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# Highly Substituted Cyclopentane–CMP Conjugates as Potent Sialyltransferase Inhibitors

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# **(5)** Supporting Information

**ABSTRACT:** Sialylconjugates on cell surfaces are involved in many biological events such as cellular recognition, signal transduction, and immune response. It has been reported that aberrant sialylation at the nonreducing end of glycoconjugates and overexpression of sialyltransferases (STs) in cells are correlated with the malignance, invasion, and metastasis of tumors. Therefore, inhibitors of STs would provide valuable leads for the discovery of antitumor drugs. On the basis of the transition



state of the enzyme-catalyzed sialylation reaction, we proposed that the cyclopentane skeleton in its two puckered conformations might mimic the planar structure of the donor (CMP-Neu5Ac) in the transition state. A series of cyclopentane-containing compounds were designed and synthesized by coupling different cyclopentane  $\alpha$ -hydroxyphosphonates with cytidine phosphoramidite. Their inhibitory activities against recombinant human ST6Gal-I were assayed, and a potent inhibitor **481** with a  $K_i$  of 0.028  $\pm$  0.006  $\mu$ M was identified. The results show that the cyclopentanoid-type compounds could become a new type of sialyltransferase inhibitors as biological probes or drug leads.

# INTRODUCTION

Sialic acids (in which the most abundant being Neu5Ac, known as N-acetylneuraminic acid) are nine-carbon  $\alpha$ -keto aldonic acids, typically located at the nonreducing terminus of glycans presented in glycoproteins and glycolipids of vertebrates.<sup>1,2</sup> They are also the components of lipooligosaccharides or capsular polysaccharides of pathogenic bacteria.<sup>3</sup> The sialylconjugates play important roles in a variety of physiological or pathological processes including cell differentiation and signaling, immunological regulation, bacterial and viral infection, cancer metastasis, etc.<sup>4</sup> In the human body, the biosynthesis of sialylconjugates is predominantly controlled by 20 sialyltransferases. It is reported<sup>5</sup> that sialyltransferases overexpression and sialylated antigen overpresentation on cell surfaces are correlated with cancer metastasis and poor prognosis in different types of carcinomas such as human colorectal cancer,<sup>6</sup> malignant glioma,<sup>7</sup> breast carcinoma,<sup>8</sup> cervical carcinoma,<sup>9</sup> melanoma,<sup>10</sup> and leukemia.<sup>11</sup>

Independent of their origins and subtypes, all sialyltransferases share the same donor substrate, cytidine monophosphate N-acetylneuraminic acid (CMP-Neu5Ac), and transfer the sialic acid residue onto the terminal nonreducing site of sugar chains in glycoproteins and glycolipids. Currently, all sialyltransferase inhibitors<sup>12–15</sup> can be classified into donor analogues,<sup>16–21</sup> acceptor analogues,<sup>22–26</sup> bisubstrate analogues,<sup>27–29</sup> and transition-state analogues<sup>30–36</sup> and also some others can be found or modified from natural products.<sup>37–41</sup> Up to now, the most potent sialyltransferase inhibitor in vitro is the transition-state analogue 1*h* ( $K_i$  of 0.029 ± 0.006  $\mu$ M against rat  $\alpha$ (2–6)-sialyltransferase) reported by Schmidt et al. in 2002 (Figure 1A),<sup>35</sup> which is consistent with Pauling's statement in 1946 that inhibitors closely imitating the structure of the transition state in an enzyme-catalyzed process could bind more tightly to the active site of the enzyme than the ground-state substrates.<sup>42</sup> Sialyltransferase inhibitors in vivo such as KI-8110,<sup>16</sup> soyasaponin I,<sup>43</sup> and Lith-O-Asp<sup>44</sup> (Figure 1B) could effectively attenuate the total sialylation on cancer cell surfaces and suppress tumor cell metastasis. Although some advances in sialyltransferase inhibitors have been achieved, it is still highly desirable to seek and develop more efficient sialyltransferase inhibitors to elucidate the biological functions of sialylation and even provide valuable leads for antitumor drug development.

The cyclopentane moiety is widespread in natural products and has been a valuable scaffold in drug discovery.<sup>45</sup> Successful examples include the anti-HIV drug abacavir,<sup>46</sup> the orally active and selective sphingosine 1-phosphate receptor-subtype 1 (S1P1) agonist VPC01091,<sup>47</sup> the experimental antiviral drug peramivir,<sup>48</sup> and so on. Generally, the cyclopentane ring tends to adopt the half-chair or envelope conformation, but it also interconverts between different half-chair and envelope conformations as well as intermediate ones due to the low energy barrier between these conformations (Figure 1C).<sup>49</sup>

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Figure 1. (A) Transition-state analogue inhibitor 1*h*. (B) Sialyltransferase inhibitors applied in vivo. (C) Transition state of sialylation reaction and two puckered conformations of the cyclopentane ring.

According to the proposed  $S_N$ 1-like mechanism of the sialyltransferase-catalyzed sialylation reactions,<sup>50–52</sup> the donor in the transition state may involve the partial dissociation of the CMP moiety and the formation of a trigonal planar oxocarbenium ion (Figure 1C). Therefore, with great flexibility and adaptability of its conformations, the substituted cyclopentane ring might mimic the planar structure of the donor in the transition state and could potentially serve as a sialyltransferase inhibitor. Herein we want to report the synthesis of cyclopentane-merged compounds as sialyltransferase inhibitors and the studies on their structure–activity relationships.

On the basis of the previous studies by Schmidt group<sup>15,31,32,53</sup> and Horenstein group,<sup>34</sup> there are several valuable clues for designing potent transition-state-based sialyltransferase inhibitors: (1) the CMP moiety is very important to maintain the affinity to the enzyme,<sup>53</sup> (2) the NeuSAc residue could be modified or even replaced by other different scaffolds,<sup>31,34,36</sup> (3) a planar structure close to the cleavage site to mimic the conformation of the oxocarbenium ion is fundamental for the inhibitory activity,<sup>31,33</sup> (4) two adjacent negative charge centers are preferred as in CMP-NeuSAc,<sup>31,36</sup> (5) the phosphonate is a better choice than the carboxylate as the negative charge center,<sup>36</sup> and (6) an increased distance between the planar anomeric carbon and the CMP leaving group benefits to mimic the slightly longer C–O bond in the transition state.<sup>31</sup> Thus, in our design (Scheme 1), the Neu5Ac moiety was replaced by cyclopentyl  $\alpha$ hydroxyphosphonates and the effects of different substituents

# Scheme 1. Design of Cyclopentanoid Inhibitors and the Retrosynthetic Analysis



on the cyclopentane ring were investigated. At the 3-position of the cyclopentane ring, the hydrophilic hydroxyl group (3 vs 2) and the hydrophobic benzyloxy group (5 vs 4) were introduced to investigate their inhibitory activities. At the 4-position of the cyclopentane ring, acetylamino group (4 and 5) was chosen as that in the NeuSAc, and the acetylaminomethyl group (6) with one more carbon was introduced to compare the difference. Additionally, the difference of diastereoisomers and the effect of the distance between the cyclopentane ring and the CMP leaving group (7 vs 6) were also considered.

As shown in Scheme 1, the target compounds 2–7 could be prepared from the masked compounds I, which were retrosynthetically disconnected into two units: the cytidine phosphoramidite building block (II) and the substituted cyclopentanoid  $\alpha$ -hydroxyphosphonates (III). Intermediates III could be obtained via transformations starting from cyclopentanemethanol or D-ribose. After the synthetic work was completed, the inhibitory activities of these target molecules against  $\alpha(2-6)$ -sialyltransferase would be evaluated.

#### RESULTS AND DISCUSSION

**1. Synthesis of \alpha-Hydroxyphosphonates.** According to the retrosynthetic analysis, the key part for obtaining target molecules is the construction of  $\alpha$ -hydroxyphosphonates (III) that are derived from differently substituted cyclopentanoid alcohols. At first, as shown in Scheme 2, the commercially





<sup>a</sup>Reagents and conditions: (a) DMSO, oxalyl chloride,  $CH_2Cl_2$ , -72 °C, 2 h, then triethylamine, 1 h; (b) dibenzyl phosphite, triethylamine,  $CH_2Cl_2$ , rt, overnight, 44% over two steps.

available cyclopentanemethanol was converted to the volatile aldehyde by Swern oxidation, which was followed by H-phosphonate addition,<sup>31</sup> affording  $\alpha$ -hydroxyphosphonate 8 as enantiomers in moderate yield (44%).

Because all the substituted cyclopentanoid alcohols could be synthesized from a common intermediate 17, we developed an efficient method to prepare 17 from commercially available Dribose based on the Dieckmann condensation.

As shown in Scheme 3, starting from D-ribose, the known enol ether 11 was obtained according to the previous reports<sup>54,55</sup> with some modifications in 74% overall yield over three steps (see Supporting Information). The ozonolysis of 11 gave the lactone, which was followed by the acidic methanolysis and subsequent Wittig reaction<sup>56</sup> to afford olefin 12 in 65% yield over three steps. The hydrogenation of the double bond in 12 provided compound 13, which was transformed to cyclopentene 14 in 73% yield by the Dieckmann condensation.<sup>57</sup> The reduction of enol 14 by NaBH<sub>4</sub> and subsequent benzoylation of the free hydroxyl group yielded cyclopentane 16 as epimers, which were separated carefully in this step and their absolute configurations were ascertained by NMR analysis (NOESY). The cross-peaks between the proton at  $\delta_{\rm H}$  3.16 (H1) and the proton at  $\delta_{\rm H}$  4.70 (H3) in 16R confirmed that they were cofacially oriented, while in 16S there was no correlation between the proton at  $\delta_{\rm H}$  3.33 (H1) and the proton at  $\delta_{\rm H}$  4.87 (H3) (Figure 2). After treatment of 16 with 1,8diazabicyclo[5.4.0]undec-7-ene (DBU), both diastereoisomers were smoothly converted to unsaturated ester 17 via  $\beta$ -elimination.

