heterogeneous conditions (Scheme 1). In a simple procedure, a mixture of reactants was vigorously stirred at room temperature in solvent-free condition or refluxed in suitable solvents for the appropriate times. The progress of the reactions was monitored by thin-layer chromatography (TLC), and the products were isolated in good to excellent yields (82–98%) in an easy workup procedure.

At first, the reaction was optimized with respect to the solvent, amount of catalyst, temperature, and reaction time. To find the best solvent, the reaction of



Scheme 1. BiCl3- and Bi(OTf)3-catalyzed N-acylation of sulfonamides.

benzenesulfonamide with acetic anhydride was carried out in different solvents. Even though the reactions in CHCl₃ and CH₂Cl₂ led to a small amount of side products, an excellent yield was obtained, and these were selected as solvents for further studies. Also, the N-acylation of benzenesulfonamide was tried with varying amounts of catalyst in solvent and solvent-free conditions. We attempted the reaction of benzenesulfonamide with 2 equivalents acetic anhydride or acetyl chloride in the presence of 5 mol% of Bi(OTf)₃ at room temperature or in refluxed CH_2Cl_2 . The reactions were completed within 1-5 min, and the pure N-acetylsulfonamide was obtained in 91–94% yield by just passing through a small silica-gel column. In a similar manner, benzenesulfonamide was also converted to N-acetylsulfonamide using 10 mol% of BiCl₃ within 13–17 min. By reducing the amount of acylating agents or catalysts, there were a considerable change in reaction time and yield in both solvent and solvent-free conditions. To evaluate the role of catalysts, we tried the N-acylation of benzenesulfonamide with acetic anhydride and acetyl chloride in the absence of $Bi(OTf)_3$ or $BiCl_3$. The reaction did not occur after 48 h. The best conditions for the N-acylation of benzenesulfonamide are summarized in Table 1.

The BiCl₃- and Bi(OTf)₃-catalyzed N-acylation reactions were applied to a range of sulfonamides, carboxylic acid anhydrides, and carboxylic acid chlorides (Tables 2 and 3). The reaction proceeded very efficiently in all cases. As shown in Table 2, benzenesulfonamide, 4-methyl benzenesulfonamide, and methanesulfonamide underwent clean and remarkably fast N-acylation reaction with several aliphatic and aromatic carboxylic acid anhydrides. The aliphatic carboxylic acid anhydrides with long chains, like pentanoic anhydride (Table 2, entries 4, 10, and 16) and also the relatively bulky isobutanoic anhydride (Table 2, entries 5, 11, and 17), were converted to the corresponding *N*-acylsulfonamides in excellent yields in

Entry	Conditions	Yield (%)
1	Bi(OTf) ₃ 5%/Solvent-free/25°C/Acetyl chloride/3 min	92
2	Bi(OTf) ₃ 5%/Solvent-free/25°C/Acetic anhydride/1 min	92
3	Bi(OTf) ₃ 5%/CH ₂ Cl ₂ /Reflux/Acetyl chloride/5 min	91
4	Bi(OTf) ₃ 5%/CH ₂ Cl ₂ /Reflux/Acetic anhydride/2 min	94
5	BiCl ₃ 10%/Solvent-free/25°C/Acetyl chloride/16 min	88
6	BiCl ₃ 10%/Solvent-free/25°C/Acetic anhydride/13 min	91
7	BiCl ₃ 10%/CHCl ₃ /Reflux/Acetyl chloride/17 min	87
8	BiCl ₃ 10%/CHCl ₃ /Reflux/Acetic anhydride/15 min	90

