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Antileishmanial Activity of a Series of N^2, N^4 -Disubstituted Quinazoline-2,4-diamines

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Abstract

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3 A series of N^2,N^4 -disubstituted quinazoline-2,4-diamines has been synthesized and tested against
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5 *Leishmania donovani* and *L. amazonensis* intracellular amastigotes. A structure-activity and structure-
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7 property relationship study was conducted in part using the Topliss operational scheme to identify new
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9 lead compounds. This study led to the identification of quinazolines with EC_{50} s in the single digit
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11 micromolar or high nanomolar range in addition to favorable physicochemical properties. Quinazoline
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13 **23** also displayed efficacy in a murine model of visceral leishmaniasis, reducing liver parasitemia by
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15 37% when given by the intraperitoneal route at 15 mg/kg/day for five consecutive days. Their
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17 antileishmanial efficacy, ease of synthesis, and favorable physicochemical properties make the N^2,N^4 -
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19 disubstituted quinazoline-2,4-diamine compound series a suitable platform for future development of
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21 antileishmanial agents.
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28 Keywords: leishmaniasis, protozoa, quinazoline, structure-activity relationship, structure-property
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30 relationship
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Introduction

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3 Leishmaniasis is a debilitating disease that is prevalent across the globe with 350 million people in 88
4 countries at risk of acquiring leishmaniasis.^{1, 2} A recent effort by the World Health Organization to
5 provide a current estimate of the incidence of leishmaniasis concluded that between 0.2 and 0.4 million
6 cases of visceral leishmaniasis occur each year, while from 0.7 to 1.2 million cases of cutaneous
7 leishmaniasis occur each year.³ This study also estimated that from 20,000 to 40,000 deaths occur each
8 year due to leishmaniasis. More than twenty *Leishmania* species are known to cause the disease in
9 humans.⁴ Leishmaniasis manifests itself in numerous forms depending on which parasite species infects
10 the host. The parasites are transferred from host to host by about thirty species of female sandfly vectors
11 of the genera *Phlebotomus* and *Lutzomyia* that infect the host when taking a blood meal.⁵ Symptoms of
12 leishmaniasis include unsightly spontaneously healing ulcers on the skin when cutaneous leishmaniasis
13 presents, non-healing lesions in the mucosa when mucocutaneous leishmaniasis is the affliction, and
14 chronic, debilitating infection of the reticuloendothelial system which is fatal if left untreated due to
15 visceral leishmaniasis.¹ The majority of cases of visceral leishmaniasis are caused by *L. donovani* in
16 East Africa and Asia, *L. infantum* in the Mediterranean region, and *L. chagasi* in Latin America.⁶ It
17 should be noted that the latter two are genetically identical.^{7, 8} *L. infantum* and *L. chagasi* mainly affect
18 children and immunocompromised individuals and are zoonotic parasites with canines being a major
19 reservoir.¹ *L. donovani*, on the other hand, is an anthroponotic parasite that affects a broad range of
20 ages.¹

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22 For nearly one hundred years, antimonials have been the drug of choice to combat leishmaniasis. In
23 1912 Gaspar Vianna first reported the use of the trivalent antimonial tartar emetic for the treatment of
24 cutaneous leishmaniasis caused by *L. braziliensis* in Brazil.⁹ Shortly thereafter McCombie Young and
25 Upendranath Brahmachari used trivalent and pentavalent antimonials to treat visceral leishmaniasis in
26 India with great success, decreasing the mortality rate of 95% to just 10% in ten years (Figure 1.A).¹⁰
27 Pentavalent antimonials such as meglumine antimoniate and sodium stibogluconate are currently the
28 first line antileishmanial drugs in many areas.^{1, 11, 12} Treatment involves daily injections for up to a thirty

1 day period.¹¹ Problems with this treatment include a high rate of resistance that has been encountered in
2 India, especially the state of Bihar, where up to 60% of infected individuals do not improve with
3 treatment.^{11, 13} The high rate of resistance to pentavalent antimonials in India has led to the increasing
4 use of amphotericin B and miltefosine against visceral leishmaniasis.¹⁴ Since the 1960's, amphotericin
5 B has been the second line treatment for visceral leishmaniasis.¹¹ It has a cure rate of over ninety
6 percent, but is often accompanied by severe side effects such as nephrotoxicity that require
7 administration in a hospital setting.^{7, 11} Lipid formulations of amphotericin B have fewer side effects and
8 are safer to use with the same cure rate.^{7, 11} Depending on the dose and formulation, the treatment
9 regimen varies from 3-5 days to eight weeks of administration on alternate days.^{6, 11} Miltefosine is the
10 first oral drug to be released for leishmaniasis and is currently available in India, Germany, and
11 Colombia.¹¹ Miltefosine is not recommended for women who are pregnant or may become pregnant
12 because it is teratogenic.^{1, 11} Miltefosine resistance has been demonstrated *in vitro*, and its long half-life
13 in the body, the twenty-eight day treatment regimen, as well as it previously being available over the
14 counter in India have led to concerns of clinical resistance.^{1, 11, 15} A recent study of 567 individuals in
15 the Bihar state of India has been performed to determine the efficacy of miltefosine since its
16 introduction in 2002.¹⁶ The six month cure rate was found to be roughly 90% and gastrointestinal
17 intolerance was encountered in 64.5% of the cases with two deaths related to drug toxicity.¹⁶ Patients
18 who did not improve with treatment were cured using amphotericin B. The authors of this study
19 concluded that the failure rate of miltefosine has increased in the ten years since its introduction for the
20 treatment of visceral leishmaniasis in India. A recent study also showed that 20% of the visceral
21 leishmaniasis patients in Nepal who were treated with miltefosine relapsed 12 months after treatment.¹⁷

22 Due to increased parasite resistance, toxicity issues, increasing failure rates of current treatments, and
23 the lack of effective clinical agents against cutaneous leishmaniasis, new drugs are needed to have an
24 effective strategy for treating leishmaniasis. Quinazolines are a class of compounds that have shown
25 potential as antileishmanials. Berman et al. reported a class of 2,4-diaminoquinazolines with EC₅₀ as
26 low as 0.04 nM against *L. major* amastigotes in human monocyte-derived macrophages (A, Figure 1.B),
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1 however the development of this compound series was abandoned due to toxicity issues.¹⁸ Bhattacharjee
2 et al. (B), Ram et al. (C), and Shakya and Gupta et al. (D and E) have also tested quinazolines as
3 antileishmanials.¹⁹⁻²² This class of compounds has been reported as being dihydrofolate reductase
4 (DHFR) inhibitors, although another mechanism of action may be involved with *Leishmania*.^{18, 23}
5 Recently, we tested a small library of structurally diverse compounds, originally designed as potential
6 anticancer probes, for antileishmanial activity in a *L. mexicana* axenic amastigote assay. Among this
7 library were *N*²,*N*⁴-disubstituted quinazolines **1** and **2** (Figure 1.B), which are structurally different from
8 quinazoline series A-E. Antileishmanial testing of quinazolines **1** and **2** against *L. mexicana* axenic
9 amastigotes revealed EC₅₀ values in the single digit micromolar range, motivating us to investigate
10 whether quinazolines structurally related to the hits **1** and **2** have potential to display potent
11 antileishmanial activity.²⁴ Herein, we report a detailed structure-activity relationship (SAR) study
12 focusing on the 2-position, the 4-position, and the quinazoline's benzenoid ring. All compounds were
13 initially examined in *L. donovani* and *L. amazonensis* intracellular amastigote assays to preselect
14 quinazoline candidates active against parasites responsible for causing visceral leishmaniasis and
15 cutaneous leishmaniasis, respectively. Promising compounds have subsequently been tested for efficacy
16 in a murine model of visceral leishmaniasis.
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44 Results and Discussion

45 **Synthetic Chemistry.** The compounds were synthesized following known procedures (Figure 2).²⁵⁻²⁷
46 Commercially available anthranilic acids (a) were cyclized with urea and the resulting quinazoline-2,4-
47 dione (b) was reacted with phosphorous oxychloride to give the 2,4-dichloroquinazoline (c).
48 Substitution with amines occurred selectively at position 4 yielding 4-amino-2-chloroquinazoline (d)
49 followed by substitution at position 2 to give the 2,4-diaminosubstituted quinazoline. In this synthetic
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sequence, only 4-amino-2-chloroquinazoline (**d**) and the final N^2, N^4 -disubstituted quinazoline-2,4-diamine have been purified and characterized.

Figure 2

***In vitro* Antileishmanial Efficacy and Cytotoxicity.** All target compounds were tested against *L. donovani* and *L. amazonensis* intracellular amastigotes to identify molecules that are broadly active against these medically important *Leishmania* species. The testing involved murine peritoneal macrophages as host cells, since compounds with potential for clinical use must be able to penetrate the infected macrophage in a human host. Antileishmanial activity was determined in an assay using transgenic parasites expressing a β -lactamase gene as outlined previously.^{28, 29} Concentration response data for each compound was fitted by a nonlinear regression model and the concentration that induces 50% inhibition was calculated as the effective concentration EC_{50} (*L. donovani* or *L. amazonensis*). Additionally, cytotoxicity against the macrophage cell line J774A.1 was determined as the effective concentration EC_{50} (J774A.1) and the selectivity index SI was calculated as the ratio of EC_{50} value for J774A.1 and the value for *L. donovani* ($SI = EC_{50}(\text{J774A.1})/EC_{50}(\text{L. donovani})$).

Structure-Activity Relationship Studies. To validate and optimize the antileishmanial activity of N^2, N^4 -disubstituted quinazoline-2,4-diamines, two compound subseries were prepared and tested. The first subseries focused mainly on the optimization of the N^2 - and N^4 -moieties (Table 1), whereas the second subseries was designed to investigate whether analogues being substituted at the quinazoline's benzenoid ring display improved antileishmanial activity (Table 2).

Starting from hit compound **2**, N^4 -furfuryl-analogues **3** and **4** were prepared in which the N^2 -*iso*-propyl group was replaced by a short trifluoroalkyl- or hydroxyalkyl-group. While the 2,2,2-trifluoroethyl-substituted quinazoline **3** was slightly less potent than compound **2**, alcohol **4** lost potency against *L. donovani* and *L. amazonensis* by a factor of 10 and >13, respectively. Testing of a small set of

1 quinazoline-2,4-diamines **5-9** with N^4 -monosubstituted or N^4 -disubstituted with alkyl groups differing in
2 size and polarity did not identify a particular structural motif improving the potency over **2**.
3 Replacement of the furfuryl and *iso*-propyl groups in **2** by two benzyl or two n-butyl groups yielded
4 quinazolines **10** and **11**, of which both analogues were more potent than reference **2**. Bis-benzyl-
5 substituted quinazoline **10** displayed EC_{50} s of 667 nM against *L. donovani* and 1.41 μ M against *L.*
6 *amazonensis*, whereas analogue **11** was approximately twofold less potent in comparison to compound
7 **10**. Consequently, a following set of six quinazolines **12-17** was designed in which one of the 2- and 4-
8 positions was substituted by one benzyl amine, while the remaining position was derivatized with an
9 aniline, n-butylamine, or methylamine. Interestingly, with the exception of the N^4 -benzyl- N^2 -methyl-
10 quinazoline-2,4-diamine **12**, all of these compounds demonstrated sub-micromolar EC_{50} s against *L.*
11 *donovani*. Among these six quinazolines tested, the N^2 -benzyl-quinazoline-2,4-diamines **15-17** appeared
12 to be at least twofold more potent against *L. donovani* than the N^4 -benzyl counterparts **12-14** suggesting
13 that an N^2 -benzyl is more favorable than an N^4 -benzyl for antileishmanial activity. This observation is
14 similar to previous results with quinazolines **2-11**. Compound **15** was the most potent compound with
15 an EC_{50} of 149 nM against *L. donovani*. For *L. amazonensis*, the set of **12-17** displayed a similar activity
16 trend, with N^2 -benzyl-quinazolines **15** and **16** being the most potent compounds with submicromolar or
17 single digit micromolar EC_{50} s.

Table 1

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 N^4 -Furfuryl- N^2 -*iso*-propyl-substituted quinazoline **2**, with EC_{50} s of 2.50 μ M against *L. donovani* and
3.71 μ M against *L. amazonensis*, was considered to be well suited as the key scaffold for a SAR study
focusing on the benzenoid moiety of the quinazoline core. In a systematic approach following the
Topliss operational scheme, compound **2** was mono-substituted with either a chlorine atom, a methyl
group, or a methoxy group in the 5-, 6-, 7-, and 8-positions to probe the benzenoid ring for steric and
electronic effects.³⁰ Overall, substitution on the benzenoid ring provided compounds with EC_{50} s in the

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single digit micromolar or sub-micromolar range against *L. donovani*. Among the chloro-substituted subseries, 5- and 6-substituted analogues **18** and **19** were less potent by a factor of two or more compared to reference **2**, while the 7- and 8-substituted quinazolines **20** and **21** displayed an activity on par with compound **2**. In contrast, the majority of the methyl- and methoxy-substituted quinazolines demonstrated a modest potency improvement over quinazoline **2**. For the methoxy-substituted quinazolines, potency increased according to substitutions in 8- < 7- < 5- \approx 6-position, whereas a potency dependence in the order of 8- \approx 6- < 7- \approx 5-positions was observed for the methyl-substituted compounds. 7-Methoxy-quinazoline **28** with an EC₅₀ of 740 nM against *L. donovani* was the most potent analogue of the compound subseries substituted at the benzenoid ring. In contrast, the testing of the benzenoid ring substituted quinazolines against *L. amazonensis* gave insight into a SAR, which differed from the one observed for *L. donovani*. 8-Methoxy-substituted quinazoline **29** was the only analogue which was marginally more potent than reference compound **2**, whereas all of the other analogues were equally or less potent than quinazoline **2**.

Table 2

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Cytotoxicity. An important quality of the quinazolines is their selectivity to inhibit parasite growth over mammalian cells. Generally, the majority of the quinazolines **2-29** exhibited EC₅₀s of 20 μ M or higher against J774A.1 (Tables 1 and 2). Quinazolines **18-20** and **22-24**, whose benzenoid ring is substituted with one chloride or one methyl in 5-, 6- or 7-position, were significantly less toxic than their reference **2**. The quinazolines displaying potent antileishmanial activity, especially the *N*²-benzyl-quinazoline-2,4-diamines **15-17**, were shown to have respectable SI values of 10 and larger indicating that they are relatively selective, nontoxic chemotypes. The SI of 100 for compound **15** was particularly enticing and led to this compound being tested *in vivo*.

Activity against Antimony-resistant *L. donovani*. The potency of selected quinazolines against antimony-resistant *Leishmania* was assessed using murine peritoneal macrophages infected with two *L.*

1 *donovani* clinical strains, BPK206/0 and BPK164/1 (Table 3).³¹ Strain BPK164/1 was isolated from a
2 bone marrow aspirate taken from a Nepalese visceral leishmaniasis patient not responding to antimonial
3 therapy; the BPK164/1 strain was originally found to be 6-fold resistant to sodium stibogluconate
4 compared to the antimony-susceptible reference strain BPK206/0.³¹ Compounds **15**, **16**, and **23**
5 displayed similar activity against the antimony-susceptible and antimony-resistant parasites. The EC₅₀s
6 against the clinical strains were 5- to 19-fold higher than their EC₅₀s against β -lactamase expressing *L.*
7 *donovani* (Table 1). There are several possible reasons for the differences in susceptibility observed
8 between the clinical *L. donovani* isolates and the β -lactamase expressing genetically modified lab strain:
9 1) the former strains are from the Indian subcontinent while the lab strain is of African origin.
10 Antileishmanial drugs display differences in efficacy for treating Indian visceral leishmaniasis
11 compared to African visceral leishmaniasis;^{32, 33} strain differences could account for such differential
12 susceptibility; 2) the clinical strains could have a greater fitness in an *in vitro* intracellular amastigote
13 model than the lab adapted strain which has been in axenic culture for many passages, leading to lower
14 compound susceptibility in the former; 3) the clinical strains were assayed using a different method
15 under different conditions compared to the β -lactamase expressing lab strain, potentially contributing to
16 distinctions in compound susceptibility. Nevertheless, these data confirm the *in vitro* activity of
17 compounds **15**, **16** and **23** against clinical strains currently circulating in endemic regions of the Indian
18 subcontinent.
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Table 3