With compound 17 in hand, the synthesis of  $\alpha$ -hydroxyphosphonate 20 was carried out (Scheme 4). Compound 17 was subjected to hydrogenation to provide methyl carboxylate 18, which was further reduced to alcohol 19 with LiBH<sub>4</sub><sup>58</sup> in 83% yield. Compound 19 was oxidized by (diacetoxyiodo) benzene (BAIB) and catalytic amount of tetramethylpiperidinooxy (TEMPO) to afford the corresponding aldehyde, which underwent H-phosphonate addition to produce  $\alpha$ -hydroxyphosphonate 20 as enantiomers.

Previous studies demonstrated the importance of a hydrophobic group in the inhibitors,<sup>35</sup> so we decided to synthesize some cyclopentane-containing molecules with hydrophobic substituents (Scheme 5).

Acidic cleavage of the isopropylidene functionality in compound 17 provided diol 21, which was selectively benzylated at the allylic position with Ag2O59 to produce compound 22. The absolute configuration of 22 was confirmed by its NMR analysis (HMBC). In the HMBC spectrum, the methylene protons at  $\delta_{\rm H}$  4.68 (PhCH<sub>2</sub>) showed correlation to the carbon at  $\delta_{\rm C}$  83.0 (C3) and also the proton at  $\delta_{\rm H}$  4.52 (H3) was correlated with the carbon at  $\delta_{\rm C}$  72.7 (PhCH<sub>2</sub>) (Figure 3). Next, selective reduction of the double bond in 22 was performed by using nickel borohydride generated in situ from NiCl<sub>2</sub>·6H<sub>2</sub>O and NaBH<sub>4</sub>,<sup>60</sup> affording two diastereoisomers 23Rand **23S** (dr = 3:1) in 92% yield. The absolute configuration of the newly formed chiral center was assigned by the NMR analysis (NOESY). The cross-peaks between the proton at  $\delta_{\rm H}$ 2.75 (H1) and the proton at  $\delta_{\rm H}$  3.80 (H3) in 23R confirmed that they were cofacially oriented (Figure 3).

Subsequently, the free hydroxyl groups of both diastereoisomers were respectively subjected to tosylation followed by treatment with sodium azide to provide 24*R* and 24*S* in high yields. Under the conditions of thioacetic acid in pyridine,<sup>61,62</sup> the azides 24*R* and 24*S* were smoothly converted to compounds 25*R* and 25*S*. Then by using the similar procedures as the preparation of 20,  $\alpha$ -hydroxyphosphonates 27*R* and 27*S* were easily synthesized.

The preparation of  $\alpha$ -hydroxyphosphonate 34 containing acetylaminomethyl group at the 4-position of the cyclopentane ring was shown in Scheme 6, starting from the major diastereoisomer 23R. After reduction with LiBH<sub>4</sub>, the newly formed primary hydroxyl in 28 was selectively protected with the tert-butyldiphenylsilyl group<sup>63</sup> and the free secondary hydroxyl was tosylated to afford compound 30. Then replacing the tosylate by a cyano group was tried. Using KCN as the reagent, conditions such as different solvents (DMF, CH<sub>3</sub>CN, DMSO), temperatures, and additives (18-crown-6 or 4 Å molecular sieves) were screened. Unfortunately, only elimination products were observed. Switching to Bu<sub>4</sub>NCN as the cyano source, cyanide 31 was finally obtained in 48% yield at 85 °C in toluene along with some elimination byproduct. The cyano group in 31 was then reduced with NaBH<sub>4</sub>/CoCl<sub>2</sub> to amine,<sup>64</sup> which was followed by acetylation to produce compound 32. Removal of the TBDPS group with tetra-nbutylammonium fluoride yielded alcohol 33 quantitatively. Then it was subjected to Swern oxidation followed by treatment with dibenzyl phosphite and NaH to produce  $\alpha$ hydroxyphosphonate 34 as a diastereoisomeric mixture in 67% vield.

#### Scheme 3. Synthesis of the Common Intermediate $17^{a}$



"Reagents and conditions: (a)  $H_2SO_4$ , HCl(g), MeOH/acetone, rt, 8 h, 85%; (b) triphenylphosphine, imidazole, I<sub>2</sub>, toluene, 70 °C, 2 h, 92%; (c) NaH, DMF, 0 °C to rt, overnight, 94%; (d) O<sub>3</sub>,  $CH_2Cl_2/MeOH$  (1:2), -72 °C, 1 h, then dimethyl sulfide, 6 h; (e) *p*-toluenesulfonic acid, MeOH, rt; (f)  $Ph_3P$ =CHCOOMe, THF, rt, overnight, 65% over three steps; (g) Pd/C,  $H_2$ , ethyl acetate, rt, 24 h, 99%; (h) NaH, MeOH, toluene, 0–90 °C, 1 h, 73%; (i) NaBH<sub>4</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub> (2:1), -72 °C, 5 h, 95%; (j) benzoyl chloride, 4-dimethylaminopyridine (DMAP), pyridine, 0 °C to rt, 4 h, 92%; (k) 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), THF, 0 °C to reflux; for 16*R*, 1.5 h, 92%; for 16*S*, 4 h, 81%.



Figure 2. Key NOESY correlations of 16R and 16S.



<sup>a</sup>Reagents and conditions: (a) Pd/C, H<sub>2</sub>, ethyl acetate, rt, 15 h, 92%; (b) LiBH<sub>4</sub>, MeOH, THF, 0 °C, 3 h, 83%; (c) (diacetoxyiodo)benzene, 2,2,6,6-tetramethylpiperidinooxy, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 6 h; (d) dibenzyl phosphite, triethylamine, CH<sub>2</sub>Cl<sub>2</sub>, rt, overnight, 71% over two steps.

Next, the cyclopentanone  $\alpha$ -hydroxyphosphonate **43** was prepared. As shown in Scheme 7, starting from intermediate **17**, the methyl ester functionality was reduced by diisobutylaluminum hydride (DIBAL-H)<sup>65</sup> to afford alcohol **35**, which was followed by tosylation with TsCl/NaOH in ether<sup>66</sup> to give compound **36** in high yield. The tosyl group in **36** was easily replaced by the azido group at room temperature. After extraction, the volatile azide was directly treated under standard hydroboration-oxidation conditions<sup>67</sup> to generate compound **37** with a secondary hydroxyl *trans* to the vicinal protected diol,

Scheme 5. Synthesis of  $\alpha$ -Hydroxyphosphonates 27R and  $27S^{a}$ 



<sup>a</sup>Reagents and conditions: (a) 90% (v/v) acetic acid, 90 °C, 2 h, 85%; (b) Ag<sub>2</sub>O, benzyl bromide, tetrabutylammonium iodide, CH<sub>2</sub>Cl<sub>2</sub>, rt, 6 h, 74%; (c) NaBH<sub>4</sub>, NiCl<sub>2</sub>·6H<sub>2</sub>O (4% in MeOH), MeOH, 0 °C, 2.5 h, 92% (**23R:23S** = 3:1); (d) tosyl chloride, DMAP, triethylamine, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, overnight; (e) NaN<sub>3</sub>, DMF, 90 °C, overnight, **24R** 96% over two steps, **24S** 97% over two steps; (f) thioacetic acid, pyridine, rt, **25R** 36 h, 88%, **25S** 48 h, 86%; (g) LiBH<sub>4</sub>, MeOH, THF, 0 °C, **26R** 1 h, 88%, **26S** 4 h, 98%; (h) (diacetoxyiodo)benzene, 2,2,6,6-tetramethylpiperidinoxy, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 17 h for **26R**, 11 h for **26S**; (i) dibenzyl phosphite, triethylamine, CH<sub>2</sub>Cl<sub>2</sub>, rt, overnight, **27R** 91% over two steps, **27S** 87% over two steps.



Figure 3. Key HMBC correlations of 22 and key NOESY correlations of 23*R* and 23*S*.

# Scheme 6. Synthesis of $\alpha$ -Hydroxyphosphonate 34<sup>*a*</sup>



"Reagents and conditions: (a) LiBH<sub>4</sub>, MeOH, THF, 0 °C, 1.5 h, 98%; (b) *tert*-butyldiphenylchlorosilane, DMAP, triethylamine, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 36 h, 88%; (c) tosyl chloride, DMAP, pyridine/CH<sub>2</sub>Cl<sub>2</sub> (3/1), 0 °C to rt, 24 h, 89%; (d) Bu<sub>4</sub>NCN, toluene, 85 °C, 24 h, 48%; (e) NaBH<sub>4</sub>, CoCl<sub>2</sub>, MeOH, 0 °C, 3 h; (f) acetic anhydride, triethylamine, CH<sub>2</sub>Cl<sub>2</sub>, rt, overnight, 60% over two steps; (g) 1 M tetra-*n*-butylammonium fluoride in THF, THF, rt, overnight, quantitative; (h) DMSO, oxalyl chloride, CH<sub>2</sub>Cl<sub>2</sub>, -72 °C, 2 h, then triethylamine, 1 h; (i) dibenzyl phosphite, NaH, THF, 0 °C to rt, overnight, 67% over two steps.

