

# Accepted Manuscript

Total Synthesis of Calothrixins A and B via Oxidative Radical Reaction of Cyclohexenone with Aminophenanthridinedione

Su Xu, Thao Nguyen, Irene Pomilio, Maria C. Vitale, Sadanandan E. Velu



PII: S0040-4020(14)00868-0

DOI: [10.1016/j.tet.2014.06.021](https://doi.org/10.1016/j.tet.2014.06.021)

Reference: TET 25687

To appear in: *Tetrahedron*

Received Date: 19 April 2014

Revised Date: 3 June 2014

Accepted Date: 4 June 2014

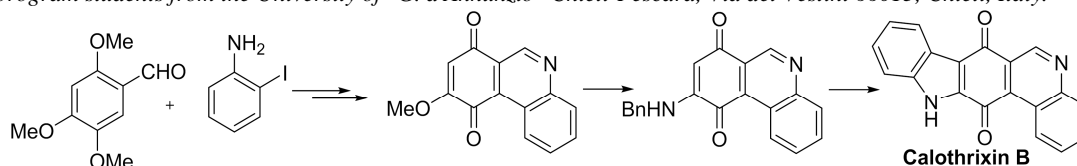
Please cite this article as: Xu S, Nguyen T, Pomilio I, Vitale MC, Velu SE, Total Synthesis of Calothrixins A and B via Oxidative Radical Reaction of Cyclohexenone with Aminophenanthridinedione, *Tetrahedron* (2014), doi: 10.1016/j.tet.2014.06.021.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

## Graphical Abstract

**Total Synthesis of Calothrixins A and B via Oxidative Radical Reaction of Cyclohexenone with Aminophenanthridinedione**

Leave this area blank for abstract info.

Su Xu<sup>a</sup>, Thao Nguyen<sup>a</sup>, Irene Pomilio<sup>c</sup>, Maria C. Vitale<sup>c</sup> and Sadanandan E. Velu<sup>\*a,b</sup><sup>a</sup>Department of Chemistry, University of Alabama at Birmingham, 901 14th Street South, Birmingham, AL 35294-1240, USA<sup>b</sup>Comprehensive Cancer Center, University of Alabama at Birmingham, 1720 2nd Avenue South, Birmingham, AL 35294-3300, USA<sup>c</sup>Exchange program students from the University of "G. d'Annunzio" Chieti-Pescara, Via dei Vestini 66013, Chieti, Italy.



Tetrahedron  
journal homepage: www.elsevier.com



# Total Synthesis of Calothrixins A and B via Oxidative Radical Reaction of Cyclohexenone with Aminophenanthridinedione

Su Xu<sup>a</sup>, Thao Nguyen<sup>a</sup>, Irene Pomilio<sup>c</sup>, Maria C. Vitale<sup>c</sup> and Sadanandan E. Velu<sup>\*a,b</sup>

<sup>a</sup>Department of Chemistry, University of Alabama at Birmingham, 901 14th Street South, Birmingham, AL 35294-1240, USA

<sup>b</sup>Comprehensive Cancer Center, University of Alabama at Birmingham, 1720 2nd Avenue South, Birmingham, AL 35294-3300, USA

<sup>c</sup>Exchange program students from the University of "G. d'Annunzio" Chieti-Pescara, Via dei Vestini 66013, Chieti, Italy.

\* Corresponding author. E-mail address: svelu@uab.edu (S.E. Velu), Phone: (205) 975 2478, Fax: (205) 934 2543.

## ARTICLE INFO

### Article history:

Received

Received in revised form

Accepted

Available online

### Keywords:

Calothrixin

Total Synthesis

Oxidative

Free Radical

Aminophenanthridinedione

## ABSTRACT

Bioactive indolo[3,2-*j*]phenanthridine alkaloids, calothrixin B and its N-oxide derivative calothrixin A have been synthesized via an oxidative free radical reaction. calothrixin B is generated from the commercially available 2,4,5-trimethoxybenzaldehyde in only 7 steps. The key step in this synthesis is the Mn(OAc)<sub>3</sub> mediated oxidative free radical reaction of 9-(benzylamino)phenanthridine -7,10-dione with cyclohexenone to form 12-benzyl-12*H*-indolo[3,2-*j*]phenanthridine-7,13-dione.

2009 Elsevier Ltd. All rights reserved.

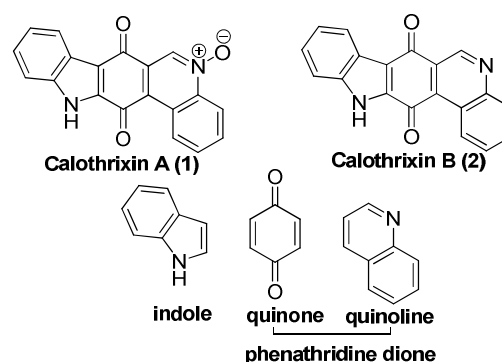
## Introduction

Natural products have traditionally played a major role in drug discovery by serving as the source for many of the earliest medicines.<sup>1</sup> In spite of the scientific advancements and the promise of alternative drug discovery strategies in the recent decades, there is still a shortage of drug leads progressing into clinical trials especially in the areas of oncology, immunosuppression, and metabolic diseases.<sup>2</sup> Natural products continue to play a major role in providing leads particularly in these areas as shown by a recent review published by Newman and Cragg which summarizes natural product derived drugs over past 30 years (1981-2010).<sup>3</sup> Even though many natural products exhibit potent biological activities, their systematic biological evaluation is precluded because they are often isolated in very minute quantities. Further, natural products are known to have unique chemical structures which provide challenging synthetic targets for organic chemists. For these reasons, development of new synthetic methods for natural products remains as a very important research area.

Calothrixin B and its N-oxide derivative, calothrixin A (Fig. 1) are two bioactive metabolites isolated from the cyanobacteria *Calothrix* in 1999.<sup>4</sup> They are also known as indolophenanthridines because they contain the unusual pentacyclic indolo[3,2-*j*]phenanthridine ring system. This ring

system consists of indole, quinone, and quinoline moieties and is unique amongst natural products (Fig. 1).

**Figure 1.** Calothrixin A and B and the three moieties present in them



Calothrixins possess a wide array of biological activities that are of interest to medicinal chemists. They display nanomolar antiproliferative activity against certain human cancer cell lines such as human cervical cancer cell, HeLa<sup>4-5</sup>, CEM leukemia cells,<sup>6</sup> and human Jurkat cancer cells.<sup>7</sup> Furthermore, calothrixins A and B act as a new class of human DNA topoisomerase I poisons. They stabilize the topoisomerase I-DNA binary complex

to prevent DNA religation and cause DNA damage which results in apoptosis.<sup>8</sup> In addition, calothrixin A binds to DNA quadruplex to inhibit DNA replication and DNA directed RNA synthesis resulting in impaired protein synthesis and cell death.<sup>9</sup> They also exhibit *in vitro* antiparasitic activity against chloroquine resistant strains of *Plasmodium falciparum*, which is the causative organism for malaria.<sup>4</sup> These biological activities make calothrixins the potential lead compounds for anticancer and antiparasitic drug discovery. Due to their unique structural features and potent bioactivity, calothrixins are notable synthetic targets.<sup>4-9</sup> The first total synthesis of calothrixins was reported by Kelly, using an *o*-lithiation strategy.<sup>10</sup> Several other syntheses of calothrixins have also been reported exploring different synthetic strategies such as metallations,<sup>11</sup> hetero Diels-Alder,<sup>12</sup> Friedel-Crafts acylation reaction,<sup>11a, 13</sup> etc. A few other syntheses of calothrixins have also been reported.<sup>14</sup>

As a part of our interest in deriving lead drug molecules from natural products,<sup>15</sup> we have been particularly interested in developing a shorter and a better yielding synthesis of calothrixins. Majority of the reported calothrixin syntheses are focused on the formation of the middle benzoquinone ring between the indole and quinoline units as the last step. Our synthetic approach relies on the construction of the indole ring on to a phenanthridine dione via a novel oxidative free radical reaction mediated by  $\text{Mn}(\text{OAc})_3$ . The method of  $\text{Mn}(\text{OAc})_3$  mediated oxidative reaction of 2-cyclohexenone with quinones was originally developed by Chuang *et al.*<sup>16</sup> None of the existing reports of calothrixin synthesis utilizes the late stage indole construction strategy and thus our synthetic approach is unique and different.

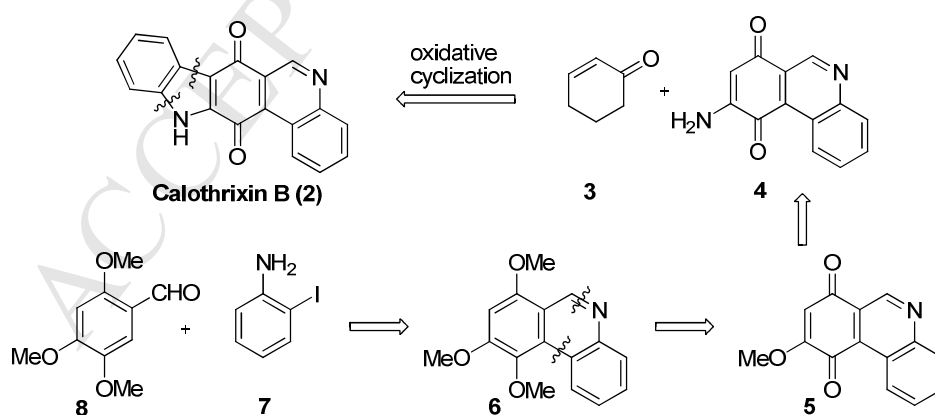
Research literature published in the recent three decades highlights the importance of oxidative free radical reactions mediated by transition metals. These reactions result in intermolecular / intramolecular formation of carbon-carbon bonds through transient electrophilic carbon radicals<sup>17</sup> and are most commonly promoted by transition metal compounds. Manganese triacetate ( $\text{Mn}(\text{OAc})_3$ ) and ceric (IV) ammonium

nitrate (CAN) are the most commonly used and efficient catalysts for the oxidative reaction between 1,3-diketones and aminoquinones. The mechanism for this type of oxidative free radical reactions has been reported.<sup>16, 17b, 17e</sup>  $\text{Mn}(\text{OAc})_3$  and CAN promoted oxidative free radical cyclizations have also been used extensively in the synthesis of naphthaquinones, which is an important skeleton of natural products.<sup>18</sup> The synthesis of several interesting compounds in our laboratory takes advantage of oxidative free radical reactions. Bispyrroloquinone, and bispyrroloiminoquinone ring systems were synthesized via CAN mediated oxidative free radical reaction of 1,3-dicarbonyl compounds with aminoquinones<sup>19</sup> while Zyzzyanones were synthesized using  $\text{Mn}(\text{OAc})_3$  mediated oxidative free radical reactions.<sup>20</sup> Apart from being a powerful tool to construct polycyclic ring systems, these reactions are generally high yielding and are relatively easy to perform. We report herein the synthesis of calothrixins taking advantage of an oxidative radical reaction between cyclohexenone and an aminophenanthridinedione derivative.

## Results and Discussion

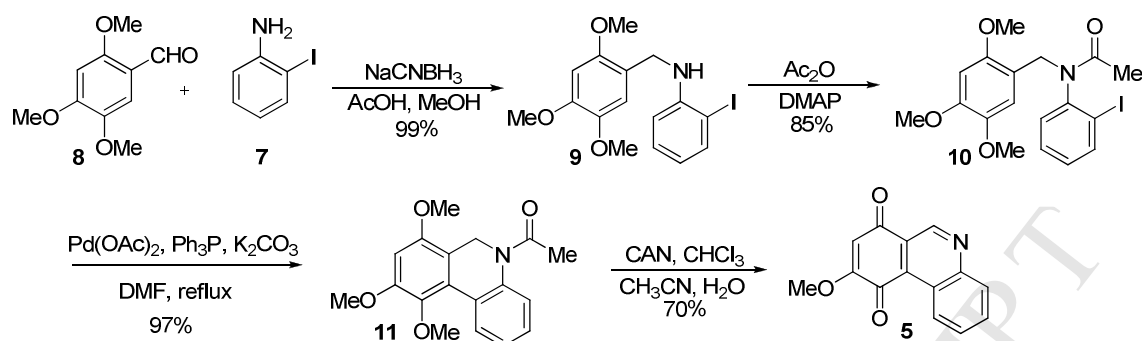
Our retrosynthetic analysis for calothrixin B utilizes an oxidative free radical reaction strategy as outlined in Scheme 1. We envisaged that the intermediate compound **6** for calothrixin synthesis could be prepared by the amination of 2,4,5-trimethoxybenzaldehyde (**8**) with *ortho*-iodoaniline (**7**) followed by a palladium catalyzed coupling. Oxidation of compound **6** with CAN can afford the corresponding methoxy quinone **5**, which could be aminated to form the aminoquinone **4**. Oxidative free radical reaction of the aminoquinone **4** with cyclohexenone (**3**) in the presence of  $\text{Mn}(\text{OAc})_3$  could give calothrixin B, which could be further oxidized with *m*-CPBA to form calothrixin A. Construction of the indole ring to the phenanthridine dione using cyclohexenone and  $\text{Mn}(\text{OAc})_3$  is the novel aspect of our calothrixin synthesis.

**Scheme 1.** Retrosynthetic analysis of calothrixin B



Our investigation started with the synthesis of the key intermediate compound **5** as outlined in Scheme 2. Reductive amination of commercially available 2,4,5-trimethoxybenzaldehyde (**8**) with *ortho*-iodoaniline (**7**) in the presence of  $\text{NaCNBH}_3$  in a mixture of MeOH and acetic acid gave the compound **9** in 99% yield. In order to avoid potential complications, the NH group present in compound **9** was

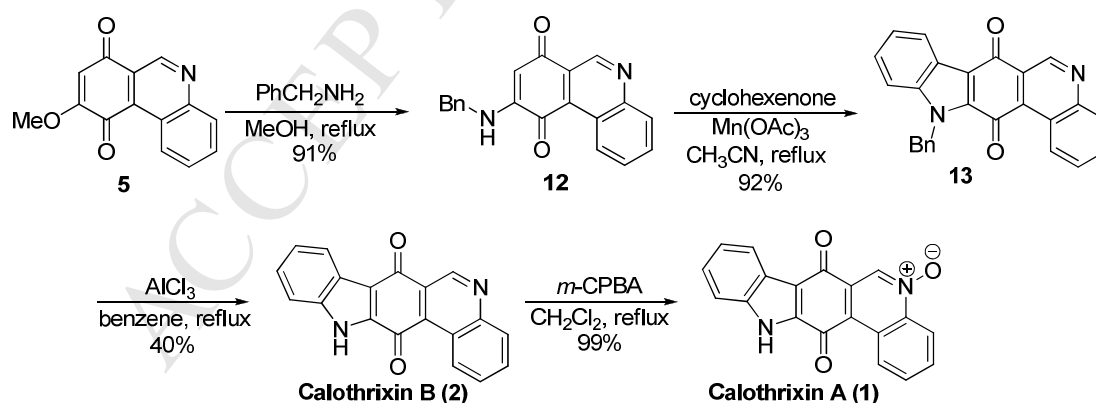
protected as an acetyl amide. This was achieved by the treatment of compound **9** with  $\text{Ac}_2\text{O}$  in the presence of catalytic amount of DMAP at room temperature for 15 hours to form compound **10** in 85% yield. Then the acetyl protected compound **10** was treated with  $\text{Pd}(\text{OAc})_2$ ,  $\text{PPh}_3$ , and  $\text{K}_2\text{CO}_3$  in DMF at reflux condition for 7 hours to generate the cyclized product **11** in 97% yield.

