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# Discovery of a novel series of nonacidic benzofuran EP<sub>1</sub> receptor antagonists

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### ABSTRACT

We describe the discovery and optimization of a novel series of benzofuran  $EP_1$  antagonists, leading to the identification of **26d**, a novel nonacidic  $EP_1$  antagonist which demonstrated efficacy in preclinical models of chronic inflammatory pain.

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There is significant interest in the identification of EP<sub>1</sub> receptor antagonists for the treatment of inflammatory pain.<sup>1</sup> Toward this end we have previously reported several classes of EP<sub>1</sub> receptor antagonists, for example, the development compound GW848687X (1)<sup>2</sup> and the related phenyl derivative GSK345931A (2)<sup>3</sup> We have also described the pyrrole series, exemplified by **3** and **4**,<sup>4</sup> and the methylene-linked pyrazole series, represented by **5** and **6**.<sup>5</sup> The latter examples demonstrated the utility of the isobutyl group (R = <sup>i</sup>Pr) as a replacement for the benzyl group (R = Ph), Figure 1. We have also described analogues of **5** and **6** where the carboxylic acid moiety was replaced by nonacidic groups such as amides, reversed amides and heterocycles.<sup>6</sup>

As part of an ongoing medicinal chemistry programme we were interested in identifying new lead series which may offer potential improvements over the compounds described in Figure 1, for example, increased affinity for the EP<sub>1</sub> receptor, increased metabolic stability or increased oral exposure.

During the course of our research activities we conducted small molecule X-ray crystallographic studies with several compounds. Analysis of these structures revealed that the alkoxy group of the left hand phenyl ring adopted a co-planar conformation with the phenyl ring as represented in Figure 2. As the solid state conformation may be a low energy conformation, we hypothesized that it could be mimicked by conformational constraint as outlined in Figure 2. In addition, the resultant benzofurans also have less rotatable bonds than the corresponding alkoxy analogues, a property which has been shown to have a beneficial effect on oral bioavailability.<sup>7</sup>

Initially, we targeted benzofuran derivatives where  $R = {}^{i}Pr$ . The synthesis is outlined in Scheme 1. 2-Methyl-3-butyn-2-ol (**8**) was



Figure 1. Selected GSK EP1 receptor antagonists.

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**Scheme 1.** Reagents and conditions: (a) TFAA, DBU, MeCN, -10 to 0 °C; (b) **9**, DBU, MeCN, CuCl<sub>2</sub>, -10 to 0 °C; (c) PhNEt<sub>2</sub>, CsF, 180 °C, 50%; (d) 1 M LiAlH<sub>4</sub> in THF, THF, rt; (e) PBr<sub>3</sub>, hexane/CH<sub>2</sub>Cl<sub>2</sub> (1:1), rt; (f) SOCl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt; (g) **16**, **14**, DMF, K<sub>2</sub>CO<sub>3</sub>, rt, 59%; (h) NaOH, EtOH, rt.

converted to the corresponding trifluoroacetate derivative **9** and treated in situ with phenol **7** in the presence of copper(II) chloride to give derivative **10**. Acetylene **10** underwent thermal rearrangement in *N*,*N*-diethylaniline to give a separable mixture of **11** and **12**, the former being the major product.<sup>8</sup> Ester reduction gave the corresponding alcohol **13** which was converted to a suitable leaving group and displaced with pyrazole derivative **16** to give two regioisomers (approximately 2:1 when X = Br), of which **17** was the major product (59%). The yield of the undesired regioisomer was 32% and the isomers where easily separated by column chromatography. Ester hydrolysis of **17** thus furnished **18a** (Scheme 1).<sup>9</sup>

We were pleased to find that compound **18a** showed good affinity for the EP<sub>1</sub> receptor in our [<sup>3</sup>H]-PGE<sub>2</sub> binding assay, Table 1.<sup>9</sup> Testing in a functional assay (FLIPR)<sup>9</sup> confirmed that compound **18a** was a potent EP<sub>1</sub> antagonist, FLIPR  $pK_i$  9.4 ± 0.1 (*n* = 3) with good selectivity over the EP<sub>3</sub> receptor subtype, EP<sub>3</sub> FLIPR  $pK_i$  6.0 (*n* = 1).

Investigation of the SAR at the 2-position of the benzofurans, Table 1, revealed that a range of alkyl groups could be introduced, with iso-propyl and cyclohexyl resulting in the highest affinity.