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50 **Mechanism of Action.** We investigated the possibility that quinazolines inhibit folate metabolism in
51 the parasite since previous studies have shown DHFR inhibition with similar structures.^{18, 23} *L. donovani*
52 axenic amastigotes and J774A.1 (mouse macrophage cell line) were adapted to grow in media deficient
53 in *p*-aminobenzoic acid (PABA) and folic acid prior to susceptibility testing. Susceptibility to
54 quinazolines and miltefosine, as well as the known antifolate drugs methotrexate and pyrimethamine as
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1 positive controls, were assessed in the presence or absence of folinic acid (Figure 3).^{34,35} The presence
2 of folinic acid dramatically increased the EC₅₀ values of each of the quinazolines as well as the
3 methotrexate and pyrimethamine. For example, quinazoline **23** was 6.4 fold less potent in the presence
4 of 488 nM folinic acid than in completely deficient media. Conversely, efficacy of miltefosine was not
5 affected by addition of folinic acid to the media. Interestingly, we saw no antagonism of activity of
6 quinazolines **10**, **12**, **13**, **15**, **16** or **23** for the mammalian cell line (J774A.1) in the presence of folinic
7 acid. For example, the EC₅₀ of quinazoline **10** was 84.8 ± 2.2 μM and 84.2 ± 2.8 μM in the presence or
8 absence of 488 nM folinic acid, respectively. In contrast, the efficacy of methotrexate and
9 pyrimethamine were antagonized significantly by folinic acid in J774A.1. These results are consistent
10 with the hypothesis that the quinazolines interfere with folate metabolism in *L. donovani*.
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26 Figure 3
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31 **Structure-Property Relationship Studies.** In parallel to testing the compounds for antileishmanial
32 activity *in vitro*, a structure-property relationship (SPR) study focusing on log D, aqueous solubility, and
33 permeability has been conducted with all compounds to assess potential physicochemical liabilities
34 (Table 4). Log D_{3.0} and Log D_{7.4}, the distribution coefficient between octanol and water at pH 3.0 and
35 pH 7.4, were experimentally determined via a previously described HPLC-based method.³⁶ Solubility at
36 pH 7.4 was determined using Biomek FX lab automation workstation with pION μSOL evolution
37 software as reported previously and at pH 2.0 using an in-house HPLC assay based on UV absorption.³⁷
38 Passive transcellular permeability was assessed in a standard parallel artificial membrane permeability
39 assay (PAMPA) at pH 7.4 and pH 4.0. Generally, the aqueous solubility, the distribution coefficient Log
40 D, and the permeability of all quinazolines display a pH dependence (exemplified on Log D or
41 permeability in Table 4). The permeability is enhanced at neutral pH while the aqueous solubility and
42 Log D are better at lower pH ranges. However, since the aqueous solubility, the distribution coefficient,
43 and the permeability are within the acceptable ranges (solubility > 20 μM, Pe > 10 × 10⁻⁶ cm·s⁻¹, 1 <
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1 Log D < 4), the quinazoline compound series is considered to be suitable for the development of
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3 bioavailable antileishmanial compounds.
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11 ***In Vivo* Antileishmanial Efficacy Studies.** For the selection of the candidates for *in vivo* efficacy
12 against *L. donovani*, only compounds which displayed submicromolar *in vitro* activity against *L.*
13 *donovani* with a SI value greater than ten and a balanced combination of good physicochemical
14 properties were chosen. Among the benzyl-substituted quinazolines **12** - **17**, **15** and **16** appeared to be
15 the best candidates for *in vivo* evaluation due to their combination of potency, selectivity, and favorable
16 physicochemical properties. From the compound subseries substituted at the benzenoid ring, the
17 physicochemical properties were not discriminatory and hence compound **23** was chosen as a viable
18 candidate due to its submicromolar EC₅₀ against *L. donovani* as well as the outstanding SI value of this
19 compound (> 40).
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33 The three compounds mentioned above were dissolved in an appropriate vehicle (**15** and **23** dissolved
34 in 0.5% methyl cellulose and 0.1% Tween80 in water, **16** dissolved in 45% (w/v) (2-hydroxypropyl)-β-
35 cyclodextrin solution (HPβCD)) and administered to uninfected BALB/c mice intraperitoneally to
36 determine a tolerated dose for *in vivo* efficacy studies. While **15** was well tolerated when given at 30
37 mg/kg ip for five consecutive days, **16** (slowed breathing) and **23** (hypoactivity) were toxic to animals
38 when given at the same dosing regimen. Considering the toxicity of **16** and **23** when given at 30 mg/kg
39 i.p., lower doses of these compounds were administered in subsequent *in vivo* efficacy studies in a
40 murine visceral leishmaniasis model (Figure 4). When tested at 5 × 15 mg/kg ip, **23** inhibited liver
41 parasitemia by 37% compared to the vehicle control. However, **15** did not show significant
42 antileishmanial efficacy when tested at 5 × 30 mg/kg i.p. There was also no significant difference in the
43 parasite burden between mice in the group treated with compound **16** (5 × 10 mg/kg i.p.) and the vehicle
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1 control group ($P > 0.05$). As expected, the 45% HP β CD vehicle used to solubilize **16** itself resulted in
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3 18% inhibition of liver parasitemia, consistent with our previous report.²⁹
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8 Figure 4
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11 **Pharmacokinetics of 16 and 23 After p.o. and i.p. Administration in Mice.** Pharmacokinetic
12 studies were conducted to determine the systemic and target tissue exposures of **16** and **23**. The mean
13 plasma and tissue concentration–time profiles of **16** and **23** after p.o. and i.p. administration are shown
14 in Figures 5 and 6, respectively. The relevant pharmacokinetic outcomes are listed in Table 5. After p.o.
15 administration at 100 $\mu\text{mol/kg}$ (or approximately 30 mg/kg), both compounds were absorbed from the
16 gastrointestinal tract of mice and plasma concentration reached a C_{max} of 0.44 and 0.25 μM for **16** and
17 **23**, respectively. The systemic and tissue exposure of **16** (AUC and C_{max}) were considerably greater than
18 those of **23** (Table 4). After i.p. administration, plasma concentration reached a C_{max} of 5.2 and 2.7 μM
19 for **16** and **23**, respectively, before decreasing rapidly in the first 4 h, followed by a slower elimination
20 process until 24 h. The rapid decline in plasma concentration was likely due to extensive tissue
21 redistribution after absorption, as indicated by the high target tissue concentrations. The plasma and
22 tissue exposures after i.p. administration were markedly greater than those after p.o. administration
23 (Table 4), suggesting significant first-pass metabolism and/or partial gastrointestinal absorption after
24 oral administration of **16** and **23**. The terminal elimination half-life ranged from 5 to 20 h. Minor
25 reversible overt toxicity (hypoactivity) was observed after i.p. administration of **16**, however more
26 severe toxicity was observed, unexpectedly, after a single i.p. administration of **23**, which warranted
27 euthanization of some mice within 15 min post dose. As efficacy was evaluated at lower doses than 30
28 mg/kg to avoid toxicity and dose linearity for pharmacokinetic outcomes were unknown, it is not
29 possible to directly correlate drug exposure with antileishmanial activity in mice. In addition, it is worth
30 pointing out that the terminal half-life of **23** after i.p. administration was considerably shorter than that
31 after p.o. administration (5 h vs. 20 h; Table 5), and the liver concentration of **23** was detectable at 12
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1 and 24 h after p.o. administration, whereas it was below detection limit after i.p. administration (Figure
2 6). These observations suggest flip-flop pharmacokinetics, where the rate of gastrointestinal absorption
3 is slower than the rate of elimination due to dosage formulations (e.g. sustained-release), excipients,
4 physiological factors (e.g. intestinal mobility and pH), and/or drug characteristics.^{38, 39} The dramatic
5 decrease (~12-fold) in the permeability of **23** from neutral to acidic pH (Table 4) could contribute to the
6 slowed absorption of this compound in the upper gastrointestinal tract. In contrast, the permeability of
7 **16** decreased only 3-fold from neutral to acidic pH (Table 4). Furthermore, **23** was metabolized quickly
8 in the mouse liver microsomes ($t_{1/2} = 9.4$ min; Table 5), suggesting that it is possible that the rate of
9 elimination was greater than the rate of gastrointestinal absorption for **23**.
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24 Figure 5

25 Figure 6

26 Table 5
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33 Conclusions

34 N^2, N^4 -disubstituted quinazoline-2,4-diamines **1** and **2** were found to display antileishmanial activity
35 in the single digit micromolar range. Subsequently a total of 28 molecules have been synthesized
36 systematically by varying the substitutions in the 2-, 4-, 5-, 6-, 7-, and/or 8-positions. All quinazolines
37 have been tested with the aim to further optimize hit compounds **1** and **2** and to conduct a detailed SAR
38 study against *L. donovani* and *L. amazonensis*. The most potent activities with EC_{50} s in the
39 submicromolar range against *L. donovani* were obtained with quinazoline-2,4-diamine scaffolds bearing
40 either a N^2 -benzyl- N^4 -alkyl/phenyl or a N^2 -isopropyl- N^4 -furfuryl substituent combination. Furthermore,
41 although the benzenoid ring of the quinazoline-2,4-diamine scaffold has been identified to play a
42 secondary role for efficacy, quinazolines substituted at the 5- or 6-position with one methyl or one
43 methoxy group have also been identified to possess submicromolar EC_{50} s against *L. donovani*. Although
44 the quinazoline-2,4-diamines appear to display some cytotoxicity against the macrophage cell line
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J774A.1 yielding modest SI values, the SI values of the best antileishmanial quinazolines were larger than 10. In addition, assessment of key physicochemical properties confirmed that the quinazoline's aqueous solubility, distribution coefficient, and passive transcellular permeability were in acceptable ranges. These promising results led to efficacy testing of the lead compounds **15**, **16** and **23** in an *in vivo* murine visceral leishmaniasis model. While compounds **15** and **16** did not have activity translate from *in vitro* to *in vivo*, quinazoline **23** reduced parasitemia by 37% when 15 mg/kg/day were given via the intraperitoneal route for five consecutive days. Pharmacokinetic studies of compound **23** revealed a maximum plasma concentration that was threefold higher than the EC₅₀ and a terminal half-life of 5 hours after i.p. administration. Although a clear correlation between *in vitro* activity, *in vitro* physicochemical properties, and *in vivo* activity is not clearly observed, the potencies of frontrunner compounds **15**, **16** and **23** in conjunction with favorable physicochemical properties make *N*²,*N*⁴-disubstituted quinazoline-2,4-diamines a suitable platform for the future development of antileishmanial agents. For future compound design to be successful, optimization will not only focus on improving antileishmanial activity and key physicochemical properties, but also on improving the pharmacokinetics and SI values for the entire quinazoline compound series.

Experimental Section

Chemistry

General. All reagents and solvents were obtained from Aldrich Chemical Co. and used without further purification. Anthranilic acids were purchased from Sigma-Aldrich, Oakwood Products, Inc. or TCI America. NMR spectra were recorded at ambient temperature on a 250 MHz Bruker, 400 MHz Varian or 500 MHz Varian NMR spectrometer in the solvent indicated. All ¹H NMR experiments are reported in δ units, parts per million (ppm) downfield of TMS and were measured relative to the signals for chloroform (7.26 ppm), methanol (3.31 ppm) and dimethylsulfoxide (2.50 ppm). All ¹³C NMR spectra were reported in ppm relative to the signals for chloroform (77 ppm), methanol (49 ppm) and

1 dimethylsulfoxide (39.5 ppm) with ^1H decoupled observation. Data for ^1H NMR are reported as follows:
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3 chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, sext =
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5 sextet, sept = septet, oct = octet m = multiplet), integration and coupling constant (Hz), whereas ^{13}C
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7 NMR analyses were reported in terms of chemical shift. NMR data was analyzed by using MestReNova
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9 Software ver. 5.3.2-4936. The purity of the final compounds was determined to be $\geq 95\%$ by high
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11 pressure liquid chromatography (HPLC) using an Agilent 1100 LC/MSD-VL with electrospray
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13 ionization. Melting points were determined using a MEL-TEMP 3.0 instrument and are uncorrected.
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15 Low resolution mass spectra were performed on an Agilent 1100 LC/MSD-VL with electrospray
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17 ionization. High resolution mass spectra (HRMS) were performed on an Agilent LC/MSD TOF system
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19 G3250AA. Analytical thin layer chromatography (TLC) was performed on silica gel 60 F254 pre-coated
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21 plates (0.25 mm) from EMD Chemical Inc. and components were visualized by ultraviolet light (254
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23 nm). Reported R_f was determined for TLC. Silicycle silica gel 230-400 (particle size 40-63 μm) mesh
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25 was used for all flash column chromatography.
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31 **General Procedure A: Cyclization of Anthranilic Acids to the Corresponding Quinazoline-2,4-**
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33 **diones:** 1 equivalent of anthranilic acid and 3.5 equivalents of urea were mortar and pestled to a powder
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35 and heated to 200 $^\circ\text{C}$ in a round bottom flask open to the atmosphere. After two hours, the mixture was
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37 cooled, triturated with water, and filtered to give the product as crude. No further purification was
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39 performed.
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42 **General Procedure B: Chlorination of Quinazoline-2,4-diones to the Corresponding 2,4-**
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44 **Dichloroquinazolines:** 1 equivalent of quinazoline-2,4-dione and 1 equivalent of *N,N*-dimethylaniline
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46 were mixed in 12 equivalents of phosphorous oxychloride and the mixture refluxed under an argon
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48 atmosphere until starting material was no longer present by TLC (3-16 hours). The mixture was then
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50 cooled and added to ice in the amount of ten times the reaction volume. The solution was filtered to give
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52 crude product.
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56 **General Procedure C: Amine Substitution of 2,4-Dichloroquinazolines to Yield 4-Amino-**
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58 **substituted 2-Chloroquinazolines:** 1.1 equivalents of amine and sodium acetate were mixed with 1
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1 equivalent of 2,4-dichloroquinazoline at 0.1 M concentration in a 3 to 1 mix of tetrahydrofuran and water and heated to 65 °C. When the reaction was observed to be finished by TLC, the solution was diluted with ethyl acetate, the layers were separated, and the organic phase washed three times with an equal amount of water and dried over Na₂SO₄. The crude was then purified by either method Ca or Cb:

Purification Method Ca: the compound was recrystallized with ethanol and water, filtered, and rinsed with cold ethanol to yield pure product.

Purification Method Cb: the crude was purified by flash chromatography using hexanes and ethyl acetate.

General Procedure D: Amine Substitution of 4-Aminosubstituted-2-chloroquinazolines to Yield 2,4-Diamino-substituted Quinazolines: 1.5 equivalents of amine was mixed with 1 equivalent of 4-amino-substituted 2-chloroquinazoline at 0.2 M concentration in ethanol in a sealed tube and heated to 150 °C. When the reaction was finished as observed by TLC, the compound was purified by either method Da or Db:

Purification Method Da: compound crystallized out of the cool solution, was filtered, and rinsed with cold ethanol to yield pure product.

Purification Method Db: solvent was evaporated and the crude mixture was purified by flash chromatography using dichloromethane and methanol.

2,4-Dichloroquinazoline (1c): commercially available benzoyleneurea (0.12 mol) and *N,N*-dimethylaniline (0.12 mol) were mixed in 60 mL of phosphorous oxychloride and heated to reflux under an atmosphere of argon. After 5 hours the solution was cooled and slowly added to 300 mL of ice. Once quenching was finished, the compound was extracted with chloroform (4 x 125 mL) and purified by flash chromatography using hexanes and ethyl acetate to yield the title compound in 61% yield (14.5 g, 73 mmol). ¹H NMR (500 MHz, CDCl₃) δ 8.29 – 8.24 (m, 1H), 8.02 – 7.99 (m, 2H), 7.74 (ddd, *J* = 6.3, 5.0, 3.2, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 164.25, 155.36, 152.60, 136.41, 129.61, 128.25, 126.42,

1 122.59. Mass (ESI): $[M+H]^+$ 199, 201; found 198.9 (100%), 200.9 (64%). $R_f = 0.56$ (hexanes to ethyl
2 acetate 4:1).
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5 **2-Chloro-*N*-(furan-2-ylmethyl)quinazolin-4-amine (2d)**: 1g (5.0 mmol) of **1c** was reacted with
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7 furfurylamine and purified according to method Ca to furnish 1.21 g (4.7 mmol) of the title compound
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9 in 93% yield. ^1H NMR (250 MHz, CDCl_3) δ 7.75 – 7.55 (m, 3H), 7.34 (ddd, $J = 8.1, 5.4, 2.9$ Hz, 1H),
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11 6.48 (s, 1H), 6.30-6.22 (m, 2H), 4.77 (d, $J = 5.1$ Hz, 2H). ^{13}C NMR (63 MHz, CDCl_3) δ 160.57, 157.54,
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13 150.82, 150.24, 142.56, 133.62, 127.66, 126.32, 121.09, 113.24, 110.65, 108.66, 38.43. Mass (ESI):
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15 $[M+H]^+$ 260; found 260.1. $R_f = 0.14$ (hexanes to ethyl acetate 4:1).
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19 ***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropylquinazoline-2,4-diamine (2)**: 1.05 g (4.04 mmol) of **2d** was
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21 reacted with isopropylamine and purified according to method Da to furnish 0.47 g (1.66 mmol) of the
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23 title compound as a light yellow solid in 41% yield. ^1H NMR (500 MHz, CD_3OD) δ 7.94 (d, $J = 8.1$ Hz,
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25 1H), 7.53 – 7.46 (m, 1H), 7.44 – 7.40 (m, 1H), 7.31 (s, 1H), 7.17 – 7.09 (m, 1H), 6.38 – 6.31 (m, 2H),
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27 4.78 (s, 2H), 4.26 (sept, $J = 6.5$ Hz, 1H), 1.27 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (126 MHz, CD_3OD) δ
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29 160.14, 154.22, 151.24, 142.01, 141.97, 134.05, 123.14, 123.04, 118.38, 110.17, 109.87, 107.44, 43.29,
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31 37.65, 21.62. HRMS: m/z calcd for $\text{C}_{16}\text{H}_{19}\text{N}_4\text{O}$ $[M+H]^+$ 283.1553; found 283.1558. $R_f = 0.44$ (9:1
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33 dichloromethane to methanol). Melting point 190-192 °C.
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38 ***N*⁴-(Furan-2-ylmethyl)-*N*²-(2,2,2-trifluoroethyl)quinazoline-2,4-diamine (3)**: 0.045 g (0.17 mmol)
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40 of **2d** was reacted with 2,2,2-trifluoroethylamine and purified according to method Da to furnish 0.041 g
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42 (0.13 mmol) of the title compound as a white crystalline solid in 76% yield. ^1H NMR (500 MHz,
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44 CD_3OD) δ 8.17 (d, $J = 8.2$, 1H), 7.79 (dd, $J = 8.4, 7.3$, 1H), 7.51 (d, $J = 7.1$, 1H), 7.48 – 7.45 (m, 1H),
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46 7.44 (dd, $J = 7.3, 1.0$, 1H), 6.39 (m, 2H), 4.89 (s, 2H), 4.38 (q, $J = 9.0$, 2H). ^{13}C NMR (126 MHz,
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48 CD_3OD) δ 160.64, 153.51, 150.23, 142.31, 138.72, 135.45, 125.29, 124.36 (q, $J = 278.71$), 123.64,
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50 116.87, 110.16, 110.05, 107.89, 41.73 (q, $J = 35.03$), 38.11. HRMS: m/z calcd for $\text{C}_{15}\text{H}_{14}\text{F}_3\text{N}_4\text{O}$
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52 $[M+H]^+$ 323.1114; found 323.1122. $R_f = 0.14$ (dichloromethane). Decomposed at 225 °C.
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57 **2-(4-(Furan-2-ylmethylamino)quinazolin-2-ylamino)ethanol (4)**: 0.12 g (0.46 mmol) of **2d** was
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59 reacted with 2-aminoethanol and purified according to method Da to furnish 0.048 g (0.17 mmol) of the
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title compound in 37% yield. ^1H NMR (500 MHz, CD_3OD) δ 8.11 (d, $J = 7.6$ Hz, 1H), 7.76 (ddd, $J = 8.4, 7.3, 1.3$ Hz, 1H), 7.46 (d, $J = 0.8$ Hz, 1H), 7.40 (t, $J = 7.5$ Hz, 2H), 6.41 (d, $J = 2.9$ Hz, 1H), 6.40 – 6.36 (m, 1H), 3.77 (s, 2H), 3.70 (s, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.87, 159.63, 151.51, 150.07, 142.05, 132.90, 124.21, 121.55, 121.39, 110.88, 110.45, 107.64, 63.77, 44.88, 37.89. HRMS: m/z calcd for $\text{C}_{15}\text{H}_{17}\text{N}_4\text{O}_2$ $[\text{M}+\text{H}]^+$ 285.1346; found 285.1350. $R_f = 0.24$ (dichloromethane to methanol 9:1). Melting point 219-221 $^\circ\text{C}$.