**Scheme 2.** Synthesis of intermediate quinone **5**

Complications occurred with the assignment of CH<sub>2</sub> proton signal within the <sup>1</sup>H-NMR spectra of compound **11** as seen by the broad multiplet ranging from 4.2 to 5.5 ppm. These protons were almost invisible in the spectra. This might be due to the fact that the molecule exists as a pair of amide rotamers that are in dynamic equilibrium. In order to confirm the structure of compound **11** unambiguously, we recorded a <sup>1</sup>H-NMR in DMSO-*d*<sub>6</sub> at 90 °C. The CH<sub>2</sub> proton signal was clearly visible as a sharp singlet in the high temperature NMR, which confirms the structure of compound **11**. This might be due to the faster equilibration of the amide rotamers at higher temperature. Compound **11** was then oxidized using CAN in a mixture of CH<sub>3</sub>CN, chloroform and water to afford the quinone **5** in 70% yield. Surprisingly, the deacetylation and aromatization of the N containing ring also occurred under the same reaction conditions.

Conversion of quinone **5** to calothrixins A and B is outlined in Scheme 3. In order to construct the indole ring on to the quinone **5** by oxidative free radical reaction we needed to

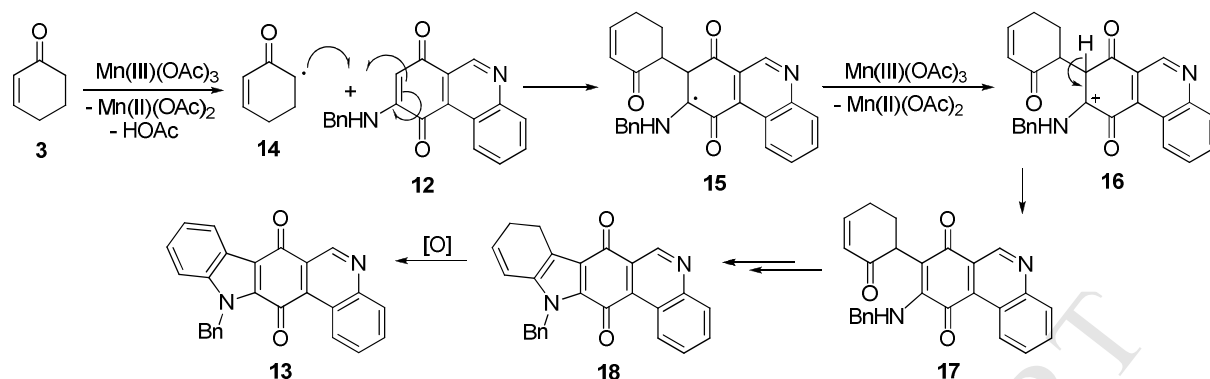
substitute the methoxy group in **5** with an amino functionality. We chose a benzyl amino group in this case as the benzyl group could serve as a protecting group and could be removed at the end of the synthesis. Reaction of the quinone **5** with benzyl amine in anhydrous MeOH under reflux conditions resulted in the formation of benzylamino phenanthridine dione **12** in 91% yield. Then the oxidative free radical reaction was performed by refluxing the mixture of benzylamino phenanthridine dione **12** and cyclohexenone in the presence of Mn(OAc)<sub>3</sub> in anhydrous CH<sub>3</sub>CN for three days to afford compound **13** in 92% yield. Finally, the debenzoylation of compound **13** was carried out by following a previously reported literature procedure using AlCl<sub>3</sub> and anhydrous benzene to furnish calothrixin B (**2**) in 40 % yield.<sup>12b</sup> In order to improve the yield of this step, we attempted other debenzoylation reaction conditions such as H<sub>2</sub> in the presence of Pd/C or Pd black/HCOONH<sub>4</sub>. But, these reactions resulted in lower yields than what we obtained with AlCl<sub>3</sub>. Calothrixin B (**2**) was oxidized to calothrixin A (**1**) in 99 % yield using *m*-CPBA in DCM following the literature procedure.<sup>10</sup>

**Scheme 3.** Conversion of quinone **5** to calothrixin A and B

Based on the previous literature reports on the mechanism of oxidative free radical reactions,<sup>16, 17e, 21</sup> a plausible mechanism for the conversion of compound **12** to **13** is outlined in Scheme 4. Initiation of the reaction occurs with the interaction of Mn(OAc)<sub>3</sub> with 2-cyclohexenone to generate the radical **14** by a one-electron oxidation. Intermolecular addition of radical **14** to the

quinone **12** generates another radical **15**. Oxidation of **15** by another molecule of Mn(OAc)<sub>3</sub> produces the corresponding cation **16**, which is deprotonated to form the intermediate **17**. Intramolecular condensation of **17** forms the compound **18** which up on aromatization yields the product **13**.

**Scheme 4.** Mechanism of oxidative radical reaction of 2-cyclohexenone with aminophenanthridinedione.



## Conclusions

In summary, we have described a new and efficient synthesis of calothrixin A and B with good overall yields. Synthesis of calothrixin B was achieved from 2,4,5-trimethoxybenzaldehyde in only 7 steps. This novel synthetic strategy involves the construction of an indole ring on to a phenanthridine-7,10-dione to form the calothrixin core. The key step in this synthesis is Mn(OAc)<sub>3</sub> mediated oxidative free radical reaction of 9-(benzylamino)phenanthridine -7,10-dione to form 12-benzyl-12H-indolo[3,2-*j*]phenanthridine-7,13-dione in excellent yield.

## Experimental Section

**General Methods for Synthesis:** Solvent evaporations were carried out *in vacuo* using a rotary evaporator. Thin layer chromatography (TLC) was performed on silica gel plates with fluorescent indicator (Dynamic Adsorbents, Inc., Aluminum backed TLC, 20 X 20 cm F-254, 200  $\mu\text{m}$ ). Spots were visualized by UV light (254 and 365 nm). Purification by column and flash chromatography was carried out using silica gel (32-63  $\mu\text{m}$ ) from Dynamic Adsorbent in the solvent systems indicated. The amount (weight) of silica gel for column chromatography was in the range of 50-100 times the amount (weight) of the crude compounds being separated. Melting points were determined on a Mel-Temp II melting point apparatus and are uncorrected. The NMR spectra were recorded on Bruker DPX 300 spectrometer. Chemical shifts are reported in ppm relative to TMS or  $\text{CDCl}_3$  as internal standard. The values of chemical shifts ( $\delta$ ) and coupling constants  $J$  were given in parts per million and in Hz, respectively. Mass spectra were recorded on a MicroMass Platform LCC instrument. HRMS were obtained on a Waters AutoSpec-Ultima<sup>TM</sup> NT mass spectrometer with an EI source. Anhydrous solvents used for reactions were purchased in Sure-Seal<sup>TM</sup> bottles from Aldrich chemical company. Other reagents were purchased from Aldrich, Lancaster or Fisher chemical companies and used as received.

***N*-(2,4,5-trimethoxybenzyl)-2-iodobenzenamine (9):** To a stirred solution of *o*-iodoaniline **7** (3.85 g, 17.5 mmol) in MeOH (50 mL), 2,4,5-trimethoxybenzaldehyde **8** (3.45 g, 17.5 mmol) and a solution of acetic acid (1.58 g, 26.3 mmol) in MeOH (10 mL) were added and the reaction mixture was stirred at rt under N<sub>2</sub> atm for 30 min. The reaction mixture was cooled to 0 °C using an ice bath and NaCNBH<sub>3</sub> (1.44 g, 22.8 mmol) was added in 4 portions over a period of 15 min and the solution was stirred for another 20 min at 0 °C. The reaction mixture was then stirred at rt for 1 h. TLC examination (30% EtOAc in hexanes) revealed that

the reaction was complete. Reaction mixture was then quenched with sat.  $\text{NaHCO}_3$  (100 mL) and the solvent was removed to obtain a brown residue. The residue was diluted with water (30 mL) and filtrated to afford the pure *N*-(2,4,5-trimethoxybenzyl)-2-iodobenzenamine **9** (6.95 g, 99%); Mp: 84-86 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  3.80 (s, 3H), 3.87 (s, 3H), 3.90 (s, 3H), 4.32 (d,  $J$  = 5.5 Hz, 2H), 4.63 (t,  $J$  = 5.5 Hz, 1H), 6.43 (t,  $J$  = 7.6 Hz, 1H), 6.56 (s, 1H), 6.63 (d,  $J$  = 8.4 Hz, 1H), 6.86 (s, 1H), 7.17 (t,  $J$  = 7.6 Hz, 1H), 7.66 (d,  $J$  = 8.1 Hz, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  43.6, 56.5 (2C), 57.0, 85.8, 97.9, 111.5, 113.5, 118.3, 119.0, 129.6, 139.2, 143.3, 147.7, 149.2, 151.9; MS (ES+)  $m/z$  400 ( $M + H$ ) and HRMS calcd for  $\text{C}_{16}\text{H}_{18}\text{INO}_3$  399.0331, found 399.0343.

***N*-(2,4,5-trimethoxybenzyl)-*N*-(2-iodophenyl)acetamide (**10**):** To a stirred solution of *N*-(2,4,5-trimethoxybenzyl)-2-iodobenzenamine **9** (6.42 g, 16.1 mmol) in acetic anhydride (35 mL), a cat amount of DMAP (196 mg, 1.6 mmol) was added and the reaction mixture was stirred at rt under N<sub>2</sub> atm for 15 h. TLC examination (50% EtOAc in hexanes) revealed that the reaction was complete. The solvent was removed and the residue was dissolved in EtOAc (400 mL), which was washed with water (3 × 100 mL) and brine (1 × 100 mL) and dried over Na<sub>2</sub>SO<sub>4</sub>. The drying agent was filtered off and the filtrate was concentrated on a rotary evaporator to afford the pure *N*-(2,4,5-trimethoxybenzyl)-*N*-(2-iodophenyl)acetamide **10** (6.04 g, 85%); Mp: 126-127 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 1.79 (s, 3H), 3.41 (s, 3H), 3.79 (s, 3H), 3.83 (s, 3H), 4.36 (d, J = 13.8 Hz, 1H), 5.28 (d, J = 13.8 Hz, 1H), 6.33 (s, 1H), 6.73 (d, J = 7.8 Hz, 1H), 6.91 (s, 1H), 6.98 (t, J = 7.2 Hz, 1H), 7.17 (t, J = 7.5 Hz, 1H), 7.87 (d, J = 7.8 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 23.1, 44.9, 56.2 (2C), 56.7, 97.2, 100.6, 115.3, 116.6, 129.0, 129.5, 130.6, 139.8, 143.1, 144.8, 149.3, 152.4, 170.1; MS (ES<sup>+</sup>) *m/z* 442 (M + H) and HRMS calcd for C<sub>18</sub>H<sub>20</sub>INO<sub>4</sub> 441.0437, found 441.0432.

**1-(7,9,10-trimethoxyphenanthridin-5(6*H*)-yl)ethanone (11):** To a stirred solution of *N*-(2,4,5-trimethoxybenzyl)-*N*-(2-iodophenyl)acetamide **10** (300 mg, 0.68 mmol) in anhyd DMF (5 mL), PPh<sub>3</sub> (52 mg, 0.20 mmol), anhyd K<sub>2</sub>CO<sub>3</sub> (136 mg, 0.99 mmol) and Pd(OAc)<sub>2</sub> (15 mg, 0.06 mmol) were added and the reaction mixture was refluxed under N<sub>2</sub> atm for 7 h. TLC examination (50% EtOAc in hexanes) revealed that the reaction was complete. The reaction mixture was cooled to rt and filtered through celite and washed with EtOAc (3 × 50 mL). The combined filtrates were washed with water (3 × 50 mL) and brine (1 × 50 mL) and dried over Na<sub>2</sub>SO<sub>4</sub>. The drying agent was filtered off and the filtrate was concentrated to afford the crude product, which was purified by column chromatography over Si gel (20 × 2 cm) using EtOAc / hexanes (1:3) as eluent to afford the pure 1-(7,9,10-trimethoxyphenanthridin-5(6*H*)-yl)ethanone



**11** (210 mg, 97%); Mp: 173-175 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  2.15-2.39 (m, 3H), 3.68 (s, 3H), 3.84 (s, 3H), 3.90 (s, 3H) 4.14-5.52 (brs, 2H), 6.50 (s, 1H), 7.12-7.34 (m, 3H), 8.38-8.50 (m, 1H);  $^1\text{H}$  NMR (90 °C,  $\text{DMSO}-d_6$ )  $\delta$  2.15 (s, 3H), 3.71 (s, 3H), 3.91 (s, 3H), 3.94 (s, 3H), 4.72 (s, 2H), 6.83 (s, 1H), 7.31-7.43 (m, 2H), 7.53 (d,  $J = 7.7$  Hz, 1H), 8.35 (d,  $J = 7.7$  Hz, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  22.2, 39.1, 56.1, 56.3, 60.5, 97.0, 117.9, 124.5, 125.7, 126.0, 127.7, 128.3, 128.7, 138.7, 140.8, 151.4, 152.9, 169.0; MS (ES<sup>+</sup>)  $m/z$  314 (M + H) and HRMS calcd for  $\text{C}_{18}\text{H}_{19}\text{NO}_4$  313.1314, found 313.1315.

**9-Methoxyphenanthridine-7,10-dione (5):** To a stirred solution of 1-(7,9,10-trimethoxyphenanthridin-5(6H)-yl)ethanone **11** (400 mg, 1.3 mmol) in a mixture of  $\text{CH}_3\text{CN}$  (100 mL) and  $\text{CHCl}_3$  (5 mL), a solution of CAN (2.79 g, 5.1 mmol) in water (100 mL) was added. The reaction mixture was stirred at rt for 7 h. TLC examination (50% EtOAc in hexanes) revealed that the reaction was complete.  $\text{CH}_3\text{CN}$  was removed under reduced pressure and the residue obtained was extracted with  $\text{CHCl}_3$  (4  $\times$  100 mL). The combined organic layers were washed with water (3  $\times$  100 mL), brine (2  $\times$  100 mL) and dried over  $\text{Na}_2\text{SO}_4$ . The drying agent was filtered off and the filtrate was concentrated to obtain the crude product, which was purified by column chromatography over Si gel (20  $\times$  2 cm) using  $\text{CHCl}_3$  as eluent to afford the pure 9-methoxyphenanthridine-7,10-dione **5** (214 mg, 70%); Mp: 243-245 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  3.97 (s, 3H), 6.20 (s, 1H), 7.81 (t,  $J = 7.8$  Hz, 1H), 7.89 (t,  $J = 7.2$  Hz, 1H), 8.21 (d,  $J = 8.7$  Hz, 1H), 9.41 (d,  $J = 8.4$  Hz, 1H), 9.66 (s, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  57.0, 107.9, 122.3, 122.9, 127.5, 130.7, 131.0, 131.1, 132.1, 147.8, 152.1, 160.7, 183.0, 185.1; MS (ES<sup>+</sup>)  $m/z$  240 (M + H) and HRMS calcd for  $\text{C}_{14}\text{H}_9\text{NO}_3$  239.0582, found 239.0592.