Table 1. Best reaction conditions in the N-acylation of benzenesulfonamide

	0 R-S- 0	NH ₂ + R'		R' —	BiCl ₃ or Solvent- or Solve	Bi(OTf) free, RT ent, Reflu	¹ 3 - R	0 - - - N- H	0 -II -C—R'	
			Ca	ıtalyst: Bi	(OTf) ₃ (5	i%)	C	Catalyst: H	BiCl ₃ (10%	/0)
			Solver	nt-free	СН	2Cl ₂	Solver	nt-free	CH	ICl ₃
Entry	R	R ′	Time ^a	Yield ^b	Time ^a	Yield ^b	Time ^a	Yield ^b	Time ^a	Yield ^b
1	CH ₃	CH ₃	1	90	2	90	10	88	10	90
2	CH ₃	C ₂ H ₅	1	90	2	91	10	85	12	88
3	CH ₃	$C_{3}H_{7}$	1	90	1	91	8	85	10	88
4	CH ₃	C ₄ H ₉	1	91	5	90	10	88	13	90
5	CH ₃	CH(CH ₃) ₂	1	90	2	91	10	90	13	91
6	CH ₃	Ph	15	90	30	91	60	82	60	87
7	CH ₃ -Ph	CH ₃	2	90	2	94	15	89	20	90
8	CH ₃ -Ph	C_2H_5	3	91	2	93	17	90	20	92
9	CH ₃ -Ph	C_3H_7	3	91	3	90	20	88	20	90
10	CH ₃ -Ph	C_4H_9	3	92	3	96	22	90	20	88
11	CH ₃ -Ph	$CH(CH_3)_2$	1	92	1	91	17	88	20	89
12	CH ₃ -Ph	Ph	50	90	30	88	70	80	80	82
13	Ph	CH ₃	1	92	2	94	13	91	15	90
14	Ph	C_2H_5	2	93	2	92	17	90	17	89
15	Ph	C_3H_7	3	90	2	95	12	85	17	90
16	Ph	C_4H_9	3	94	3	92	16	86	20	90
17	Ph	$CH(CH_3)_2$	1	94	1	96	15	88	20	92
18	Ph	Ph	30	90	35	90	70	88	70	85
19	Ph	NO ₂ -Ph	10^{c}	15	10^{c}	10	12^{c}	10	12^{c}	10

Table 2. N-Acylation of sulfonamides with carboxylic acid anhydride in solvent or solvent-free conditions

^aMinutes.

^bIsolated yield (%).

^cIn hours.

both solvent and solvent-free conditions. These results show that there was no steric effect from the substituents at anhydride moiety. Comparisons between the results of Tables 2 and 3 show that carboxylic acid anhydrides were found to be more reactive than carboxylic acid chlorides. Also, N-acylation of sulfonamides with electron-withdrawing groups containing carboxylic acid anhydrides needed a longer reaction time to form *N*-acylsulfonamides. For example, the reaction of benzenesulfonamide with 4-nitrobenzoic anhydride did not proceed well, and only 10-15% of product was obtained after 10-12 h (Table 2, entry 19). These results show that the acylinium ion was formed and then reacted with sulfonamide. This fact was further supported by N-acylation of benzenesulfonamide with benzoic-4-nitrobenzoic anhydride. After completion of the reaction, we observed that the only product is *N*-benzoyl benzensulfonamide. In fact, sulfonamide reacted with a carbonyl group to form more stable acylinium ions (Scheme 2). On the other hand, comparison between the results obtained in solution and those under solvent-free conditions show that reaction

Table 3. N-Acylation of sulfonamides with carboxylic acid chloride in solvent or solvent-free conditions

			С	atalyst: Bi	(OTf) ₃ (59	%)	(Catalyst: E	BiCl ₃ (10%)
			Solver	nt-free	СН	₂ Cl ₂	Solver	nt-free	CH	Cl ₃
Entry	R	R ″	Time ^a	Yield ^b	Time ^a	Yield ^b	Time ^a	Yield ^b	Time ^a	Yield ^t
1	Ph	CH ₃	3	92	5	91	16	88	17	87
2	Ph	C_2H_5	5	95	5	96	14	88	15	90
3	Ph	Ph	15	91	40	95	45	90	45	85
4	CH ₃ -Ph	CH ₃	3	90	5	86	10	89	12	90
5	CH ₃ -Ph	C_2H_5	7	95	5	92	10	89	10	89
6	CH ₃ -Ph	Ph	20	90	45	85	50	80	55	83
7	CH ₃	CH ₃	1	94	5	95	10	90	12	88
8	CH ₃	C_2H_5	1	95	1	94	10	92	10	90
9	CH ₃	Ph	10	98	25	93	30	85	35	88

^aMinutes.