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***N*-2,2,2-Trifluoroethyl-2-chloroquinazolin-4-amine (5d)**: 0.050 g (0.25 mmol) of **1c** was reacted with 2,2,2-trifluoroethylamine and purified according to method Cb to furnish 0.017 g (0.065 mmol) of the title compound in 26% yield. ^1H NMR (400 MHz, CD_3OD) δ 8.16 (d, $J = 7.2$ Hz, 1H), 7.82 (t, $J = 7.4$ Hz, 1H), 7.68 (d, $J = 7.5$ Hz, 1H), 7.58 (t, $J = 7.7$ Hz, 1H), 4.39 (q, $J = 8.9$ Hz, 2H). Mass (ESI): $[\text{M}+\text{H}]^+$ 235, 237; found 235.0 (100%), 237.1 (33%). $R_f = 0.11$ (hexanes to ethyl acetate 4:1).

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***N*²-Isopropyl-*N*⁴-(2,2,2-trifluoroethyl)quinazoline-2,4-diamine (5)**: 0.017 g (0.065 mmol) of **5d** was reacted with isopropylamine and purified according to method Db to furnish 0.005 g (0.018 mmol) of the title compound as a white crystalline solid in 28% yield. ^1H NMR (500 MHz, CD_3OD) δ 7.91 (dd, $J = 8.2, 1.1$, 1H), 7.60 (ddd, $J = 8.4, 7.0, 1.4$, 1H), 7.38 (d, $J = 8.4$, 1H), 7.17 (ddd, $J = 8.1, 7.0, 1.0$, 1H), 4.38 (q, $J = 9.3$, 2H), 4.24 (sept, $J = 6.5$, 1H), 1.26 (d, $J = 6.5$, 6H). ^{13}C NMR (126 MHz, CD_3OD) δ 160.87, 157.71, 149.57, 133.28, 124.83 (q, $J = 278.55$), 122.38, 122.15, 121.53, 110.32, 42.57, 40.98 (q, $J = 34.34$), 21.71. HRMS: m/z calcd for $\text{C}_{13}\text{H}_{16}\text{F}_3\text{N}_4$ $[\text{M}+\text{H}]^+$ 285.1322; found 285.1325. Melting point 103-108 $^\circ\text{C}$.

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2-Chloro-*N*-(2-(methylthio)ethyl)quinazolin-4-amine (6d): 0.20 g (1.0 mmol) of **1c** was reacted with 2-(methylthio)ethylamine and purified according to method Cb to furnish 0.15 g (0.59 mmol) of the title compound in 59% yield. ^1H NMR (250 MHz, CD_3OD) δ 7.93 (dd, $J = 8.3, 0.8$ Hz, 1H), 7.65 (ddd, $J = 8.4, 7.0, 1.4$ Hz, 1H), 7.50 – 7.44 (m, 1H), 7.38 (ddd, $J = 8.3, 7.0, 1.3$ Hz, 1H), 3.70 (dd, $J = 7.5, 6.6$ Hz, 2H), 2.75 – 2.65 (m, 2H), 2.08 (s, 3H). Mass (ESI): $[\text{M}+\text{H}]^+$ 254, 256; found 254.0 (100%), 256.0 (33%). $R_f = 0.17$ (hexanes to ethyl acetate 4:1).

1 ***N*²-Isopropyl-*N*⁴-(2-(methylthio)ethyl)quinazoline-2,4-diamine (6)**: 0.12 g (0.47 mmol) of **6d** was
2 reacted with isopropylamine and purified according to method Db to furnish 0.056 g (0.20 mmol) of the
3 title compound as a white solid in 43% yield. ¹H NMR (500 MHz, CD₃OD) δ 7.81 (d, *J* = 8.1 Hz, 1H),
4 7.47 (t, *J* = 7.5 Hz, 1H), 7.28 (d, *J* = 8.2 Hz, 1H), 7.06 (t, *J* = 7.6 Hz, 1H), 4.20 (sept, *J* = 6.5 Hz, 1H),
5 3.76 – 3.71 (m, 2H), 2.79 – 2.74 (m, 2H), 2.11 (s, 3H), 1.22 (d, *J* = 6.5 Hz, 6H). ¹³C NMR (126 MHz,
6 CD₃OD) δ 160.39, 157.49, 148.19, 132.93, 122.28, 121.64, 121.48, 110.56, 42.62, 40.15, 32.36, 22.03,
7 14.11. HRMS: *m/z* calcd for C₁₄H₂₁N₄S [M+H]⁺ 277.1481; found 277.1488. *R*_f = 0.52 (dichloromethane
8 to methanol 9:1). Melting point 87-90 °C.
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19 **Methyl-2-(2-chloroquinazolin-4-ylamino)acetate (7d)**: 0.198 g (0.99 mmol) of **1c** was reacted with
20 glycine methyl ester and purified according to method Ca to furnish 0.18 g (0.72 mmol) of the title
21 compound in 73% yield. ¹H NMR (400 MHz, CD₃OD) δ 7.85 (d, *J* = 8.1 Hz, 1H), 7.62 (dd, *J* = 11.3, 4.1
22 Hz, 1H), 7.45 (d, *J* = 8.3 Hz, 1H), 7.33 (t, *J* = 7.6 Hz, 1H), 4.26 (s, 2H), 3.71 (s, 3H). ¹³C NMR (101
23 MHz, CD₃OD) δ 170.77, 161.67, 157.12, 150.04, 133.76, 126.38, 125.93, 122.25, 113.24, 51.72, 42.35.
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33 **Ethyl-2-(2-(isopropylamino)quinazolin-4-ylamino)acetate (7)**: 0.12 g (0.47 mmol) of **7d** was
34 reacted with isopropylamine and purified according to method Db to furnish 0.060 g (0.21 mmol) of the
35 title compound as a white crystalline solid in 44% yield. A transesterification reaction occurred leading
36 to the ethylester product. ¹H NMR (400 MHz, CD₃OD) δ 7.84 (d, *J* = 8.2 Hz, 1H), 7.57 – 7.51 (t, *J* = 7.8
37 Hz, 1H), 7.32 (d, *J* = 8.2 Hz, 1H), 7.10 (t, *J* = 7.8 Hz, 1H), 4.26 – 4.14 (m, 5H), 1.25 (t, *J* = 7.1 Hz, 3H),
38 1.21 (d, *J* = 6.5 Hz, 6H). ¹³C NMR (101 MHz, CD₃OD) δ 172.23, 162.13, 159.29, 150.86, 134.31,
39 123.71, 123.57, 122.61, 111.84, 62.22, 43.87, 43.66, 23.25, 14.52. HRMS: *m/z* calcd for C₁₅H₂₁N₄O₂
40 [M+H]⁺ 289.1659; found 289.1668. *R*_f = 0.34 (dichloromethane to methanol 9:1).
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52 **2-Chloro-*N*-cyclohexylquinazolin-4-amine (8d)**: 0.16 g (0.80 mmol) of **1c** was reacted with
53 cyclohexylamine and purified according to method Cb to furnish 0.15 g (0.55 mmol) of the title
54 compound in 69% yield. ¹H NMR (250 MHz, CD₃OD) δ 8.05 – 7.99 (m, 1H), 7.60 (ddd, *J* = 8.4, 7.0,
55 1.4 Hz, 1H), 7.46 – 7.41 (m, 1H), 7.33 (ddd, *J* = 8.3, 7.0, 1.3 Hz, 1H), 4.17 – 4.01 (m, 1H), 1.97-1.84
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(m, 2H), 1.70 (d, $J = 2.6$ Hz, 2H), 1.65 – 1.52 (m, 1H), 1.38 – 1.25 (m, 4H), 1.20 – 1.00 (m, 1H). ^{13}C NMR (63 MHz, CD_3OD) δ 161.94, 159.05, 151.35, 134.67, 127.25, 126.90, 123.83, 114.81, 51.68, 33.32, 26.71, 26.48. Mass (ESI): $[\text{M}+\text{H}]^+$ 262, 264; found 262.1 (100%), 264.1 (33%). $R_f = 0.43$ (hexanes to ethyl acetate 2:1).

***N*⁴-Cyclohexyl-*N*²-isopropylquinazoline-2,4-diamine (8):** 0.10 g (0.40 mmol) of **8d** was reacted with isopropylamine and purified according to method Db to furnish 0.060 g (0.21 mmol) of the title compound as a white crystalline solid in 53% yield. ^1H NMR (400 MHz, CD_3OD) δ 7.95 (d, $J = 8.0$ Hz, 1H), 7.55 (t, $J = 7.5$ Hz, 1H), 7.31 (d, $J = 8.3$ Hz, 1H), 7.14 (t, $J = 7.6$ Hz, 1H), 4.20 (sept, $J = 6.5$ Hz, 2H), 2.04 (d, $J = 2.2$ Hz, 2H), 1.83 (d, $J = 4.5$ Hz, 2H), 1.70 (d, $J = 12.4$ Hz, 1H), 1.41 (m, 5H), 1.25 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101 MHz, CD_3OD) δ 159.82, 157.34, 147.57, 133.16, 122.65, 121.83, 121.25, 110.84, 50.46, 42.93, 32.26, 25.37, 21.93. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{25}\text{N}_4$ $[\text{M}+\text{H}]^+$ 285.2074; found 285.2077. $R_f = 0.38$ (dichloromethane to methanol 9:1). Decomposed at 345 °C.

***N,N*-Dimethyl-2-chloroquinazolin-4-amine (9d):** 0.054 g (0.27 mmol) of **1c** was reacted with dimethylamine and purified according to method Ca to furnish 0.037 g (0.18 mmol) of the title compound in 69% yield. ^1H NMR (400 MHz, CD_3OD) δ 8.06 (d, $J = 8.5$ Hz, 1H), 7.68 – 7.62 (m, 1H), 7.52 (d, $J = 8.4$ Hz, 1H), 7.39 – 7.33 (m, 1H), 3.35 (s, 6H). ^{13}C NMR (101 MHz, CD_3OD) δ 163.76, 156.03, 152.57, 133.10, 126.41, 125.89, 124.83, 114.07, 41.20. $R_f = 0.35$ (hexanes to ethyl acetate 4:1).

***N*²-Isopropyl-*N*⁴,*N*⁴-dimethylquinazoline-2,4-diamine (9):** 0.030 g (0.14 mmol) of **9d** was reacted with isopropylamine and purified according to method Db to furnish 0.015 g (0.065 mmol) of the title compound as a white solid in 46% yield. ^1H NMR (400 MHz, CD_3OD) δ 8.11 (d, $J = 8.3$ Hz, 1H), 7.69 (t, $J = 8.2$ Hz, 1H), 7.44 (d, $J = 8.2$ Hz, 1H), 7.31 (t, $J = 8.2$ Hz, 1H), 4.31 – 4.19 (m, 1H), 3.48 (s, 6H), 1.29 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101 MHz, CD_3OD) δ 162.97, 152.20, 143.16, 134.25, 127.59, 122.97, 117.82, 110.60, 43.69, 41.45, 21.46. HRMS: m/z calcd for $\text{C}_{13}\text{H}_{19}\text{N}_4$ $[\text{M}+\text{H}]^+$ 231.1604; found 231.1608. $R_f = 0.33$ (dichloromethane to methanol 9:1). Melting point 110-115 °C.

***N*-Benzyl-2-chloroquinazolin-4-amine (10d):** 0.30g (1.5 mmol) of **1c** was reacted with benzylamine and purified according to method Ca to furnish 0.345 g (1.28 mmol) of the title compound in 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.70 (ddd, *J* = 11.3, 8.4, 4.9 Hz, 3H), 7.48 – 7.26 (m, 5H), 6.27 (s, 1H), 4.83 (d, *J* = 5.3 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 160.93, 157.93, 151.09, 137.48, 133.76, 129.11, 128.51, 128.21, 128.00, 126.46, 121.07, 113.38, 45.94. *R_f* = 0.21 (hexanes to ethyl acetate 4:1).

***N*²,*N*⁴-Dibenzylquinazoline-2,4-diamine (10):** 0.10 g (0.37 mmol) of **10d** was reacted with benzylamine and purified according to method Da to furnish 0.101 g (0.297 mmol) of the title compound as a white crystalline solid in 80% yield. ¹H NMR (500 MHz, DMSO-d₆) δ 10.46 (s, NH), 8.64 (s, NH), 8.45 (d, *J* = 8.0 Hz, 1H), 7.74 (dt, *J* = 7.8, 1.5 Hz, 1H), 7.44 (d, *J* = 8.5 Hz, 1H), 7.36 (dt, *J* = 7.8, 1.3 Hz, 1H), 7.36-7.12 (m, 10H), 4.75 (d, *J* = 5.5 Hz, 2H), 4.63 (d, *J* = 4.8 Hz, 2H). ¹³C NMR (126 MHz, DMSO-d₆) δ 160.26, 153.49, 139.44, 138.85, 138.29, 135.67, 129.93, 128.85, 128.80, 128.24, 128.12, 127.72, 127.57, 124.93, 124.64, 117.19, 110.22, 110.12, 44.86, 44.35. HRMS: *m/z* calcd for C₂₂H₂₁N₄ [M+H]⁺ 341.1761; found 341.1751. *R_f* = 0.45 (dichloromethane to methanol 9:1). Melting point 237-240 °C.

***N*-Butyl-2-chloroquinazolin-4-amine (11d):** 0.48 g (2.4 mmol) of **1c** was reacted with *n*-butylamine and purified according to method Ca to furnish 0.49 g (2.1 mmol) of the title compound in 86% yield. ¹H NMR (250 MHz, CDCl₃) δ 7.77 – 7.53 (m, 3H), 7.44 – 7.31 (m, 1H), 5.82 (s, NH), 3.61 (td, *J* = 7.1, 5.6, 2H), 1.64 (dt, *J* = 14.7, 7.2, 2H), 1.40 (td, *J* = 14.5, 7.2, 2H), 0.92 (t, *J* = 7.3, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 161.14, 150.99, 133.58, 128.10, 126.29, 121.19, 120.79, 113.42, 41.57, 31.47, 20.35, 14.02. Mass (ESI): [M+H]⁺ 236, 238; found 236.0 (100%), 238.0 (34%). *R_f* = 0.21 (hexanes to ethyl acetate 4:1). Melting point 103-105°C.

***N*²,*N*⁴-Dibutylquinazoline-2,4-diamine (11):** 0.080 g (0.34 mmol) of **11d** was reacted with *n*-butylamine and purified according to method Da to furnish 0.031 g (0.114 mmol) of the title compound as a white solid in 34% yield. ¹H NMR (500 MHz, DMSO-d₆) δ 9.78 (s, NH), 8.36 (d, *J* = 8.5 Hz, 1H), 8.14 (s, NH), 7.76 (t, *J* = 8.0 Hz, 1H), 7.42 (d, *J* = 6.6 Hz, 1H), 7.37 (t, *J* = 6.2 Hz, 1H), 3.58 (m, 2H), 3.45 (m, 2H), 1.65 (p, *J* = 7.5 Hz, 2H), 1.57 (p, *J* = 7.5, 2H), 1.36 (sept, *J* = 7.3 Hz, 4H), 0.92 (td, *J* = 7.3, 4.7 Hz, 6H). ¹³C NMR (101 MHz, DMSO-d₆) δ 160.10, 153.50, 139.26, 135.43, 124.83, 124.36,

116.97, 110.02, 41.37, 40.72, 31.49, 30.69, 20.16, 19.93, 14.12, 14.07. HRMS: m/z calcd for $C_{16}H_{25}N_4$ $[M+H]^+$ 273.2074; found 273.2072. Melting point 151-154 °C.

***N*⁴-Benzyl-*N*²-methylquinazoline-2,4-diamine (12):** 0.088 g (0.326 mmol) of **10d** was reacted with 2.0 M methylamine in methanol and purified according to method Da to furnish 0.085 g (0.321 mmol) of the title compound as a cream colored solid in 98% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.29 (s, 1H), 7.42 (d, *J* = 7.7 Hz, 1H), 7.37 (d, *J* = 8.0 Hz, 2H), 7.27 (d, *J* = 9.0 Hz, 1H), 7.20 – 7.11 (m, 3H), 7.06 (t, *J* = 7.6 Hz, 1H), 4.80 (s, 2H), 2.93 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 160.11, 156.03, 142.69, 138.07, 133.96, 128.35, 127.94, 127.26, 123.74, 123.10, 119.20, 110.28, 45.06, 28.01. HRMS: m/z calcd for $C_{16}H_{17}N_4$ $[M+H]^+$ 265.1448; found 265.1454. *R*_f = 0.47 (dichloromethane to methanol 9:1).

***N*⁴-Benzyl-*N*²-butylquinazoline-2,4-diamine (13):** 0.10 g (0.37 mmol) of **10d** was reacted with *n*-butylamine and purified according to method Da to furnish 0.047 g (0.15 mmol) of the title compound as a white solid in 41% yield. ¹H NMR (400 MHz, CDCl₃) δ 9.54 (s, 1H), 8.43 (d, *J* = 8.1 Hz, 1H), 7.82 (s, 1H), 7.44 – 7.37 (m, 3H), 7.25 – 7.11 (m, 5H), 4.83 (d, *J* = 5.7 Hz, 2H), 3.39 (q, *J* = 6.8 Hz, 2H), 1.49 (p, *J* = 7.3 Hz, 2H), 1.31 (sext, *J* = 7.3 Hz, 2H), 0.85 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 160.08, 153.46, 137.43, 134.65, 128.41, 127.83, 127.40, 124.41, 124.12, 116.62, 109.99, 109.68, 45.31, 41.02, 31.30, 19.95, 13.66. HRMS: m/z calcd for $C_{19}H_{23}N_4$ $[M+H]^+$ 307.1917; found 307.1926. *R*_f = 0.33 (dichloromethane to methanol 9:1). Melting point 210-212°C.