We next sought to investigate benzofuran replacements with the goal of lowering the lipophilicity of the series. Results from this Table 1SAR of C-2 alkyl benzofurans<sup>a</sup>



Compd.	А	$EP_1$ binding $pIC_{50}^{b}$
18a	<sup>i</sup> Pr	8.3 <sup>c</sup>
18b	cPr	7.3
18c	Pr	7.6
18d	Cyhex	8.4
18e	t-Bu	7.2

<sup>a</sup> Data from [<sup>3</sup>H]-GW875240X binding assay.

<sup>b</sup> Values are means of at least three experiments.

<sup>c</sup> Data from [<sup>3</sup>H]-PGE<sub>2</sub> binding assay.

investigation are summarized in Table 2. Disappointingly, all derivatives showed a significant decrease in affinity. Surprisingly, the

Table 2SAR of bicyclic benzofurans replacements<sup>a</sup>



## Values are means of at least three experiments.

<sup>b</sup> Data from [<sup>3</sup>H]-GW875240X binding assay.

<sup>c</sup> Data from [<sup>3</sup>H]-PGE<sub>2</sub> binding assay.

regioisomeric benzofuran **19a** also displayed a 50-fold decrease in activity. More polar heterocycles (**19c–e**) showed very weak affinity.

Retaining the iso-propyl group in the 2-postion of the benzofuran ring, we investigated several replacements for the carboxylic acid, Table 3.

Results in Table 3 indicated that it was possible to incorporate a weakly basic centre to the compounds, compound 20g retained reasonable affinity whilst displaying modest solubility. Compound **20g** was profiled further, a summary of the data generated is listed in Table 4. The compound was found to have good in vitro metabolic stability, poor aqueous solubility, but moderate solubility in fed state simulated intestinal fluid. Compound 20g was also found to be highly permeable in MDCK cells and was not a Pgp substrate (efflux ratio 0.9). Based on this data, compound 20g was profiled in the rat CFA model of inflammatory pain<sup>10</sup> where it demonstrated an oral ED<sub>50</sub> of 14.7 mg/kg, with a dose of 30 mg/kg showing comparable reversal of hypersensitivity to the standard (celecoxib at a dose of 10 mg/kg po). Disappointingly, compound 20g was found to have low oral bioavailability in the rat, which we attributed to its high blood clearance, Table 4. The intrinsic clearance in rat liver microsomes predicts low to moderate metabolic stability for **20g**. however in vivo the clearance is high, equal to or in excess of liver blood flow. The high in vivo clearance may be suggestive of non-cytochrome P450 metabolism also occurring. Despite the high in vivo clearance, bioanalysis from the CFA study revealed that compound **20g** exhibited high blood concentrations 1 h post-dose; 3 mg/kg blood concn  $1.945 \,\mu\text{M}$  (*n* = 7),  $10 \,\text{mg/kg}$  blood concn 5.614  $\mu$ M (*n* = 7), 30 mg/kg blood concn 10.166  $\mu$ M (*n* = 7). The

### Table 3

SAR of various novel carboxylic acid replacements<sup>a</sup>





<sup>a</sup> Values are means of at least three experiments.

<sup>b</sup> Data from [<sup>3</sup>H]-PGE<sub>2</sub> binding assay.

<sup>c</sup> Data from [<sup>3</sup>H]-GW875240X binding assay.

CNS penetration was also good (as measured from the 30 mg/kg dose group); brain concn 11.315  $\mu$ M (*n* = 3) with a blood concn of 10.818  $\mu$ M (*n* = 3) for the corresponding rats, giving a Br:Bl of 1.05. Free fractions in blood and brain tissue were not determined for this compound but are likely to be low in line with that of close analogues such as 26a and 26d, vide infra. The high blood concentrations do not exclude the fact that the efficacy is mediated by a metabolite, however in a separate study compound 20g was dosed orally in the rat CFA model at 5 mg/kg (in 1% methylcellulose, as before) and efficacy was assessed at 1 and 3 h post-dose. At this dose, there was a small but significant reversal of hypersensitivity at the 1 h time point but no effect at the 3 hour time point supporting the fact that the efficacy is mediated by the parent compound, but not excluding a contribution from the 2-isopropenyl metabolite, vide infra, which is likely to have EP<sub>1</sub> activity. The blood and brain concentrations of 20g 3 h post-dose were 0.694 and 0.804 µM giving a Br:Bl of 1.15.

