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Orally Active Docetaxel Analogue: Synthesis of 10-Deoxy-10-C-morpholinoethyl Docetaxel Analogues

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Abstract—To improve cytotoxicity of 10-deoxy-10-*C*-morpholinoethyl docetaxel analogues against various tumor cell lines including resistant cells expressing P-glycoprotein (P-gp), we modified the 7-hydroxyl group to hydrophobic groups (methoxy, deoxy, 6,7olefin, α -F, 7- β -8- β -methano, fluoromethoxy). Among these analogues, the 7-methoxy analogue showed the strongest cytotoxicity. This analogue showed potent activity against B16 melanoma BL6 in vivo by oral administration. © 2001 Elsevier Science Ltd. All rights reserved.

Paclitaxel (1, Taxol[®])¹ and docetaxel (2, Taxotere[®])² are currently considered to be some of the most important drugs in cancer chemotherapy. However, their low water-solubility requires co-injection of a detergent, Cremophor[®] EL or Tween[®] 80. These detergents frequently cause untoward hypersensitivity reactions, and patients receiving these drugs require premedication.³ To resolve these problems, we previously described the synthesis of non-prodrug water-soluble docetaxel analogues, such as 10-*O-sec*-aminoethyl docetaxel analogues⁴ and 10-deoxy-10-*C-sec*-aminoalkyl docetaxel analogues.⁵ Among these analogues, 10-deoxy-10-*C*morpholinoethyl docetaxel analogue (3) exhibited cytotoxicity equal to that of docetaxel (2) (Table 1).

However, the cytotoxicity of **3** against drug-resistant cell lines expressing P-gp was not sufficiently potent.



Figure 1. Structure of paclitaxel analogues.

Many patients whose initial response is good go into relapse because of the development of drug resistance after treatment.⁶ To overcome this problem, we modified the 7-hydroxyl group of the 10-C-morpholinoethyl docetaxel analogue (3) to the groups that have been reported to be able to improve the cytotoxicity against cancer cell lines such as methoxy, deoxy, 6,7-olefin, etc. We found that the 7-methoxy analogue showed the most potent activity, regardless of P-gp expression. It has been reported that the poor oral bioavailability of docetaxel is caused by P-gp, which is present in the gastrointestinal tract.7 Consequently, the oral bioavailability of paclitaxel could be increased markedly with Pgp inhibitors.⁸ Therefore, a taxane that is a poor substrate of P-gp showed oral bioavailability.9 On the basis of these observations, the 7-methoxy analogue, a poor substrate of P-gp, appeared to have good bioavailability and showed antitumor activity in vivo by oral administration. Here we report modification of the 7-hydroxyl group of 10-deoxy-10-C-morpholinoethyl docetaxel analogues, and the results of in vivo studies.

Chemistry

10-Deoxy-7-*O*-methyl-10-*C*-morpholinoethyl docetaxel (7) was synthesized from 10-deacetoxy-10-*C*-morpholinoethyl-7,13-*O*-bis-TES baccatin III (4)⁵ (Scheme 1). Selective desilylation and substitution with MTM and

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following desulfurylation¹⁰ gave the key intermediate (6). Introduction of the phenylisoserine side chain using protected β -lactam (8)¹¹ as a side-chain precursor and deprotection using the reported method¹¹ gave 7.¹²

Synthesis of 7,10-di-deoxy-10-*C*-morpholinoethyl docetaxel (12) is described in Scheme 2. The key intermediate (11) was synthesized from 9^{13} in the reported manner.⁵ Introduction of the phenylisoserine side chain,

 Table 1. Cytotoxicity of 10-C-morpholinoethyl docetaxel analogues

 modified 7-hydroxyl group^a

	7-group	Cytoto	Cytotoxic activity GI50 (ng/mL) ^b			
		PC-6	PC-12	PC-6/VCR29-9		
2		0.408-2.55	11.7-72.7	39.6-230		
3	OH	1.72	21.5	76.3		
7	OMe	0.266	0.756	9.01		
12	Deoxy	0.761	1.47	20.4		
15	olefin	0.500	0.824	18.0		
18	α-F	0.379	2.55	16.2		
21	Methano	0.873	3.56	38.6		
27	OCH_2F	0.246	1.50	40.4		

^aThe in vitro experiments were performed with three different cell lines: PC-6, a human small cell lung cancer,²³ and its variant, PC-6/VCR29-9, vincristine-resistant cell line expressing P-gp,²⁴ PC-12, a human non-small cell lung cancer cell line.²³ Determination of GI50 was performed using the MTT assay.²⁵ The cells were exposed continuously to the test compounds for 72 h.