**9-(Benzylamino)phenanthridine-7,10-dione (12):** To a stirred solution of 9-methoxyphenanthridine-7,10-dione **5** (200 mg, 0.83 mmol) in MeOH (50 mL), a solution of benzyl amine (358 mg, 3.3 mmol) in MeOH (25 mL) was added in 2 portions over 5 h and the reaction mixture was refluxed under  $\text{N}_2$  atm for another 6 h. TLC analysis (50% EtOAc in hexanes) revealed that the reaction was complete. The solvent was removed by a rotary evaporator to afford the crude product, which was purified by column chromatography over Si gel (20  $\times$  2 cm) using EtOAc / hexanes (1:1) as eluent to afford the pure 9-(benzylamino)phenanthridine-7,10-dione **12** (239 mg, 91%); Mp: 183-185 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  4.43(d,  $J = 6.0$  Hz, 2H), 5.82 (s, 1H), 6.30 (brs, 1H), 7.28-7.48 (m, 5H), 7.73-7.90 (m, 2H), 8.20 (d,  $J = 8.1$  Hz, 1H), 9.37 (d,  $J = 8.4$  Hz, 1H), 9.71 (s, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  47.3, 100.2, 122.3, 124.1, 127.0, 127.9, 128.6, 129.4, 130.2, 130.8 (2C), 131.5, 135.9, 148.2, 148.52, 151.6, 183.1, 185.0; MS (ES<sup>+</sup>)  $m/z$  315 (M + H) and HRMS calcd for  $\text{C}_{20}\text{H}_{14}\text{N}_2\text{O}_2$  314.1055, found 314.1067.

**12-benzyl-12H-indolo[3,2-*j*]phenanthridine-7,13-dione (13):** To a stirred solution of 9-(benzylamino)phenanthridine-7,10-dione **12** (12 mg, 0.04 mmol) in  $\text{CH}_3\text{CN}$  (16 mL),  $\text{Mn}(\text{OAc})_3 \cdot 2\text{H}_2\text{O}$  (142 mg, 0.53 mmol) and a solution of 2-cyclohexen-1-one **3** (15 mg, 0.15 mmol) in  $\text{CH}_3\text{CN}$  (4 mL) were added. The reaction mixture was refluxed under  $\text{N}_2$  atm for 3 days. TLC examination (30% EtOAc in hexanes) revealed that the reaction was complete. The reaction mixture was then allowed to attain rt and the solvent was removed under reduced pressure. The residue obtained was then dissolved in EtOAc (50 mL) and washed with saturated  $\text{NaHSO}_3$  (3  $\times$  30 mL), water (3  $\times$  30 mL) and brine (2  $\times$  25 mL) and dried over  $\text{Na}_2\text{SO}_4$ . The drying agent was filtered off and the filtrate was concentrated to afford the crude product, which was purified by column chromatography over Si gel using EtOAc / hexanes (1:19) as

eluent to afford the pure 12-benzyl-12H-indolo[3,2-*j*]phenanthridine-7,13-dione **13** (14 mg, 92 %); Mp: 255-257 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  6.02 (s, 2H), 7.17-7.37 (m, 5H), 7.38-7.51 (m, 3H), 7.74 (t,  $J = 8.4$  Hz, 1H), 7.83(t,  $J = 6.9$  Hz, 1H), 8.18 (d,  $J = 7.5$  Hz, 1H), 8.43-8.50 (m, 1H), 9.53 (d,  $J = 8.4$  Hz, 1H), 9.80 (s, 1H);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  48.8, 111.8, 118.0, 123.4, 123.6, 124.2, 124.8, 125.4, 126.8, 127.9, 128.1, 128.2, 129.1, 130.3, 130.6, 131.6, 133.6, 135.4, 136.5, 140.4, 148.2, 152.5, 181.3, 182.3; MS (ES<sup>+</sup>)  $m/z$  389 (M + H) and HRMS calcd for  $\text{C}_{26}\text{H}_{16}\text{N}_2\text{O}_2$  388.1212, found 388.1218.

**Calothrixin B (1):** To a solution of 12-benzyl-12H-indolo[3,2-*j*]phenanthridine-7,13-dione **13** (10 mg, 0.03 mmol) in anhydrous benzene (10 mL),  $\text{AlCl}_3$  (17 mg, 0.13 mmol) was added and the reaction mixture was refluxed for 5 h. TLC examination (20 % EtOAc in hexanes) revealed that the reaction was complete. The reaction mixture was then allowed to attain rt, quenched with water (10 mL), and extracted by DCM (2  $\times$  30 mL). The combined organic layers were dried over  $\text{Na}_2\text{SO}_4$ . The drying agent was filtered off and the filtrate was concentrated on a rotary evaporator to afford the crude product, which was purified by column chromatography over Si gel (20  $\times$  2 cm) using EtOAc/hexanes (1:9) as eluent to afford the pure calothrixin B (3 mg, 40 %); Mp: 298-300 °C;  $^1\text{H}$  NMR ( $\text{DMSO}-d_6$ )  $\delta$  7.42 (t,  $J = 8.1$  Hz, 1H), 7.50 (t,  $J = 8.4$  Hz, 1H), 7.63 (d,  $J = 8.1$  Hz, 1H), 7.89 (t,  $J = 6.9$  Hz, 1H), 7.97 (t,  $J = 6.9$  Hz, 1H), 8.19 (m, 2H), 9.59 (d,  $J = 8.7$  Hz, 1H), 9.63(s, 1H), 13.18 (bs, 1H);  $^{13}\text{C}$  NMR ( $\text{DMSO}-d_6$ )  $\delta$  113.9, 115.5, 122.3, 122.6, 123.3, 124.3, 124.9, 127.2, 129.8, 130.2, 131.6 (2C), 132.6, 138.0, 138.4, 147.5, 151.2, 180.4, 180.8; MS (ES<sup>+</sup>)  $m/z$  299 (M + H) and HRMS calcd for  $\text{C}_{19}\text{H}_{10}\text{N}_2\text{O}_2$  298.0742, found 298.0745.

**Calothrixin A (2):** To a suspension of calothrixin B (5 mg, 0.02 mmol) in DCM (10 mL), under nitrogen, *m*-CPBA (77 %, 13 mg, 0.08 mmol) in DCM (10 mL) was added and the reaction mixture was refluxed overnight. TLC (50 % EtOAc in hexanes) examination revealed that the reaction was complete. The reaction mixture was diluted with DCM (50 mL) to afford a red orange solution, which was washed with saturated  $\text{K}_2\text{CO}_3$  (3  $\times$  15 mL), water (3  $\times$  15 mL), brine (15 mL) and dried over  $\text{Na}_2\text{SO}_4$ . The drying agent was filtered off and the filtrate was concentrated on a rotary evaporator to afford the crude product, which was purified by column chromatography over Si gel (20  $\times$  2 cm) using EtOAc / hexanes (3:17) as eluent to afford the pure calothrixin A (5 mg, 99 %); Mp: 283-285 °C;  $^1\text{H}$  NMR ( $\text{DMSO}-d_6$ )  $\delta$  7.38 (t,  $J = 7.2$  Hz, 1H), 7.46 (t,  $J = 7.8$  Hz, 1H), 7.61 (d,  $J = 7.8$  Hz, 1H), 7.99-7.97 (m, 2H), 8.13 (d,  $J = 7.5$  Hz, 1H), 8.61 (d,  $J = 9.9$  Hz, 1H), 8.89 (s, 1H), 9.68 (d,  $J = 9.3$  Hz, 1H), 13.22 (s, 1H);  $^{13}\text{C}$  NMR ( $\text{DMSO}-d_6$ )  $\delta$  114.0, 115.1, 119.1, 121.9, 122.0, 123.4, 124.5, 126.8, 127.0, 128.1, 129.9, 131.8, 131.9, 132.0, 138.1, 138.7, 143.1, 177.8, 178.3; MS (ES<sup>+</sup>)  $m/z$  315 (M + H) and HRMS calcd for  $\text{C}_{19}\text{H}_{10}\text{N}_2\text{O}_3$  314.0691, found 314.0701.

## Acknowledgements

Authors also wish to acknowledge the financial support by the Collaborative Programmatic Development Grant from the University of Alabama at Birmingham (UAB) Comprehensive Cancer Center. Beginning Grant-in-Aid (AHA0865323E) from American Heart Association Greater Southeast Affiliate is also acknowledged. Authors would like to thank Ms. Bhavitavya Nijampatnam for the proof reading of this manuscript.

## References and notes

1. (a) Brown, D. G.; Lister, T.; May-Dracka, T. L., *Bioorg. Med. Chem. Lett.* **2014**, 24, 413; (b) Camp, D.; Davis, R. A.; Evans-Illidge, E. A.; Quinn, R. J., *Future Med. Chem.* **2012**, 4, 1067; (c) Cragg, G. M.; Newman, D. J., *Expert Opin. Invest. Drugs* **2000**, 9, 2783; (d) Newman, D. J.; Cragg, G. M.; Snader, K. M., *Nat. Prod. Rep.* **2000**, 17, 215; (e) Ojima, I., *J. Med. Chem.* **2008**, 51, 2587.
2. Butler, M. S., *Nat. Prod. Rep.* **2005**, 22, 162.
3. Newman, D. J.; Cragg, G. M., *J. Nat. Prod.* **2012**, 75, 311.
4. Rickards, R. W.; Rothschild, J. M.; Willis, A. C.; de Chazal, N. M.; Kirk, J.; Kirk, K.; Saliba, K. J.; Smith, G. D., *Tetrahedron* **1999**, 55, 13513.
5. Bernardo, P. H.; Chai, C. L.; Le Guen, M.; Smith, G. D.; Waring, P., *Bioorg. Med. Chem. Lett.* **2007**, 17, 82.
6. (a) Khan, Q. A.; Lu, J.; Hecht, S. M., *J. Nat. Prod.* **2009**, 72, 438; (b) Costa, M.; Costa-Rodrigues, J.; Fernandes, M. H.; Barros, P.; Vasconcelos, V.; Martins, R., *Marine drugs* **2012**, 10, 2181.
7. Chen, X. X.; Smith, G. D.; Waring, P., *J. Appl. Phycol.* **2003**, 15, 269.
8. (a) Doan, N. T.; Rickards, R. W.; Rothschild, J. M.; Smith, G. D., *J. Appl. Phycol.* **2000**, 12, 409; (b) Doan, N. T.; Stewart, P. R.; Smith, G. D., *FEMS Microbiol. Lett.* **2001**, 196, 135.
9. Owen, E. A.; Keniry, M. A., *Aus. J. Chem.* **2009**, 62, 1544.
10. Kelly, T. R.; Zhao, Y.; Cavero, M.; Torneiro, M., *Org. Lett.* **2000**, 2, 3735.
11. (a) Bernardo, P. H.; Chai, C. L. L.; Elix, J. A., *Tetrahedron Lett.* **2002**, 43, 2939; (b) Abe, T.; Ikeda, T.; Yanada, R.; Ishikura, M., *Org. Lett.* **2011**, 13, 3356; (c) Bhosale, S. M.; Gawade, R. L.; Puranik, V. G.; Kusurkar, R. S., *Tetrahedron Lett.* **2012**, 53, 2894.
12. (a) Sissouma, D.; Collet, S. C.; Guingant, A. Y., *Synlett* **2004**, 2612; (b) Sissouma, D.; Maingot, L.; Collet, S.; Guingant, A., *J. Org. Chem.* **2006**, 71, 8384.
13. (a) Bernardo, P. H.; Chai, C. L., *J. Org. Chem.* **2003**, 68, 8906; (b) Ramkumar, N.; Nagarajan, R., *J. Org. Chem.* **2014**, 79, 736.
14. (a) Bernardo, P. H.; Chai, C. L.; Heath, G. A.; Mahon, P. J.; Smith, G. D.; Waring, P.; Wilkes, B. A., *J. Med. Chem.* **2004**, 47, 4958; (b) Bennasar, M. L.; Roca, T.; Ferrando, F., *Org. Lett.* **2006**, 8, 561; (c) Ramkumar, N.; Nagarajan, R., *J. Org. Chem.* **2013**, 78, 2802.
15. (a) Zhang, X.; Xu, H.; Zhang, X.; Voruganti, S.; Murugesan, S.; Nadkarni, D. H.; Velu, S. E.; Wang, M. H.; Wang, W.; Zhang, R., *Marine drugs* **2012**, 10, 1138; (b) Nadkarni, D. H.; Wang, F.; Wang, W.; Rayburn, E. R.; Ezell, S. J.; Murugesan, S.; Velu, S. E.; Zhang, R., *Med. Chem. (Shariqah (United Arab Emirates))* **2009**, 5, 227; (c) Wang, W.; Rayburn, E. R.; Velu, S. E.; Nadkarni, D. H.; Murugesan, S.; Zhang, R., *Clin. Cancer Res.* **2009**, 15, 3511; (d) Chen, T.; Xu, Y.; Guo, H.; Liu, Y.; Hu, P.; Yang, X.; Li, X.; Ge, S.; Velu, S. E.; Nadkarni, D. H.; Wang, W.; Zhang, R.; Wang, H., *PLoS one* **2011**, 6, e20729.
16. Tseng, C.-M.; Wu, Y.-L.; Chuang, C.-P., *Tetrahedron* **2004**, 60, 12249.
17. (a) Iqbal, J.; Bhatia, B.; Nayyar, N. K., *Chem. Rev.* **1994**, 94, 519; (b) Tseng, C.-C.; Wu, Y.-L.; Chuang, C.-P., *Tetrahedron* **2002**, 58, 7625; (c) Chuang, C.-P.; Wu, Y.-L., *Tetrahedron Lett.* **2001**, 42, 1717; (d) Chuang, C.-P.; Tsai, A. I., *Tetrahedron* **2007**, 63, 11911; (e) Wu, Y.-L.; Chuang, C.-P.; Lin, P.-Y., *Tetrahedron* **2001**, 57, 5543.
18. Patai, S.; Rappoport, Z., *The Chemistry of Functional Groups: The Chemistry of The Quinoid Compounds*. Wiley: New York, 1988.
19. Murugesan, S.; Nadkarni, D. H.; Velu, S. E., *Tetrahedron Lett.* **2009**, 50, 3074.
20. Nadkarni, D. H.; Murugesan, S.; Velu, S. E., *Tetrahedron* **2013**, 69, 4105.
21. Kurz, M. E.; Baru, V.; Nguyen, P. N., *J. Org. Chem.* **1984**, 49, 1603.

### Supplementary material

Copies of  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR spectra of all compounds are available as Supplementary Material.