To further optimize this series and improve the in vivo metabolic stability, a study was conducted to assess the sites of metabolism and the vulnerability of this compound to form reactive metabolites. One of the major sites of metabolism was found to be the iso-propyl group in the 2-position of the benzofuran which appeared to undergo oxidation and elimination. In order to address this liability we sought to replace the iso-propyl group with a phenyl group.

#### Table 4

Summary of data for compound 20g



Metabolic stability <sup>a</sup>	Human CL <sub>i</sub> (mL/min/g liver) Rat CL <sub>i</sub> (mL/min/g liver)	1.1 2.8
Solubility <sup>b</sup> (1 h)	Water (µg/mL) SGF (µg/mL) FaSSIF (µg/mL) FeSSIF (µg/mL)	0 1 5 71
Lipophilicity <sup>c</sup>	Log D (pH 7.4)	2.6
Permeability <sup>d</sup>	Pexact Efflux ratio	671 nm/sec 0.9
In vivo activity <sup>e</sup> (CFA)	Doses 3,10 and 30 mg/kg	ED <sub>50</sub> 14.7 mg/kg
Rat pharmacokinetics	$\begin{array}{l} CL_{b}^{f}\left(mL/min/kg\right)\\ V_{ss}^{f}\left(L/kg\right)\\ t_{\gamma_{2}^{f}}\left(h\right)\\ F_{po}^{g}\\ t_{\gamma_{2}^{g}}\left(h\right) \end{array}$	$85 \pm 83.0 \pm 0.30.5 \pm 119 \pm 6\%1.7 \pm 0.2$

<sup>a</sup> See Ref. 11 for details.

<sup>b</sup> Solubility of solid material in each fluid determined at several time points, only 1 h time point quoted. See Ref. 12 for details, SGF (simulated gastric fluid), FaSSIF (fasted state simulated intestinal fluid), FeSSIF (fed state simulated intestinal fluid).

<sup>c</sup> Shake flask method.

<sup>d</sup> MDCK cells.

<sup>e</sup> Efficacy at 1 h post-dose (oral).

 $^{\rm f}$  1 mg/kg intravenous dose, compound dissolved in 2% (v/v) DMSO then added to 0.9% (w/v) saline containing 10% (w/v) Kleptose.

<sup>g</sup> 3 mg/kg oral dose, vehicle = 1% (w/v) methylcelluose aq.

Our initial route to prepare the 2-alkyl benzofurans, Scheme 1, was clearly unsuitable for the preparation of 2-aryl benzofurans. As a result, a new route was devised, Scheme 2. Starting from phenol 7, bromination using *N*-bromosuccinimide afforded **21**. Bromide **21** was subjected to a modified Castro reaction<sup>13</sup> to provide the benzofuran intermediate **22** in one step. Subsequent reduction of ester **22**, and coversion of the intermediate alcohol to a suitable leaving group gave mesylate **23** in good yield. The mesylate was reacted

 Table 5

 Summary of data for composition

Summary of data for compound 26a



EP1 binding affinity	pIC <sub>50</sub>	8.0
Metabolic stability <sup>a</sup>	Human CL <sub>i</sub> (mL/min/g liver) Rat CL <sub>i</sub> (mL/min/g liver)	1.1 1.3
Solubility <sup>b</sup> (1 h)	Water (µg/mL) SGF (µg/mL) FaSSIF (µg/mL) FeSSIF (µg/mL)	171 227 382 0
Lipophilicity <sup>c</sup>	Log D (pH 7.4)	3.0
In vivo activity <sup>d</sup> (CFA)	Dose 10 mg/kg	49% reversal
Rat pharmacokinetics	$\begin{array}{l} CL_{b}^{e} \ (mL/min/kg) \\ V_{ss}^{e} \ (L/kg) \\ t_{y_{2}}^{e} \ (h) \end{array}$	22 3.5 2.1

<sup>a</sup> See Ref. 11 for details.

<sup>b</sup> Solubility of solid material in each fluid determined at several time points, only 1 h time point quoted. See Ref. 12 for details, SGF (simulated gastric fluid), FaSSIF

(fasted state simulated intestinal fluid), FeSSIF (fed state simulated intestinal fluid). <sup>c</sup> Shake flask method.