^bGrowth inhibition of 50%: the concentration required to obtain half of the maximal inhibition for cell growth.



Scheme 1. Reagents and conditions: (a) TsOH, MeOH (71%); (b) Ac_2O , DMSO (42%); (c) Raney-Ni, EtOH, reflux (67%); (d) HF-Py (82%); (e) 8, NaHMDS, THF, $-78 \degree C$ (72%); (f) HF-Py (82%).



Scheme 2. Reagents and conditions: (a) hydrazine (53%); (b) CS₂, MeI, *n*-BuLi (55%); (c) *n*-Bu₃SnH, AIBN, acrolein (30%); (d) NaBH₄ (97%); (e) *o*-nitrophenylseleno cyanide, *n*-Bu₃P (75%); (f) mCPBA (89%); (g) 8, NaH, THF (94%); (h) 1) OsO₄, *N*-methylmorpholine *N*oxide, 2) NaIO₄ (63%); (i) 1) morpholine, H₂, Pd–C, 2) HF–Py (47%).

oxidation, reductive amination, and deprotection gave 12. 14

Synthesis of 6,7-dehydro-7,10-di-deoxy-10-*C*-morpholinoethyl docetaxel (15) is described in Scheme 3. The key intermediate (14) was synthesized from 13^5 in high yield.¹⁵ 14 was converted to 15^{16} by reactions similar to those used for preparation of 12.

Synthesis of 7,10-di-deoxy-7- α -fluoro-10-*C*-morpholinoethyl docetaxel (**18**) is described in Scheme 4. The key intermediate (**17**) was synthesized from **16** in moderately high yield.¹⁷ Compound **17** was converted to **18**.¹⁸



Scheme 3. Reagents and conditions: (a) cat. *p*-TsOH, MeOH (75%); (b) Tf₂O, Py (92%); (c) DBU (98%); (d) HF–Py (96%); (e) 8, LiHMDS, THF (100%); (f) HF–Py (94%); (g) 1) OsO₄, *N*-methylmorpholine *N*-oxide, 2) NaIO₄, 3) morpholine, AcOH, NaBH₃CN (95%).



Scheme 4. Reagents and conditions: (a) DAST (50%); (b) HF–Py (50%); (c) 8, LiHMDS, THF (62%); (d) 1) OsO₄, *N*-methylmorpholine *N*-oxide, 2) NaIO₄, 3) morpholine, AcOH, NaBH₃CN (51%); (e) HF–Py (84%).



Scheme 5. Reagents and conditions: (a) silica gel, $CH_3CN-THF$, 55°C (72%); (b) HF–Py (q.y.); (c) 8, LiHMDS, THF (89%); (d) 1) OsO₄, *N*-methylmorpholine *N*-oxide, 2) NaIO₄, 3) morpholine, AcOH, NaBH₃CN (76%); (e) HF–Py (91%).

Synthesis of 7,10-di-deoxy-7- β -8- β -methano-10-*C*-morpholinoethyl docetaxel (21) is described in Scheme 5. The key intermediate (20) was synthesized from 19 in high yield.¹⁹ Compound 20 was converted to 21.²⁰

Synthesis of 10-deoxy-7-*O*-fluoromethyl-10-*C*-morpholinoethyl docetaxel (27) is described in Scheme 6. The key intermediate (26) was synthesized from 22. We found that oxidative fluorination of 7-*O*-methylthiomethyl ether (23) using NIS and DAST afforded fluoromethyl ether (24) in good yield.²¹ The 10-*C*-alkyl group was introduced in the reported manner.⁵ Compound 26 was converted to 27.²²



Scheme 6. Reagents and conditions: (a) cat. TsOH, MeOH (80%); (b) Ac_2O , DMSO (49%); (c) NIS, DAST, CH_2Cl_2 , 0°C (64%); (d) HF–Py (80%); (e) hydrazine, EtOH (97%); (f) *n*-BuLi, carbon disulfide, methyl iodide, THF (91%); (g) acrolein, (TMS)₃SiH, AIBN, toluene (50%); (h) NaBH₄, THF (77%); (i) *o*-nitrophenylseleno cyanide, *n*-Bu₃P (79%); (j) mCPBA (71%); (k) & LiHMDS, THF (92%); (l) HF–Py (84%); (m) 1) OsO₄, *N*-methylmorpholine *N*-oxide, 2) NaIO₄, 3) morpholine, AcOH, NaBH₃CN (68%).