#### Scheme 7. Synthesis of $\alpha$ -Hydroxyphosphonate 43<sup>a</sup>



"Reagents and conditions: (a) diisobutylaluminum hydride (DIBAL-H), toluene, -78 °C, 4 h, 98%; (b) TsCl, NaOH, ether, 0 °C to rt, 24 h, 97%; (c) NaN<sub>3</sub>, DMF, rt, overnight; (d) 1 M B<sub>2</sub>H<sub>6</sub>·THF, THF, 0 °C, 5 h, then 3 N NaOH, 27% H<sub>2</sub>O<sub>2</sub> (aq), 0–40 °C, overnight, 77% over two steps; (e) BnBr, NaH, DMF, 0 °C to rt, overnight, 87%; (f) 90% (v/v) acetic acid (aq), 70 °C, 5 h, 98%; (g) Im<sub>2</sub>SO<sub>2</sub>, NaH, THF, -20 °C, 45 min, 80%; (h) *t*BuOK, THF, rt, 40 min, 5% H<sub>2</sub>SO<sub>4</sub>, rt, overnight, 90%; (i) thioacetic acid, pyridine, rt, 4 h, 74%; (j) dibenzyl phosphite, NaH, THF, 0 °C to rt, overnight, 88%.



Figure 4. Key NOESY correlations of 37, 43R, 43S, and key HMBC correlations of 42.

whose absolute configuration was assigned by its NMR analysis (NOESY). The cross-peaks between the proton at  $\delta_{\rm H}$  2.18 (H5) and the protons at  $\delta_{\rm H}$  4.40 (H2) and 4.71 (H3) in 37 demonstrated that they were cofacially oriented. In addition, the proton at  $\delta_{\rm H}$  2.51 (OH) was correlated with the proton at  $\delta_{\rm H}$  4.40 (H2), therefore the hydroxyl group was believed to be  $\beta$ -oriented (Figure 4).

Benzylation of 37 and subsequent removal of the isopropylidene group produced diol 39, which was converted

to cyclic sulfate **40** with sulfonyl diimidazole  $(Im_2SO_2)^{68}$  and sodium hydride. Compound **40** was then treated with *t*BuOK to give the vinyl sulfate intermediate, which was tautomerized in acidic conditions.<sup>69</sup> However, it provided **41** instead of the desired ketone **41'** in 90% yield with the carbonyl group on the 2-position. Reductive acetylation of azide **41** with thioacetic acid afforded cyclopentanone **42** in 74% yield, and the absolute configuration was confirmed by the NMR analysis (H–H COSY, HSQC and HMBC). As in the HMBC spectrum, the methylene protons at  $\delta_{\rm H}$  5.12 and 4.67 (PhC $H_2$ ) were correlated with the carbon at  $\delta_{\rm C}$  84.5 (C2), so the doublet proton at  $\delta_{\rm H}$  3.62 (H2) showed that the carbonyl group was on the position near the benzyloxy group (Figure 4). H-Phosphonate addition to ketone 42 produced  $\alpha$ -hydroxyphosphonate 43 as a mixture of two diastereoisomers (R:S =1:1.6, determined by <sup>1</sup>H NMR). The mixture could be partially separated, and their absolute configuration was confirmed by the NMR analysis (NOESY). The cross-peaks between the proton at  $\delta_{\rm H}$  3.54 (OH) and the proton at  $\delta_{\rm H}$  3.84 (H2) in 43*R* demonstrated that they were cofacially oriented, while in 43*S* the proton at  $\delta_{\rm H}$  3.26 (OH) did not show any correlations with the proton at  $\delta_{\rm H}$  3.79 (H2) (Figure 4).

Because of its good inhibitory activity against rat  $\alpha(2-6)$ sialyltransferase, the 3-phenoxybenzyl-derived compound 491 prepared by Schmidt et al.<sup>36</sup> was selected as the control compound for the following biological assay. Thus, 3phenoxybenzaldehyde was transformed to  $\alpha$ -hydroxyphosphonate 44 as enantiomers by treatment with dibenzyl phosphite and triethylamine in THF (Scheme 8).

Scheme 8. Synthesis of  $\alpha$ -Hydroxyphosphonate 44<sup>*a*</sup>



"Reagents and conditions: dibenzyl phosphite, triethylamine, THF, rt, overnight, 84%.

2. Synthesis of Cytidine Phosphoramidite. After finishing the preparation of various  $\alpha$ -hydroxyphosphonates, the benzyl and benzyloxycarbonyl (Cbz) protected cytidine phosphoramidite 46 was obtained based on the previous report.<sup>70</sup> The Cbz-protected cytidine 45 was prepared from cytidine according to the literature procedure.<sup>71</sup> Compound 45 was condensed with benzyloxybis(diisopropylamino)-phosphine, which was prepared from bis(N,N-diisopropylamino)chlorophosphine and benzyl alcohol,<sup>72</sup> affording 46 in high yield (Scheme 9).

**3.** Synthesis of Target Molecules. With  $\alpha$ -hydroxyphosphonates and building block 46 in hand, the coupling reaction was performed (Scheme 10). Condensation of 8 with cytidine phosphoramidite 46 in the presence of *H*-tetrazole and

# Scheme 10. Synthesis of Target Molecules<sup>4</sup>



<sup>a</sup>Reagents and conditions: (a) *H*-tetrazole,  $CH_2Cl_2$ , rt, then *tert*-butyl hydroperoxide, rt; (b) (i) 10% Pd/C, 1,4-cyclohexadiene, DMF, rt, (ii) RP-HPLC, (iii) IR 120 Na<sup>+</sup>; (c) (i) 1 M BCl<sub>3</sub> in heptane,  $CH_2Cl_2$ , 0 °C to rt, (ii) RP-HPLC, (iii) IR 120 Na<sup>+</sup>; (d) (i) 10% Pd/C, 1,4-cyclohexadiene, EtOH, rt, (ii) RP-HPLC, (iii) IR 120 Na<sup>+</sup>.

subsequent oxidation of the phosphite with *tert*-butyl hydroperoxide provided benzyl- and Cbz-protected target molecule **47a** as a mixture of four diastereoisomers. The global deprotection of **47a** was achieved by using 10% Pd/C and 1,4-cyclohexadiene in DMF.<sup>71</sup> After filtration and purification by C18 reverse-phase column chromatography ( $H_2O \rightarrow H_2O/MeOH 5:1$ ), the crude product **2** was obtained in 75% yield. The two diastereoisomers were further separated by preparative RP-HPLC, converted to their corresponding sodium salt by ion-exchange (IR 120 Na<sup>+</sup>), and lyophilized from water, yielding the final products **2s** and **2l** as white powder (**s** stands for the component with a longer retention time) (Table 1, entry 1).

Following the same procedure as the preparation of 47a, masked target molecules 47b-g were successfully synthesized. To obtain products 3s and 3l, the isopropylidene, benzyl, and Cbz protective groups in 47b were all removed smoothly by treatment with BCl<sub>3</sub> in CH<sub>2</sub>Cl<sub>2</sub> (Table 1, entry 2).<sup>73,74</sup> The full deprotection of the Cbz and benzyl groups in 47c was realized by catalytic transfer hydrogenation in DMF, affording 4S in 79% yield (Table 1, entry 3). When the solvent was changed from DMF to EtOH,<sup>75</sup> the reaction showed some selectivity between the ether benzyl and ester benzyl, thus compound 5S was obtained in moderate yield (Table 1, entry 3). Finally,



"Reagents and conditions: (a) BnOH, triethylamine, ether, 0 °C to rt, 1 h; (b) H-tetrazole, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 4 h, 92% over two steps.

Table 1. Coupling of Different  $\alpha$ -Hydroxyphosphonates and the Deprotection



 $^{a}$ The data are the isolated yields of the deprotection procedures.

using the same procedures mentioned above, compounds 47d– g were transformed to their corresponding target molecules.

**4. Biological Assay.** Measurement of the inhibition data and kinetic constants of  $K_i$  against recombinant human ST6Gal-I (aa 44-406)<sup>76</sup> was carried out based on the UV/ RP-HPLC method developed by Schmidt group<sup>30</sup> with some modifications. CMP-NeuSAc and *p*-nitrophenyl LacNAc were used as the donor and acceptor, and the product trisaccharide was detected by analytical RP-HPLC at 300 nm. Compound **491** with known  $K_i^{36}$  was selected as the control (Figure 5).