***N*⁴-Benzyl-*N*²-phenylquinazoline-2,4-diamine (14):** 0.10 g (0.37 mmol) of **10d** was reacted with aniline and purified according to method Da to furnish 0.117 g (0.358 mmol) of the title compound as a white crystalline solid in 97% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.46 (s, 2H), 8.46 (d, *J* = 8.3 Hz, 1H), 7.84 (ddd, *J* = 8.4, 7.3, 1.1 Hz, 1H), 7.56 (d, *J* = 8.3 Hz, 1H), 7.48 (td, *J* = 7.8, 1.0 Hz, 1H), 7.45 – 7.41 (m, 2H), 7.37 – 7.29 (m, 6H), 7.29 – 7.22 (m, 1H), 7.18 (t, *J* = 7.4 Hz, 1H), 4.76 (d, *J* = 5.9 Hz, 2H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 160.71, 151.94, 139.36, 138.02, 137.13, 135.93, 129.41, 128.89, 127.78, 127.67, 125.45, 125.31, 124.81, 122.71, 117.94, 110.70, 45.14. HRMS: m/z calcd for $C_{21}H_{19}N_4$ $[M+H]^+$ 327.1604; found 327.1613. *R*_f = 0.56 (dichloromethane to methanol 9:1). Decomposed at 310°C.

***N*-Methyl-2-chloroquinazolin-4-amine (15d):** 1.0 g (5.0 mmol) of **1c** was reacted with methylamine and purified according to method Ca to furnish 0.89 g (4.6 mmol) of the title compound in 92% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.74-7.66 (m, 3H), 7.41 (t, *J* = 6.9 Hz, 1H), 6.28 (s, NH), 3.20 (d, *J* = 4.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 161.82, 158.02, 150.80, 133.64, 127.90, 126.41, 121.02, 113.60, 28.74. Mass (ESI): [M+H]⁺ 194; found 194. *R*_f = 0.12 (hexanes to ethyl acetate 4:1).

***N*²-Benzyl-*N*⁴-methylquinazoline-2,4-diamine (15):** 0.287 g (1.48 mmol) of **15d** was reacted with benzylamine and purified according to method Da to furnish 0.193 g (0.730 mmol) of the title compound as a white crystalline solid in 49% yield. ¹H NMR (250 MHz, CD₃OD) δ 8.06 (d, *J* = 8.1 Hz, 1H), 7.73 (t, *J* = 8.4 Hz, 1H), 7.50 – 7.20 (m, 7H), 4.75 (s, 2H), 3.14 (s, 3H). ¹³C NMR (63 MHz, CD₃OD) δ 162.17, 154.54, 140.08, 139.51, 136.27, 129.70, 128.69, 128.59, 125.92, 124.85, 117.86, 111.30, 45.91, 29.04. HRMS: *m/z* calcd for C₁₆H₁₇N₄ [M+H]⁺ 265.1453; found 265.1457. *R*_f = 0.31 (dichloromethane to methanol 9:1). Decomposed at 240°C.

***N*²-Benzyl-*N*⁴-butylquinazoline-2,4-diamine (16):** 0.48 g (2.0 mmol) of **11d** was reacted with benzylamine and purified according to method Da to furnish 0.40 g (1.3 mmol) of the title compound as a white crystalline solid in 65% yield. ¹H NMR (500 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 8.64 (s, 1H), 8.37 (d, *J* = 7.7 Hz, 1H), 7.76 (ddd, *J* = 8.4, 7.3, 1.1 Hz, 1H), 7.46 (d, *J* = 7.9 Hz, 1H), 7.4-7.3 (m, 5H), 7.24 (t, *J* = 7.1 Hz, 1H), 4.66 (d, *J* = 4.4 Hz, 2H), 3.49 (d, *J* = 6.1 Hz, 2H), 1.61-1.43 (m, 2H), 1.33-1.17 (m, 2H), 0.80 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 160.11, 153.48, 139.24, 139.11, 135.49, 128.83, 127.61, 127.51, 124.83, 124.54, 117.14, 110.15, 44.46, 41.46, 30.62, 20.11, 14.09. HRMS: *m/z* calcd for C₁₉H₂₃N₄ [M+H]⁺ 307.1923; found 307.1921. *R*_f = 0.48 (dichloromethane to methanol 9:1). Melting point 194-196°C.

2-Chloro-*N*-phenyl-quinazolin-4-amine (17d): 0.30 g (1.5 mmol) of **1c** was reacted with aniline and purified according to method Ca to furnish 0.36 g (1.4 mmol) of the title compound as a white solid in 93% yield. ¹H NMR (400 MHz, CD₃OD) δ 8.33 (d, *J* = 8.3 Hz, 1H), 7.80 (ddd, *J* = 8.3, 7.0, 1.2 Hz, 1H), 7.78 – 7.73 (m, 2H), 7.64 (dd, *J* = 8.4, 0.9 Hz, 1H), 7.56 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 7.40 – 7.34 (m,

2H), 7.17 (t, $J = 7.4$ Hz, 1H). ^{13}C NMR (101 MHz, CD_3OD) δ 159.61, 156.96, 150.58, 137.99, 133.70, 128.28, 126.50, 126.00, 124.67, 122.55, 122.40, 113.65. $R_f = 0.19$ (hexanes to ethyl acetate 4:1).

***N*²-Benzyl-*N*⁴-phenylquinazoline-2,4-diamine (17):** 0.10 g (0.39 mmol) of **17d** was reacted with benzylamine and purified according to method Da to furnish 0.010 g (0.031 mmol) of the title compound as a white crystalline solid in 10% yield. ^1H NMR (500 MHz, DMSO-d_6) δ 9.56 (s, 1H), 9.13 (s, 1H), 8.37 (d, $J = 7.5$ Hz, 1H), 7.92 (d, $J = 7.5$ Hz, 2H), 7.86 (d, $J = 8.1$ Hz, 2H), 7.66 (ddd, $J = 8.3, 6.9, 1.3$ Hz, 1H), 7.48 (d, $J = 8.4$ Hz, 1H), 7.39 (dd, $J = 8.4, 7.4$ Hz, 2H), 7.27 (ddd, $J = 8.1, 7.0, 1.2$ Hz, 1H), 7.22 (dd, $J = 8.4, 7.4$ Hz, 2H), 7.13 (t, $J = 7.4$ Hz, 1H), 6.89 (t, $J = 7.3$ Hz, 1H), 1.89 (s, 2H). ^{13}C NMR (101 MHz, DMSO-d_6) δ 159.01, 157.09, 152.33, 141.80, 140.10, 133.64, 129.12, 128.92, 126.26, 124.14, 123.77, 123.10, 122.52, 121.45, 119.58, 112.43, 21.76. HRMS: m/z calcd for $\text{C}_{21}\text{H}_{19}\text{N}_4$ $[\text{M}+\text{H}]^+$ 327.1604; found 327.1614. $R_f = 0.53$ (dichloromethane to methanol 9:1).

2,4,8-Trichloroquinazoline (18c): 0.5 g (2.9 mmol) of commercially available 2-amino-3-chlorobenzoic acid was reacted according to general procedure A to give crude **18b**. Without further purification, **18b** was reacted according to general procedure B to give 0.15 g (0.6 mmol) of the crude title compound (20% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 233, 235; found 232.9 (100%), 235.0 (86%).

2,8-Dichloro-*N*⁴-(furan-2-ylmethyl)-quinazolin-4-amine (18d): 0.10 g (0.43 mmol) of **18c** was reacted with furfurylamine according to general procedure Cb to give 0.11 g (0.37 mmol) of **18d** in 87% yield. ^1H NMR (250 MHz, CD_3OD) δ 7.89 (dd, $J = 8.3, 1.2$ Hz, 1H), 7.69 (dd, $J = 7.7, 1.2$ Hz, 1H), 7.33 (dd, $J = 1.7, 1.0$ Hz, 1H), 7.30 – 7.21 (m, 1H), 6.28 – 6.21 (m, 2H), 4.67 (s, 2H). ^{13}C NMR (63 MHz, CD_3OD) δ 162.61, 159.88, 152.48, 148.51, 143.44, 134.77, 132.04, 127.17, 122.63, 116.31, 111.47, 109.13, 39.00. Mass (ESI): $[\text{M}+\text{H}]^+$ 294, 296; found 294.0 (100%), 296.0 (64%).

8-Chloro-*N*⁴-(furan-2-ylmethyl)-*N*²-isopropylquinazoline-2,4-diamine (18): 0.047 g (0.16 mmol) of **18d** was reacted with isopropylamine according to general procedure Db to give 0.030 g (0.095 mmol) of the title compound as a yellow solid in 59% yield. ^1H NMR (400 MHz, DMSO-d_6) δ 8.38 (s, NH), 7.97 (d, $J = 8.0$ Hz, 1H), 7.63 (d, $J = 7.5$ Hz, 1H), 7.56 (s, 1H), 6.95 (t, $J = 7.8$ Hz, 1H), 6.65 (s, NH), 6.42 – 6.36 (m, 1H), 6.32 (s, 1H), 4.70 (d, $J = 5.5$ Hz, 2H), 4.28-4.14 (m, 1H), 1.17 (d, $J = 5.5$ Hz,

6H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.87, 158.98, 151.43, 148.83, 142.16, 132.55, 128.98, 119.93, 119.78, 111.79, 110.46, 107.66, 42.93, 38.15, 22.99. HRMS: m/z calcd for $\text{C}_{16}\text{H}_{17}\text{ClN}_4\text{O}$ $[\text{M}+\text{H}]^+$ 317.1164; found 317.1173. Decomposed at 150 °C.

2,4,7-Trichloroquinazoline (19c): 0.75 g (4.4 mmol) of commercially available 2-amino-4-chlorobenzoic acid was reacted according to general procedure A to give crude **19b**. Without further purification, **19b** was reacted according to general procedure B to give 0.64 g (2.7 mmol) of the crude title compound (61% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 233, 235; found 232.9 (100%), 235.0 (94%). R_f = 0.68 (hexanes to ethyl acetate 4:1).

2,7-Dichloro- N^4 -(furan-2-ylmethyl)-quinazolin-4-amine (19d): 0.30 g (1.3 mmol) of **19c** was reacted with furfurylamine according to general procedure Cb to give 0.35 g (1.2 mmol) of **19d** in 92% yield. ^1H NMR (250 MHz, CD_3OD) δ 8.13 – 8.06 (m, 1H), 7.62 – 7.58 (m, 1H), 7.49 (d, J = 2.1 Hz, 1H), 7.47 – 7.42 (m, 1H), 6.37 – 6.35 (m, 2H), 4.80 (s, 2H). ^{13}C NMR (63 MHz, CD_3OD) δ 163.24, 160.01, 156.93, 152.44, 143.49, 140.90, 128.01, 126.34, 125.69, 113.46, 111.47, 109.11, 38.87. Mass (ESI): $[\text{M}+\text{H}]^+$ 294, 296; found 294.0 (100%), 296.0 (63%). R_f = 0.27 (hexanes to ethyl acetate 4:1).

7-Chloro- N^4 -(furan-2-ylmethyl)- N^2 -isopropylquinazoline-2,4-diamine (19): 0.113 g (0.38 mmol) of **19d** was reacted with isopropylamine according to general procedure Db to give 0.085 g (0.27 mmol) of the title compound in 71% yield. ^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 8.37 (s, NH), 8.03 (d, J = 8.7 Hz, 1H), 7.56 – 7.55 (m, 1H), 7.23 (s, 1H), 7.02 (dd, J = 8.7, 1.8 Hz, 1H), 6.57 (s, NH), 6.46 – 6.37 (m, 1H), 6.34 (s, 1H), 4.70 (d, J = 5.6 Hz, 2H), 4.16 (oct, J = 6.6 Hz, 1H), 1.15 (d, J = 6.5 Hz, 6H). ^{13}C NMR (126 MHz, $\text{DMSO}-d_6$) δ 159.87, 153.82, 152.94, 142.36, 142.32, 137.40, 125.34, 123.78, 120.26, 110.95, 110.90, 107.56, 42.22, 37.32, 23.15. HRMS: m/z calcd for $\text{C}_{16}\text{H}_{18}\text{ClN}_4\text{O}$ $[\text{M}+\text{H}]^+$ 317.1164; found 317.1161. R_f = 0.63 (dichloromethane to methanol 9:1).

2,4,6-Trichloroquinazoline (20c): 1 g (5.8 mmol) of commercially available 2-amino-5-chlorobenzoic acid was reacted according to general procedure A to give crude **20b**. Without further purification, **20b** was reacted according to general procedure B to give 0.55 g (2.7 mmol) of the crude title compound (47% over two steps). ^1H NMR (250 MHz, CDCl_3) δ 8.17 (d, J = 8.9 Hz, 1H), 7.94 (d, J

= 2.0 Hz, 1H), 7.66 (dd, $J = 8.9, 2.0$ Hz, 1H). ^{13}C NMR (63 MHz, CDCl_3) δ 163.85, 156.31, 152.68, 142.97, 130.45, 127.37, 127.07, 120.77. Mass (ESI): $[\text{M}+\text{H}]^+$ 233, 235; found 232.9 (100%), 235.0 (98%). $R_f = 0.68$ (hexanes to ethyl acetate 4:1).

2,6-Dichloro- N^4 -(furan-2-ylmethyl)-quinazolin-4-amine (20d): 0.30 g (1.3 mmol) of **20c** was reacted with furfurylamine according to general procedure Cb to give 0.072 g (0.24 mmol) of **20d** in 18% yield. ^1H NMR (400 MHz, DMSO-d_6) δ 9.23 (t, $J = 5.1$ Hz, NH), 8.44 (d, $J = 2.1$ Hz, 1H), 7.78 (dd, $J = 8.9, 2.1$ Hz, 1H), 7.65 – 7.54 (m, 2H), 6.42 – 6.34 (m, 2H), 4.69 (d, $J = 5.4$ Hz, 2H). ^{13}C NMR (63 MHz, CDCl_3) δ 160.42, 158.82, 151.67, 139.79, 136.95, 128.95, 128.39, 128.14, 127.04, 126.91, 122.42, 111.58, 45.86. Mass (ESI): $[\text{M}+\text{H}]^+$ 294, 296; found 294.0 (100%), 296.0 (67%). $R_f = 0.27$ (hexanes to ethyl acetate 4:1).

6-Chloro- N^4 -(furan-2-ylmethyl)- N^2 -isopropylquinazoline-2,4-diamine (20): 0.041 g (0.14 mmol) of **20d** was reacted with isopropylamine according to general procedure Db to give 0.026 g (0.082 mmol) of the title compound in 59% yield. ^1H NMR (400 MHz, DMSO-d_6) δ 8.36 (s, NH), 8.15 (d, $J = 2.3$ Hz, 1H), 7.57 (d, $J = 0.7$ Hz, 1H), 7.46 (dd, $J = 8.9, 2.3$ Hz, 1H), 7.23 (d, $J = 8.4$ Hz, 1H), 6.56 (s, NH), 6.39 (dd, $J = 3.0, 1.9$ Hz, 1H), 6.34 (s, 1H), 4.68 (d, $J = 5.4$ Hz, 2H), 4.14 (oct, $J = 6.6$ Hz, 1H), 1.14 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.22, 158.96, 151.64, 150.51, 142.01, 133.02, 126.36, 125.48, 120.96, 111.58, 110.43, 107.59, 42.72, 37.84, 23.16. HRMS: m/z calcd for $\text{C}_{16}\text{H}_{17}\text{ClN}_4\text{O}$ $[\text{M}+\text{H}]^+$ 317.1164; found 317.1165.

2,4,5-Trichloroquinazoline (21c): 0.36 g (2.1 mmol) of commercially available 2-amino-6-chlorobenzoic acid was reacted according to general procedure A to give crude **21b**. Without further purification, **21b** was reacted according to general procedure B to give 0.12 g (0.51 mmol) of the crude title compound (24% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 233, 235; found 235.0 (100%), 232.9 (84%). $R_f = 0.68$ (hexanes to ethyl acetate 4:1).

5-Chloro- N^4 -(furan-2-ylmethyl)- N^2 -isopropylquinazoline-2,4-diamine (21): 0.045 g (0.15 mmol) of **21c** was reacted with furfurylamine according to general procedure Cb. **21d** was reacted with isopropylamine according to general procedure Db to give 0.006 g (0.019 mmol) of the title compound

1 in 13% yield over two steps. ^1H NMR (400 MHz, CDCl_3) δ 7.78 (s, 1H), 7.35 (dd, $J = 1.8, 0.8$ Hz, 1H),
2 7.28 – 7.24 (m, 2H), 6.98 – 6.88 (m, 1H), 6.31 (dd, $J = 3.2, 1.9$ Hz, 1H), 6.26 (dd, $J = 3.2, 0.8$ Hz, 1H),
3 4.93 (s, 1H), 4.73 (d, $J = 5.0$ Hz, 2H), 4.22 (oct, $J = 6.5$ Hz, 1H), 1.21 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101
4 MHz, CDCl_3) δ 159.17, 158.25, 155.04, 151.58, 142.07, 131.75, 128.92, 124.92, 122.93, 110.40,
5 108.71, 107.17, 42.66, 38.54, 23.17. HRMS: m/z calcd for $\text{C}_{16}\text{H}_{18}\text{ClN}_4\text{O}$ $[\text{M}+\text{H}]^+$ 317.1164; found
6 317.1169.

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14 **2,4-Dichloro-8-methylquinazoline (22c)**: 1.0 g (6.6 mmol) of commercially available 2-amino-3-
15 methylbenzoic acid was reacted according to general procedure A to give crude **22b**. Without further
16 purification, **22b** was reacted according to general procedure B to give 1.1 g (5.2 mmol) of the crude
17 title compound (78% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 213, 215; found 213.0 (100%), 215.0 (73%).
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19 $R_f = 0.67$ (hexanes to ethyl acetate 4:1).