<sup>d</sup> Efficacy at 1 h post-dose (oral).

 $e^{-1}$  mg/kg intravenous dose, compound dissolved in 2% (v/v) DMSO then added to 0.9% (w/v) saline containing 10% (w/v) kleptose.

with commercially available pyrazole **16** to give **24** as a mixture of regiosiomers which were easily separated using column chromatography. Finally, ester hydrolysis of **24** gave carboxylic acid **25**.<sup>14</sup>

Gratifyingly, this structural change, to give **26a**, Table 5, resulted in increased affinity,  $EP_1$  binding  $pIC_{50}$  8.0, maintained in vitro metabolic stability and improved in vivo metabolic stability (rat  $CL_b$  22 mL/min/kg), supporting the hypothesis that the isopropyl group of **20g** was a major site of metabolism. Surprisingly,



Scheme 2. Reagents and conditions: (a) NBS, DMF, rt; (b) phenylacetylene, CuI, PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub>, Et<sub>3</sub>N, DMF, 75 °C; (c) 1 M LiAlH<sub>4</sub> in THF, THF 0 °C; (d) MsCl, Et<sub>3</sub>N, DCM, 0 °C; (e) 16, DMF, K<sub>2</sub>CO<sub>3</sub>, rt; (h) NaOH, EtOH, 50 °C.

#### Table 6

SAR of various novel carboxylic acid replacements<sup>a</sup>

Compd.	R	$EP_1$ binding <sup>b</sup> pIC <sub>50</sub>
25	CO <sub>2</sub> H	8.7
26b	N N N N N N N N N N N N N N N N N N N	8.0
26c	O NH <sub>2</sub>	8.3
26d	NH H	8.0
26e		8.3
26f	NH NH	7.7
26g		7.2
26h	H NH	7.0
26i		7.4
26j	H N CF <sub>3</sub>	7.6
26k	O NH	8.1
261		8.2
26m		7.6

<sup>a</sup> Data from [<sup>3</sup>H]-GW875240X binding assay.

<sup>b</sup> Values are means of at least three experiments.

despite the increased lipophilicity (log *D* 3.0 @ pH 7.4) this analogue displayed higher solubility in all media tested apart from FaSSIF. When tested in the rat CFA model of inflammatory pain,<sup>10</sup> compound **26a** showed moderate reversal of hypersensitivity (using a weight bearing readout) following an oral dose of 10 mg/kg. Bioanalysis indicated good exposure in both blood and brain (blood concn 2.340  $\mu$ M, *n* = 6, brain concn 3.969  $\mu$ M, *n* = 3, 1 h post-dose, which equates to a Br:Bl of 1.70, *n* = 3).<sup>15</sup> In blood

#### Table 7

Summary of data for compound 26d



EP <sub>1</sub> binding affinity	pIC <sub>50</sub>	8.0
Lipophilicity <sup>a</sup>	Log D (pH 7.4)	2.9
In vivo activity <sup>b</sup> (CFA)	Doses 1, 3, 10 mg/kg	ED <sub>50</sub> 5 mg/kg
Rat pharmacokinetics	CL <sub>b</sub> <sup>c</sup> (mL/min/kg)	28
	$V_{\rm ss}^{\rm c}$ (L/kg)	4.3
	$t_{\frac{1}{2}}^{c}(h)$	4.2

<sup>a</sup> Shake flask method.

<sup>b</sup> Efficacy at 1 h post-dose (oral).

<sup>c</sup> 1 mg/kg intravenous dose, compound dissolved in 2% (v/v) DMSO then added to 0.9% (w/v) saline containing 10% (w/v) kleptose.

and brain tissue binding assays<sup>16</sup> **26a** exhibits high binding with 0.662% free in blood and 0.170% free in brain, that is, Bl Fu 0.0066, Br Fu 0.0017, which results in a calculated free blood concn of 15 nM and free brain concn of 7 nM, which are in line with the binding IC<sub>50</sub> (10 nM). Following this encouraging data, compound **26a** was profiled in the joint pain model of chronic inflammatory pain<sup>17</sup> at doses of 1, 3, 10 and 30 mg/kg po b.i.d.. Disappointingly, a statistically significant reversal (p < 0.001) was only observed in the 30 mg/kg dose group. Thus, although we had made a significant improvement of optimizing the pharmacokinetic parameters of the series, the in vivo efficacy required improvement.