Table 2. Antitumor activity against B16 melanoma BL-6^a

	Route	Dose (mg/kg)	IR (%) ^b	BWL max. (%) ^c	Mortality
2	iv	100.0	95.1	< 0	0/6
	ро	600.0	6.2	< 0	0/6
7	po	112.5	99	20.0	4/6
		75.0	98.3	5.0	0/6
		50.0	96.2	1.7	0/6
		33.3	82.8	0.9	0/6
		22.2	81.7	< 0	0/6
	iv	75.0	99.4	19.2	5/6
		50.0	98.6	18.0	0/6
		33.3	94.4	9.4	0/6
		22.2	93.7	5.7	0/6

^aB16 melanoma BL6 was kindly provided by Dr. Tsuruo (Institute of Molecular and Cellular Biosciences, University of Tokyo) by courtesy of Dr. Fidler (The University of Texas M. D. Anderson Cancer Center).²⁶ B16 melanoma BL6 cells were subcutaneously inoculated into C57BL/6 mice on day 0. Compounds were administered intravenously or orally on day 4 (single administration). Tumor masses were weighed on day 15.

^bIR ($^{\circ}$)=(1-TWt/TWc)×100 TWt: the mean tumor weight of the treated group. TWc: the mean tumor weight of the control group. ^cBWL max. ($^{\circ}$): Maximum rate of body weight loss (<0 indicates no body weight loss). These analogues (7, 12, 15, 18, 21, 27) having a morpholinoethyl group are more soluble than paclitaxel and docetaxel in acidic solution.

Results and Discussion

Antitumor activities of the 10-deoxy-10-*C*-morpholinoethyl docetaxel analogues modified at the 7-hydroxyl group were determined in vitro and in vivo. The cytotoxicities of 7-modified docetaxel analogues (7, 12, 15, 18, 21, 27) against two human lung cancers and resistant cell lines expressing P-gp were improved in comparison with those of 7-hydroxy analogue (3) (Table 1).

Among these analogues, 7-methoxy analogue (7) showed the most potent cytotoxic activity. We selected compound 7 as a candidate for further in vivo investigation. To evaluate the antitumor effects of compound 7 in vivo, we used B16 melanoma BL6 subcutaneously implanted into mice, and the activity of 7 was compared with that of docetaxel (2). Furthermore, we investigated whether compound 7 exhibits antitumor effects by oral administration or not. Compound 7 showed potent antitumor effects over a wide dose range by both iv and po administration. However, five of the six mice that received compound 7 intravenously at 75 mg/kg died from toxicity. The same dosage by oral administration exhibited potent antitumor activity with an IR value of 98.3% with body weight loss of only 5.0%. The near dosage (112.5 mg/kg) by oral administration gave death. Thus, we considered compound 7 to have excellent oral bioavailability in mice. In contrast, docetaxel at 600.0 mg/kg given po exhibited no antitumor activity, and showed no body weight loss. This result clearly showed that docetaxel has very poor bioavailability (Table 2).

In conclusion, we synthesized several 10-*C*-morpholinoethyl docetaxel analogues modified at the 7-hydroxyl group. The 7-methoxy analogue was the most potent of the analogues prepared and was highly active against B16 melanoma BL6 by both iv and po administration. Further investigations of orally active docetaxel analogues will be reported in the near future.

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16. ¹H NMR of **15** (400 MHz, CDCl₃) δ 1.10 (s, 3H), 1.22 (s, 3H), 1.33 (s, 9H), 1.45–1.55 (m, 2H), 1.72 (s, 3H), 1.83 (s, 3H), 2.23 (dd, J = 8 Hz, 15 Hz, 1H), 2.39 (s, 3H), 2.30–2.60 (m, 8H), 3.68 (t, J = 4.7 Hz, 4H), 3.78 (d, J = 5 Hz, 7 Hz, 1H), 4.19 (d, J = 7 Hz, 1H), 4.31 (d, J = 8 Hz, 1H), 4.42 (d, J = 8 Hz, 1H), 4.60 (s, 1H), 5.11 (d, J = 6 Hz, 1H), 5.27 (d, J = 9 Hz, 1H), 5.41 (d, J = 9 Hz, 1H), 5.72 (d, J = 10 Hz, 1H), 5.83 (d, J = 7 Hz, 1H), 6.02 (dd, J = 6 Hz, 10 Hz, 1H), 6.18 (t, J = 9 Hz, 1H), 7.51 (t, J = 7 Hz, 2H), 7.62 (t, J = 7 Hz, 1H), 8.16 (d, J = 7 Hz, 2H).