In our assay, when the acceptor *p*-nitrophenyl LacNAc was set at 1 mM, the  $K_m$  for CMP-Neu5Ac was determined to be

41.63  $\pm$  6.66  $\mu$ M (see Supporting Information, Figure S1), which is comparable to the reported  $K_{\rm m}$  data ranging from 4.77<sup>41</sup> to 18  $\mu$ M.<sup>77</sup> The inhibition rates of all the target compounds were first obtained at 10  $\mu$ M level, then the  $K_i$  values were determined for the inhibitors whose inhibitory rates are superior to 50%. The Lineweaver–Burk double reciprocal plots as well as comparison among different inhibition modes of the inhibitors in GraphPad Prism 6 suggest competitive inhibition of the inhibitors (see Supporting Information,  $K_i$  data in Figures S2–S10). As shown in Table 2, the nonsubstituted cyclopentyl inhibitor **2***s* exhibits about 7-fold stronger affinity to the enzyme rhST6Gal-I than the natural



**Figure 5.** Enzymatic sialylation of acceptor *p*-nitrophenyl LacNAc and the structure of control compound **49***l*.

Table 2. Affinity of CMP-Neu5Ac  $(K_m)$  to Recombinant Human ST6Gal-I and Inhibition Data of the Cyclopentanoid Inhibitors<sup>*a*</sup>

compd $[\alpha]$	inhibition at 10 $\mu M$ (%)	$K_{\rm m}$ or $K_{\rm i}~(\mu{ m M})$	$K_{\rm m}/K_{\rm i}$
CMP-Neu5Ac		41.63 ± 6.66	
<b>2s</b> (+17.0)	43.25	$5.852 \pm 0.908$	7.1
<b>2l</b> (+0.9)	12.82		
<b>3s</b> (+6.7)	30.99		
3l (+2.9)	12.98		
<b>4Ss</b> (+4.0)	50.30	4.314 ± 0.968	9.6
4Sl (-3.9)	9.99		
5Ss (-1.7)	63.86	$1.629 \pm 0.285$	25.6
<b>5Sl</b> (+0.5)	75.29	$0.436 \pm 0.117$	95.5
5Rs (+0.7)	65.87	$1.510 \pm 0.334$	27.6
5Rl (-3.7)	22.08		
<b>6s</b> (-3.6)	70.83	$0.865 \pm 0.141$	48.1
<b>6</b> <i>l</i> (-7.1)	46.27		
<b>48</b> <i>s</i> (-22.2)	74.13	$0.477 \pm 0.087$	87.3
<b>48</b> <i>l</i> (-11.5)	97.50	$0.028 \pm 0.006$	1486.8
49 <i>l</i> <sup>36</sup>	98.12	$0.019 \pm 0.005$	2191.0
<sup>4</sup> For details of the procedures, see Supporting Information.			

substrate CMP-NeuSAc, indicating that the cyclopentane ring could mimic the NeuSAc residue or its planar structure in the transition state. It seems that the hydrophilic hydroxyl group on the cyclopentane ring does not contribute to the inhibition and even results in erosion of the activity (3s, 3l vs 2s, 2l). Comparing 4Ss and 4Sl with 5Ss and 5Sl, the hydrophobic benzyl group on the 3-position of the cyclopentane ring is really appreciated, which may possess some useful interaction with the hydrophobic area in the catalytic site of the enzyme and it is also consistent with the previous results reported by Schmidt et al. in 2002.<sup>35</sup> On the 4-position of the cyclopentane ring, adding one more carbon is somewhat useful for the activity (6s, 6l vs 5Rs, 5Rl). As 5Ss and 5Sl are better inhibitors than 5Rs

and **5***Rl*, the configuration of the C-1 position of the cyclopentane ring is also an important factor, and the *S* isomer might be a better choice. To our delight, the cyclopentanoid inhibitor **48***l* that is the surrogate of the designed target molecule 71 shows the best inhibitory activity ( $K_i = 0.028 \pm 0.006 \ \mu$ M), which is 15-fold stronger than the best cyclopentanemethyl-containing inhibitor **5***Sl* and is almost equivalent to the control inhibitor **49***l* ( $K_i = 0.019 \pm 0.005 \ \mu$ M). Perhaps the cyclopentane ring with a quaternary carbon center could adopt a more planar conformation, thus better mimicking the NeuSAc residue in the transition state.

**5. Configuration Assignment of 48***l*. Since the new stereogenic center in 43*R* was assigned, the condensation of the pure compound 43*R* with 46 was carried out, which was followed by the oxidation under the same conditions as mentioned above to produce 47*fR*. After selective removal of the ester benzyl groups in 47*fR*, purification, and conversion to the corresponding sodium salt, the final product 48*R* was obtained. It was found that the <sup>1</sup>H NMR spectrum of 48*R* was the same as that of 48*l*. Therefore, the absolute configuration of the new stereogenic center in 48*l* was determined to be *R*.

## CONCLUSIONS

In conclusion, with the proposal that the comformations of cyclopentane ring may mimic the planar character of the oxocarbenium ion in the transition state of the enzymecatalyzed sialylation, we designed and synthesized 14 cyclopentane-containing compounds by coupling the highly substituted cyclopentane  $\alpha$ -hydroxyphosphonates with cytidine phosphoramidite. The synthetic compounds were evaluated against the sialyltransferase (recombinant human ST6Gal-I), and several compounds with inhibitory activities were identified. Especially, among them, a potent transition statebased inhibitor 48l with  $K_i$  of 0.028  $\pm$  0.006  $\mu$ M was obtained. These results show that the cyclopentane moiety could be introduced as a key structural motif for the discovery of new sialyltransferase inhibitors. Further structural modifications and their applications in the development of antitumor leads and synthesis of biological probes are now under investigation.

# EXPERIMENTAL SECTION

General. All chemicals purchased were reagent grade and used without further purification unless otherwise stated. Dichloromethane  $(CH_2Cl_2)$  and pyridine were distilled over calcium hydride  $(CaH_2)$ . N,N-Dimethylformamide (DMF) was stirred with CaH<sub>2</sub> and distilled under reduced pressure. Methanol and ethanol were distilled from magnesium and iodine. Toluene, tetrahydrofuran (THF), and ether were distilled over sodium/benzophenone. All reactions were carried out under anhydrous conditions using flame-dried glassware with freshly distilled solvents unless otherwise noted. Reactions were monitored by analytical thin-layer chromatography (TLC) on silica gel 60 F<sub>254</sub> precoated on aluminum plates (E. Merck). Spots were detected under UV (254 nm) light and/or by staining with acidic ceric ammonium molybdate. Solvents were evaporated under reduced pressure and below 40 °C (water bath). Column chromatography was performed on silica gel (200-300 mesh) or C18 reverse phase silica gel. <sup>1</sup>H NMR, <sup>13</sup>C NMR, <sup>31</sup>P NMR, and 2D NMR spectra were recorded on an Avance III Bruker-600 or a Varian INOVA-500 or an Avance DRX Bruker-400 spectrometer at 25 °C. Chemical shifts (in ppm) were calibrated with the solvent residual peak. Mass spectra were recorded by using a Waters Xevo G2 Q-TOF spectrometer. Elemental analysis data were recorded on a Vario EL-III elemental analyzer. Optical rotation was determined by a Rudolph Research Analytical Autopol IV polarimeter. Preparative HPLC was performed with Waters 2545 autopurification system and detected at 254 nm.

Columns: (A) SunFire Prep C18 OBD (Waters, 5  $\mu$ m, 19 mm × 150 mm), (B) XBridge Prep C18 OBD (Waters, 5  $\mu$ m, 19 mm × 150 mm). The purity of all the final compounds was determined to be  $\geq$ 95% (see Supporting Information Table S1 ) by HPLC analysis, which was performed on an Agilent 1260 Infinity system equipped with VWD detector and the data were collected at 254 nm.

Dibenzyl  $\alpha$ -Hydroxycyclopentylmethylphosphonate (8). To a solution of DMSO (176.2 mg, 160 µL, 2.25 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6.0 mL) was added oxalyl chloride (253.2 mg, 174 µL, 2.00 mmol) dropwise at -78 °C under argon atmosphere. After stirring at the same temperature for 30 min, a solution of cyclopentanemethanol (50.0 mg, 54  $\mu$ L, 0.50 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL) was slowly added. After 2 h, triethylamine (455.4 mg, 626  $\mu$ L, 4.50 mmol) was added and the reaction mixture was stirred at -78 °C for another 1 h. Then the mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O. The combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (3.0 mL), and dibenzyl phosphite (261.1 mg, 0.22 mL, 0.90 mmol, 90%) and triethylamine (101.8 mg, 0.14 mL, 1.00 mmol) were added. The reaction mixture was stirred overnight and concentrated. The crude product was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 3:1) to give 8 (78.7 mg, 44% yield) as white solid.  $R_f = 0.37$  (petroleum ether/ethyl acetate 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.33–7.27 (m, 10H), 5.11–5.01 (m, 4H), 4.09 (dd, J = 6.8, 4.4 Hz, 1H), 3.82 (td, J = 7.2, 4.8 Hz, 1H), 2.32-2.24 (m, 1H), 1.90–1.87 (m, 1H), 1.75–1.70 (m, 1H), 1.60–1.38 (m, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  136.52 (d, J = 2.0 Hz), 136.46 (d, J = 2.0 Hz), 128.6, 128.4, 128.0, 71.5 (d, J = 156.0 Hz), 68.0 (d, J = 7.0 Hz), 67.9 (d, J = 7.0 Hz), 41.4 (d, J = 1.0 Hz), 29.4 (d, J = 6.0 Hz), 28.8 (d, J = 10.0 Hz), 25.6, 25.3. <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$ 25.96. HRMS (ESI) calcd for C<sub>20</sub>H<sub>26</sub>O<sub>4</sub>P [M + H]<sup>+</sup>, 361.1569; found, 361.1564.