Toward this end, we opted to make further amide analogues, data summarized in Table 6. A selection of neutral amides (**26b**–**e**) showed good affinity. Various groups were assessed in the amide or reversed amide series to try to lower the lipophilicity of the series either via the introduction of a basic amine (**26f**–**j**) or polarity (**26k**–**m**). It was found that basic amines were relatively well tolerated in this region of the molecule, however it appeared that affinity could be increased by decreasing the amine  $pK_a$ , compare **26h** with **26i** and **26j**.

A range of compounds were assessed in the rat CFA model of inflammatory pain<sup>10</sup> (oral dose of 10 mg/kg in 1% methylcellulose, efficacy reading 1 h postdose, when blood and brain samples were also collected) to screen for efficacy and exposure in both blood and brain. Disappointingly, most compounds displayed low efficacy, which could generally be attributed to low exposure. However, compound 26d, displayed promising efficacy in this screen (95% reversal of hypersensitivity, as measured by weight bearing, at 10 mg/kg po) and good CNS penetration (blood concn = 381 nM, brain concn = 112 nM, Br:Bl 0.3). Based on this data, the compound was progressed to a dose-response study in CFA-treated rats (doses 1, 3 and 10 mg/kg po) where it gave an ED<sub>50</sub> of 5 mg/kg. Bioanalysis of the 10 mg/kg dose group showed similar blood exposure to the previous study (blood concn = 493 nM) but higher brain exposure (brain concn = 407nM) which resulted in a Br:Bl of 0.8. The slightly higher Br:Bl in the second experiment is within the range of variability of these assays (<3-fold variation), the same strain and sex of rat and administration vehicle were used in both studies. In blood and brain tissue binding assays<sup>16</sup> 26d exhibits high binding with 0.028% free in blood and 0.017% free in brain, which results in calculated free blood and free brain concentrations considerably lower than the in vitro binding IC<sub>50</sub> value of this compound. The reason for this is unknown, however, there are likely to be errors in the measurements of the free fractions in both tissues with such lipophilic compounds. It is also feasible that the affinity at the rat receptor is higher than that at the human receptor. However without an assessment of in vivo/ex vivo receptor binding it is not possible to say whether the concentrations achieved in these studies were sufficient to block the receptor or whether the effect is on-target. Assessment in the joint pain model of chronic inflammatory pain<sup>17</sup> (at doses of 0.3, 1, 3 and 10 mg/kg po b.i.d.) produced a dose-related reversal of hypersensitivity with a calculated ED<sub>50</sub> of ~0.6 mg/kg and a corresponding calculated blood concentration of 0.06  $\mu$ M. Full reversal of hypersensitivity was achieved at 10 mg/kg. Profiling of the compound in a rat PK study demonstrated good metabolic stability with satisfactory half-life, Table 7.

In conclusion we have described the identification of a new series of  $EP_1$  antagonists which enriches the chemical tools available for the investigation of  $EP_1$  pharmacology. We have also described the use of metabolite identification in lead optimisation which resulted in compounds such as **26a** and **26d**, which, to the best of our knowledge, are the first nonacidic  $EP_1$  antagonists to demonstrate efficacy in preclinical models of chronic inflammatory pain.

#### **References and notes**

- 1. Hall, A.; Billinton, A.; Giblin, G. M. P. Curr. Opin. Drug Discovery Dev. 2007, 10, 597. and references cited therein.
- Giblin, G. M. P.; Bit, R. A.; Brown, S. H.; Chaignot, H. M.; Chowdhury, A.; Chessell, I. P.; Clayton, N. M.; Coleman, T.; Hall, A.; Hammond, B.; Hurst, D. N.; Michel, A. D.; Naylor, A.; Novelli, R.; Scoccitti, T.; Spalding, D.; Tang, S. P.; Wilson, A. W.; Wilson, R. Bioorg. Med. Chem. Lett. 2007, 17, 385.
- Hall, A.; Brown, S. H.; Budd, C.; Clayton, N. M.; Giblin, G. M. P.; Goldsmith, P.; Hayhow, T. G.; Hurst, D. N.; Naylor, A.; Rawlings, D. A.; Scoccitti, T.; Wilson, A. W.; Winchester, W. J. Bioorg. Med. Chem. Lett. 2009, 19, 497.