## Supporting Information

### Total Synthesis of Calothrixins A and B via Oxidative Radical Reaction of Cyclohexenone with Aminophenanthridinedione

Su Xu<sup>a</sup>, Thao Nguyen<sup>a</sup>, Irene Pomilio<sup>c</sup>, Maria C. Vitale<sup>c</sup> and Sadanandan E. Velu<sup>\*a,b</sup>

<sup>a</sup>*Department of Chemistry, University of Alabama at Birmingham, 901 14th Street South, Birmingham, AL 35294-1240, USA*

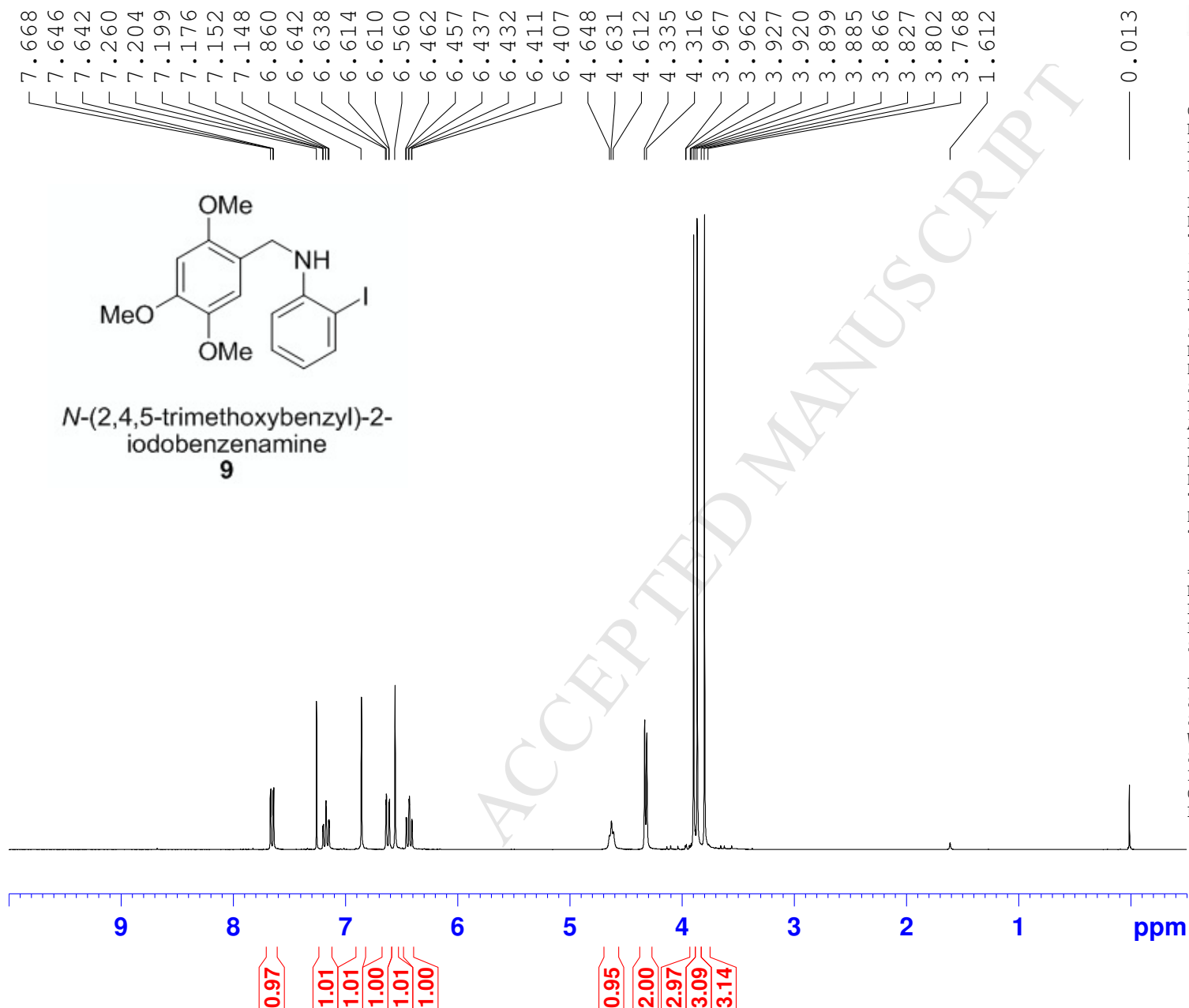
<sup>b</sup>*Comprehensive Cancer Center, University of Alabama at Birmingham, 1720 2nd Avenue South, Birmingham, AL 35294-3300, USA*

<sup>c</sup>*Exchange program students from the University of "G. d'Annunzio" Chieti-Pescara, Via dei Vestini 66013, Chieti, Italy.*

#### Table of Contents

<sup>1</sup>H-NMR, <sup>13</sup>C-NMR for all compounds S1-S17

SX-I-248-Pure 1HNMR CDC13

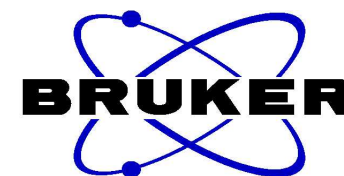


Current Data Parameters  
 NAME SX-I-248-Pure  
 EXPNO 1  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130107  
 Time 12.19  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 16  
 DS 2  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 5.2953587 sec  
 RG 143.7  
 DW 80.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 15.00 usec  
 PL1 -2.50 dB  
 SFO1 299.8818519 MHz

F2 - Processing parameters  
 SI 32768  
 SF 299.8800035 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.00

SX-I-248-Pure <sup>13</sup>CNMR CDCl<sub>3</sub>

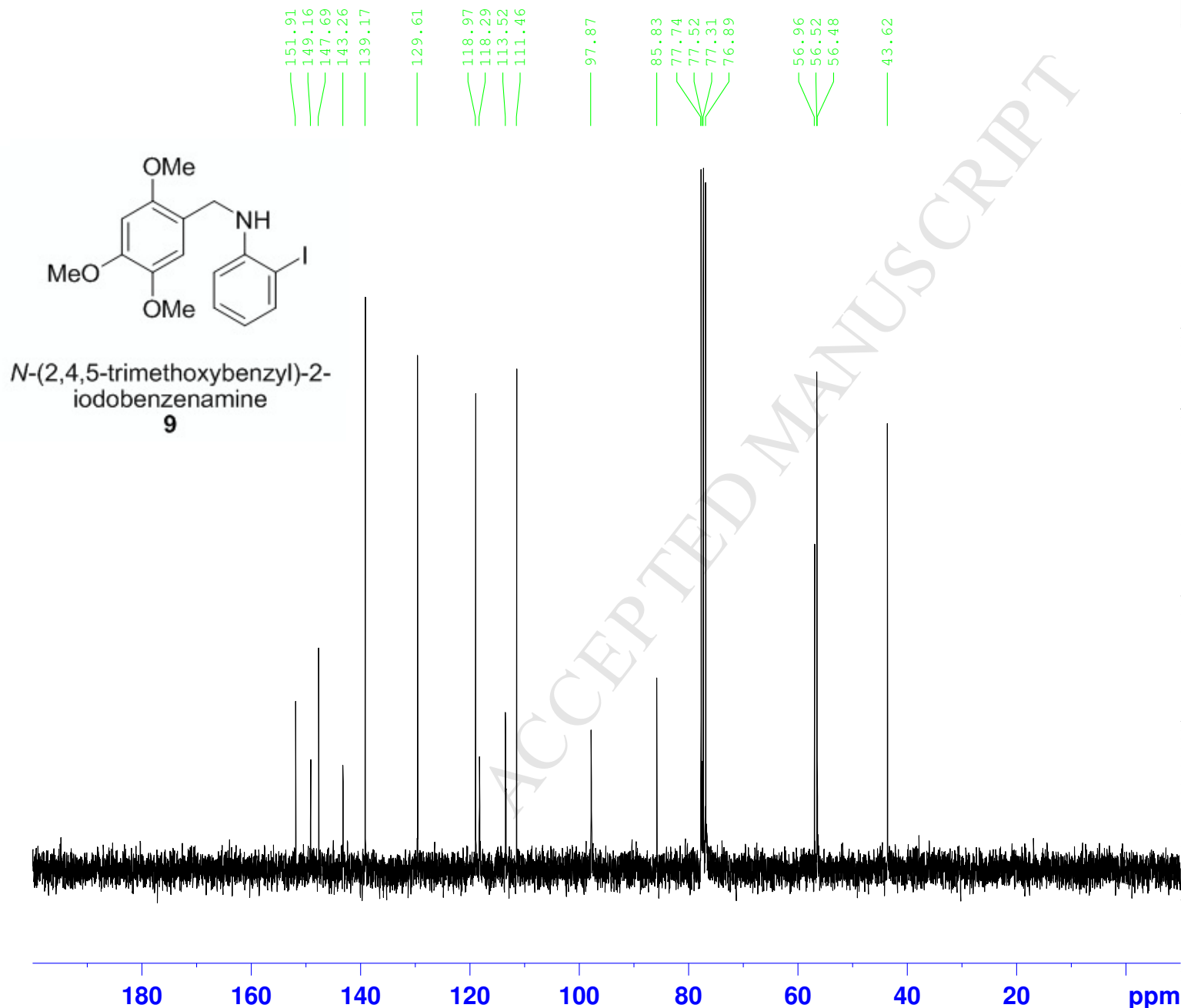
Current Data Parameters  
 NAME SX-I-248-Pure  
 EXPNO 2  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130107  
 Time 12.25  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 68  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

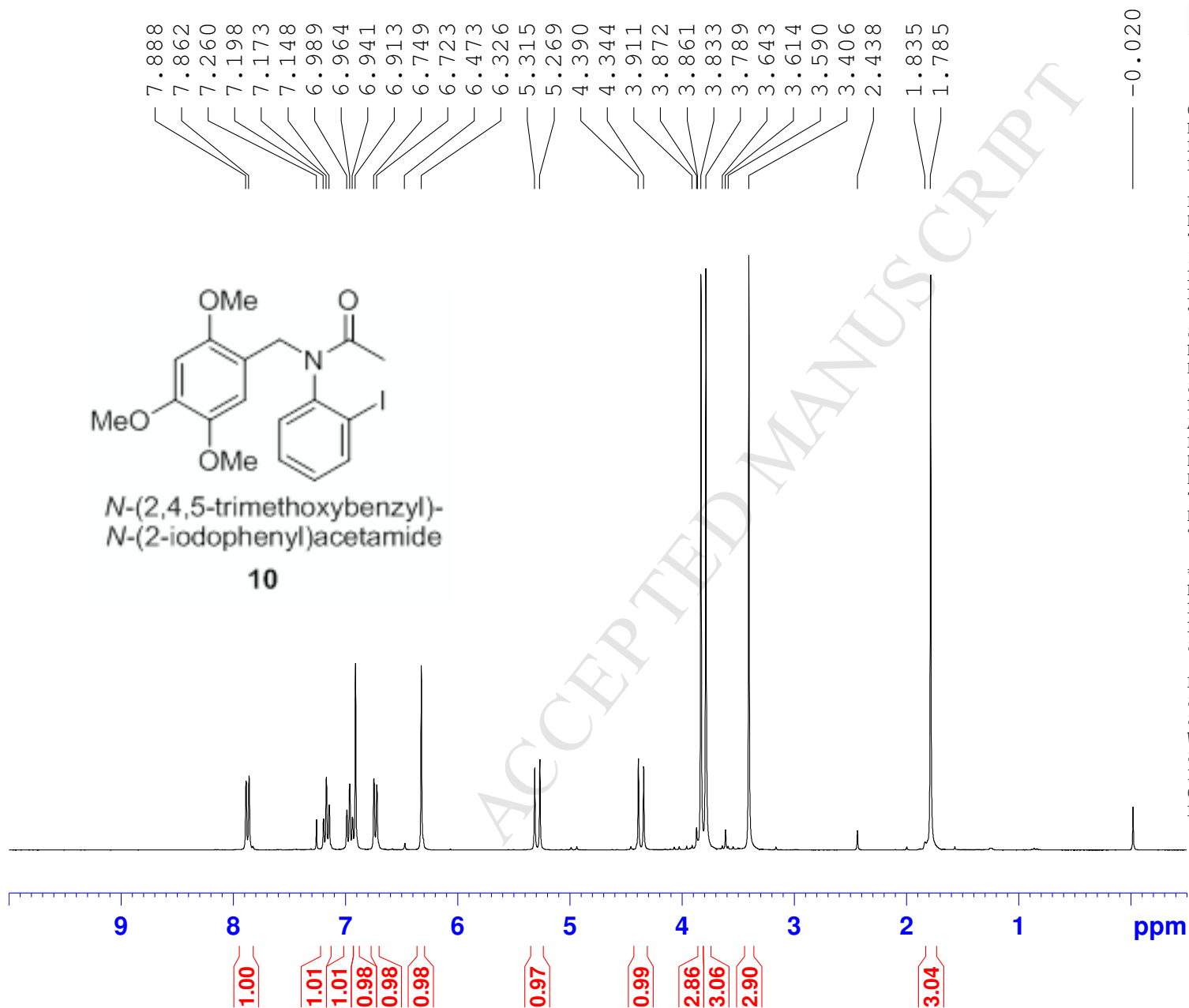
===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4048693 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.00



SX-I-250-pure 1HNMR CDC13

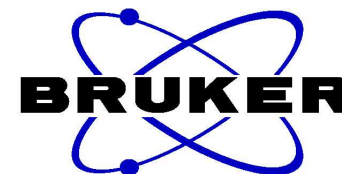


Current Data Parameters  
NAME SX-I-250-pure  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20130131  
Time 10.59  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 16  
DS 2  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953587 sec  
RG 228.1  
DW 80.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 15.00 usec  
PL1 -2.50 dB  
SFO1 299.8818519 MHz

F2 - Processing parameters  
SI 32768  
SF 299.8800034 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

SX-I-250-pure <sup>13</sup>CNMR CDC13

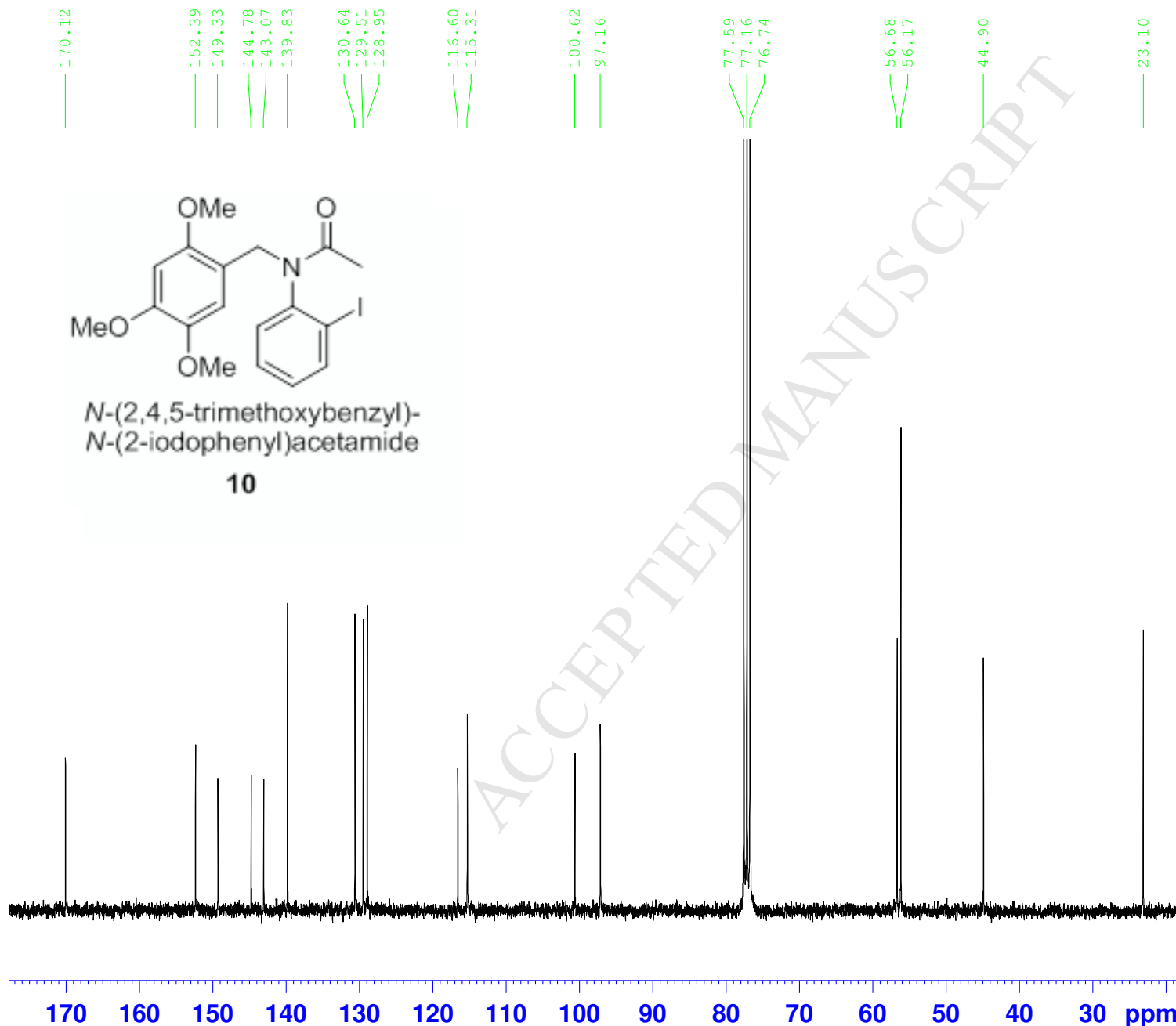
Current Data Parameters  
 NAME SX-I-250-pure  
 EXPNO 3  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130131  
 Time 11.03  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 1207  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