^bIsolated yield (%).

green chemistry. Furthermore, the experimental results show that $Bi(OTf)_3$ is more reactive and works better than $BiCl_3$ in terms of amount of catalyst, temperature, reaction times, and the yields of products (Tables 1–3).

Antimicrobial Activity

The test compounds were evaluated for their antibacterial activity against three Gram-positive cocci [*Staphylococcus aureus* ATCC 25922, clinical strains of *Staphylococcus aureus* VISA (vancomycin intermediate *Staphylococcus aureus*), and *Enterococcus faesium*), three Gram-negative bacilli (*Pseudomonas aeruginosa* ATCC 27853, clinical strains of *Klebsiella pneumonia*, and *Escherichia coli*), and the yeast (clinical strains of *Candida albicans*) using the microbroth-dilution



Scheme 2. N-Acylation of benzenesulfonamide with benzenoic-4-nitrobenzoic anhydride.

T	ble 4. MICs	(in μg/mL) va	lues of N-acylsulfo	namides against bac	teria and fungi		
Compound	<i>E. coli</i> (clinically isolated)	<i>E. faecium</i> (clinically isolated)	K. pneumoniae (clinically isolated)	S. aureus VISA (clinically isolated)	S. aureus ATCC 25922	P. aeruginosa ATCC 27853	Candida albicans (clinically isolated)
H ₃ C-C-SO ₂ NHCOCH ₃	256	32	512	l	32	512	64
So ₂ NHCOC ₅ H ₁₁ - <i>n</i>	512	32	512		32	256	128
H ₃ C SO ₂ NHCOC ₃ H ₇ -n	512	64	128		32	512	256
H ₃ C-C-SO2NHCOC ₃ H _{11-n}	512	32			32	512	512
CH ₃ SO ₂ NHCOCH ₃	512	64		512	128	512	512
H ₃ C So ₂ NHCOC ₃ H ₇ -iso	128	512	512	512	256	256	512
SO2NHCOC3H7-iso	512	64			256	512	512
CH ₃ SO ₂ NHCOC ₃ H ₇ -iso	512	512			256	512	1024
SO2NHCOC3H7-n	512	256	I	I	512	512	512
Fluconazole							4
Ciprofloxacin	4	4	8	2	4	4	
Gentamicin	4	8	7	8	4	4	
Sulfamethoxazole	7	4	8	4	2	8	
Ceftriaxone	32	8	16	8	4	16	

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L	Fable 5. Inhib	ition zone of <i>l</i>	V-acylsulfonamide	s against bacteria an	d fungi (in mm)		
Compound	<i>E. coli</i> (clinically isolated)	E. faecium (clinically isolated)	K. pneumoniae (clinically isolated)	S. aureus VISA (clinically isolated)	S. aureus ATCC 25922	P. aeruginosa ATCC 27853	Candida albicans (clinically isolated)
H ₃ C Solution	15	10	10	15	13	10	Π
SO2NHCOC5H11-n	18	15	12	10	25	14	13
H ₃ C SO ₂ NHCOC ₃ H ₇ -n	14	20	10	I	18	12	17
H ₃ C - SO ₂ NHCOC ₅ H ₁₁ - <i>n</i>	15	14	6		30		15
CH ₃ SO ₂ NHCOCH ₃	14	17			10	10	13
H ₃ C Solution Solution Solution		15		14	30		14
SO2NHCOC3H7-iso	12	13	8	15	18		14
CH ₃ SO ₂ NHCOC ₃ H ₇ -iso	15	14	15	15	20		15
\bigvee SO ₂ NHCOC ₃ H ₇ -n	10	13	12	I	29	12	10
Fluconazole							17
Gentamicin	20	20	20	20	25	18	
Co-trimoxazole	21			20	18	22	
Cefoxitin	20	18	18	14	16	15	
Imipenem	30	25	25	23	30	25	
Azithromycin	27		17	27	27	2	
Tetracycline			15	25	30	20	

method.^[31] The obtained MIC (minimum inhibitory concentration) values were compared to gentamicin, ciprofloxacin, sulfamethoxazole, ceftriaxone, and fluconazole as the reference drugs and are presented in Table 4. Inhibition zones were determined by the disc diffusion method^[32] and compared with fluconazole ($25 \mu g$), gentamicin ($10 \mu g$), co-trimoxazole ($1.25:23.75 \mu g$), cefoxitin ($30 \mu g$), imipenem ($10 \mu g$), tetracycline ($30 \mu g$), and azithromycin ($15 \mu g$) as the reference drugs (Table 5).