Dimethyl (4S,5S)-4,5-Isopropylidenedioxy-2-hexene-1,6-dioate (12). To a solution of enol ether 11 (see the Supporting Information) (10.18 g, 54.67 mmol) in MeOH (100 mL) and CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was bubbled with  $O_3$  at -72 °C until the starting material was completely consumed. Then the O<sub>3</sub> dissolved in the reaction mixture was replaced by argon, and the reaction was quenched with dimethyl sulfide (12 mL) at the same temperature for 6 h. After the reaction mixture was concentrated, the residue was dissolved in MeOH (470 mL) and followed by adding p-toluenesulfonic acid (470.1 mg, 2.73 mmol). Then the reaction was stirred at room temperature until the starting material was completely consumed and quenched with triethylamine (0.46 mL). The solvent was evaporated under reduced pressure, and the residue was dissolved in THF (150 mL) under argon atmosphere.  $Ph_3P$ =CHCOOMe<sup>78,79</sup> (21.93 g, 65.60 mmol) was added, and the reaction was stirred overnight at room temperature. The reaction mixture was concentrated and the residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 8:1) to give 12 (8.72 g, 65% yield) as colorless oil. The Z-isomer:  $R_f = 0.59$ (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25} = +195.0$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  6.20 (dd, J = 11.5, 7.0 Hz, 1H, H3), 5.95 (dd, J = 11.5, 1.5 Hz, 1H, H2), 5.85 (td, J = 7.5, 2.0 Hz, 1H, H4), 4.94 (d, J = 7.0 Hz, 1H, H5), 3.76 (s, 3H, OMe), 3.66 (s, 3H, OMe), 1.64 (s, 3H, CH<sub>3</sub>), 1.42 (s, 3H, CH<sub>3</sub>). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$ 170.1, 165.7, 143.7, 122.3, 111.6, 77.1, 75.1, 51.8, 51.6, 26.8, 25.5. HRMS (ESI) calcd for  $C_{11}H_{16}O_6Na$  [M + Na]<sup>+</sup>, 267.0845; found, 267.0841. The *E*-isomer:  $R_f = 0.49$  (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25} = +44.6$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 6.82 (dd, J = 15.6, 5.2 Hz, 1H, H3), 6.15 (dd, J = 15.6, 1.2 Hz, 1H, H2), 4.97 (ddd, J = 6.8, 5.6, 1.2 Hz, 1H, H4), 4.79 (d, J = 7.2 Hz, 1H, H5), 3.74 (s, 3H, OMe), 3.69 (s, 3H, OMe), 1.66 (s, 3H, Me), 1.42 (s, 3H, Me).  $^{13}\mathrm{C}$  NMR (100 MHz, CDCl\_3)  $\delta$  169.4, 166.2, 140.9, 123.2, 112.1, 77.6, 76.5, 52.2, 51.9, 27.0, 25.7. HRMS (ESI) calcd for  $C_{11}H_{16}O_6Na [M + Na]^+$ , 267.0845; found, 267.0842.

Dimethyl (25,35)-2,3-lsopropylidenedioxy-1,6-hexanedioate (13). To a solution of 12 (9.27 g, 37.95 mmol) in dry ethyl acetate (30 mL) was added Pd/C (404.4 mg, 0.38 mmol, 10%), and the reaction mixture was stirred at room temperature for 24 h under hydrogen atmosphere (4 atm). Then the mixture was filtered through a pad of

Celite and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 8:1) to give **13** (9.25 g, 99% yield) as colorless oil.  $R_{\rm f}$  = 0.40 (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25}$  = +1.2 (c = 0.8, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.62 (d, J = 6.5 Hz, 1H), 4.38–4.34 (m, 1H), 3.77 (s, 3H), 3.68 (s, 3H), 2.56–2.43 (m, 2H), 1.94–1.88 (m, 1H), 1.74–1.66 (m, 1H), 1.59 (s, 3H), 1.37 (s, 3H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  173.2, 170.4, 110.6, 77.0, 76.5, 52.0, 51.6, 30.5, 27.0, 25.6, 25.5. HRMS (ESI) calcd for C<sub>11</sub>H<sub>18</sub>O<sub>6</sub>Na [M + Na]<sup>+</sup>, 269.1001; found, 269.0993.

Methyl (35,45)-2-Hydroxy-3,4-isopropylidenedioxy-1-cyclopentene-1-carboxylate (14). To a solution of 13 (3.86 g, 15.67 mmol) in toluene (77 mL) were added NaH (1.88 g, 47.00 mmol, 60% in mineral oil) in portions and methanol (0.4 mL) at 0 °C under argon atmosphere. After 10 min, the mixture was stirred at 90 °C for about 1 h. Then the reaction mixture was concentrated and diluted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL). After the pH of the solution was adjusted to 5-6 with 1 N HCl, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> then the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The crude product was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 10:1) to give 14 (2.44 g, 73% yield) as colorless oil.  $R_f = 0.44$  (petroleum ether/ethyl acetate = 2/1);  $[\alpha]_{D}^{25} = +37.8$  (c = 0.8, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.93 (s, 1H, OH), 4.97 (d, I = 6.0 Hz, 1H, H3), 4.69 (t, J = 5.2 Hz, 1H, H4), 3.79 (s, 3H, OMe), 2.70 (dd, J = 15.6 Hz, 5.2 Hz, 1H, H5 $\alpha$ ), 2.62 (d, I = 16.0 Hz, 1H, H5 $\beta$ ), 1.45 (s, 3H, CH<sub>2</sub>), 1.37 (s, 3H, CH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.9, 169.8, 111.4, 99.9, 81.5, 75.0, 51.6, 32.3, 27.4, 25.6. Anal. Calcd for C<sub>10</sub>H<sub>14</sub>O<sub>5</sub>: C, 56.09; H, 6.57. Found: C, 56.11; H, 6.50. HRMS (ESI) calcd for  $C_{10}H_{15}O_5 [M + H]^+$ , 215.0919; found, 215.0920.

Methyl (2R,3R,4S)-2-Hydroxy-3,4-isopropylidenedioxy-cyclopentane-1-carboxylate (15). To a solution of 14 (1.17 g, 5.46 mmol) in methanol (30 mL) and CH<sub>2</sub>Cl<sub>2</sub> (15 mL) was added NaBH<sub>4</sub> (0.62 g, 16.38 mmol) in portions at -72 °C under argon atmosphere. After stirring at the same temperature for 5 h, 1 N HCl was added to adjust the pH to 7. Then the mixture was concentrated and extracted with CH<sub>2</sub>Cl<sub>2</sub>, the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The crude product was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 10:1 to 4:1) to give 15 (1.12 g, 95% yield) as colorless oil.  $R_f = 0.31$ (petroleum ether/ethyl acetate 2:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 4.66 (t, J = 5.2 Hz, 0.8H, H4'), 4.62 (td, J = 6.0, 2.4 Hz, 1H, H4), 4.51 (t, J = 5.6 Hz, 0.8H, H3'), 4.49 (dd, J = 6.4, 5.2 Hz, 1H, H3), 4.12 (dt, J = 7.6, 5.2 Hz, 1H, H2), 4.04 (td, J = 9.6, 5.6 Hz, 0.8H, H2'), 3.91 (d, J = 7.6 Hz, 1H, OH), 3.74 (s, 2.4H, OMe'), 3.73 (s, 3H, OMe), 2.86-2.78 (m, 1.8H, H1 and H1'), 2.60 (d, J = 10.0 Hz, 0.8H, OH'), 2.42 (ddd, J = 14.4, 6.0, 2.8 Hz, 1H, H5α), 2.07 (dd, J = 14.4, 6.4 Hz, 0.8H,  $H5\alpha'$ ), 1.92 (ddd,  $J = 14.0, 7.6, 6.0 \text{ Hz}, 1\text{H}, H5\beta$ ), 1.74 (ddd, J = 14.4, 14.412.8, 4.8 Hz, 0.8H, H5 $\beta'$ ), 1.50 (s, 2.4H, Me'), 1.45 (s, 3H, Me), 1.34 (s, 2.4H, Me'), 1.33 (s, 3H, Me).  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 174.3, 173.0, 112.1, 110.5, 79.4, 78.6, 78.2, 78.0, 76.0, 72.8, 52.0, 51.8, 47.5, 45.4, 32.0, 30.8, 25.9, 25.3, 24.0. Anal. Calcd for C10H16Os: C, 55.55; H, 7.46. Found: C, 55.40; H, 7.21. HRMS (ESI) calcd for  $C_{10}H_{16}O_5Na [M + Na]^+$ , 239.0890; found, 239.0891.

Methyl (2R,3S,4S)-2-Benzoyloxy-3,4-isopropylidenedioxy-cyclopentane-1-carboxylate (16). To a solution of 15 (5.36 g, 24.79 mmol) in pyridine (80 mL) were added DMAP (0.60 g, 4.96 mmol) and benzoyl chloride (5.20 g, 4.3 mL, 37.18 mmol) dropwise at 0 °C under argon atmosphere. After stirring at room temperature for 4 h, the reaction mixture was concentrated and partitioned between CH<sub>2</sub>Cl<sub>2</sub> and saturated NaHCO<sub>3</sub> solution. The combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 10:1 to 8:1) to give 16R (3.91 g, 49%yield) as white solid and 16S (3.44 g, 43% yield) as colorless oil. 16R:  $R_{\rm f} = 0.29$  (petroleum ether/ethyl acetate 2:1);  $[\alpha]_{\rm D}^{25} = +45.0$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.08–8.06 (m, 2H, Ar), 7.58-7.54 (m, 1H, Ar), 7.46-7.42 (m, 2H, Ar), 5.57 (t, J = 4.4 Hz, 1H, H2), 4.77 (td, J = 6.8 Hz, 4.8 Hz 1H, H4), 4.70 (dd, J = 6.8, 4.4 Hz, 1H, H3), 3.62 (s, 3H, OMe), 3.16 (ddd, J = 11.6, 7.6, 4.4 Hz, 1H, H1), 2.56 (ddd, J = 14.0, 11.6, 4.8 Hz, 1H, H5 $\alpha$ ), 2.38 (dt, J = 14.0, 7.2