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26 **2-Chloro- N^4 -(furan-2-ylmethyl)-8-methylquinazolin-4-amine (22d)**: 0.50 g (2.3 mmol) of **22c** was
27 reacted with furfurylamine according to general procedure Cb to give 0.43 g (1.6 mmol) of **22d** in 70%
28 yield. ^1H NMR (400 MHz, DMSO-d_6) δ 9.08 (t, $J = 5.5$ Hz, NH), 8.07 (d, $J = 8.2$ Hz, 1H), 7.58 – 7.53
29 (m, 2H), 7.33 (t, $J = 7.7$ Hz, 1H), 6.39 – 6.35 (m, 1H), 6.32 (d, $J = 3.0$ Hz, 1H), 4.70 (d, $J = 5.5$ Hz, 2H),
30 2.45 (s, 3H). ^{13}C NMR (101 MHz, DMSO-d_6) δ 161.92, 156.75, 152.08, 149.93, 142.85, 135.39,
31 134.26, 126.11, 121.26, 113.88, 111.21, 108.30, 38.10, 17.88. Mass (ESI): $[\text{M}+\text{H}]^+$ 274, 276; found
32 274.1 (100%), 276.1 (35%). $R_f = 0.33$ (hexanes to ethyl acetate 4:1).

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43 **N^4 -(Furan-2-ylmethyl)- N^2 -isopropyl-8-methylquinazoline-2,4-diamine (22)**: 0.13 g (0.47 mmol)
44 of **22d** was reacted with isopropylamine according to general procedure Db to give 0.10 g (0.34 mmol)
45 of the title compound as a yellow solid in 74% yield. ^1H NMR (400 MHz, CDCl_3) δ 8.87 (s, 1H), 8.53
46 (s, 1H), 7.94 (s, 1H), 7.35 (d, $J = 7.3$ Hz, 1H), 7.22 – 7.21 (m, 1H), 7.04 (t, $J = 7.7$ Hz, 1H), 6.27 – 6.19
47 (m, 2H), 4.80 (d, $J = 5.2$ Hz, 2H), 4.22 (oct, $J = 6.7$ Hz, 1H), 1.25 (d, $J = 6.6$ Hz, 6H). ^{13}C NMR (101
48 MHz, CDCl_3) δ 160.37, 153.04, 150.27, 142.20, 138.12, 135.91, 126.12, 123.57, 121.16, 110.46,
49 109.19, 108.09, 43.89, 38.50, 22.34, 18.07. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}$ $[\text{M}+\text{H}]^+$ 297.1710; found
50 297.1712. $R_f = 0.50$ (dichloromethane to methanol 9:1). Decomposed at 150 °C.
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2,4-Dichloro-7-methylquinazoline (23c): 0.5 g (3.3 mmol) of commercially available 2-amino-4-methylbenzoic acid was reacted according to general procedure A to give crude **23b**. Without further purification, **23b** was reacted according to general procedure B to give 0.34 g (1.6 mmol) of the crude title compound (48% over two steps). Mass (ESI): $[M+H]^+$ 213, 215; found 213.0 (100%), 215.0 (63%). $R_f = 0.55$ (hexanes to ethyl acetate 4:1).

2-Chloro-*N*⁴-(furan-2-ylmethyl)-7-methylquinazolin-4-amine (23d): 0.20 g (0.94 mmol) of **23c** was reacted with furfurylamine according to general procedure Cb to give 0.091 g (0.33 mmol) of **23d** in 35% yield. ¹H NMR (400 MHz, CD₃OD) δ 7.91 (d, $J = 8.4$ Hz, 1H), 7.40 (s, 1H), 7.34 (s, 1H), 7.27 (d, $J = 8.5$ Hz, 1H), 6.32 (d, $J = 1.2$ Hz, 2H), 4.74 (s, 2H), 2.43 (s, 3H). ¹³C NMR (101 MHz, CD₃OD) δ 161.28, 157.55, 151.56, 150.46, 144.94, 142.11, 128.03, 125.11, 122.27, 111.37, 110.21, 107.66, 37.53, 20.59. Mass (ESI): $[M+H]^+$ 274, 276; found 274.1 (100%), 276.1 (36%). $R_f = 0.32$ (hexanes to ethyl acetate 4:1).

***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-7-methylquinazoline-2,4-diamine (23):** 0.068 g (0.25 mmol) of **23d** was reacted with isopropylamine according to general procedure Db to give 0.030 g (0.10 mmol) of the title compound in 40% yield. ¹H NMR (250 MHz, DMSO-*d*₆) δ 10.00 (s, NH), 8.21 (d, $J = 7.6$ Hz, 1H), 8.04 (s, 1H), 7.60 (d, $J = 1.3$ Hz, 1H), 7.16 (d, $J = 8.0$ Hz, 1H), 6.43-6.32 (m, 2H), 4.77 (d, $J = 4.9$ Hz, 2H), 4.32-4.16 (m, 1H), 2.41 (s, 3H), 1.22 (d, $J = 6.5$ Hz, 6H). ¹³C NMR (63 MHz, DMSO-*d*₆) δ 159.59, 151.02, 145.75, 142.30, 138.22, 135.88, 125.02, 124.02, 116.51, 112.23, 110.54, 107.67, 42.88, 37.55, 22.21, 21.34. HRMS: m/z calcd for C₁₇H₂₁N₄O $[M+H]^+$ 297.1710; found 297.1716. $R_f = 0.44$ (dichloromethane to methanol 9:1).

2,4-Dichloro-6-methylquinazoline (24c): 1 g (6.6 mmol) of commercially available 2-amino-5-methylbenzoic acid was reacted according to general procedure A to give crude **24b**. Without further purification, **24b** was reacted according to general procedure B to give 0.34 g (1.6 mmol) of the crude title compound (24% over two steps). Mass (ESI): $[M+H]^+$ 213; found 213.

2-Chloro-*N*⁴-(furan-2-ylmethyl)-6-methylquinazolin-4-amine (24d): 0.32 g (1.5 mmol) of **24c** was reacted with furfurylamine according to general procedure Cb to give 0.19 g (0.69 mmol) of **24d** in 46%

yield. ^1H NMR (400 MHz, CD_3OD) δ 7.79 (s, 1H), 7.50 (dd, $J = 8.5, 1.3$ Hz, 1H), 7.41 (d, $J = 8.3$ Hz, 2H), 6.33 (d, $J = 1.0$ Hz, 2H), 4.74 (s, 2H), 2.41 (s, 3H). ^{13}C NMR (101 MHz, CD_3OD) δ 161.06, 156.69, 151.55, 148.37, 142.11, 136.74, 135.25, 125.55, 121.59, 113.35, 110.21, 107.68, 37.56, 20.30. Mass (ESI): $[\text{M}+\text{H}]^+$ 274, 276; found 274.1 (100%), 276.1 (33%).

***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-6-methylquinazoline-2,4-diamine (24)**: 0.114 g (0.42 mmol) of **24d** was reacted with isopropylamine according to general procedure Db to give 0.105 g (0.35 mmol) of the title compound as a yellow solid in 83% yield. ^1H NMR (400 MHz, CDCl_3) δ 7.51 (s, 1H), 7.40 – 7.29 (m, 3H), 6.33 (dd, $J = 3.2, 1.9$ Hz, 1H), 6.31 (d, $J = 3.1$ Hz, 1H), 4.82 (s, 2H), 4.27 (oct, $J = 6.6$ Hz, 1H), 2.36 (s, 3H), 1.27 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.79, 154.12, 150.67, 142.21, 135.81, 133.04, 122.06, 119.30, 110.46, 109.44, 107.96, 43.45, 38.27, 22.63, 21.05. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}$ $[\text{M}+\text{H}]^+$ 297.1710; found 297.1712. $R_f = 0.43$ (dichloromethane to methanol 9:1). Melting point 200–204 °C.

2,4,-Dichloro-5-methylquinazoline (25c): 1 g (6.6 mmol) of commercially available 2-amino-6-methylbenzoic acid was reacted according to general procedure A to give crude **25b**. Without further purification, **25b** was reacted according to general procedure B to give 0.60 g (2.8 mmol) of the crude title compound (42% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 213, 215; found 213.0 (100%), 215.0 (71%). $R_f = 0.77$ (hexanes to ethyl acetate 1:1).

2-Chloro-*N*⁴-(furan-2-ylmethyl)-5-methylquinazolin-4-amine (25d): 0.30 g (1.4 mmol) of **25c** was reacted with furfurylamine according to general procedure Cb to give 0.055 g (0.20 mmol) of **25d** in 14% yield. ^1H NMR (400 MHz, CD_3OD) δ 7.59 – 7.54 (m, 1H), 7.45 – 7.34 (m, 2H), 7.25 (d, $J = 7.1$ Hz, 1H), 6.36 (s, 2H), 4.79 (s, 2H), 2.82 (s, 3H). ^{13}C NMR (101 MHz, CD_3OD) δ 162.25, 152.30, 142.08, 134.99, 134.03, 132.97, 129.84, 129.37, 126.90, 124.28, 110.25, 107.61, 38.32, 22.20. Mass (ESI): $[\text{M}+\text{H}]^+$ 274, 276; found 274.1 (100%), 276.1 (33%). $R_f = 0.26$ (hexanes to ethyl acetate 4:1).

***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-5-methylquinazoline-2,4-diamine (25)**: 0.10 g (0.37 mmol) of **25d** was reacted with isopropylamine according to general procedure Db to give 0.066 g (0.22 mmol) of the title compound as a yellow solid in 60% yield. ^1H NMR (400 MHz, CD_3OD) δ 7.41 (d, $J = 1.0$

1 Hz, 1H), 7.28 (t, $J = 7.8$ Hz, 1H), 7.14 (d, $J = 8.3$ Hz, 1H), 6.78 (d, $J = 7.1$ Hz, 1H), 6.34 (dd, $J = 3.0, 1.9$
2 Hz, 1H), 6.27 (d, $J = 2.8$ Hz, 1H), 4.74 (s, 2H), 4.21 (sept, $J = 6.5$ Hz, 1H), 2.69 (s, 3H), 1.21 (d, $J = 6.5$
3 Hz, 6H). ^{13}C NMR (101 MHz, CD_3OD) δ 161.42, 158.32, 153.28, 152.67, 141.75, 134.10, 131.88,
4 124.40, 122.13, 111.02, 110.20, 106.52, 42.50, 38.16, 22.56, 22.30. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}$
5 $[\text{M}+\text{H}]^+$ 297.1710; found 297.1710. $R_f = 0.16$ (Ethyl acetate). Melting point 89-92 °C.
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11 **2,4,-Dichloro-8-methoxyquinazoline (26c):** 1 g (6.0 mmol) of commercially available 2-amino-3-
12 methoxybenzoic acid was reacted according to general procedure A to give crude **26b**. Without further
13 purification, **26b** was reacted according to general procedure B to give 0.15 g (0.65 mmol) of the crude
14 title compound (11% over two steps). Mass (ESI): $[\text{M}+\text{H}]^+$ 229, 231; found 229.0 (100%), 230.9 (68%).
15 $R_f = 0.26$ (hexanes to ethyl acetate 4:1).
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23 **2-Chloro- N^4 -(furan-2-ylmethyl)-8-methoxyquinazolin-4-amine (26d):** 0.10 g (0.46 mmol) of **26c**
24 was reacted with furfurylamine according to general procedure Cb to give 0.083 g (0.29 mmol) of **20d**
25 in 63% yield. Mass (ESI): $[\text{M}+\text{H}]^+$ 290, 292; found 290.1 (100%), 292.0 (68%). $R_f = 0.72$
26 (dichloromethane to methanol 19:1).
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33 **N^4 -(Furan-2-ylmethyl)- N^2 -isopropyl-8-methoxyquinazoline-2,4-diamine (26):** 0.069 g (0.24
34 mmol) of **26d** was reacted with isopropylamine according to general procedure Db to give 0.023 g
35 (0.073 mmol) of the title compound as a yellow crystalline solid in 30% yield. ^1H NMR (500 MHz,
36 CD_3OD) δ 7.54 (dd, $J = 7.2, 1.8$ Hz, 1H), 7.44 – 7.42 (m, 1H), 7.17 – 7.09 (m, 2H), 6.37 – 6.33 (m, 2H),
37 4.81 (s, 2H), 4.30 (hept, $J = 6.6$ Hz, 1H), 3.92 (s, 3H), 1.29 (d, $J = 6.6$ Hz, 6H). ^{13}C NMR (126 MHz,
38 CD_3OD) δ 160.21, 153.10, 150.93, 147.67, 142.12, 131.49, 123.46, 114.22, 113.55, 110.15, 109.86,
39 107.53, 55.58, 43.46, 37.85, 21.51. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_2$ $[\text{M}+\text{H}]^+$ 313.1659; found
40 313.1654. $R_f = 0.22$ (dichloromethane to methanol 19:1). Melting point 157-162 °C.
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52 **2,4,-Dichloro-7-methoxyquinazoline (27c):** 0.98 g (5.9 mmol) of commercially available 2-amino-4-
53 methoxybenzoic acid was reacted according to general procedure A to give crude **27b**. Without further
54 purification, **27b** was reacted according to general procedure B to give 0.09 g (0.39 mmol) of the crude
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1 title compound (7% over two steps). Mass (ESI): $[M+H]^+$ 229, 231; found 228.9 (100%), 231.0 (68%).

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3 $R_f = 0.71$ (hexanes to ethyl acetate 2:1).

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5 **2-Chloro-*N*⁴-(furan-2-ylmethyl)-7-methoxyquinazolin-4-amine (27d)**: 0.075 g (0.33 mmol) of **27c**
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7 was reacted with furfurylamine according to general procedure Cb to give 0.083 g (0.29 mmol) of **27d**
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9 in 88% yield. ¹H NMR (500 MHz, CD₃OD) δ 7.92 (d, *J* = 9.1 Hz, 1H), 7.43 (d, *J* = 1.0 Hz, 1H), 7.01
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11 (dd, *J* = 9.1, 2.5 Hz, 1H), 6.91 (d, *J* = 2.3 Hz, 1H), 6.35 (s, 2H), 4.76 (s, 2H), 3.88 (s, 3H). ¹³C NMR
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13 (126 MHz, CD₃OD) δ 163.95, 160.75, 157.72, 152.39, 151.46, 141.90, 123.70, 117.16, 109.99, 107.40,
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15 107.26, 105.06, 54.76, 37.32. Mass (ESI): $[M+H]^+$ 290, 292; found 290.1 (100%), 292.0 (33%). $R_f =$
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17 0.44 (hexanes to ethyl acetate 2:1).
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21 ***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-7-methoxyquinazoline-2,4-diamine (27)**: 0.065 g (0.29
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23 mmol) of **27d** was reacted with isopropylamine according to general procedure Db to give 0.007 g
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25 (0.022 mmol) of the title compound in 8% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.70 (d, *J* = 21.0 Hz,
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27 1H), 7.37 – 7.31 (m, 1H), 6.77 (s, 1H), 6.70 (dd, *J* = 9.0, 2.4 Hz, 1H), 6.32 (dd, *J* = 3.1, 1.9 Hz, 1H),
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29 6.29 (d, *J* = 3.2 Hz, 1H), 4.80 (s, 2H), 4.27 (oct, *J* = 6.6 Hz, 1H), 3.81 (s, 3H), 1.27 (d, *J* = 6.6 Hz, 6H).
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31 ¹³C NMR (101 MHz, CDCl₃) δ 163.99, 159.57, 156.47, 151.19, 142.15, 127.72, 123.50, 113.25, 110.44,
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33 107.73, 103.92, 101.78, 55.57, 43.17, 38.07, 22.83. HRMS: *m/z* calcd for C₁₇H₂₁N₄O₂ $[M+H]^+$
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35 313.1659; found 313.1667.
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40 **2,4,-Dichloro-6-methoxyquinazoline (28c)**: 0.20 g (1.2 mmol) of commercially available 2-amino-4-
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42 methoxybenzoic acid was reacted according to general procedure A to give crude **28b**. Without further
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44 purification, **28b** was reacted according to general procedure B to give 0.14 g (0.61 mmol) of the crude
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46 title compound (50% over two steps). Mass (ESI): $[M+H]^+$ 229, 231; found 229.0 (100%), 231.0 (67%).
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48 $R_f = 0.53$ (hexanes to ethyl acetate 4:1).
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52 **2-Chloro-*N*⁴-(furan-2-ylmethyl)-6-methoxyquinazolin-4-amine (28d)**: 0.061 g (0.27 mmol) of **28c**
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54 was reacted with furfurylamine according to general procedure Cb to give 0.031 g (0.11 mmol) of **28d**
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56 in 41% yield. ¹H NMR (500 MHz, CD₃OD) δ 7.45 (m, 2H), 7.40 (s, 1H), 7.28 (d, *J* = 9.0 Hz, 1H), 6.37
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58 (d, *J* = 3.8 Hz, 2H), 4.77 (s, 2H), 3.86 (s, 3H). ¹³C NMR (126 MHz, CD₃OD) δ 160.52, 158.00, 155.06,
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151.38, 144.99, 141.90, 127.01, 124.51, 113.88, 109.99, 107.59, 101.40, 54.99, 37.43. Mass (ESI): [M+H]⁺ 290, 292; found 290.0 (100%), 292.0 (33%). R_f = 0.85 (dichloromethane to methanol 9:1).

***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-6-methoxyquinazoline-2,4-diamine (28)**: 0.14 g (0.48 mmol) of **28d** was reacted with isopropylamine according to general procedure Db to give 0.045 g (0.14 mmol) of the title compound as a yellow solid in 29% yield. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.42 (s, 1H), 7.58 (dd, J = 1.8, 0.9 Hz, 1H), 7.54 (d, J = 2.4 Hz, 1H), 7.24 (d, J = 9.0 Hz, 1H), 7.18 (dd, J = 9.0, 2.5 Hz, 1H), 6.40 (dd, J = 3.1, 1.8 Hz, 1H), 6.35 (d, J = 2.8 Hz, 1H), 4.72 (d, J = 5.3 Hz, 2H), 4.13 (oct, J = 6.5 Hz, 1H), 3.79 (s, 3H), 1.15 (d, J = 6.5 Hz, 6H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 159.63, 157.27, 153.87, 152.97, 145.81, 142.36, 125.50, 123.66, 110.91, 107.59, 103.66, 56.05, 42.25, 37.36, 23.19. HRMS: m/z calcd for C₁₇H₂₁N₄O₂ [M+H]⁺ 313.1659; found 313.1659. R_f = 0.28 (dichloromethane to methanol 9:1). Melting point 138-142 °C.