Microbiological results showed that the tested compounds possessed a broad spectrum of antibacterial activity against the screened microorganisms; however, they exhibited less antibacterial potency than the control drugs. The results reported in Table 4 indicate that the tested compounds are able to inhibit growth of the screened pathogens, showing MIC values between 32 and $1024 \mu g/mL$. Among the synthesized compounds, *N*-ethanoyl methansulfonamide and *N*-isobutanoyl-4-methyl benzenesulfonamide were also potent against clinical VISA strains, showing an MIC value of $512 \mu g/mL$. All the other compounds exhibited no antibacterial activity against the pathogens.

Moreover, the tested compounds also possessed antimycotic activity against the yeast *Candida albicans*, showing MIC values between 64 and $1024 \mu g/mL$, whereas the antifungal potency of the compared control drug fluconazole was observed to be better than the corresponding compounds, showing an MIC value of $4 \mu g/mL$. As seen in Table 5, *N*-butanoyl-4-methyl benzenesulfonamide shows the best antifungal activity against *C. albicans*, and its potency compares to that of the reference antibiotic (fluconazole).

CONCLUSION

In summary, we found that $Bi(OTf)_3$ and $BiCl_3$ are efficient catalysts for the Nacylation of different sulfonamides. This method offers several advantages, including mild reaction conditions, use of green catalysts, and no formation of bis-acylated byproducts. Furthermore, this solvent-free protocol is a simple, inexpensive, safe, and ecofriendly alternative to the other methods for the N-acylation of sulfonamides. Also, antimicrobial activity of some of the synthesized sulfonamides demonstrated moderate to good inhibition against all the strains tested. Finally, these preliminary results are promising and are beneficial for further study in developing new compounds.

EXPERIMENTAL

General

All chemicals were purchased from Merck and Fluka chemical companies. The products were characterized by comparing the physical data with those of known samples or by their spectral data. Infrared (IR) spectra were recorded on Nicollet (impact 400D model) Fourier transform (FT)-IR spectrophotometer. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker DRX 300 Avance spectrophotometer in dimethylsulfoxide (DMSO- d_6) as the solvent and tetramethylsilane

(TMS) as internal standard. Column chromatography was performed using silica gel 60 (230-400 mesh). All yields refer to isolated yield.

General Procedure for N-Acylation of Sulfonamides Under Solvent-Free Conditions

Carboxylic acid chloride (2 mmol) or carboxylic acid anhydride (2 mmol) were added to a vigorously stirred mixture of sulfonamide (1 mmol) and BiCl₃ (0.1 mmol) or $Bi(OTf)_3$ (0.05 mmol) at room temperature. The progress of the reaction was monitored by thin-layer chromatography (TLC). After completion of the reaction, ethyl acetate (20 mL) was added, and the solid catalyst was removed by filtration. The filtrate was washed with water and dried over MgSO₄. After evaporation of the solvent, the crude product was purified by column chromatography on silica gel (mixed EtOAc-petroleum ether as eluent) or recrystallized (toluene or ethyl acetate/n-hexane mixed solvent) to afforded the corresponding N-acyl sulfonamide in good to excellent yields.

General Procedure for N-Acylation of Sulfonamides in Solvent

A mixture of sulfonamide (1 mmol), carboxylic acid chloride (2 mmol), or carboxylic acid anhydride (2 mmol) was refluxed in $CHCl_3$ or CH_2Cl_2 (5 mL) in the presence of BiCl₃ (0.1 mmol) or Bi(OTf)₃ (0.05 mmol) for an appropriate time (Table 1). The progress of the reaction was monitored by TLC. After completion of the reaction, CHCl₃ or CH₂Cl₂ (20 mL) was added, and the solid catalyst was removed by filtration. The filtrate was washed with water and dried over MgSO₄. After evaporation of the solvent, the crude product was purified as described for the solvent-free method.