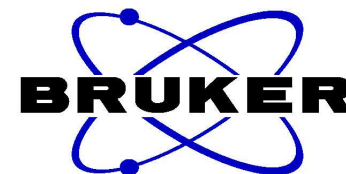
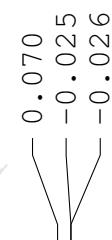
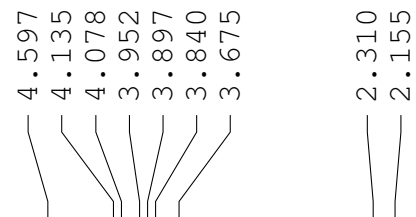
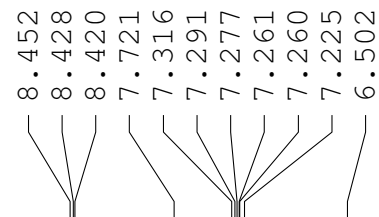
===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4048767 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40





SX-I-254B-WhiteSolid 1HNMR CDCl3

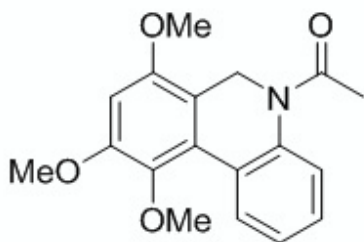


Current Data Parameters  
NAME SX-I-254B-WhiteSolid  
EXPNO 1  
PROCNO 1

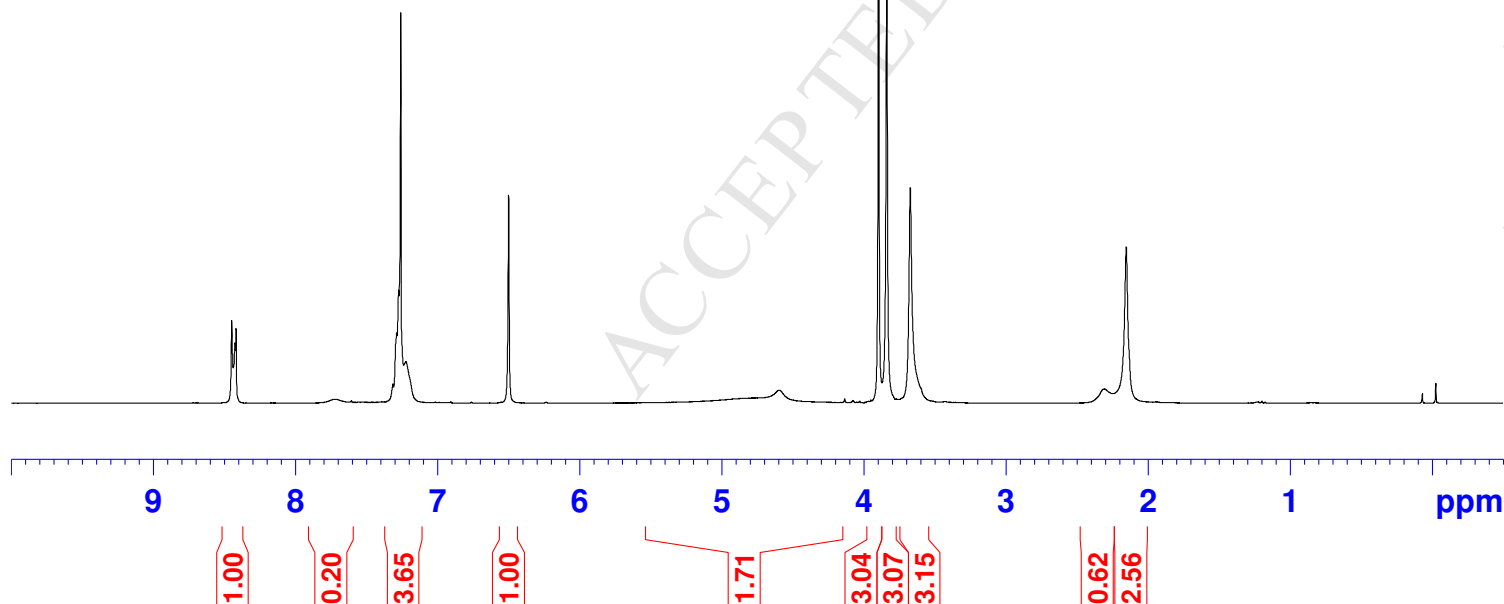
F2 - Acquisition Parameters  
Date\_ 20130115  
Time 10.47  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953587 sec  
RG 80.6  
DW 80.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

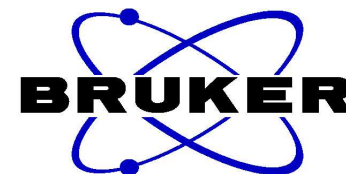
===== CHANNEL f1 =====  
NUC1 1H  
P1 15.00 usec  
PL1 -2.50 dB  
SFO1 299.8818519 MHz

F2 - Processing parameters  
SI 32768  
SF 299.8800029 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



1-(7,9,10-trimethoxyphenanthridin-  
5(6H)-yl)ethanone  
**11**



SX-I-254B-WhiteSolid <sup>13</sup>CNMR CDCl<sub>3</sub>

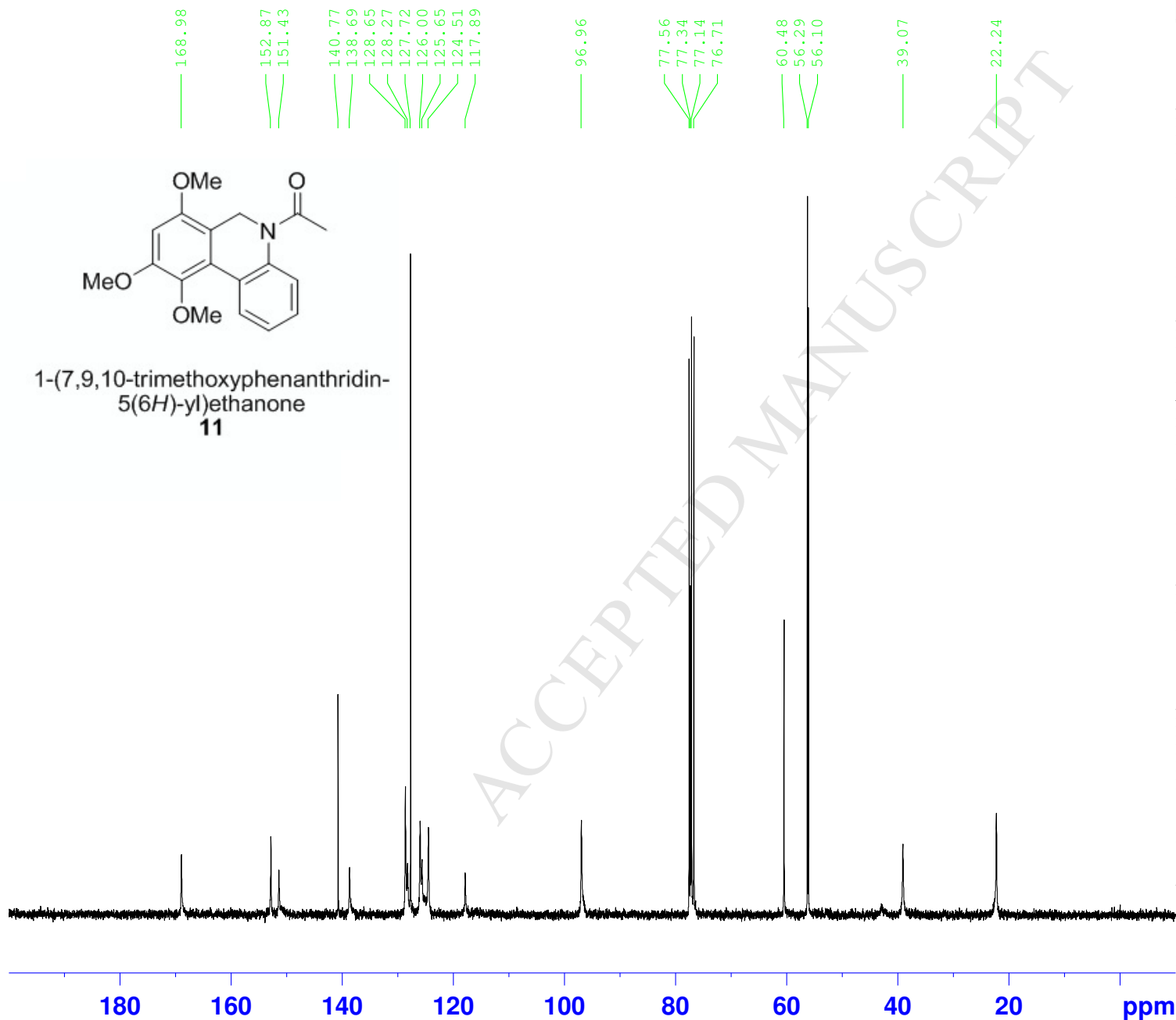
Current Data Parameters  
 NAME SX-I-254B-WhiteSolid  
 EXPNO 2  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130115  
 Time 10.50  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 782  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

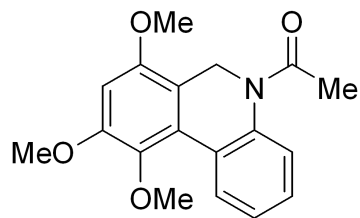
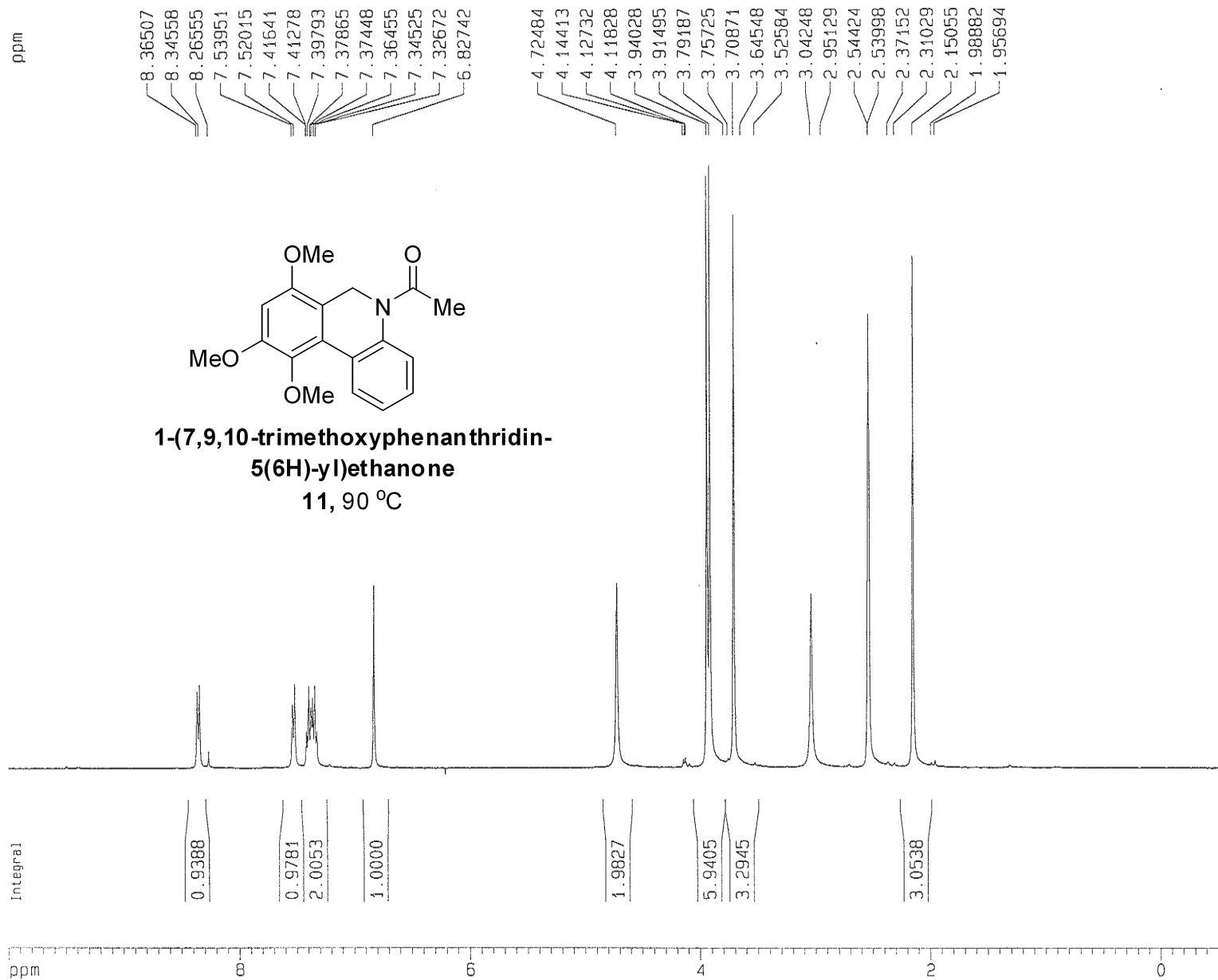
===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4048875 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



SX-I-254-90oC 1HNMR DMSO-d6



**1-(7,9,10-trimethoxyphenanthridin-5(6H)-yl)ethanone**  
11, 90 °C

Current Data Parameters

NAME SX-I-254-temp4  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters

Date\_ 20140328  
Time 10.27  
INSTRUM drx400  
PROBHD 5 mm Multinu  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 16  
DS 2  
SWH 8278.146 Hz  
FIDRES 0.126314 Hz  
AQ 3.9584243 sec  
RG 456.1  
DW 60.400 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec

===== CHANNEL f1 =====

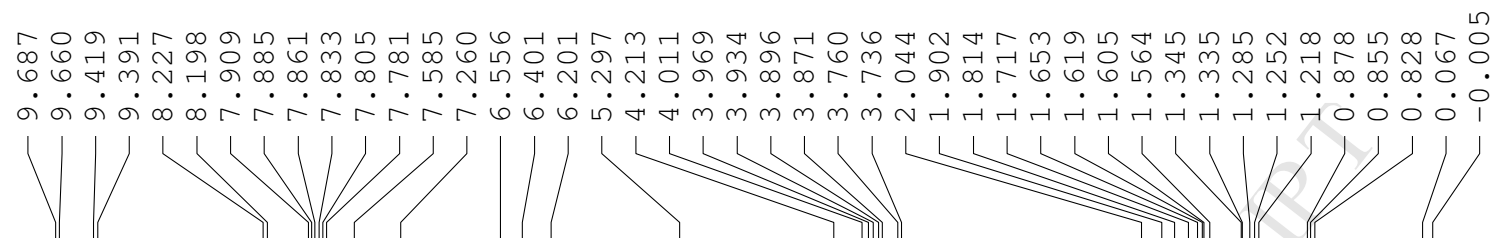
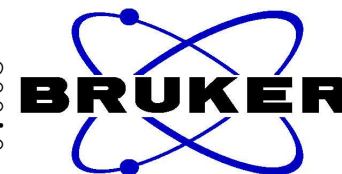
NUC1 1H  
P1 6.50 usec  
PL1 0.00 dB  
SFO1 400.1324710 MHz

F2 - Processing parameters

SI 131072  
SF 400.1299859 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

1D NMR plot parameters

CX 20.00 cm  
F1P 10.000 ppm  
F1 4001.30 Hz  
F2P -0.500 ppm  
F2 -200.06 Hz  
PPMCM 0.52500 ppm/cm  
HZCM 210.06824 Hz/cm

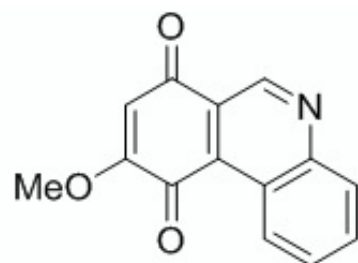
SX-I-246-Column-F1 1H NMR CDCl<sub>3</sub>

Current Data Parameters  
NAME SX-I-246-Column-F1  
EXPNO 1  
PROCNO 1

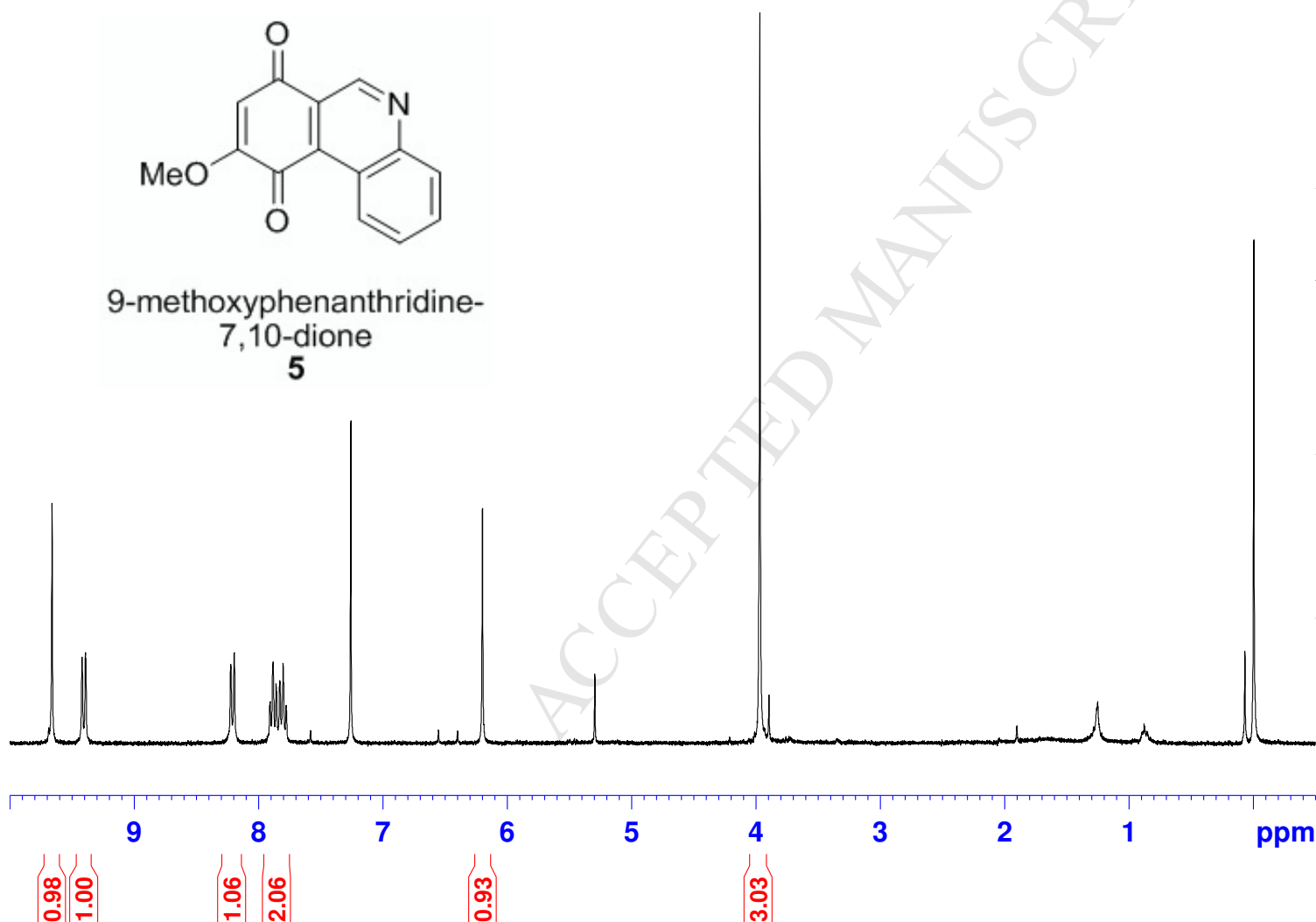
F2 - Acquisition Parameters  
Date\_ 20130103  
Time 21.54  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 16  
DS 2  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953587 sec  
RG 812.7  
DW 80.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

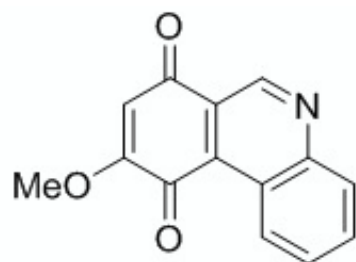
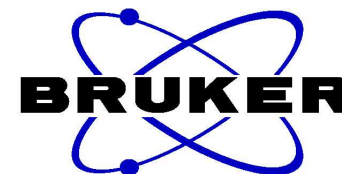
===== CHANNEL f1 =====  
NUC1 1H  
P1 15.00 usec  
PL1 -2.50 dB  
SFO1 299.8818519 MHz

F2 - Processing parameters  
SI 32768  
SF 299.8800036 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00

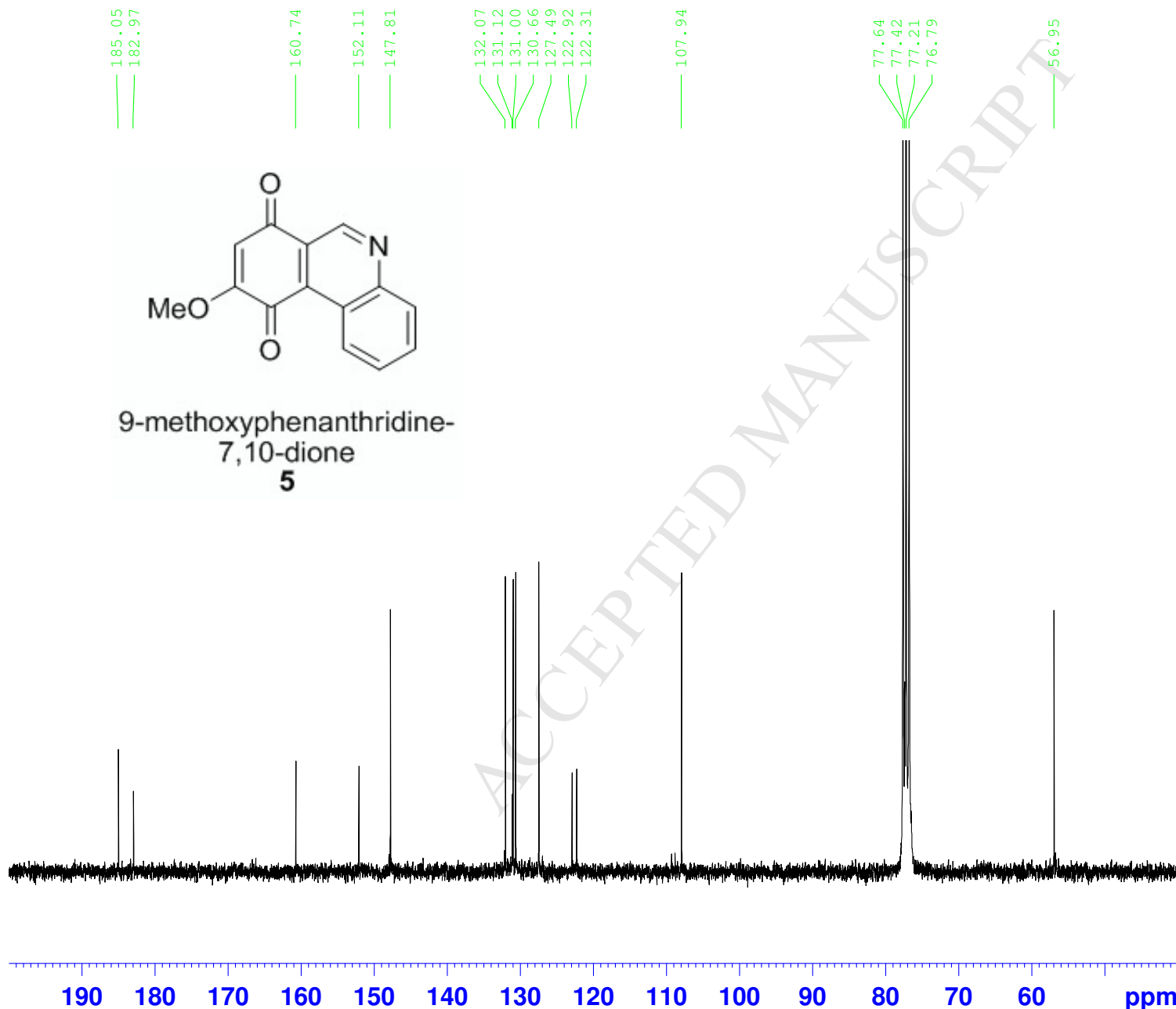


9-methoxyphenanthridine-  
7,10-dione  
5



SX-I-246-Column-F1 <sup>13</sup>CNMR CDCl<sub>3</sub>

9-methoxyphenanthridine-  
7,10-dione  
**5**



Current Data Parameters  
NAME SX-I-246-Column-F1  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20130103  
Time 22.02  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 9418  
DS 4  
SWH 17985.611 Hz  
FIDRES 0.274439 Hz  
AQ 1.8219508 sec  
RG 32768  
DW 27.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
d11 0.03000000 sec  
DELTA 1.89999998 sec  
TD0 1

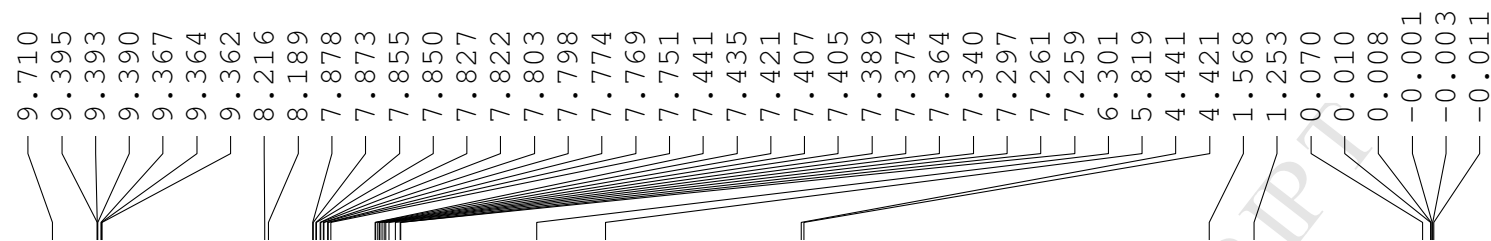
===== CHANNEL f1 =====  
NUC1 13C  
P1 8.00 usec  
PL1 -1.80 dB  
SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 75.00 usec  
PL2 -2.50 dB  
PL12 11.48 dB  
PL13 12.00 dB  
SFO2 299.8811995 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4048695 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



SX-I-252-Column 1HNMR CDCl3

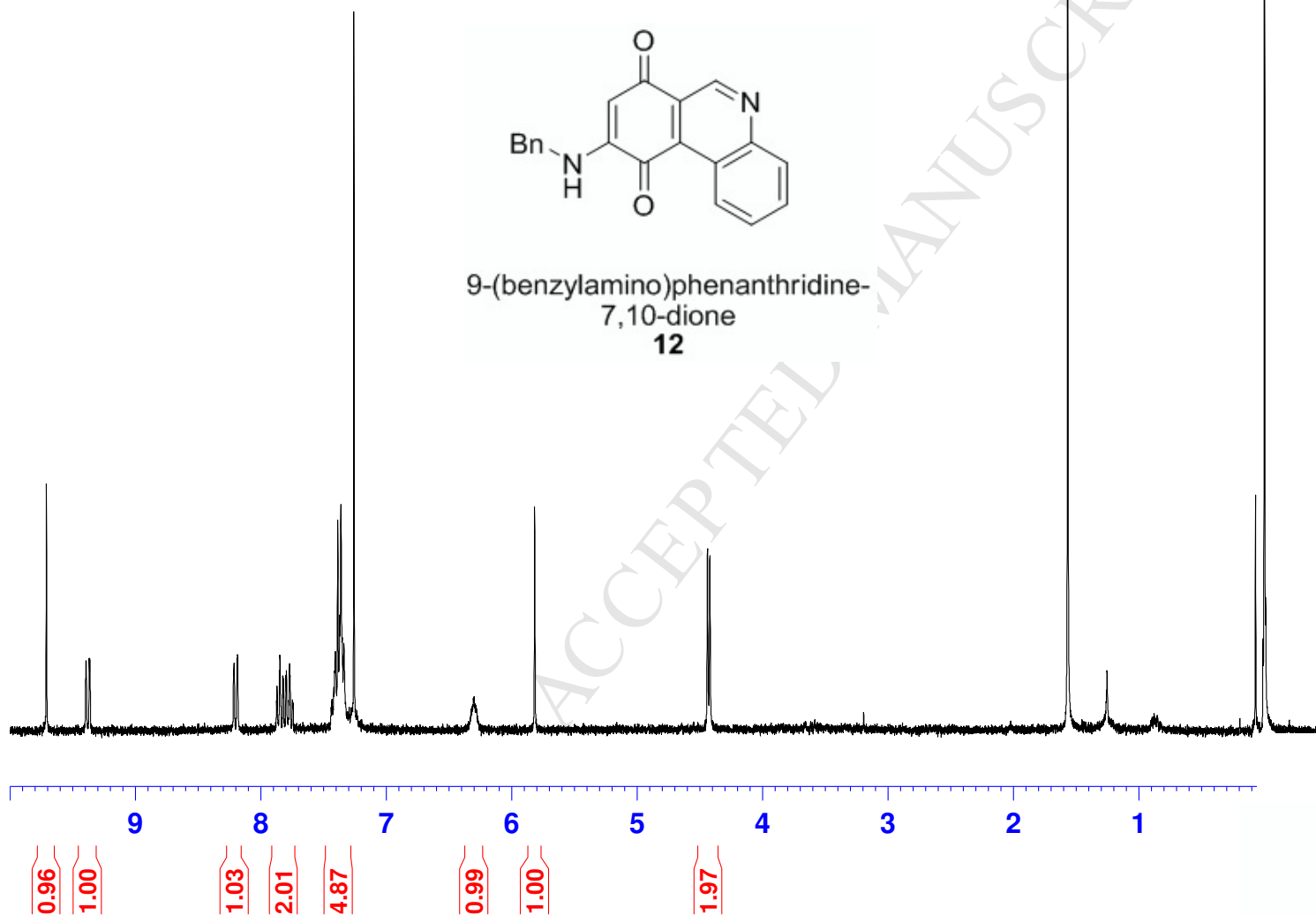
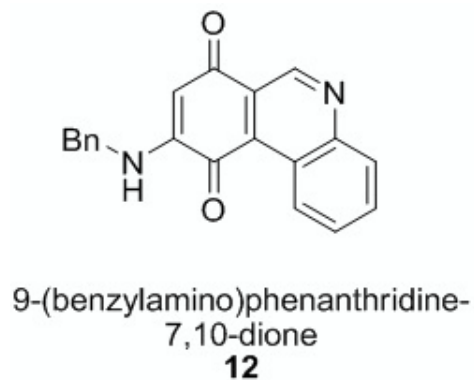