2,4-Dichloro-5-methoxyquinazoline (29c): 1.0 g (6.2 mmol) of commercially available 2-amino-4-methoxybenzoic acid was reacted according to general procedure A to give crude **29b**. Without further purification, **29b** was reacted according to general procedure B to give 0.036 g (0.16 mmol) of the crude title compound (3% over two steps). Mass (ESI): [M+H]⁺ 229, 231; found 229.0 (100%), 231.0 (70%). R_f = 0.53 (hexanes to ethyl acetate 2:1).

2-Chloro-*N*⁴-(furan-2-ylmethyl)-5-methoxyquinazolin-4-amine (29d): 0.0094 g (41 μmol) of **29c** was reacted with furfurylamine according to general procedure Cb to give 0.0045 g (0.11 mmol) of **29d** in 38% yield. ¹H NMR (250 MHz, CD₃OD) δ 7.58 (t, J = 8.3 Hz, 1H), 7.36 (d, J = 1.0 Hz, 1H), 7.08 (d, J = 8.2 Hz, 1H), 6.93 (d, J = 7.9 Hz, 1H), 6.33 – 6.20 (m, 2H), 4.70 (s, 2H), 3.95 (s, 3H). Mass (ESI): [M+H]⁺ 290, 292; found 290.0 (100%), 292.0 (37%). R_f = 0.40 (hexanes to ethyl acetate 2:1).

***N*⁴-(Furan-2-ylmethyl)-*N*²-isopropyl-5-methoxyquinazoline-2,4-diamine (29)**: 0.0019 g (6.6 μmol) of **29d** was reacted with isopropylamine according to general procedure Db to give 0.0012 g (3.8 μmol) of the title compound in 58% yield. ¹H NMR (400 MHz, CD₃OD) δ 7.53 (t, J = 8.3 Hz, 1H), 7.41 (t, J = 6.7 Hz, 1H), 6.93 (d, J = 8.1 Hz, 1H), 6.78 (d, J = 8.2 Hz, 1H), 6.38 – 6.32 (m, 1H), 6.29 (s, 1H), 4.79

(s, 2H), 4.30 – 4.16 (m, 1H), 4.00 (s, 3H), 1.23 (d, $J = 6.5$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 160.20, 157.19, 150.24, 142.45, 142.31, 134.94, 127.73, 113.87, 110.56, 107.75, 104.07, 100.31, 56.50, 43.55, 38.44, 22.48. HRMS: m/z calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_2$ $[\text{M}+\text{H}]^+$ 313.1659; found 313.1654. $R_f = 0.31$ (dichloromethane to methanol 9:1).

Physicochemical Assays

Permeability Assay. Permeability P_e has been determined by a standard parallel artificial membrane permeability assay (PAMPA by pION) as reported previously.³⁷

Aqueous Solubility Assay. Solubility at pH 7.4 has been determined using Biomek FX lab automation workstation with pION μSOL evolution software as reported previously and at pH 2.0 using an in-house HPLC assay.³⁷ For pH 2.0, a calibration curve was made by plotting the area under the curve at 254 nm (uv by HPLC) against the concentration of each compound injected after performing a serial dilution (25 μM to 0.781 μM) using a solvent in which the compound is soluble (DMSO). A 100 μM solution was then made for each compound in a buffer at 2.0 by performing a 1:100 serial dilution using a 10 mM DMSO stock solution of each compound. This solution was incubated at 21 $^\circ\text{C}$ for 18 hours, filtered using a filter plate, and injected into the HPLC to compare the area found at wavelength 254 nm with the previously made calibration curve.

Partition Coefficient Assay. Log D was also determined in-house via an HPLC method adapted from the strategy reported by Donovan and Pescatore.³⁶ Two buffers were made at a concentration of 50 μM each: ammonium acetate at pH 7.4 and ammonium formate at pH 3.0. A set of compounds with known log D values between -0.36 and 5.68 were used to make a calibration curve at each pH by using a linear gradient between 0 and 100% acetonitrile, with buffer at the specific pH used as the second solvent. The curve was made by plotting the log D value against the retention time. Quinazolines were then injected into the HPLC and the retention time compared to the calibration curve previously determined.

Efficacy studies

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Intracellular amastigote assays. *In vitro* antileishmanial potency of compounds against intracellular *L. donovani* MHOM/ET/67/L82²⁹ and *L. amazonensis* LV78²⁵ parasites was measured using the colorimetric assay as described previously. The potency of compounds **15**, **16** and **23** against intracellular antimony-resistant clinical isolate *L. donovani* MHOM/NP/02/BPK164/1³¹ and antimony-susceptible isolate *L. donovani* MHOM/NP/03/BPK206/0³¹ was also assessed. Briefly, 2nd-3rd day stationary phase promastigotes grown in HOMEM (Invitrogen) supplemented with 20% fetal calf serum were used to infect murine primary peritoneal macrophages at a ratio of 10 parasites to 1 host cell and subsequently exposed to four concentrations ranging between 0.31-25 μ M for compound **15** or 0.31-10 μ M for compounds **16** and **23**. After four days of exposure to compounds, slides were fixed with methanol and stained with Giemsa to manually assess parasite survival (the percentage of infected macrophages) using conventional light microscopy. A standardized antimony-susceptibility test³¹ was performed in parallel to confirm the validity of the test and the clinical isolates' antimony-susceptibility profiles.

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Cytotoxicity assays. Toxicity of compounds against the murine macrophage cell line J774A.1 was conducted using the 3-(4,5)-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay as previously described⁴⁰ except that cells were incubated with compounds for 72 h.

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Murine visceral leishmaniasis model. The *in vivo* antileishmanial efficacy of compounds was evaluated in a murine model of visceral leishmaniasis by the general method described earlier²⁵ with minor modifications. Briefly, BALB/c mice were inoculated with LV82 promastigotes on day 0, and then weighed and marked individually on day 6 in order to calculate the volume of solution to be administered to each mouse. In these studies, 45% (w/v) (2-hydroxypropyl)- β -cyclodextrin (HP β CD) vehicle was used to formulate **16** and 0.5% methyl cellulose/0.1% Tween80 in water (MC) was used to dissolve **15** and **23**; all compounds and vehicles were administered intraperitoneally. After 5 consecutive days of treatment from days 7 to 11, animals were sacrificed on day 14. Liver smear slides from each animal were made and stained, and the liver parasitemia expressed in Leishman-Donovan units (LDU)

1 was determined by microscopy.²⁵ The efficacy of each compound was calculated according to the
2 method published previously.²⁵
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5 **Pharmacokinetics**

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7 **Animals.** Protocols for pharmacokinetics studies in mice were approved by the Institutional Animal
8 Care and Use Committee of the University of Kansas. Male Swiss Webster mice (weighing 20–25 g)
9 were purchased from the Charles River Laboratories (Raleigh, NC). Mice were housed in a clean-room
10 under filtered, pathogen-free air, in a 12-h light/dark cycle, and with food pellets and water available *ad*
11 *libitum*.
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19 **Pharmacokinetics and metabolic stability of 16 and 23 in mice.** **16** (dissolved in 50% DMA) and
20 **23** [dissolved in PEG400: DMA: water = 5: 3: 2 (v/v)] were administered at a dose of 100 $\mu\text{mol/kg}$ body
21 weight (or approximately 30 mg/kg; dose volume = 5 mL/kg) via oral gavage (p.o.) or intraperitoneal
22 (i.p.) administration to Swiss Webster mice. Blood sampling time schedules were 1, 2, 4, 8, 12, and 24 h
23 ($n = 3$ at each blood sampling time point) after dosing. Blood and tissue samples were collected and
24 processed for drug concentration determination as previously described.⁴¹ Metabolic stability of **16** and
25 **23** was evaluated using pooled mouse liver microsomes (XenoTech LLC, Lenexa, KS) supplemented
26 with NADPH. Microsomal incubations were carried out according to a protocol described previously.⁴²
27 Substrate and microsomal protein concentrations during incubations were 3 μM and 0.5 mg/mL,
28 respectively. Substrate depletion over a 60-min incubation time was quantified using high-performance
29 liquid chromatography–tandem mass spectrometry (HPLC-MS/MS) as described below.
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45 **Analytical assays.** The processed samples (plasma, tissue homogenates, and microsomal incubation
46 mixtures) were analyzed for drug concentration using an API3000 triple quadrupole mass spectrometer
47 equipped with a Turbo IonSpray interface (AB Sciex, Foster City, CA). Samples (5 μL) were introduced
48 to the mass spectrometer using a thermostatic (4°C) autosampler (CTC PAL, LEAP Technologies,
49 Carrboro, NC), and a Shimadzu HPLC system (Shimadzu, Kyoto, Japan). All equipment was controlled
50 using Analyst software (Version 1.3). Compounds were separated on a ZORBAX Bonus-RP C₁₈ HPLC
51 column (2.1 x 50 mm, 3.5 μm particle size; Agilent, Milford, MA). HPLC mobile phases consisted of
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1 water containing 0.1% formic acid (A) and methanol containing 0.1% formic acid (B) with a flow rate
2 of 0.35 mL/min. After a 0.4-min initial hold at 15% B, mobile phase composition started with 15% B
3 and was increased to 95% B over 1.1 min. Then the column was washed with 95% B for 1.5 min and
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5 and was increased to 95% B over 1.1 min. Then the column was washed with 95% B for 1.5 min and
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7 was re-equilibrated with 15% B for 1.0 min before injecting the next sample. The characteristic single
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9 reaction monitoring (SRM) transitions for **16** and **23** were m/z 297.1→81.0 and 307.1→91.0 under
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11 positive ion mode. They were used as internal standard for each other during quantification. The
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13 calibration curves for **16** and **23** ranged from 0.01 to 10 μM for plasma and from 0.10 to 50 μM for
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15 tissue homogenates. For microsomal stability samples, substrate depletion was calculated based on the
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17 relative MS signal of the analyte normalized by the internal standard.
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21 **Data analysis.** Microsomal half-life ($t_{1/2}$) values were obtained by fitting the one-phase
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23 exponential decay equation to the percentage of substrate remaining versus time curves (GraphPad
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25 Prism, Version 5.04, San. Diego, CA). Noncompartmental pharmacokinetic analysis of plasma
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27 concentration versus time curves was performed to obtain the area under the concentration-time curve
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29 (AUC), C_{max} , T_{max} , and terminal half-life ($t_{1/2}$) (WinNonlin 6.3, Pharsight, Mountain View, CA). Tissue
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31 AUC was calculated using the trapezoid rule from 1 h (first sampling time) to the last time point with
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33 quantifiable drug levels without extrapolating back to $t = 0$ h or beyond the last quantifiable time point
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35 (GraphPad Prism, Version 5.04).
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5 Tampa, FL 33620
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8 **Abbreviations:** dimethyl acetamide (DMA), 3-(4,5)-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium
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10 bromide (MTT), (2-hydroxypropyl)- β -cyclodextrin (HP β CD), maximum concentration (C_{\max}),
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12 microsomal half-life (Mic $t_{1/2}$), *para*-aminobenzoic acid (PABA), permeability (Pe), selectivity index
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14 (SI) single reaction monitoring (SRM), structure-property relationship (SAR), weight per unit volume
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16 (w/v).
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Figure 1: Antileishmanial compounds. A) Structures of currently used antileishmanial drugs.¹² B) Reported structures of quinazolines displaying antileishmanial activity and including the hit compounds **1** and **2** and SAR studies targeting the major quinazoline sites.

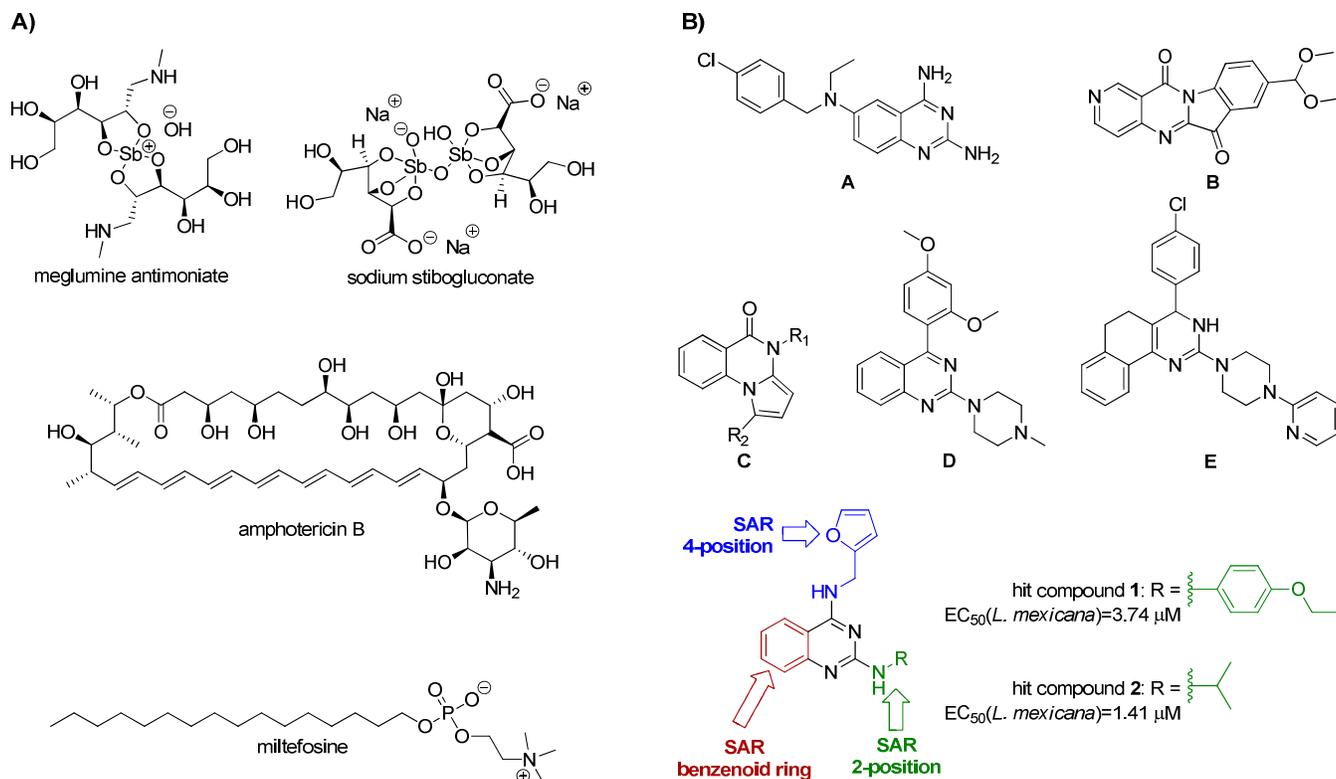


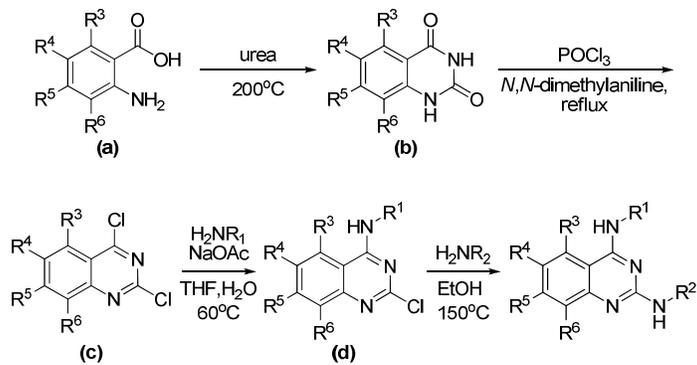
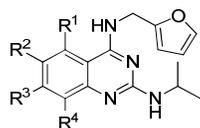
Figure 2: Synthesis of N^2,N^4 -disubstituted quinazoline-2,4-diamines.

Table 1: SAR study focusing on 2- and 4-positions^a

Compound	R ¹	R ²	<i>L. donovani</i>	<i>L. amazonensis</i>	J774A.1	SI
			EC ₅₀ μM	EC ₅₀ μM	EC ₅₀ μM	
2			2.5 ± 0.4	3.7 ± 1.6	17 ± 6	6.8
3			3.6 ± 1.1	5.8 ± 2.3	23 ± 10	6.4
4			25 ± 18	> 50	> 33	> 1.3
5			3.7 ± 0.3	7.6 ± 3.2	> 33	> 8.9
6			4.3 ± 1.2	5.1 ± 0.9	20 ± 6	4.7
7			6.9 ± 1.8	20 ± 3	> 33	4.8
8			>50.0	> 50.0	n.d.	n.d.
9			8.9 ± 1.0	19 ± 3	> 33	> 3.7
10			0.67 ± 0.27	1.4 ± 0.5	5.5 ± 1.4	8.2
11			1.8 ± 0.4	2.1 ± 1.1	5.4 ± 1.4	3.0
12			1.5 ± 0.5	13 ± 3	18 ± 8	12
13			0.65 ± 0.10	1.8 ± 0.2	5.1 ± 1.5	7.8
14			0.64 ± 0.10	2.6 ± 0.7	6.8 ± 0.2	11
15			0.15 ± 0.02	0.90 ± 0.27	15 ± 1	100
16			0.34 ± 0.12	1.4 ± 0.2	4.9 ± 1.3	14
17			0.26 ± 0.15	2.2 ± 0.3	5.2 ± 1.3	20

^aAmphotericin B is the internal control for the *in vitro* antileishmanial activity assay with EC₅₀ = 40 ± 9 nM against *L. donovani* and EC₅₀ = 89 ± 16 nM against *L. amazonensis*. Podophyllotoxin is the internal control for the *in vitro* cytotoxicity assay with EC₅₀ = 250 ± 10 nM against J774A.1.

Table 2: SAR study focusing on the benzenoid ring of the quinazoline scaffold.