Structural assignments of the products are based on their IR, ¹H NMR, ¹³C NMR, and CHN analysis and also by comparison of their melting points with those of known compounds.

Spectral Data of Isolated N-Acylsulfonamides

N-Ethanoyl benzenesulfonamide (Table 2, entry 1). Mp 120–122°C; IR v (cm⁻¹): 3125, 2901, 1699, 1468, 1356, 1243, 1169, 867, 689; ¹H NMR (DMSO-d₆) δ (ppm) 1.92 (s, 3H), 7.59–7.73 (m, 3H), 7.91 (2H, dd, $J_1 = 7.2$ Hz, $J_2 = 1.5$ Hz) 12.10 (s, 1H);¹³C NMR (DMSO-d₆) δ (ppm) 23.68, 127.90, 129.58, 134.08, 139.81, 169.23. (Found: C, 48.61; H, 4.77; N, 6.88. C₈H₉NO₃S requires C, 48.23; H, 4.55; N, 7.03%.)

N-Propanoyl benzenesulfonamide (Table 2, entry 2). Mp 75–76°C; IR ν (cm⁻¹): 3246, 2986, 1710, 1461, 1341, 1156, 845, 766, 595 cm¹; ¹H NMR (DMSO-d₆) δ (ppm) 0.88 (t, 3H, J=7.5 Hz), 2.22 (q, 2H, J=7.5 Hz), 7.59–7.70 (m, 3H), 7.91-7.94 (m, 2H), 12.06 (s, 1H);¹³C NMR (DMSO-d₆) δ (ppm) 8.68, 29.15, 126.04, 127.87, 129.37, 129.56, 172.72. (Found: C, 50.88; H, 5.29; N, 6.68. C₉H₁₁NO₃S requires C, 50.69; H, 5.20; N, 6.57%.)

N-Butanoyl benzenesulfonamide (Table 2, entry 3). Mp 99–101°C; IR ν (cm⁻¹) 3216, 1687, 1608, 1330, 1171; ¹H NMR (DMSO-d₆) δ (ppm) 0.73 (t, 3H, J = 7.3 Hz), 1.41 (sext, 2H, J = 7.3 Hz), 2.16 (t, 2H, J = 7.3 Hz), 7.59–7.92 (m, 5H), 12.05 (s, 1H); ¹³C NMR (DMSO-d₆) δ (ppm) 13.59, 17.94, 37.59, 127.86, 129.56, 134.05, 139.90, 171.96. (Found: C, 52.65; H, 5.59; N, 6.36. C₁₀H₁₃NO₃S requires C, 52.85; H, 5.77; N, 6.16%.)

N-Pentanoyl benzenesulfonamide (Table 2, entry 4). Mp 82–84°C; IR ν (cm⁻¹) 3101, 1688, 1608, 1350, 1164; ¹H NMR (DMSO-d₆) δ (ppm) 0.77 (t, 3H, J=7.2 Hz), 1.13 (sext, 2H, J=7.4 Hz), 1.37 (quin, 2H, J=7.4 Hz), 2.19 (t, 2H, J=7.3 Hz), 7.59–7.91 (m, 5H), 12.03 (s, 1H); ¹³C NMR (DMSO-d₆) δ (ppm) 13.47, 21.31, 26.02, 34.97, 127.36, 129.04, 133.55, 139.38, 171.57. (Found: C, 54.92; H, 6.07; N, 5.68. C₁₁H₁₅NO₃S requires C, 54.75; H, 6.27; N, 5.80%.)

N-Isobutanoyl benzenesulfonamide (Table 2, entry 5). Mp 120–122°C; IR ν (cm⁻¹) 3257, 2979, 2880, 1719, 1434, 1335, 1176, 1090, 891, 758; ¹H NMR (DMSO-d₆) δ (ppm) 0.92 (d, 6H, J=6.8 Hz), 2.45 (sept, 1H, J=6.8 Hz), 7.59–7.91 (m, 5H), 12.07 (s, 1H); ¹³C NMR (DMSO-d₆) δ (ppm) 18.83, 34.71, 127.81, 129.56, 134.04, 139.79, 175.75. (Found: C, 52.71; H, 5.89; N, 6.28. C₁₀H₁₃NO₃S requires C, 52.85; H, 5.77; N, 6.16%.)