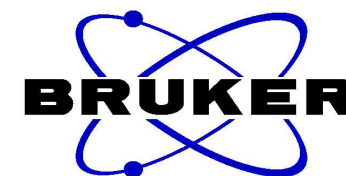
Current Data Parameters  
NAME SX-I-252-Column  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20130110  
Time 17.20  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953587 sec  
RG 912.3  
DW 80.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 15.00 usec  
PL1 -2.50 dB  
SFO1 299.8818519 MHz

F2 - Processing parameters  
SI 32768  
SF 299.8800036 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



SX-I-252-Column <sup>13</sup>CNMR CDC13

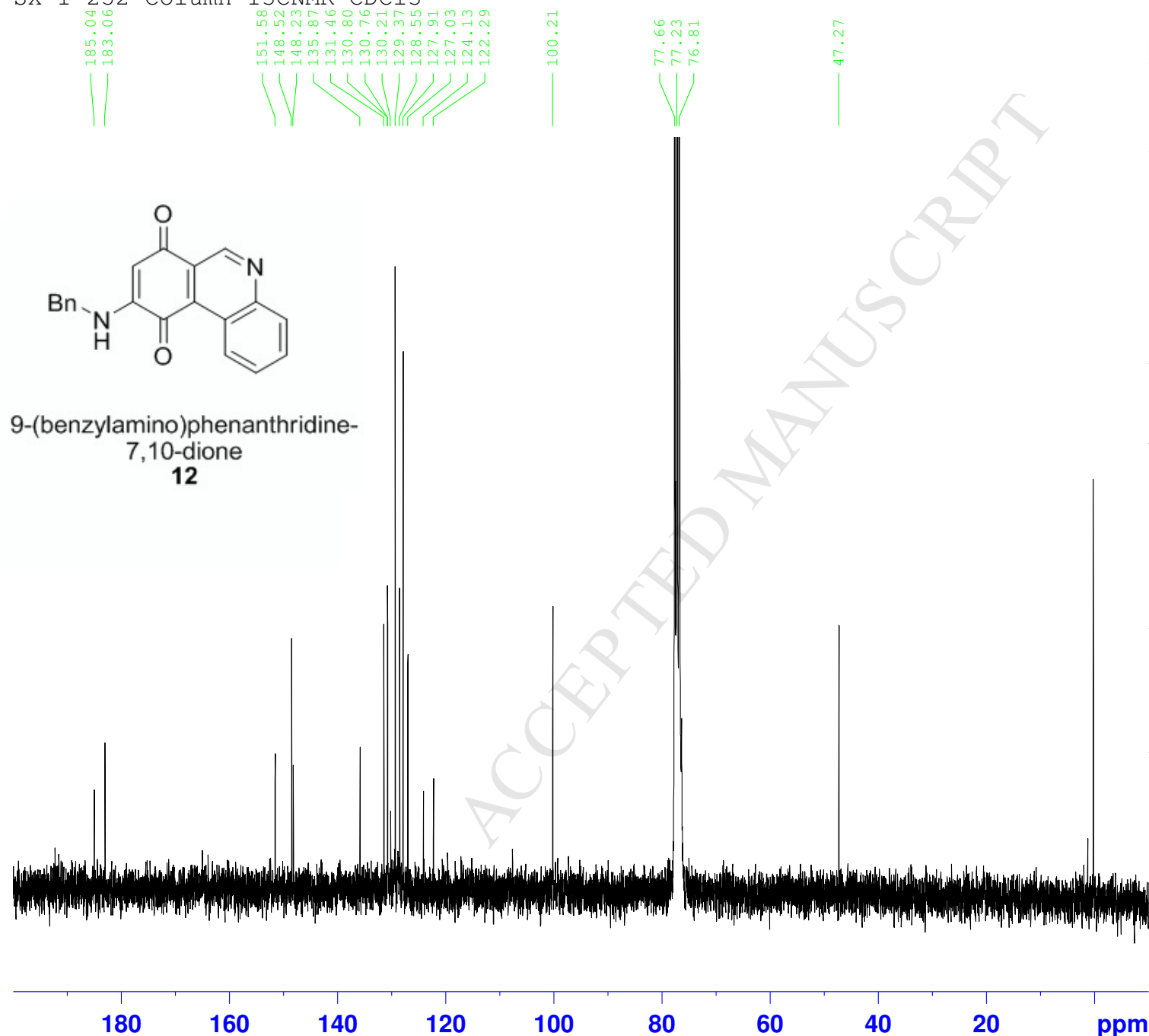
Current Data Parameters  
 NAME SX-I-252-Column  
 EXPNO 2  
 PROCNO 1

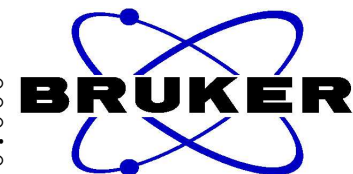
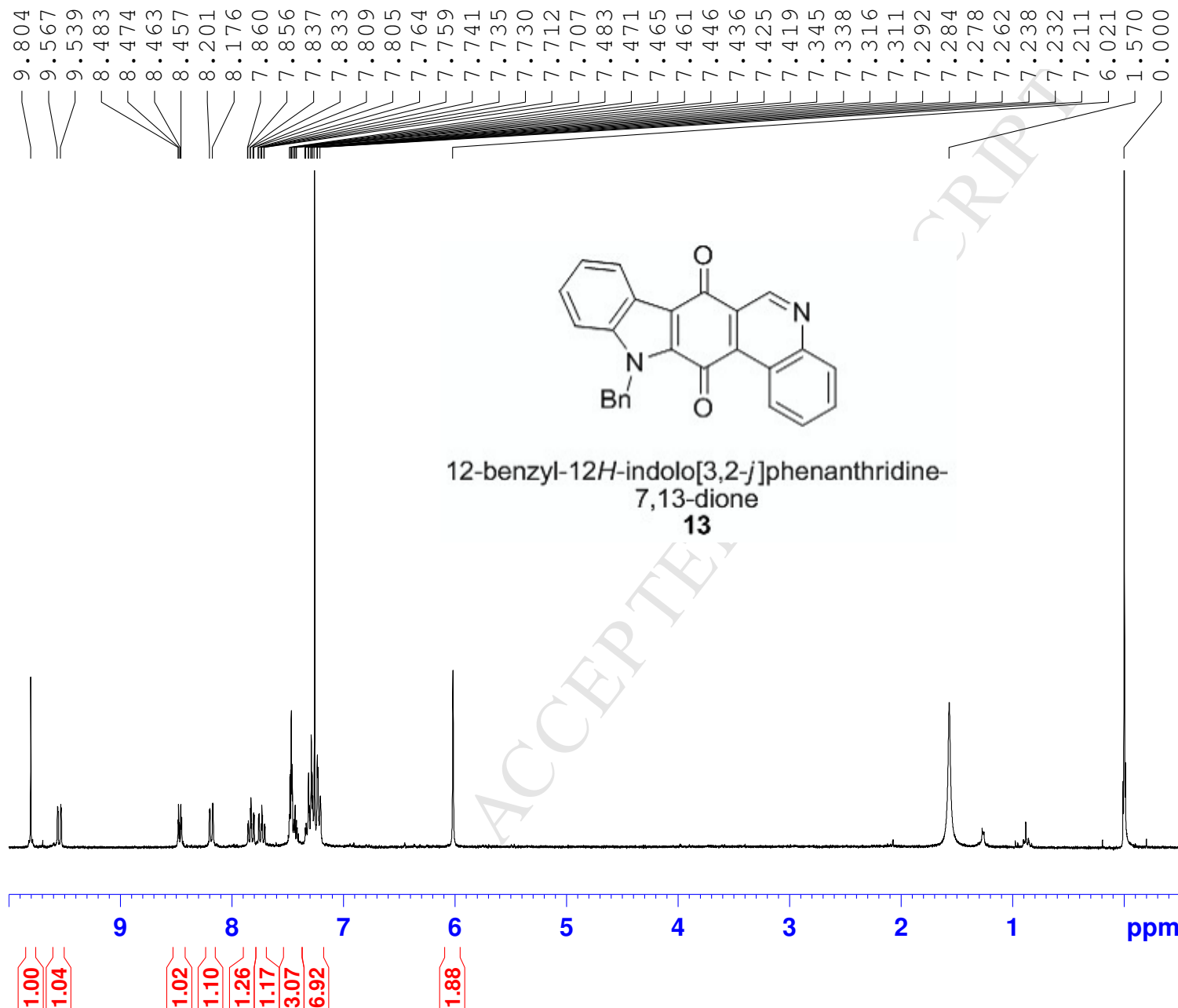
F2 - Acquisition Parameters  
 Date\_ 20130110  
 Time 17.28  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 13843  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4048677 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



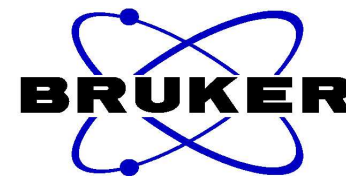
SX-I-256-recrystallization 1H NMR CDCl<sub>3</sub>

Current Data Parameters  
 NAME SX-I-258-recrystallization  
 EXPNO 2  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130117  
 Time 16.07  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 125  
 DS 2  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 5.2953587 sec  
 RG 1024  
 DW 80.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 15.00 usec  
 PL1 -2.50 dB  
 SFO1 299.8818519 MHz

F2 - Processing parameters  
 SI 32768  
 SF 299.8800035 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.00

SX-I-256-recrystallization <sup>13</sup>CNMR CDCl<sub>3</sub>

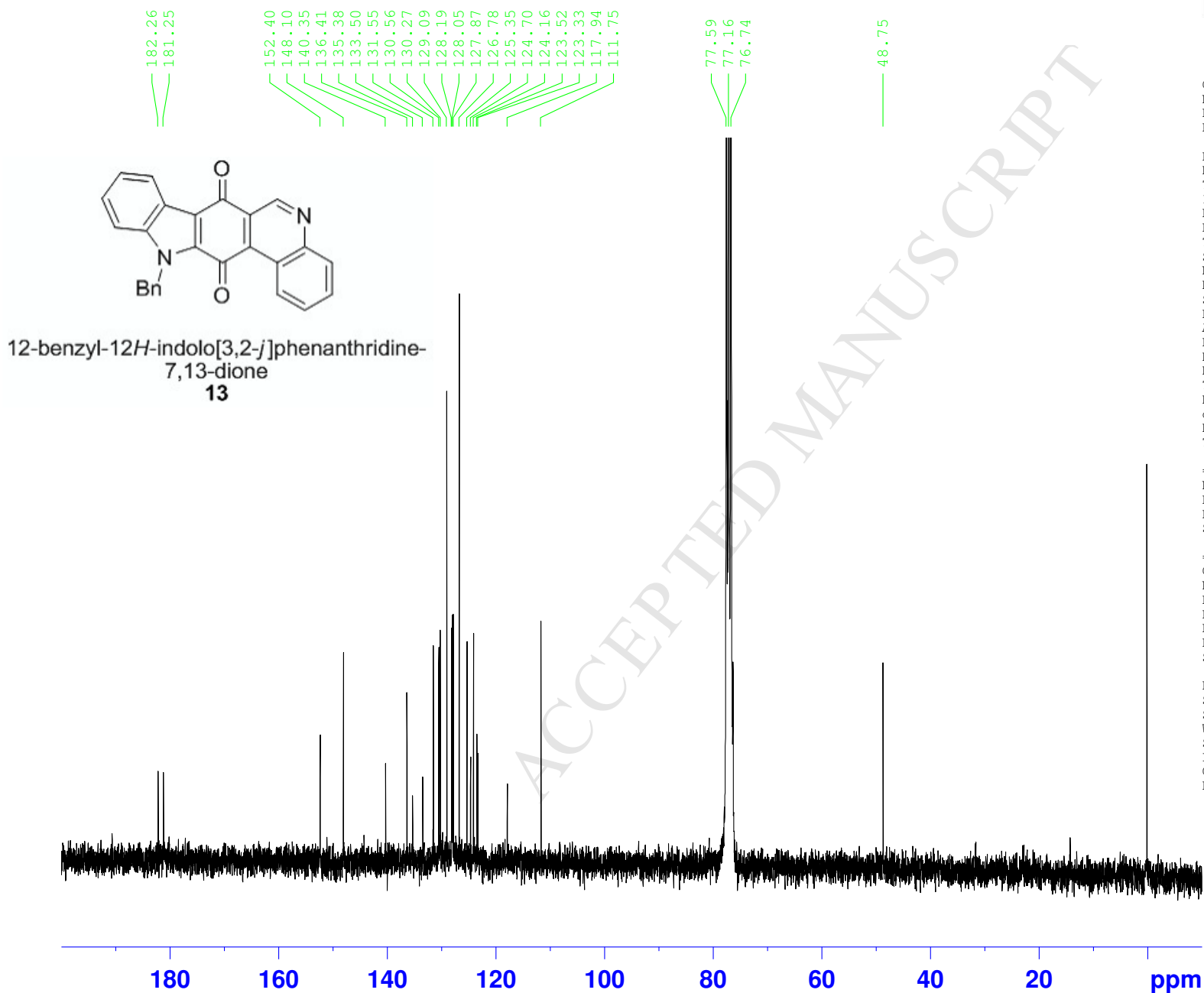
Current Data Parameters  
 NAME SX-I-258-recrystallization  
 EXPNO 5  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20130119  
 Time 15.48  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 37109  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