Hz, 1H, H5β), 1.29 (s, 3H, CH<sub>3</sub>), 1.21 (s, 3H, CH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 170.6, 165.5, 133.2, 130.0, 129.9, 128.5, 114.4, 80.6, 78.6, 73.5, 52.1, 46.9, 31.6, 25.4, 24.8. Anal. Calcd for  $C_{17}H_{20}O_6$ : C, 63.73; H, 6.29. Found: C, 63.74; H, 6.10. HRMS (ESI) calcd for  $C_{17}H_{20}O_6$ Na [M + Na]<sup>+</sup>, 343.1158; found, 343.1164. **165**:  $R_f = 0.44$  (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25} = +114.6$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.08–8.06 (m, 2H, Ar), 7.58–7.54 (m, 1H, Ar), 7.46–7.42 (m, 2H, Ar), 5.10 (dd, J = 10.8, 5.6 Hz, 1H, H2), 4.87 (t, J = 5.6 Hz, 1H, H3), 4.72 (t, J = 4.8 Hz, 1H, H4), 3.70 (s, 3H, OMe), 3.33 (ddd, J = 12.8, 10.4, 6.8 Hz, 1H, H1), 2.22 (dd, J = 14.4, 6.8 Hz, 1H, H5α), 1.86 (ddd, J = 14.0, 12.8, 4.8 Hz, 1H, H5β), 1.46 (s, 3H, CH<sub>3</sub>), 1.29 (s, 3H, CH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 173.4, 166.0, 133.2, 130.0, 129.9, 128.4, 111.0, 78.4, 77.3, 76.9, 52.2, 44.5, 31.9, 26.1, 24.4. HRMS (ESI) calcd for  $C_{17}H_{21}O_6$  [M + H]<sup>+</sup>, 321.1333; found, 321.1338.

Methyl (3R,4S)-3,4-Isopropylidenedioxy-1-cyclopentene-1-carboxylate (17). To a solution of 16R (3.89 g, 12.14 mmol) in THF (40 mL) was added DBU (3.67 g, 3.6 mL, 24.29 mmol) at 0 °C under argon atmosphere. After refluxing for 1.5 h, the reaction mixture was concentrated and partitioned between ethyl acetate and saturated NaHCO3 solution. The combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ ethyl acetate = 9:1) to give 17 (2.22 g, 92% yield) as colorless oil.  $R_f$  = 0.53 (petroleum ether/ethyl acetate = 2/1). <sup>1</sup>H NMR (400 MHz,  $CDCl_3$ )  $\delta$  6.63 (dd, J = 3.6, 1.6 Hz, 1H), 5.19–5.17 (m, 1H), 4.82– 4.80 (m, 1H), 3.76 (s, 3H), 2.79-2.77 (m, 2H), 1.40 (s, 3H), 1.35 (s, 3H). The spectroscopic data coincide with the previous report. Compound 17 was also prepared from 16S in the same manner as described above. Quantities: 16S (3.15 g, 9.83 mmol), DBU (2.96 g, 2.9 mL, 19.66 mmol), THF (40 mL), 4 h, affording 17 (1.58 g, 81% vield).

*Methyl c-3,c-4-lsopropylidenedioxy-r-1-cyclopentanecarboxylate* (18). To a solution of 17 (115.7 mg, 0.58 mmol) in dry ethyl acetate (5 mL) was added Pd/C (15.6 mg, 0.02 mmol, 10%), and the reaction mixture was stirred at room temperature for 15 h under hydrogen atmosphere (4 atm). Then the mixture was filtered through a pad of Celite and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 6:1) to give 18 (108.1 mg, 92% yield) as colorless oil.  $R_f$  = 0.38 (petroleum ether/ethyl acetate 2:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.65–4.61 (m, 2H), 3.70 (s, 3H), 2.81 (tt, *J* = 8.4, 2.8 Hz, 1H), 2.47 (dd, *J* = 14.4, 2.8 Hz, 2H), 1.91–1.85 (m, 2H), 1.38 (s, 3H), 1.27 (s, 3H). The spectroscopic data coincide with the previous report.<sup>81</sup>

*c*-3,*c*-4-Isopropylidenedioxy-*r*-1-cyclopentylmethanol (**19**). To a solution of **18** (196.5 mg, 0.98 mmol) in THF (10 mL) were added LiBH<sub>4</sub> (85.3 mg, 3.92 mmol) and methanol (0.2 mL) at 0 °C under argon atmosphere. After stirring at the same temperature for 3 h, 1 N HCl was added to adjust the pH to 7. Then the mixture was extracted with ethyl acetate, and the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 2:1) to give **19** (140.9 mg, 83% yield) as colorless oil.  $R_f = 0.38$  (petroleum ether/ethyl acetate 1:2). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.66–4.65 (m, 2H), 3.70 (d, *J* = 6.8 Hz, 2H), 2.29–2.20 (m, 1H), 1.93–1.79 (m, 4H), 1.70 (brs, 1H), 1.49 (s, 3H), 1.30 (s, 3H). The spectroscopic data coincide with the previous report.<sup>82</sup>

Dibenzyl  $\alpha$ -Hydroxy-(c-3,c-4-isopropylidenedioxy-r-1-cyclopentylmethyl)-phosphonate (**20**). To a solution of **19** (88.6 mg, 0.51 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) were added TEMPO (8.0 mg, 0.05 mmol) and BAIB (199.7 mg, 0.62 mmol) at 0 °C under argon atmosphere. After stirring at room temperature for 6 h, the reaction was quenched with saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution and partitioned between CH<sub>2</sub>Cl<sub>2</sub> and saturated NaHCO<sub>3</sub> solution. The combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (4 mL), and dibenzyl phosphite (297.2 mg, 250  $\mu$ L, 1.02 mmol, 90%) and triethylamine (103.2 mg, 142  $\mu$ L, 1.02 mmol) were added. The reaction mixture was stirred overnight and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 1:2) to give **20** (157.2 mg, 71% yield) as colorless oil.  $R_{\rm f}$  = 0.19 (petroleum ether/ethyl acetate 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33–7.30 (m, 12H), 5.14–5.01 (m, 4.8H), 4.63–4.57 (m, 2.4H), 4.06 (dd, *J* = 12.8, 5.6 Hz, 1H), 3.86 (dd, *J* = 11.6, 6.0 Hz, 0.2H), 3.37 (dd, *J* = 8.8, 6.0 Hz, 1H), 3.29 (t, *J* = 5.2 Hz, 0.2H), 2.67–2.58 (m, 0.2H), 2.55–2.44 (m, 1H), 2.18–2.13 (m, 1H), 2.10–2.09 (m, 0.2H), 2.01–1.88 (m, 3.6H), 1.47 (s, 3H), 1.40 (s, 0.6H), 1.29 (s, 3H), 1.27 (s, 0.6H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  25.60–25.36 (m), 25.00–24.77 (m). HRMS (ESI) calcd for C<sub>23</sub>H<sub>30</sub>O<sub>6</sub>P [M + H]<sup>+</sup>, 433.1780; found, 433.1771.

*Methyl* (3*R*,4*S*)-3,4-*Dihydroxy-1-cyclopentene-1-carboxylate* (21). Compound 17 (100.1 mg, 0.50 mmol) was dissolved in 90% (v/v) acetic acid solution (2 mL) and stirred at 90 °C for 2 h. After the starting material was completely consumed, the mixture was concentrated and the residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 2:3) to give 21 (67.6 mg, 85% yield) as white solid.  $R_f = 0.36$  (petroleum ether/ethyl acetate 1:5);  $[\alpha]_{25}^{D5} = -66.8 (c = 0.9, CHCl_3)$ . <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.64 (dd, J = 3.6, 2.0 Hz, 1H), 4.71 (brs, 1H), 4.39 (t, J = 5.6 Hz, 1H), 3.76 (s, 3H), 3.65 (brs, 1H), 3.42 (brs, 1H), 2.81–2.74 (m, 1H), 2.64–2.59 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  165.6, 141.8, 135.8, 76.6, 71.3, 52.0, 38.8. Anal. Calcd for  $C_7H_{10}O_4$ : C, 53.16; H, 6.37. Found: C, 53.25; H, 6.49. HRMS (ESI) calcd for  $C_7H_{14}NO_4$ [M + NH<sub>4</sub>]<sup>+</sup>, 176.0917; found, 176.0918.

*Methyl* (3R,4S)-3-Benzyloxy-4-hydroxy-cyclopentene-1-carboxylate (22). To a solution of 21 (220.2 mg, 1.39 mmol) in  $CH_2Cl_2$ (80 mL) were added Ag<sub>2</sub>O (354.3 mg, 1.53 mmol), BnBr (285.6 mg, 0.2 mL, 1.67 mmol), and TBAI (102.7 mg, 0.28 mmol) under argon atmosphere, and the reaction bottle was protected from the daylight. After stirring at room temperature for 6 h, the mixture was filtered through a pad of Celite and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 4:1) to give 22 (255.0 mg, 74% yield) as colorless oil.  $R_f = 0.41$ (petroleum ether/ethyl acetate 1:1);  $[\alpha]_D^{28} = -43.3$  (c = 1.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.40–7.31 (m, 5H, Ar), 6.68 (dd, J = 3.6, 2.0 Hz, 1H, H2), 4.68 (s, 2H, PhCH<sub>2</sub>), 4.52 (ddd, J = 5.2, 3.2, 1.2 Hz, 1H, H3), 4.43 (td, J = 5.6, 3.2 Hz, 1H, H4), 3.75 (s, 3H, OMe), 2.80 (brs, 1H, OH), 2.73 (dddd, J = 16.8, 5.6, 2.4, 1.2 Hz, 1H, H5 $\alpha$ ), 2.65 (ddt, J = 17.2, 2.8, 1.6 Hz, 1H, H5 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 165.3 (CO<sub>2</sub>Me), 139.2 (C2), 137.4 (C1), 137.1 (Ar), 128.8 (Ar), 128.3 (Ar), 128.1 (Ar), 83.0 (C3), 72.7 (PhCH<sub>2</sub>), 70.7 (C4), 51.9 (OMe), 39.1 (C5). HRMS (ESI) calcd for C14H20NO4[M + NH<sub>4</sub>]<sup>+</sup>, 266.1387; found, 266.1383.