N-Benzoyl benzenesulfonamide (Table 2, entry 6). Mp 137–139°C; IR ν (cm⁻¹): 3277, 3059, 1699, 1580, 1455, 1342, 1177, 1072, 900, 834, 575; ¹H NMR (DMSO- d_6) δ (ppm) 7.36–7.53 (m, 3H), 7.56–7.58 (m, 2H), 7.83–7.87 (m, 3H), 8.01–8.03 (m, 2H), 12.56 (s,1H);¹³C NMR (DMSO- d_6) δ (ppm) 126.03, 128.12, 128.87, 129.07, 129.38, 129.60, 133.77, 134.12, 165.91. (Found: C, 59.95; H, 4.38; N, 5.21. C₁₃H₁₁NO₃S requires C, 59.76; H, 4.24; N, 5.36%.)

N-Ethanoyl-4-methyl benzenesulfonamide (Table 2, entry 7). Mp 136–138°C; IR ν (cm⁻¹) 3290, 1725, 1607, 1441, 1336, 1217, 1171, 867, 669 cm¹; ¹H NMR (DMSO-*d*₆) δ (ppm) 1.90 (s, 3H), 2.38 (s, 3H), 7.40 (d, 2H, *J*=8 Hz), 7.79 (d, 2H, *J*=8.0 Hz), 12.01 (s, 1H); ¹H NMR (DMSO-*d*₆) δ (ppm) 21.50, 23.64, 128.00, 129.96, 136.96, 144.62, 169.13. (Found: C, 51.01; H, 5.37; N, 6.57. C₉H₁₁NO₃S requires C, 50.69; H, 5.20; N, 6.57%.)

N-Propanoyl-4-methyl benzenesulfonamide (Table 2, entry 8). Mp 104–106°C; IR ν (cm⁻¹) 3143, 1715, 1596, 1344, 1191; ¹H NMR (DMSO- d_6) δ (ppm) 0.87 (t, 3H, J=7.5 Hz), 2.20 (q, 2H, J=7.5 Hz), 2.38 (s, 3H), 7.41 (d, 2H, J=8.1 Hz), 7.79 (d, 2H, J=8.1 Hz), 11.96 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm) 8.70, 21.50, 29.10, 127.97, 129.93, 137.09, 144.56, 172.61. (Found: C, 52.61; H, 5.87; N, 6.36. C₁₀H₁₃NO₃S requires C, 52.85; H, 5.77; N, 6.16%.)

N-Butanoyl-4-methyl benzenesulfonamide (Table 2, entry 9). Mp 79–81°C; IR ν (cm⁻¹) 3090, 1680, 1601, 1349, 1158; ¹H NMR (DMSO-d₆) δ (ppm) 0.74 (t, 3H, J=7.3 Hz), 1.40 (sext, 2H, J=7.3 Hz), 2.15 (t, 2H, J=7.3 Hz), 2.38 (s, 3H), 7.4 (d, 2H, J=8.1 Hz), 7.79 (d, 2H, J=8.1 Hz), 11.97 (s, 1H); ¹³C NMR (DMSO-d₆) δ (ppm) 13.60, 17.96, 21.49, 37.59, 127.95, 129.93, 137.08, 144.55, 171.85. (Found: C, 54.63; H, 6.21; N, 5.97. C₁₁H₁₅NO₃S requires C, 54.75; H, 6.27; N, 5.80%.)

N-Pentanoyl-4-methyl benzenesulfonamide (Table 2, entry 10). Mp 88–89°C; IR ν (cm⁻¹) 3271, 2960, 2868, 1719, 1593, 1428, 1342, 1151, 854; ¹H NMR (DMSO- d_6) δ (ppm) 0.76 (t, 3H, J=7.3 Hz), 1.15 (sext, 2H, J=7.5 Hz), 1.36 (quint, 2H, J=7.5 Hz), 2.17 (t, 2H, J=7.3 Hz), 2.37 (s, 3H), 7.4 (d, 2H, J=8 Hz), 7.79 (d, 2H, J=8 Hz), 11.96 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm); ¹³C NMR (DMSO- d_6) δ (ppm) 13.94, 21.47, 21.83, 26.52, 35.46, 127.95, 129.90, 137.07, 144.51, 171.95. (Found: C, 56.78; H, 6.89; N, 5.33. C₁₂H₁₇NO₃S requires C, 56.45; H, 6.71; N, 5.49%.)