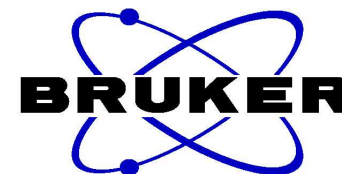
===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4048735 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



SX-II-8-Solid 1HNMR DMSO



Current Data Parameters  
NAME SX-II-8-Solid  
EXPNO 2  
PROCNO 1

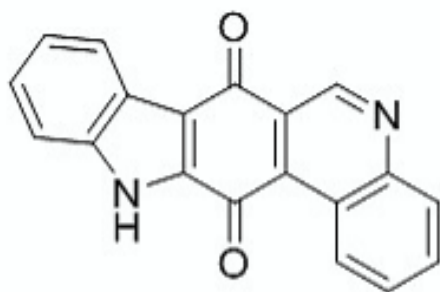
F2 - Acquisition Parameters  
Date\_ 20131011  
Time 23.02  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zg30  
TD 65536  
SOLVENT DMSO  
NS 40  
DS 2  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953587 sec  
RG 812.7  
DW 80.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 15.00 usec  
PL1 -2.50 dB  
SFO1 299.8818519 MHz

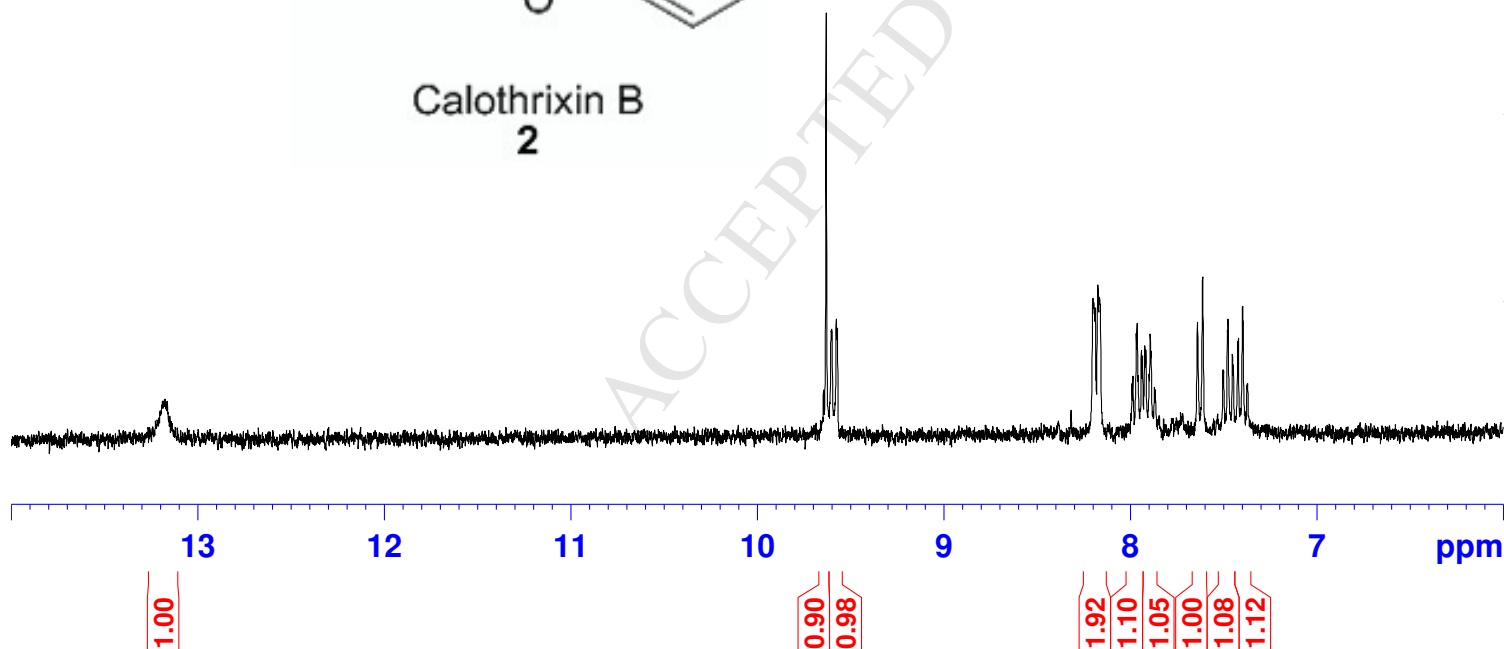
F2 - Processing parameters  
SI 32768  
SF 299.8799971 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

13.175

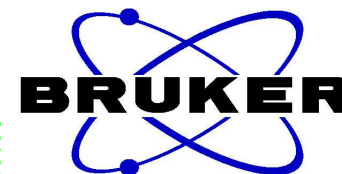
9.645  
9.631  
9.601  
9.576  
8.200  
8.174  
7.991  
7.986  
7.968  
7.963  
7.940  
7.935  
7.922  
7.917  
7.893  
7.870  
7.640  
7.612  
7.503  
7.476  
7.453  
7.421  
7.398



Calothrixin B  
2





SX-II-Calothrixin B <sup>13</sup>CNMR DMSO

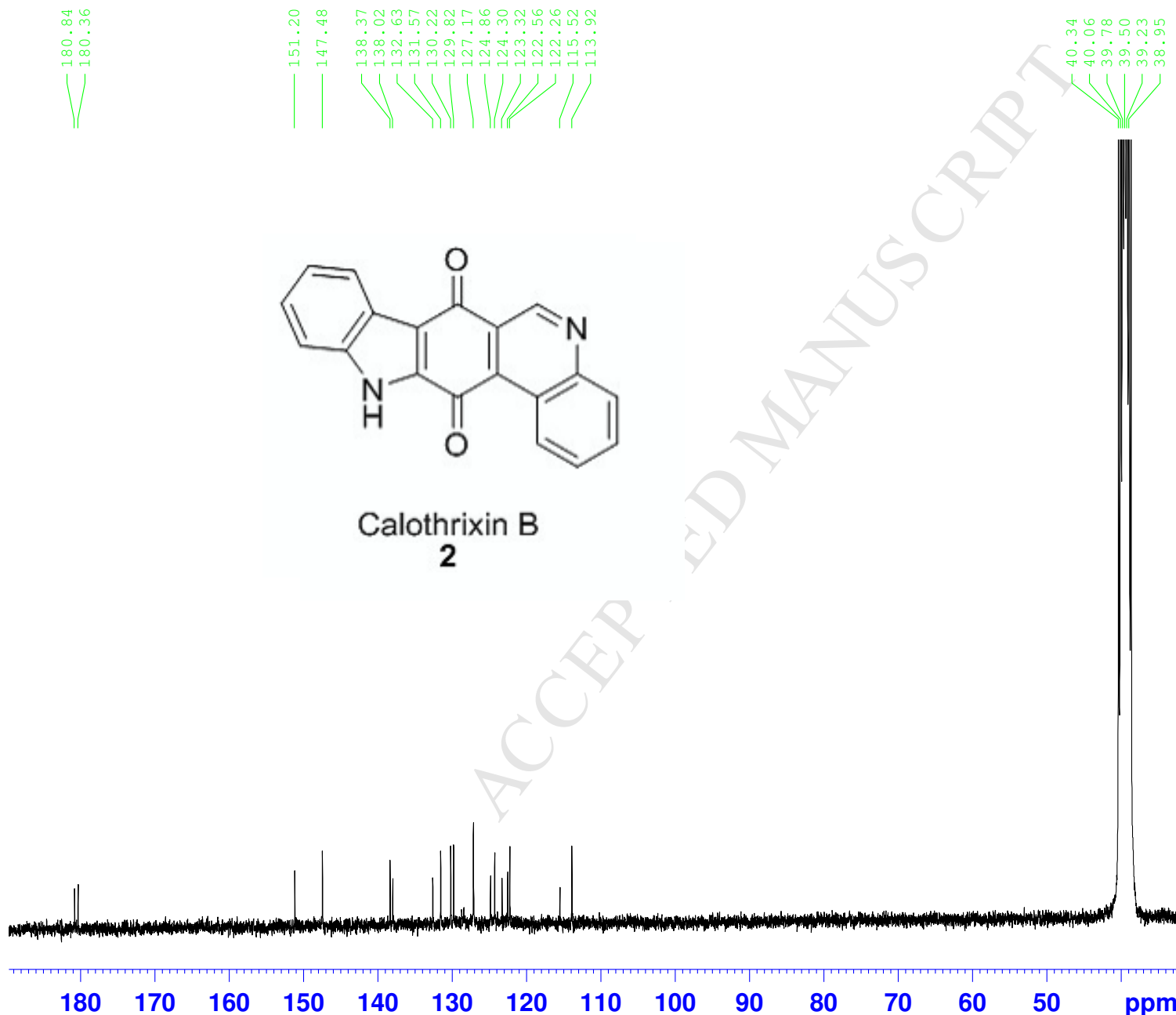
Current Data Parameters  
 NAME SX-II-Calothrixin B  
 EXPNO 6  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20140214  
 Time 21.29  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT DMSO  
 NS 45218  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 2.00000000 sec  
 d11 0.03000000 sec  
 DELTA 1.89999998 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 8.00 usec  
 PL1 -1.80 dB  
 SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 75.00 usec  
 PL2 -2.50 dB  
 PL12 11.48 dB  
 PL13 12.00 dB  
 SFO2 299.8811995 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4049241 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



SX-II-41 1HNMR DMSO-d6

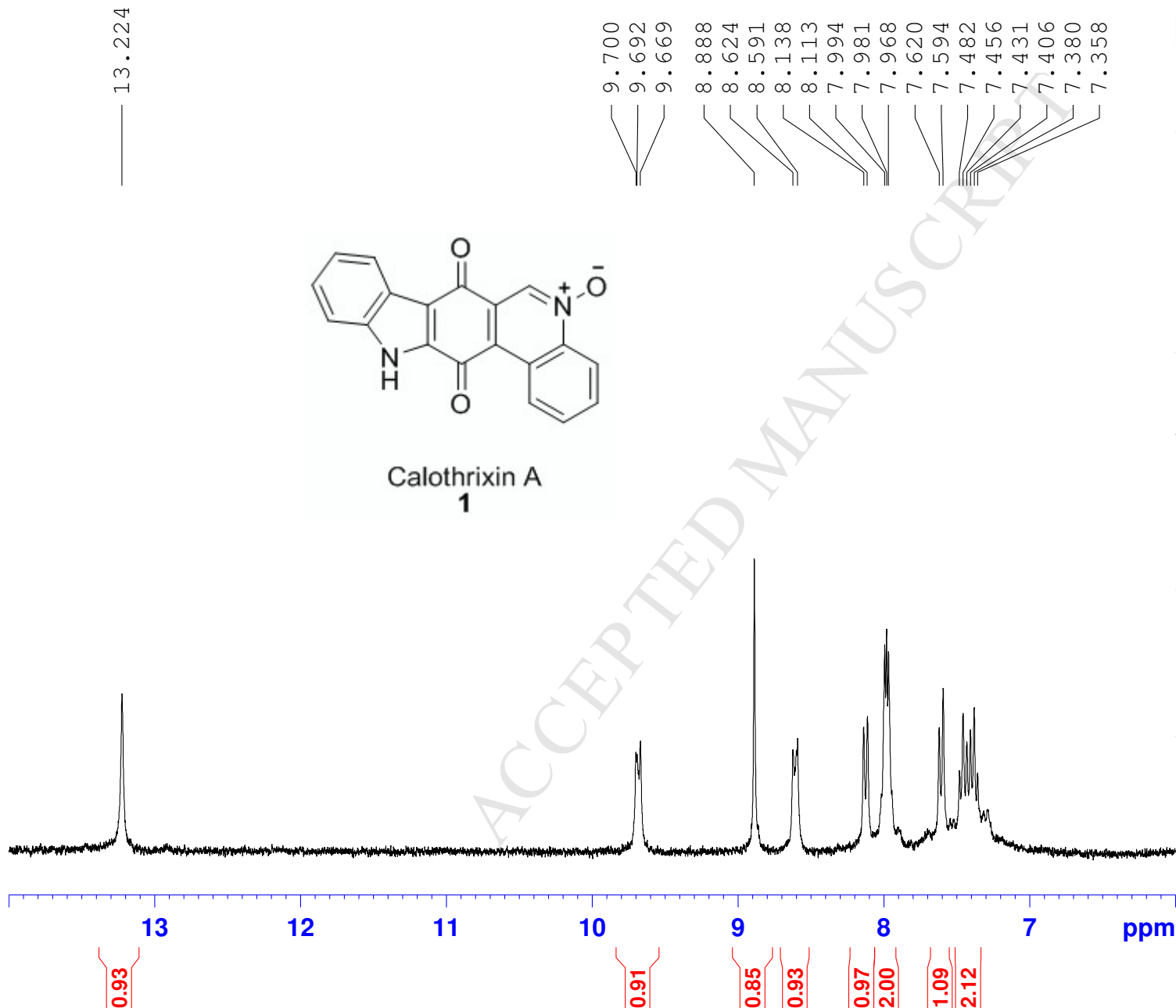


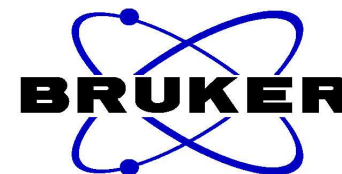
Current Data Parameters  
 NAME SX-II-41  
 EXPNO 1  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20140228  
 Time 9.39  
 INSTRUM spect  
 PROBHD 5 mm DUL 13C-1  
 PULPROG zg30  
 TD 65536  
 SOLVENT DMSO  
 NS 64  
 DS 2  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 5.2953587 sec  
 RG 1024  
 DW 80.800 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 15.00 usec  
 PL1 -2.50 dB  
 SFO1 299.8818519 MHz

F2 - Processing parameters  
 SI 32768  
 SF 299.879979 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



SX-II-41 <sup>13</sup>CNMR DMSO-d<sub>6</sub>

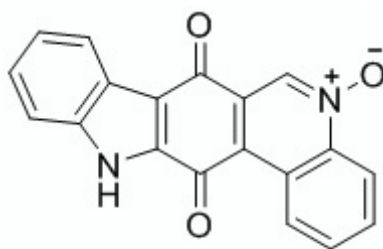
Current Data Parameters  
NAME SX-II-41  
EXPNO 3  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20140301  
Time 8.31  
INSTRUM spect  
PROBHD 5 mm DUL 13C-1  
PULPROG zgpg30  
TD 65536  
SOLVENT DMSO  
NS 44265  
DS 4  
SWH 17985.611 Hz  
FIDRES 0.274439 Hz  
AQ 1.8219508 sec  
RG 32768  
DW 27.800 usec  
DE 6.00 usec  
TE 300.0 K  
D1 2.00000000 sec  
d11 0.03000000 sec  
DELTA 1.89999998 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 8.00 usec  
PL1 -1.80 dB  
SFO1 75.4124265 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 75.00 usec  
PL2 -2.50 dB  
PL12 11.48 dB  
PL13 12.00 dB  
SFO2 299.8811995 MHz

F2 - Processing parameters  
SI 32768  
SF 75.4049227 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



Calothrixin A  
1