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18. ¹H NMR of **18** (400 MHz, CDCl₃) δ 1.09 (s, 3H), 1.21 (s, 3H), 1.33 (s, 9H), 1.68 (s, 3H), 1.77 (s, 3H), 2.40 (s, 3H), 2.14–2.65 (m, 13H), 3.68 (m, 4H), 4.20–4.24 (m, 2H), 4.33, 4.36 (each d, J=9 Hz, total 2H), 4.52 (dd, J=3 Hz, 47 Hz, 1H), 4.61 (s, 1H), 5.02 (dd, J=2 Hz, 9 Hz, 1H), 5.30 (d, J=10 Hz, 1H), 5.41 (d, J=10 Hz, 1H), 5.74 (d, J=7 Hz, 1H), 6.19 (t, J=8 Hz, 1H), 7.40–7.42 (m, 5H), 7.51 (t, J=8 Hz, 2H), 7.61 (t, J=8 Hz, 1H), 8.14 (d, J=8 Hz, 2H).

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20. ¹H NMR of **21** (400 MHz, CDCl₃) δ 1.20 (s, 3H), 1.22 (s, 3H), 1.27 (s, 9H), 1.56 (t, J = 7 Hz, 1H), 1.79 (s, 3H), 1.88 (brs, 1H), 2.12 (m, 1H), 2.20 (m, 1H), 2.39 (m, 4H), 2.43 (m, 6H), 2.55 (m, 1H), 3.71 (m, 4H), 3.75 (m, 1H), 4.04 (d, J = 9 Hz, 1H), 4.27 (d, J = 7 Hz, 1H), 4.29 (d, J = 9 Hz, 1H), 4.60 (brs, 1H), 4.75 (d, J = 3 Hz, 1H), 5.29 (m, 1H), 5.35 (d, J = 10 Hz, 1H), 5.65 (d, J = 7 Hz, 1H), 6.22 (m, 1H), 7.30–7.40 (m, 5H), 7.50 (t, J = 8 Hz, 2H), 7.60 (t, J = 8 Hz, 1H), 8.16 (d, J = 8 Hz, 2H).

21. Experimental procedure of fluorination: To a cooled (icewater) solution of 7-O-methylthiomethyl-13-O-TES baccatin III (100 mg, 0.13 mmol) in CH₂Cl₂, was added N-iodosuccinimide (45 mg, 0.2 mmol) and DAST (35 μ L, 0.20 mmol). Stirring was continued for 1 h at 0 °C. The reaction mixture was poured into satd. NaHCO₃ and the mixture was stirred vigorously for 5 min. The organic phase was separated and dried over anhydrous MgSO₄ and evaporated. The residue was purified by TLC to give **24** as colorless solids (62 mg, 64%).

22. ¹H NMR of **27** (400 MHz, CDCl₃) δ 1.15 (s, 3H), 1.20 (s, 3H), 1.34 (s, 9H), 1.72 (s, 3H), 1.85 (s, 3H), 2.38 (s, 3H), 1.60–2.70 (m, 12H), 3.68 (m, 4H), 4.00–4.06 (m, 2H), 4.19 (d, J=8 Hz, 1H), 4.31 (d, J=8 Hz, 1H), 4.41 (dd, J=7 Hz, 10 Hz, 1H), 4.60 (s, 1H), 4.96 (d, J=8 Hz, 1H), 5.11 (dd, J=8 Hz, 28 Hz, 1H), 5.22–5.28 (m, 2H), 5.41 (d, J=10 Hz, 1H), 5.65 (d, J=7 Hz, 1H), 6.17 (t, J=8 Hz, 1H), 7.30–7.42 (m, 5H), 7.49 (t, J=8 Hz, 2H), 7.61 (t, J=8 Hz, 1H), 8.10 (d, J=8 Hz, 2H).

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