N-Isobutanoyl-4-methyl benzenesulfonamide (Table 2, entry 11). Mp 110–112°C; IR ν (cm⁻¹) 3259, 1731, 1329, 1171; ¹H NMR (DMSO-d₆) δ (ppm) 0.92 (d, 6H, J = 6.8 Hz), 2.38 (s, 3H), 2.41–2.46 (m, 1H,), 7.40 (d, 2H, J = 7.8 Hz), 7.78 (d, 2H, J = 7.8 Hz), 11.99 (s, 1H); ¹³C NMR (DMSO-d₆) δ (ppm) 16.92, 19.57, 32.75, 125.97, 128.03, 135.03, 142.61, 173.72. (Found: C, 54.60; H, 6.41; N, 5.62. C₁₁H₁₅NO₃S requires C, 54.75; H, 6.27; N, 5.80%.)

N-Benzoyl-4-methyl benzenesulfonamide (Table 2, entry 12). Mp 145 – 147°C; IR ν (cm⁻¹) 3292, 3065, 1699, 1605, 1500, 1421, 1342, 1176, 897, 712, ¹H NMR (DMSO- d_6) δ (ppm) 2.38 (s, 3H), 7.42–7.63(m, 5H), 7.84–7.91 (m, 4H), 12.48 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm) 21.53, 128.22, 128.84, 129.06, 129.99, 131.94, 133.71, 137.02, 144.72, 165.83. (Found: C, 59.98; H, 4.61; N, 5.35. C₁₄H₁₃NO₃S requires C, 61.07; H, 4.76; N, 5.09%.)

N-Ethanoyl methanesulfonamide (Table 2, entry 13). Mp 97–98°C; IR ν (cm⁻¹) 3138, 2907, 1706, 1468, 1341, 1243, 1158, 874, 755, 590, 509; ¹H NMR (DMSO- d_6) δ (ppm) 1.99 (s, 3H), 3.21 (s, 3H), 11.67 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm) 23.70, 41.38, 170.25. (Found: C, 26.22; H, 5.21; N, 10.01. C₃H₇NO₃S requires C, 26.27; H, 5.14; N, 10.21%.)

N-Propanoyl methanesulfonamide (Table 2, entry 14). Mp 68–70°C; IR ν (cm⁻¹) 3143, 1687, 1343, 1158; ¹H NMR (DMSO-*d*₆) δ (ppm) 0.99 (t, 3H, J=7.5 Hz), 2.27 (q, 2H, J=7.5 Hz), 3.21 (s, 3H), 11.63 (s, 1H); ¹³C NMR (DMSO-*d*₆) δ (ppm) 8.84, 29.16, 43.58, 173.72. (Found: C, 31.54; H, 6.21; N, 9.08. C₄H₉NO₃S requires C, 31.78; H, 6.00; N, 9.26%.)

N-Butanoyl methanesulfonamide (Table 2, entry 15). Mp 68–70°C; IR ν (cm⁻¹) 3203, 1700, 1330, 1151; ¹H NMR (DMSO- d_6) δ (ppm) 0.86 (t, 3H, J = 7.3 Hz), 1.53 (sext, 2H, J = 7.3 Hz), 2.23 (t, 2H, J = 7.3 Hz), 3.22 (s, 3H), 11.64 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm) 13.22, 17.50, 37.14, 40.88, 172.46. (Found: C, 36.61; H, 6.51; N, 8.78. C₅H₁₁NO₃S requires C, 36.35; H, 6.71; N, 8.84%.)

N-Pentanoyl methanesulfonamide (Table 2, entry 16). Mp 69–71°C; IR ν (cm⁻¹) 3269, 1720, 1330, 1158; ¹H NMR (300 MHz, DMSO-*d*₆) δ (ppm) 0.86 (t, 3H, J = 7.3 Hz), 1.26 (sext, 2H, J = 7.4 Hz), 1.48 (quin, 2H, J = 7.4 Hz), 2.25 (t, 2H, J = 7.3 Hz), 3.2 (s, 3H), 11.62 (s, 1H); ¹³C NMR (75 MHz, DMSO-*d*₆) δ (ppm) 14.06, 21.96, 26.62, 35.52, 43.55, 173.09. (Found: C, 39.98; H, 7.22; N, 7.97. C₆H₁₃NO₃S requires C, 40.21; H, 7.31; N, 7.81%.)