Compound	R ¹	R ²	R ³	R ⁴	<i>L. donovani</i> EC ₅₀ μM	<i>L. amazonensis</i> EC ₅₀ μM	J774A.1 EC ₅₀ μM	SI
2	-H	-H	-H	-H	2.5 ± 0.4	3.7 ± 1.6	17 ± 6	6.8
18	-H	-H	-H	-Cl	4.4 ± 1.3	18 ± 6	> 33	> 7.5
19	-H	-H	-Cl	-H	9.0 ± 3.7	23 ± 5	>100	> 11
20	-H	-Cl	-H	-H	2.6 ± 1.9	25 ± 4	> 33	> 13
21	-Cl	-H	-H	-H	2.3 ± 0.6	12 ± 3	25 ± 3	11
22	-H	-H	-H	-CH ₃	4.2 ± 2.1	4.4 ± 1.9	> 33	> 7.9
23	-H	-H	-CH ₃	-H	0.83 ± 0.32	4.1 ± 1.2	> 33	> 40
24	-H	-CH ₃	-H	-H	4.7 ± 2.5	15 ± 6	> 33	> 7.0
25	-CH ₃	-H	-H	-H	0.95 ± 0.27	3.6 ± 1.5	20 ± 9	21
26	-H	-H	-H	-OCH ₃	3.2 ± 1.2	7.3 ± 2.5	30 ± 5	9.4
27	-H	-H	-OCH ₃	-H	1.2 ± 0.5	10 ± 2	16 ± 2	13
28	-H	-OCH ₃	-H	-H	0.74 ± 0.37	9.6 ± 3.0	14 ± 1	19
29	-OCH ₃	-H	-H	-H	0.97 ± 0.26	2.7 ± 1.4	18 ± 6	19

^aAmphotericin B is the internal control for the *in vitro* antileishmanial activity assay with EC₅₀ = 40 ± 9 nM against *L. donovani* and EC₅₀ = 89 ± 16 nM against *L. amazonensis*. Podophyllotoxin is the internal control for the *in vitro* cytotoxicity assay with EC₅₀ = 250 ± 10 nM against J774A.1.

Figure 3: *In vitro* efficacy of quinazolines, methotrexate (MTX), pyrimethamine (PYR), and miltefosine (MILT) for axenic amastigotes of *Leishmania donovani* (A-C) and J774.A1 macrophages (D-F) in the absence or presence of *d,l*-folinic acid (FNA). Results are presented as the EC₅₀ (μM) in the absence or presence of increasing concentrations of FNA.

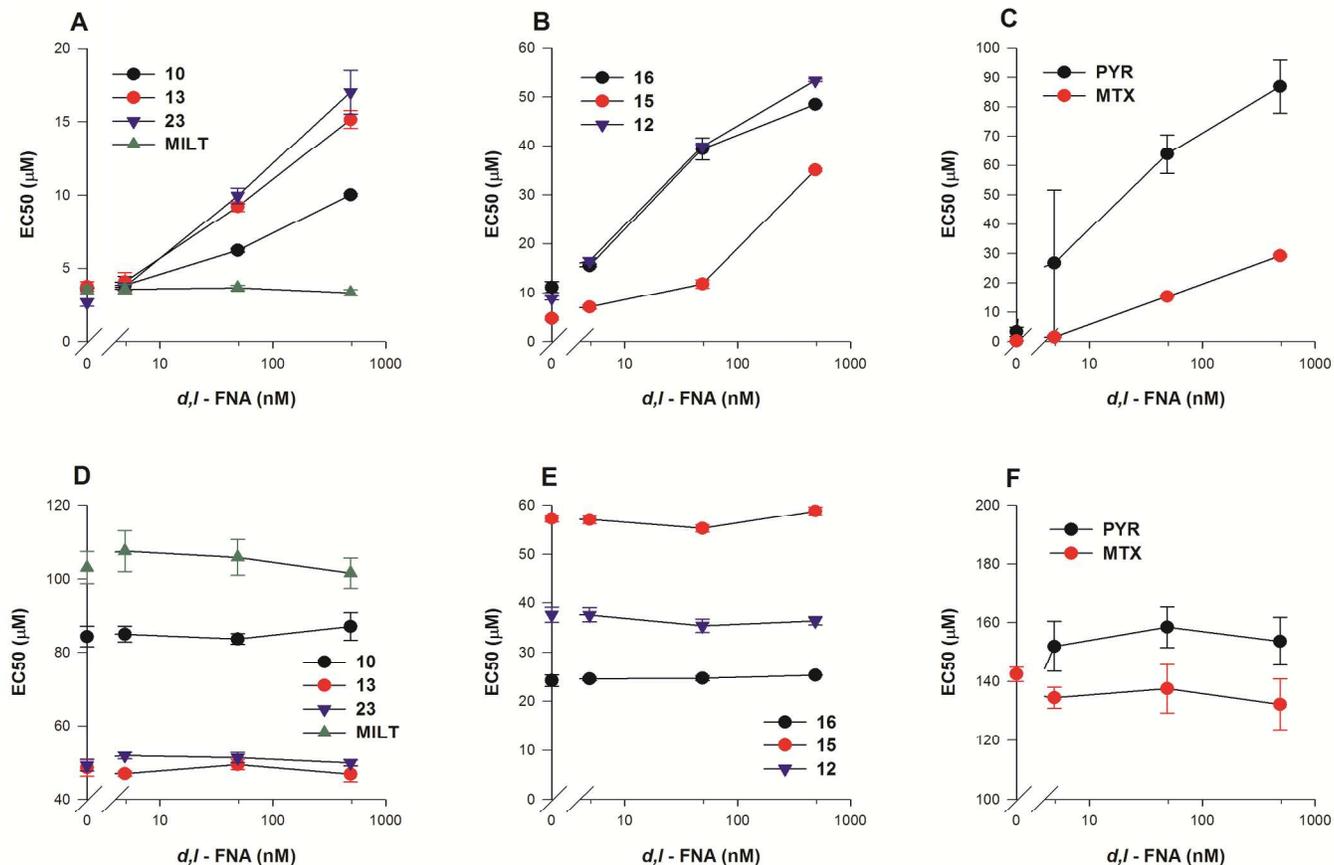


Table 3: Activities of compounds against *L. donovani* clinical isolates (μM)^a

Compound	BPK206/0 EC ₅₀ μM	BPK164/1 EC ₅₀ μM
Sodium stibogluconate ^b	24 \pm 6	>60
15	2.8 \pm 0.4	2.0 \pm 0.4
16	2.0 \pm 0.7	1.4 \pm 0.3
23	4.2 \pm 0.0	3.8 \pm 0.6

^aMean \pm range of two independent measurements

^bValues for sodium stibogluconate are given in micrograms of pentavalent antimony per milliliter

Table 4: Physicochemical properties of quinazolines

Compound	Solubility ^a		Permeability Pe · 10 ⁻⁶ (cm/s) ^b		Log D	
	pH 7.4	pH 2.0	pH 7.4	pH 4.0	pH 7.4	pH 3.0
2	*****	*****	883	62.2	3.15	2.18
3	***	*****	979	67.7	4.20	2.02
4	****	*****	1130	26.4	2.19	1.48
5	*****	*****	780	19.7	3.19	2.40
6	****	*****	1480	28.8	3.02	2.06
7	****	*****	744	38.8	2.54	1.73
8	****	*****	783	112	4.69	3.56
9	***	*****	1120	72.3	3.13	1.64
10	**	****	383	312	3.82	2.82
11	***	*****	880	485	3.89	2.91
12	*****	*****	n/d	n/d	3.14	2.11
13	***	*****	1250	161	3.80	2.84
14	*	**	1480	271	4.17	2.78
15	***	*****	1720	57.1	2.96	2.00
16	***	*****	1020	341	3.84	2.86
17	*	*****	663	412	4.25	2.67
18	**	*****	1330	67.6	3.96	2.38
19	****	*****	1380	103	3.91	2.51
20	****	*****	1630	170	3.99	2.60
21	***	*****	1240	169	4.44	2.62
22	****	*****	666	45.9	3.55	1.57
23	****	*****	798	67.2	3.61	1.58
24	****	*****	789	67.2	3.63	1.55
25	***	*****	812	45.9	3.55	2.44
26	****	*****	1060	38.4	3.40	2.99
27	****	*****	1330	59.2	3.70	3.11
28	*****	*****	1010	80.2	3.58	2.95
29	****	*****	624	87.0	3.35	2.53

^a (*) for solubility ≤ 5 μM, (**) for 5 μM < solubility ≤ 10 μM, (***) for 10 μM < solubility ≤ 20 μM, (****) for 20 μM < solubility ≤ 30 μM, (*****) for solubility ≥ 30 μM. n/d: not determined. ^b For the determination of the Pe values, the following internal controls were utilized: carbazepine pH 4.0 permeability Pe = 108·10⁻⁶ (cm/s) and pH 7.4 permeability Pe = 130·10⁻⁶ (cm/s); ranitidine·HCl pH 4.0 permeability Pe = 5.2·10⁻⁶ (cm/s) and pH 7.4 permeability Pe = 2.2·10⁻⁶ (cm/s); verapamil·HCl pH 4.0 permeability Pe = 20.6·10⁻⁶ (cm/s) and pH 7.4 permeability Pe = 1360·10⁻⁶ (cm/s).

Figure 4: In vivo efficacy of quinazolines against LV82 in *L. donovani* infected BALB/c mice. Results are presented as the liver parasitemia (LDU) for each mouse (♦) and the average LDU in each group (-) determined by microscopy (n = 4). (A), LDU for mice treated with **16** and **23**; (B), LDU for mice treated with **15**. All treatments were given by the i.p. route. Compounds **15** and **23** were dissolved in 0.5% methyl cellulose and 0.1% Tween80 (MC), while compound **16** was dissolved in 45% (w/v) (2-hydroxypropyl)- β -cyclodextrin solution (HP β CD). (*): $p < 0.05$, compared with untreated control.

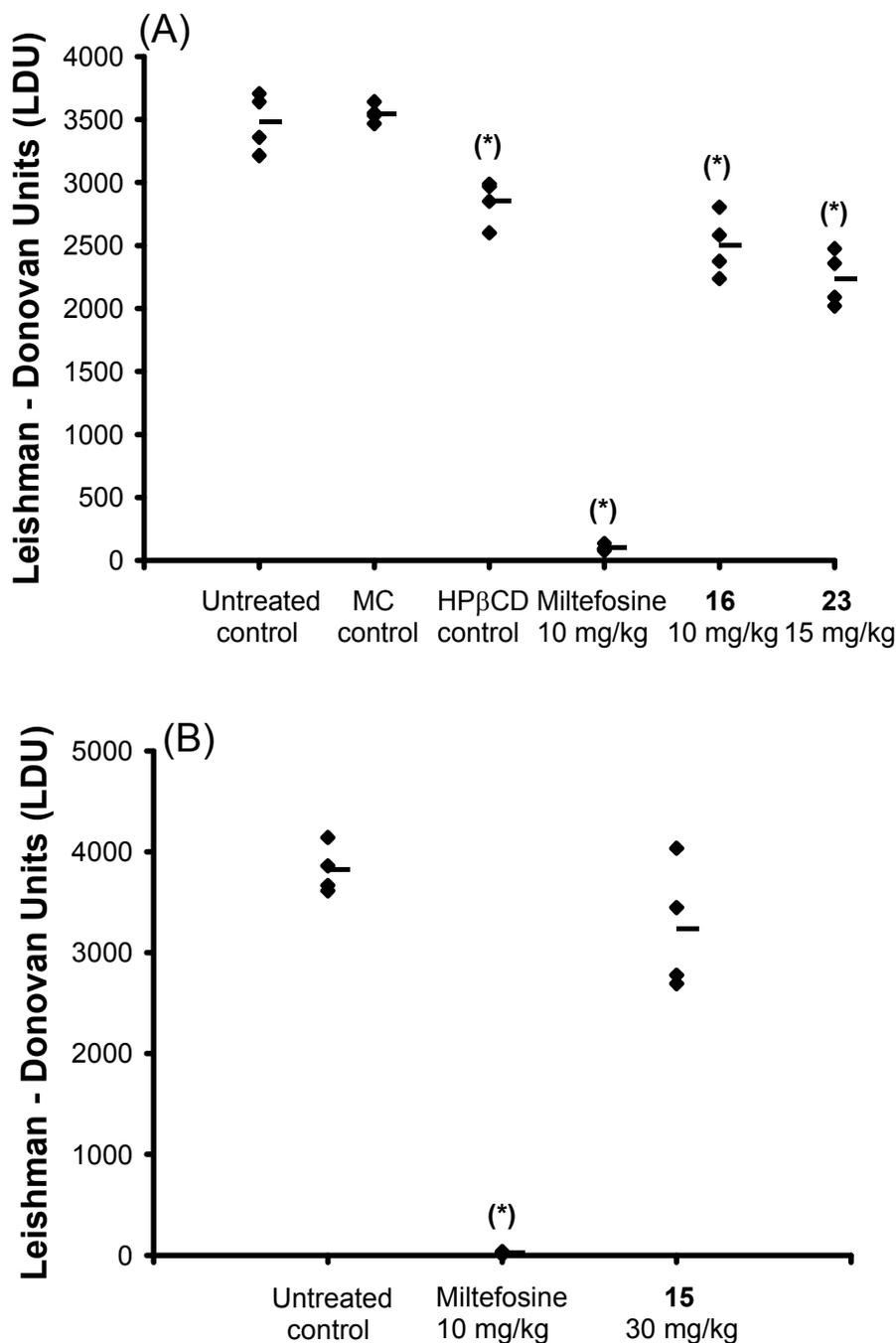


Figure 5. Plasma (open circles) and tissue (squares for liver and triangles for spleen) concentration-time profiles after p.o. (A) and i.p. (B) administration of **16** in mice at a dose level of 100 $\mu\text{mol/kg}$ (~ 30 mg/kg). Symbols and error bars represent the mean and standard error of triplicate determinations, except those labeled with asterisks where only one or two determinations were obtained due to sample loss.

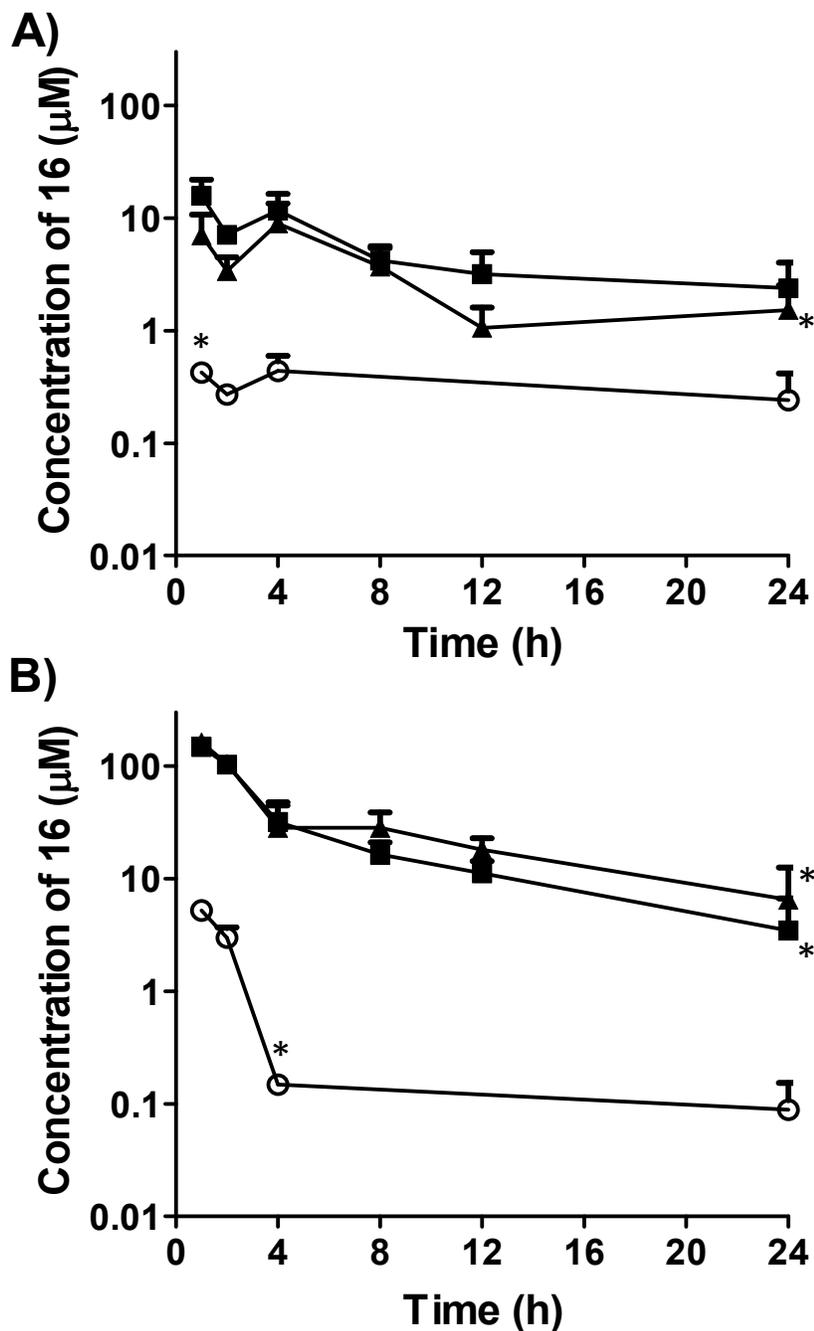


Figure 6. Plasma (open circles) and tissue (squares for liver and triangles for spleen) concentration-time profiles after p.o. (A) and i.p. (B) administration of **23** in mice at a dose level of 100 $\mu\text{mol/kg}$ (~ 30 mg/kg). Symbols and error bars represent the mean and standard error of triplicate determinations, except those labeled with asterisks where only one or two determinations were obtained due to sample loss. **23** was below the detection limit ($0.1 \mu\text{M}$) in the liver and spleen 12 and 24 h after i.p. administration.