Methyl (3R,4S)-3-Benzyloxy-4-hydroxy-cyclopentane-1-carboxylate (23). To a solution of 22 (531.7 mg, 2.14 mmol) in methanol (25 mL) was added a solution of NiCl<sub>2</sub>·6H<sub>2</sub>O in methanol (21 mL, 3.64 mmol, 4% in methanol) at 0 °C under argon atmosphere. After 30 min, NaBH<sub>4</sub> (323.8 mg, 8.56 mmol) was added in portions. After stirring at 0 °C for another 2 h, the reaction was quenched with a drop of acetic acid and the mixture was filtered through a pad of Celite and concentrated. The residue was partitioned between ethyl acetate and saturated NaHCO<sub>3</sub> solution, and the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 5:1) to give 23R (371.7 mg, 69%) and 23S (121.4 mg, 23%) both as colorless oil. 23R:  $R_{\rm f} = 0.40$ (petroleum ether/ethyl acetate 1:1);  $[\alpha]_D^{25} = -23.1$  (c = 0.6, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.37–7.27 (m, 5H, Ar), 4.63 (d, J = 11.6 Hz, 1H, PhC $H_2$ ), 4.52 (d, J = 11.6 Hz, 1H, PhC $H_2$ ), 4.08 (dt, J =10.0, 5.2 Hz, 1H, H4), 3.80 (td, J = 6.8, 4.8 Hz, 1H, H3), 3.68 (s, 1H, OMe), 2.75 (ddd, J = 16.4, 9.2, 7.2 Hz, 1H, H1), 2.63 (d, J = 5.6 Hz, 1H, OH), 2.21 (td, J = 14.0, 7.6 Hz, 1H, H2 $\alpha$ ), 2.15–2.02 (m, 3H,  $H2\beta$ ,  $H5\alpha$ ,  $H5\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  175.9 (CO<sub>2</sub>Me), 137.9 (Ph), 128.6 (Ph), 127.9 (Ph), 127.8 (Ph), 80.2 (C3), 72.1 (C4), 71.4 (PhCH<sub>2</sub>), 52.1 (OMe), 38.5 (C1), 34.7 (C5), 31.2 (C2). HRMS (ESI) calcd for  $C_{14}H_{22}NO_4[M + NH_4]^+$ , 268.1543; found, 268.1538. **23S:**  $R_f = 0.47$  (petroleum ether/ethyl acetate 1:1);  $[\alpha]_D^{25} = +3.1$  (c =0.3, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.39–7.30 (m, 5H, Ar), 4.61 (d, *J* = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.54 (d, *J* = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.22 (dt, J = 8.0, 3.6 Hz, 1H, H4), 3.99 (td, J = 6.8, 4.0 Hz, 1H, H3), 3.67 (s, 3H, OMe), 3.20-3.11 (m, 1H, H1), 2.50 (d, J = 2.8 Hz, 1H,

OH), 2.18–2.02 (m, 3H, H2 $\alpha$ , H2 $\beta$ , H5 $\alpha$ ), 1.95 (ddd, J = 13.6, 7.6, 5.6 Hz, 1H, H5 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  176.9, 137.9, 128.7, 128.1, 127.9, 80.9, 72.1, 72.0, 52.0, 39.1, 35.1, 31.7. HRMS (ESI) calcd for C<sub>14</sub>H<sub>22</sub>NO<sub>4</sub>[M + NH<sub>4</sub>]<sup>+</sup>, 268.1543; found, 268.1539.

Methyl (1R,3R,4R)-3-Benzyloxy-4-azido-cyclopentane-1-carboxylate (24R). To a solution of 23R (96.1 mg, 0.38 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (3 mL) were added TsCl (217.3 mg, 1.14 mmol), DMAP (23.2 mg, 0.19 mmol), and triethylamine (115.4 mg, 158 µL, 1.14 mmol) at 0 °C under argon atmosphere. After stirring overnight at room temperature, the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and saturated NaHCO3 solution. The combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was dissolved in DMF (2 mL), and NaN3 (247.1 mg, 3.80 mmol) was added. After stirring at 90 °C for 12 h, the reaction mixture was partitioned between ethyl acetate and H<sub>2</sub>O, and the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 15:1) to give 24R (101.2 mg, 96% yield) as colorless oil.  $R_{\rm f} = 0.41$  (petroleum ether/ethyl acetate 4:1);  $[\alpha]_D^{25} = -19.6$  (c = 1.0, CHCl<sub>3</sub>).<sup>1</sup>H NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.37–7.27 (m, 5H), 4.57 (d, J = 12.0 Hz, 1H), 4.52 (d, J = 12.0 Hz, 1H), 4.01-3.97 (m, 1H), 3.85-3.81 (m, 1H), 3.68 (s, 3H), 2.92 (dt, J = 16.4, 8.0 Hz, 1H), 2.39–2.27 (m, 2H), 2.01 (ddd, J =13.6, 7.6, 5.6 Hz, 1H), 1.90 (ddd, J = 14.0, 8.8, 4.8 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 175.2, 138.0, 128.6, 127.9, 127.8, 83.6, 71.7, 66.0, 52.1, 39.6, 33.6, 32.3. HRMS (ESI) calcd for C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>3</sub>Na [M + Na]<sup>+</sup>, 298.1162; found, 298.1166.

*Methyl* (15,3*R*,4*R*)-3-Benzyloxy-4-azido-cyclopentane-1-carboxylate (245). This compound was prepared in the same manner as described in the preparation of 24*R*. Quantities: 23*S* (28.5 mg, 0.11 mmol), CH<sub>2</sub>Cl<sub>2</sub> (1 mL), TsCl (41.9 mg, 0.22 mmol), DMAP (6.7 mg, 0.055 mmol), triethylamine (22.3 mg, 31 μL, 0.22 mmol), NaN<sub>3</sub> (71.5 mg, 1.10 mmol), DMF (1 mL), and eluent (petroleum ether/ethyl acetate = 20:1), affording 24*S* (30.4 mg, 97% yield) as colorless oil. *R*<sub>f</sub> = 0.45 (petroleum ether/ethyl acetate 4:1);  $[\alpha]_D^{25} = -16.4$  (*c* = 0.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.38–7.28 (m, 5H), 4.54 (s, 2H), 3.92–3.88 (m, 2H), 3.70 (s, 1H), 3.03 (dt, *J* = 16.8, 8.4 Hz, 1H), 2.37 (ddd, *J* = 13.6, 8.8, 6.8 Hz, 1H), 2.22 (ddd, *J* = 14.0, 8.0, 6.0 Hz, 1H), 2.06–2.00 (m, 1H), 1.96–1.89 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 175.4, 137.9, 128.6, 128.0, 127.8, 83.6, 71.8, 66.1, 52.2, 39.8, 33.4, 32.8. HRMS (ESI) calcd for C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>3</sub>Na [M + Na]<sup>+</sup>, 298.1168; found, 298.1165.

Methyl (1R,3R,4R)-3-Benzyloxy-4-acetamido-cyclopentane-1-carboxylate (25R). To a solution of 24R (32.7 mg, 0.12 mmol) in pyridine (1.5 mL) was added thioacetic acid (191.8 mg, 0.18 mL, 2.52 mmol) under argon atmosphere. After stirring at room temperature for 36 h, the reaction mixture was partitioned between ethyl acetate and saturated NaHCO3 solution. The combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 1:5) to give 25R (30.5 mg, 88% yield) as colorless oil.  $R_{\rm f} = 0.13$  (petroleum ether/ethyl acetate 1:5);  $[\alpha]_{\rm D}^{25} =$ +31.9 (c = 0.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.36–7.26 (m, 5H), 5.42 (d, J = 5.2 Hz, 1H), 4.59 (s, 2H), 4.24-4.18 (m, 1H), 3.86 (dd, J = 10.4, 5.6 Hz, 1H), 3.67 (s, 3H), 2.91 (dt, J = 16.8, 8.4 Hz, 1H), 2.48 (dt, J = 14.0, 7.6 Hz, 1H), 2.27 (ddd, J = 14.4, 8.8, 6.8 Hz, 1H), 2.04 (ddd, J = 13.2, 7.2, 5.6 Hz, 1H), 1.94 (s, 3H), 1.79 (ddd, J = 13.6, 8.8, 4.8 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  175.2, 169.9, 138.5, 128.5, 127.8, 127.7, 83.5, 71.4, 55.7, 52.1, 40.1, 33.8, 33.1, 23.6. HRMS (ESI) calcd for  $C_{16}H_{21}NO_4Na$  [M + Na]<sup>+</sup>, 314.1368; found, 314.1368.