N-lsobutanoyl methanesulfonamide (Table 2, entry 17). Mp 97–99°C; IR ν (cm⁻¹) 3213, 1717, 1329, 1158; ¹H NMR (DMSO- d_6) δ (ppm) 1.03 (d, 6H,

J = 6.8 Hz), 2.49 (sept, 1H, J = 6.9 Hz), 3.22 (s, 3H), 11.64 (s, 1H); ¹³C NMR (DMSO- d_6) δ (ppm) 19.05, 34.73, 41.32, 176.85. (Found: C, 36.32; H, 6.83; N, 8.67. C₅H₁₁NO₃S requires C, 36.35; H, 6.71; N, 8.84%.)

N-Benzoyl methanesulfonamide (Table 2, entry 18). Mp 153–155°C; IR ν (cm⁻¹) 3244, 1692, 1335, 1177; ¹H NMR (DMSO-*d*₆) δ (ppm) 3.38 (s, 3H), 7.49–7.95 (m, 5H), 12.14 (s, 1H); ¹³C NMR (DMSO-*d*₆) δ (ppm) 41.78, 128.90, 129.00, 132.11, 133.70, 166.92. (Found: C, 48.58; H, 4.67; N, 6.86. C₈H₉NO₃S requires C, 48.23; H, 4.55; N, 7.03%.)

Biological Assays

Turbidity of all the bacterial cultures was adjusted to 0.5 McFarland standard by preparing a bacterial suspension of three to five well-isolated colonies of the same morphological type selected from an agar plate culture. The cultures were further diluted 1000-fold to get an inoculum size of 1.5×10^5 CFU/mL. The test compounds (50 mg) were dissolved in DMSO (0.5 mL), and the solution was diluted with distilled water (4.5 mL) to get a stock solution of 10000 mg/mL of each compound. Further progressive double dilution with Muller-Hinton broth was performed to obtain the required concentrations of 1024, 512, 256, 128, 64, 32, 16, 8, 4, and 2 µg/mL.^[31] To ensure that the solvent had no effect on bacterial growth, a control test was performed with test medium supplemented with DMSO at the same dilutions as used in the experiment. Our results indicated that all the compounds tested in this study demonactivity Gram-positive strated less against both and Gram-negative microorganisms than the reference drug.

Standard antibiotics were also diluted in the same manner. Each microwell in a series of 12 microwells was inoculated with 75 μ L of the serial dilutions, and then 75 μ L of the bacterial suspention was added. After overnight incubation in 37°C, growth was surveyed. Data of MICs of the test compounds and control drugs are shown in Table 4.

Inhibition zones of compounds were determined by the disc diffusion method.^[32] The antimicrobial screening was surveyed using the sterilized (autoclaved at 120°C for 30 min) Mueller–Hinton Agar and YEPD agar for the yeast. The culture suspensions were prepared and adjusted by comparing against 0.5 McFarland turbidity tubes. Mueller–Hinton and YEPD agar were poured into each sterile Petri dish. After the media were solidified, the bacteria were cultured using sterile swabs. All of the compounds were dissolved in DMSO of 10 mg/mL. Empty sterilized discs of 6.4 mm (Padtan Teb, Iran) were impregnated with 100μ L of compounds. Discs were placed on agar plates, and the cultures were incubated at 37°C for 24 h for bacteria and 48 h for *C. albicans*. Inhibition zones formed on the medium were evaluated in millimeters. The solvent control (DMSO) did not show any antimicrobial activity. Inhibition zones were compared with those of reference discs. Reference discs used for control are as follows: fluconazole (25 µg), gentamicin (10 µg), co-trimoxazole (1.25:23.75 µg), cefoxitin (30 µg), imipenem (10 µg), tetracycline (30 µg), and azithromycin (15 µg).

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H. ADIBI ET AL.

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