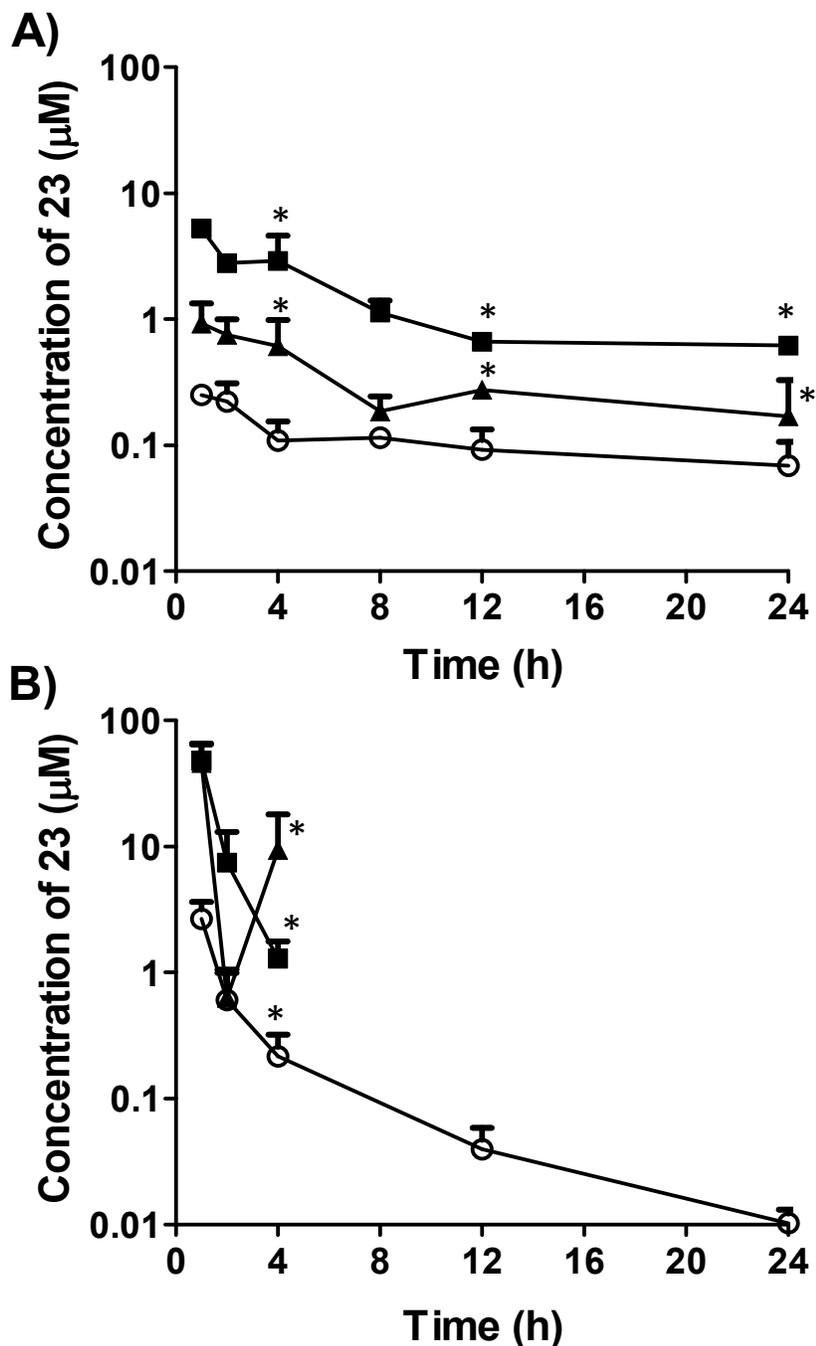


Table 5. Pharmacokinetic outcomes of **16** and **23** after p.o. and i.p. administration to mice.

Outcomes	p.o.			i.p.		
	plasma	liver	spleen	plasma	liver	spleen
16 C_{\max} (μM)	0.44	15.8	7.0	5.21	148	163
T_{\max} (h)	4	1	1	1	1	1
AUC_{last}^a ($\mu\text{M}\cdot\text{h}$)	7.8	110	68	11	500	620
$t_{1/2}$ (h)	NC ^b	ND ^c	ND	20	ND	ND
Mic $t_{1/2}^d$ (min)				27		
23 C_{\max} (μM)	0.25	5.2	0.9	2.67	48	45.8
T_{\max} (h)	1	1	1	1	1	1
AUC_{last}^a ($\mu\text{M}\cdot\text{h}$)	2.5	29	7.4	4.6	42	71
$t_{1/2}$ (h)	24	ND	ND	5.4	ND	ND
Mic $t_{1/2}^d$ (min)				9.4		

^a AUC(0-24h) was calculated for plasma using noncompartmental analysis, whereas AUC(1-24h) was calculated for tissues using trapezoid rule. AUC_{last} , AUC from the time of dose to the last measurable concentration; C_{\max} and T_{\max} , maximum concentration and time to reach C_{\max} ; $t_{1/2}$, terminal half-life. ^b NC, not calculable due to lack of a data point.

^c ND, not determined. ^d *In vitro* mouse liver microsomal half-life.

References

- 1
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4
5 1. Chappuis, F.; Sundar, S.; Hailu, A.; Ghalib, H.; Rijal, S.; Peeling, R. W.; Alvar, J.; Boelaert, M.
6
7 Visceral leishmaniasis: what are the needs for diagnosis, treatment and control? *Nature Rev. Microbiol.*
8
9 **2007**, *5*, S7-S16.
10
- 11
12
13 2. Leishmaniasis. *World Health Organization* <http://www.who.int/leishmaniasis/en/> accessed 7 January
14
15 2014.
16
17
18
19
- 20
21 3. Alvar, J.; Velez, I. D.; Bern, C.; Herrero, M.; Desjeux, P.; Cano, J.; Jannin, J.; den, B. M.; Argaw,
22
23 D.; Bhattacharya, S.; Ejov, M.; Elkhouri, A. N.; Ruiz-Postigo, J. A.; Serrano, J.; Denereaz, A.; Arana,
24
25 B.; Robledo, S. M.; Mondragon, K.; Velez, A.; Lopez, L.; Acosta, L. A. Leishmaniasis worldwide and
26
27 global estimates of its incidence. *PLoS One* **2012**, *7*, e35671.
28
29
30
31
- 32 4. Leishmaniasis Fact Sheet. *World Health Organization*
33
34 <http://www.who.int/mediacentre/factsheets/fs375/en/>, accessed 7 January 2014.
35
36
37
38
- 39 5. Desjeux, P. Leishmaniasis. Public health aspects and control. *Clin. Dermatol.* **1996**, *14*, 417-423.
40
41
42
- 43 6. Pearson, R. D.; Sousa, A. Q. Clinical spectrum of Leishmaniasis. *Clin. Infect. Dis.* **1996**, *22*, 1-13.
44
45
46
47
- 48 7. Guerin, P. J.; Olliaro, P.; Sundar, S.; Boelaert, M.; Croft, S., L.; Desjeux, P.; Wasunna, M., K.;
49
50 Bryceson, A., D. M. Visceral leishmaniasis: current status of control, diagnosis, and treatment, and a
51
52 proposed research and development agenda. *Lancet Infect. Dis.* **2002**, *2*, 494-501.
53
54
55
56
- 57 8. McCall, L.-I.; Zhang, W.-W.; Matlashewski, G. Determinants for the Development of Visceral
58
59 Leishmaniasis Disease. *PLoS Pathogens* **2013**, *9*, e1003053.
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58
59
60
9. Rey, L. Gaspar Vianna and the discovery of the treatment of leishmaniasis by antimonials. *Rev. Inst. Med. Trop. Sao Paulo* **1962**, *4*, 47-52.
10. Peters, W. The treatment of kala-azar—new approaches to an old problem. *Indian J. Med. Res.* **1981**, *73 Suppl*, 1-18.
11. Maltezou, H. C. Drug resistance in visceral leishmaniasis. *J. Biomed. Biotechnol.* **2010**, *2010*, Article ID 617521.
12. Frezard, F.; Martins, P. S.; Barbosa, M. C. M.; Pimenta, A. M. C.; Ferreira, W. A.; de Melo, J. E.; Mangrum, J. B.; Demicheli, C. New insights into the chemical structure and composition of the pentavalent antimonial drugs, meglumine antimonate and sodium stibogluconate. *J. Inorg. Biochem.* **2008**, *102*, 656-665.
13. Sundar, S.; More, D. K.; Singh, M. K.; Singh, V. P.; Sharma, S.; Makharia, A.; Kumar, P. C.; Murray, H. W. Failure of pentavalent antimony in visceral leishmaniasis in India: report from the center of the Indian epidemic. *Clin. Infect. Dis.* **2000**, *31*, 1104-1107.
14. Sundar, S.; Chakravarty, J. Leishmaniasis: an update of current pharmacotherapy. *Expert Opin. Pharmacother.* **2012**, *14*, 53-63.
15. Perez-Victoria, F. J.; Sanchez-Canete, M. P.; Seifert, K.; Croft, S. L.; Sundar, S.; Castanys, S.; Gamarro, F. Mechanisms of experimental resistance of *Leishmania* to miltefosine: implications for clinical use. *Drug Resist. Updates* **2006**, *9*, 26-39.

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60
16. Sundar, S.; Singh, A.; Rai, M.; Prajapati, V. K.; Singh, A. K.; Ostyn, B.; Boelaert, M.; Dujardin, J.-C.; Chakravarty, J. Efficacy of Miltefosine in the Treatment of Visceral Leishmaniasis in India After a Decade of Use. *Clin. Infect. Dis.* **2012**, *55*, 543-550.
17. Rijal, S.; Ostyn, B.; Uranw, S.; Rai, K.; Bhattarai, N. R.; Dorlo, T. P. C.; Beijnen, J. H.; Vanaerschot, M.; Decuypere, S.; Dhakal, S. S.; Das, M. L.; Karki, P.; Singh, R.; Boelaert, M.; Dujardin, J.-C. Increasing Failure of Miltefosine in the Treatment of Kala-azar in Nepal and the Potential Role of Parasite Drug Resistance, Reinfection, or Noncompliance. *Clin. Infect. Dis.* **2013**, *56*, 1530-1538.
18. Berman, J. D.; King, M.; Edwards, N. Antileishmanial activities of 2,4-diaminoquinazoline putative dihydrofolate reductase inhibitors. *Antimicrob. Agents Chemother.* **1989**, *33*, 1860-1863.
19. Bhattacharjee, A. K.; Skanchy, D. J.; Jennings, B.; Hudson, T. H.; Brendle, J. J.; Werbovets, K. A. Analysis of stereoelectronic properties, mechanism of action and pharmacophore of synthetic indolo[2,1-b]quinazoline-6,12-dione derivatives in relation to antileishmanial activity using quantum chemical, cyclic voltammetry and 3-D-QSAR CATALYST procedures. *Bioorg. Med. Chem.* **2002**, *10*, 1979-1989.
20. Ram, V. J.; Goel, A.; Verma, M.; Kaul, I. B.; Kapil, A. Latent leishmanicidal activity of quinazolinones and 1,2,4-triazoloquinazolinones. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 2087-2090.
21. Agarwal, K. C.; Sharma, V.; Shakya, N.; Gupta, S. Design and synthesis of novel substituted quinazoline derivatives as antileishmanial agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 5474-5477.

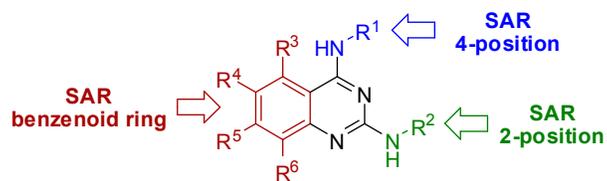
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57
58
59
60
22. Kumar, S.; Shakya, N.; Gupta, S.; Sarkar, J.; Sahu, D. P. Synthesis and biological evaluation of novel 4-(hetero)aryl-2-piperazinoquinazolines as anti-leishmanial and anti-proliferative agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2542-2545.
23. Gilbert, I. H. Inhibitors of dihydrofolate reductase in leishmania and trypanosomes. *BBA-MOL Basis Dis.* **2002**, *1587*, 249-257.
24. Van Horn, K. S.; Vesely, B.; Srivastava, A.; Kyle, D. E.; Manetsch, R. unpublished results.
25. Feng, J.; Zhang, Z.; Wallace, M. B.; Stafford, J. A.; Kaldor, S. W.; Kassel, D. B.; Navre, M.; Shi, L.; Skene, R. J.; Asakawa, T.; Takeuchi, K.; Xu, R.; Webb, D. R.; Gwaltney, S. L., II. Discovery of alogliptin: a potent, selective, bioavailable, and efficacious inhibitor of dipeptidyl peptidase IV. *J. Med. Chem.* **2007**, *50*, 2297-2300.
26. Ife, R. J.; Brown, T. H.; Blurton, P.; Keeling, D. J.; Leach, C. A.; Meeson, M. L.; Parsons, M. E.; Theobald, C. J. Reversible inhibitors of the gastric (H⁺/K⁺)-ATPase. 5. Substituted 2,4-diaminoquinazolines and thienopyrimidines. *J. Med. Chem.* **1995**, *38*, 2763-2773.
27. Kanuma, K.; Omodera, K.; Nishiguchi, M.; Funakoshi, T.; Chaki, S.; Semple, G.; Tran, T.-A.; Kramer, B.; Hsu, D.; Casper, M.; Thomsen, B.; Sekiguchi, Y. Lead optimization of 4-(dimethylamino)quinazolines, potent and selective antagonists for the melanin-concentrating hormone receptor 1. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 3853-3856.
28. Buckner, F. S.; Wilson, A. J. Colorimetric assay for screening compounds against *Leishmania* amastigotes grown in macrophages. *Am. J. Trop. Med. Hyg.* **2005**, *72*, 600-605.

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57
58
59
60
29. Zhu, X.; Pandharkar, T.; Werbovetz, K. Identification of new antileishmanial leads from hits obtained by high-throughput screening. *Antimicrob. Agents Chemother.* **2012**, *56*, 1182-1189.
30. Topliss, J. G. Utilization of operational schemes for analog synthesis in drug design. *J. Med. Chem.* **1972**, *15*, 1006-1011.
31. Rijal, S.; Yardley, V.; Chappuis, F.; Decuypere, S.; Khanal, B.; Singh, R.; Boelaert, M.; De Doncker, S.; Croft, S.; Dujardin, J.-C. Antimonial treatment of visceral leishmaniasis: are current in vitro susceptibility assays adequate for prognosis of in vivo therapy outcome? *Microbes Infect.* **2007**, *9*, 529-535.
32. Hailu, A.; Musa, A.; Wasunna, M.; Balasegaram, M.; Yifru, S.; Mengistu, G.; Hurissa, Z.; Hailu, W.; Weldegebreal, T.; Tesfaye, S.; Makonnen, E.; Khalil, E.; Ahmed, O.; Fadlalla, A.; El-Hassan, A.; Raheem, M.; Mueller, M.; Koummuki, Y.; Rashid, J.; Mbui, J.; Mucee, G.; Njoroge, S.; Manduku, V.; Musibi, A.; Mutuma, G.; Kirui, F.; Lodenyo, H.; Mutea, D.; Kirigi, G.; Edwards, T.; Smith, P.; Muthami, L.; Royce, C.; Ellis, S.; Alobo, M.; Omollo, R.; Kesusu, J.; Owiti, R.; Kinuthia, J. Geographical variation in the response of visceral leishmaniasis to paromomycin in east Africa: a multicentre, open-label, randomized trial. *PLoS Negl Trop Dis* **2010**, *4*, e709.
33. Berman, J. D.; Badaro, R.; Thakur, C. P.; Wasunna, K. M.; Behbehani, K.; Davidson, R.; Kuzoe, F.; Pang, L.; Weerasuriya, K.; Bryceson, A. D. Efficacy and safety of liposomal amphotericin B (AmBisome) for visceral leishmaniasis in endemic developing countries. *Bull. World Health Organ.* **1998**, *76*, 25-32.

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34. El Fadili, A.; Richard, D.; Kündig, C.; Ouellette, M. Effect of polyglutamylation of methotrexate on its accumulation and the development of resistance in the protozoan parasite *Leishmania*. *Biochem. Pharmacol.* **2003**, *66*, 999-1008.
35. Osorio, E.; Aguilera, C.; Naranjo, N.; Marin, M.; Muskus, C. Biochemical characterization of the bifunctional enzyme dihydrofolate reductase-thymidylate synthase from *Leishmania (Viannia)* and its evaluation as a drug target. *Biomedica* **2013**, *33*, 393-401.
36. Donovan, S. F.; Pescatore, M. C. Method for measuring the logarithm of the octanol-water partition coefficient by using short octadecyl-poly(vinyl alcohol) high-performance liquid chromatography columns. *J. Chromatogr. A* **2002**, *952*, 47-61.
37. Zhang, Y.; Clark, J. A.; Connelly, M. C.; Zhu, F.; Min, J.; Guiguemde, W. A.; Pradhan, A.; Iyer, L.; Furimsky, A.; Gow, J.; Parman, T.; El, M. F.; Phillips, M. A.; Kyle, D. E.; Mirsalis, J.; Guy, R. K. Lead optimization of 3-carboxyl-4(1H)-quinolones to deliver orally bioavailable antimalarials. *J. Med. Chem.* **2012**, *55*, 4205-4219.
38. Zornoza, T.; Cano-Cebrian, M. J.; Hipolito, L.; Granero, L.; Polache, A. Evidence of a flip-flop phenomenon in acamprosate pharmacokinetics: an in vivo study in rats. *Biopharm. Drug Dispos.* **2006**, *27*, 305-311.
39. Yanez, J. A.; Remsberg, C. M.; Sayre, C. L.; Forrest, M. L.; Davies, N. M. Flip-flop pharmacokinetics--delivering a reversal of disposition: challenges and opportunities during drug development. *Ther. Deliv.* **2011**, *2*, 643-672.

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40. Yamada, T.; Goto, M.; Punj, V.; Zaborina, O.; Kimbara, K.; Das, G. T. K.; Chakrabarty, A. M. The bacterial redox protein azurin induces apoptosis in J774 macrophages through complex formation and stabilization of the tumor suppressor protein p53. *Infect. Immun.* **2002**, *70*, 7054-7062.
41. Wang, M. Z.; Zhu, X.; Srivastava, A.; Liu, Q.; Sweat, J. M.; Pandharkar, T.; Stephens, C. E.; Riccio, E.; Parman, T.; Munde, M.; Mandal, S.; Madhubala, R.; Tidwell, R. R.; Wilson, W. D.; Boykin, D. W.; Hall, J. E.; Kyle, D. E.; Werbovetz, K. A. Novel arylimidamides for treatment of visceral leishmaniasis. *Antimicrob. Agents Chemother.* **2010**, *54*, 2507-2516.
42. Wang, M. Z.; Saulter, J. Y.; Usuki, E.; Cheung, Y. L.; Hall, M.; Bridges, A. S.; Loewen, G.; Parkinson, O. T.; Stephens, C. E.; Allen, J. L.; Zeldin, D. C.; Boykin, D. W.; Tidwell, R. R.; Parkinson, A.; Paine, M. F.; Hall, J. E. CYP4F enzymes are the major enzymes in human liver microsomes that catalyze the O-demethylation of the antiparasitic prodrug DB289 [2,5-bis(4-amidinophenyl)furan-bis-O-methylamidoxime]. *Drug Metab. Dispos.* **2006**, *34*, 1985-1994.

Table of Contents Graphic



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