- Hall, A.; Atkinson, S.; Brown, S. H.; Chessell, I. P.; Chowdhury, A.; Clayton, N. M.; Coleman, T.; Giblin, G. M. P.; Gleave, R. J.; Hammond, B.; Healy, M. P.; Johnson, M. R.; Michel, A. D.; Naylor, A.; Novelli, R.; Spalding, D. J.; Tang, S. P.; Wilson, R. J. Bioorg. Med. Chem. Lett. 2006, 16, 3657.
- McKeown, S. C.; Hall, A.; Giblin, G. M. P.; Lorthioir, O.; Blunt, R.; Lewell, X. Q.; Wilson, R. J.; Brown, S. H.; Chowdhury, A.; Coleman, T.; Watson, S. P.; Chessell, I. P.; Pipe, A.; Clayton, N.; Goldsmith, P. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4767.
- (a) Hall, A.; Billinton, A.; Brown, S. H.; Clayton, N. M.; Chowdhury, A.; Giblin, G. M. P.; Goldsmith, P.; Hayhow, T. G.; Hurst, D. N.; Kilford, I. R.; Naylor, A.; Passingham, B.; Winyard, L. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 3392; (b) Hall, A.; Billinton, A.; Bristow, A. K.; Brown, S. H.; Chowdhury, A.; Cutler, L.; Giblin, G. M. P.; Goldsmith, P.; Hayhow, T. G.; Kilford, I. R.; Naylor, A.; Passingham, B.; Rawlings, D. A. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4027.
- Veber, D. F.; Johnson, S. R.; Cheng, H.-Y.; Smith, B. R.; Ward, K. W.; Kopple, K. D. J. Med. Chem. 2002, 45, 2615.
- Attwood, M. R.; Churcher, I.; Dunsdon, R. M.; Hurst, D. N.; Jones, P. S. Tetrahedron Lett. 1991, 32, 811.
- 9. Gibson, M.; Hall, A.; Hurst, D.N.; Rawlings, D.A. WO2007113289A1.
- (a) Iadarola, M. J.; Douglass, J.; Civelli, O.; Naranjo, J. R. *Brain Res.* **1988**, 455, 205;
   (b) Hay, C. H.; Trevethick, M. A.; Wheeldon, A.; Browers, J. S.; De Belleroche, J. S. *Neuroscience* **1997**, 7, 843.
- Clarke, S. E.; Jeffrey, P. Xenobiotica 2001, 31, 591. In this assay CL<sub>i</sub> of 5 mL/min/g liver corresponds to approximately 60% turnover after 30 min incubation.
- (a) Kostewicz, E. S.; Wunderlich, M.; Brauns, U.; Becker, R.; Bock, T.; Dressman, J. B. J. Pharm. Pharmacol. 2005, 56, 43; (b) Galia, E.; Nicolaides, E.; Horter, D.; Löbenberg, R.; Reppas, C.; Dressman, J. B. Pharm. Res. 1998, 15, 698.
- (a) Castro, C. E.; Gaughan, E. J.; Owsley, D. C. J. Org. Chem. **1966**, 31, 4071; (b) Castro, C. E.; Havlin, R.; Honwad, V. K.; Malte, A.; Mojé, S. J. Am. Chem. Soc. **1969**, 91, 6464.
- 14. Eatherton, A. J.; Giblin, G. M. P.; Gibson, M.; Hurst, D. N. WO2008098978A2/A3.
- 15. Note the brain concentrations were measured in only three rats, mean brain concentration =  $3.969 \mu$ M. The average blood concentration from the same
- three rats was 2.339 μM. Thus, this equates to a Br:Bl of 1.70, n = 3.
  Summerfield, S. G.; Stevens, A. J.; Cutler, L.; del Carmen Osuna, M.; Hammond, B.; Tang, S-P.; Hersey, A.; Spalding, D. J.; Jeffrey, P. J. Pharmacol. Exp. Ther. 2006, 316, 1282.
- Wilson, A. W.; Medhurst, S. J.; Dixon, C. I.; Bontoft, N. C.; Winyard, L. A.; Brackenborough, K. T.; De Alba, J.; Clarke, C. J.; Gunthorpe, M. J.; Hicks, G. A.; Bountra, C.; McQueen, D. S.; Chessell, I. P. *Eur. J. Pain* **2006**, *10*, 537.