Methyl (15,3R,4R)-3-Benzyloxy-4-acetamido-cyclopentane-1-carboxylate (255). This compound was prepared in the same manner as described in the preparation of 25R. Quantities: 24S (102.7 mg, 0.37 mmol), pyridine (5.5 mL), and thioacetic acid (704.1 mg, 0.66 mL, 9.25 mmol), 48 h, eluent (petroleum ether/ethyl acetate = 1:2), affording 25S (93.7 mg, 86% yield) as white solid.  $R_f = 0.10$ (petroleum ether/ethyl acetate 1:1);  $[\alpha]_{D}^{25} = +24.0$  (c = 0.3, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.34–7.28 (m, 5H, Ar), 6.40 (d, J =6.4 Hz, 1H, NH), 4.67 (d, J = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.56 (d, J = 12.0 Hz, 1H, PhCH<sub>2</sub>), 4.42 (t, *J* = 6.8 Hz, 1H, H4), 3.94 (d, *J* = 5.2 Hz, 1H, H3), 3.71 (s, 3H, OMe), 3.15–3.08 (m, 1H, H1), 2.42 (ddd, *J* = 14.4, 9.6, 6.8 Hz, 1H, H5α), 2.14 (dd, *J* = 14.0, 8.8 Hz, 1H, H2α), 2.02 (ddd, *J* = 14.8, 6.8, 5.6 Hz, 1H, H2β), 1.96 (s, 3H, CH<sub>3</sub>), 1.73 (dt, *J* = 14.4, 2.8 Hz, 1H, H5β). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 179.3 (CO<sub>2</sub>Me), 169.5 (CH<sub>3</sub>CO), 138.5 (Ph), 128.5 (Ph), 127.8 (Ph), 127.7 (Ph), 84.7 (C3), 71.2 (PhCH<sub>2</sub>), 55.0 (C4), 52.4 (OMe), 40.7 (C1), 35.4 (C2), 33.4 (C5), 23.6 (CH<sub>3</sub>). Anal. Calcd for C<sub>16</sub>H<sub>21</sub>NO<sub>4</sub>: C, 65.96; H, 7.27; N, 4.81; Found: C, 65.97; H, 7.13; N, 4.65. HRMS (ESI) calcd for C<sub>16</sub>H<sub>22</sub>NO<sub>4</sub> [M + H]<sup>+</sup>, 292.1549; found, 292.1558.

(1R,3R,4R)-3-Benzyloxy-4-acetamido-1-cyclopentylmethanol (26R). This compound was prepared in the same manner as described in the preparation of 19. Quantities: 25R (319.7 mg, 1.10 mmol), LiBH<sub>4</sub> (95.8 mg, 4.40 mmol), and methanol (0.6 mL), THF (30 mL), 1 h, eluent (petroleum ether/acetone = 1:1), affording 26R (254.6 mg, 88% yield) as white solid.  $R_f = 0.21$  (petroleum ether/acetone 1:2);  $[\alpha]_{D}^{25} = +6.7$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33– 7.25 (m, 5H, Ar), 5.70 (d, J = 6.4 Hz, 1H, NH), 4.62 (d, J = 12.4 Hz, 1H, PhCH<sub>2</sub>), 4.58 (d, I = 12.0 Hz, 1H, PhCH<sub>2</sub>), 4.25 (ddd, I = 11.6, 7.6, 4.4 Hz, 1H, H4), 3.81 (dd, J = 10.4, 4.8 Hz, 1H, H3), 3.55 (d, J = 6.0 Hz, 2H, H6α, H6β), 2.66 (s, 1H, OH), 2.34–2.23 (m, 1H, H1), 2.18–2.10 (m, 1H, H2 $\alpha$ ), 2.01 (dt, J = 14.4, 7.2 Hz, 1H, H5 $\alpha$ ), 1.93 (s, 5.6 Hz, 1H, H2 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.0, 138.5, 128.5, 127.8, 127.7, 84.2, 71.3, 66.7, 55.5, 38.0, 33.8, 33.0, 23.5. HRMS (ESI) calcd for  $C_{15}H_{22}NO_3$  [M + H]<sup>+</sup>, 264.1594; found, 264.1593.

(15,3*R*,4*R*)-3-Benzyloxy-4-acetamido-1-cyclopentylmethanol (**265**). This compound was prepared in the same manner as described in the preparation of **19**. Quantities: **25S** (380.3 mg, 1.30 mmol), LiBH<sub>4</sub> (132.4 mg, 6.08 mmmol), and methanol (0.46 mL), THF (23 mL), 4 h, eluent (petroleum ether/acetone = 1:1), affording **26S** (337.4 mg, 98% yield) as colorless oil.  $R_f$  = 0.28 (petroleum ether/ acetone 1:2); [α]<sub>2</sub><sup>D5</sup> = +13.6 (*c* = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.34–7.25 (m, 5H), 6.70 (d, *J* = 7.2 Hz, 1H), 4.63 (d, *J* = 12.0 Hz, 1H), 4.54 (d, *J* = 12.4 Hz, 1H), 4.31–4.27 (m, 1H), 3.83– 3.82 (m, 1H), 3.66–3.57 (m, 2H), 2.80 (brs, 1H), 2.49–2.42 (m, 1H), 2.35 (ddd, *J* = 13.2, 10.4, 7.2 Hz, 1H), 1.90 (s, 3H), 1.88–1.75 (m, 2H), 1.36 (dt, *J* = 13.2, 4.0 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 169.5, 138.8, 128.4, 127.8, 127.6, 85.1, 70.9, 64.9, 54.6, 37.5, 32.9, 32.4, 23.6. HRMS (ESI) calcd for C<sub>15</sub>H<sub>22</sub>NO<sub>3</sub> [M + H]<sup>+</sup>, 264.1594; found, 264.1595.

Dibenzyl [α-Hydroxy-(1R,3R,4R)-3-benzyloxy-4-acetamido-1-cyclopentylmethyl]-phosphonate (27R). This compound was prepared in the same manner as described in the preparation of 20. Quantities: 26R (89.5 mg, 0.34 mmol), BAIB (164.3 mg, 0.51 mmol), TEMPO (5.3 mg, 0.034 mmol), and CH<sub>2</sub>Cl<sub>2</sub> (2.2 mL), 17 h; dibenzyl phosphite (209.8 mg, 177 µL, 0.68 mmol, 85%), triethylamine (68.8 mg, 95 µL, 0.68 mmol), and CH<sub>2</sub>Cl<sub>2</sub> (2 mL), overnight, eluent (petroleum ether/acetone = 1:1), affording 27R (161.5 mg, 91% yield) as colorless oil.  $R_f$  = 0.44 (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.31–7.26 (m, 15H), 6.07–6.00 (m, 1H), 5.10– 4.97 (m, 4H), 4.66–4.56 (m, 2H), 4.29–4.27 (m, 1H), 4.21–4.08 (m, 1H), 3.94–3.85 (m, 1H), 3.82–3.79 (m, 1H), 2.72–2.69 (m, 1H), 2.31–1.95 (m, 3H), 1.89 (s, 3H), 1.73–1.57 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 24.83, 24.56. HRMS (ESI) calcd for C<sub>29</sub>H<sub>35</sub>NO<sub>6</sub>P [M + H]<sup>+</sup>, 524.2202; found, 524.2200.

Dibenzyl [α-Hydroxy-(15,3R,4R)-3-benzyloxy-4-acetamido-1-cyclopentylmethyl]-phosphonate (275). This compound was prepared in the same manner as described in the preparation of 20. Quantities: 26S (40.2 mg, 0.15 mmol), BAIB (73.9 mg, 0.23 mmol), TEMPO (2.3 mg, 0.015 mmol), and CH<sub>2</sub>Cl<sub>2</sub> (1.0 mL), 11 h; dibenzyl phosphite (96.1 mg, 81 µL, 0.31 mmol, 85%), triethylamine (31.4 mg, 43 µL, 0.31 mmol), and CH<sub>2</sub>Cl<sub>2</sub> (1.0 mL), overnight, eluent (petroleum ether/acetone = 1:1), affording 27S (69.3 mg, 87% yield) as white solid.  $R_{\rm f}$  = 0.44 (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.32–7.26 (m, 15H), 6.52 (brs, 1H), 5.10–4.94 (m, 4H), 4.89 (brs, 1H), 4.65–4.58 (m, 1H), 4.54–4.48 (m, 1H), 4.30–4.26 (m, 1H), 3.98–3.74 (m, 2H), 2.75 (brs, 1H), 2.46–2.21 (m, 1H), 1.96–1.91 (m, 2H), 1.87–1.85 (m, 3H), 1.74–1.39 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  25.23, 25.17. HRMS (ESI) calcd for C<sub>29</sub>H<sub>35</sub>NO<sub>6</sub>P [M + H]<sup>+</sup>, 524.2202; found, 524.2198.

(15,2*R*,4*R*)-2-Benzyloxy-4-hydroxymethyl-1-cyclopentanol (28). This compound was prepared in the same manner as described in the preparation of 19. Quantities: 23*R* (270.9 mg, 1.08 mmol), LiBH<sub>4</sub> (94.3 mg, 4.33 mmmol), methanol (0.25 mL), and THF (12 mL), 1.5 h, eluent (petroleum ether/acetone = 3:2), affording 28 (235.0 mg, 98% yield) as colorless oil.  $R_f = 0.31$  (petroleum ether/acetone 1:1);  $[\alpha]_{D}^{25} = -12.0$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.36–7.30 (m, 5H), 4.60 (d, J = 12.0 Hz, 1H), 4.53 (d, J = 11.6 Hz, 1H), 4.08 (dt, J = 7.6, 3.6 Hz, 1H), 3.81 (ddd, J = 8.0, 7.2, 4.4 Hz, 1H), 3.56 (brs, 2H), 2.84 (s, 1H), 2.77 (s, 1H), 1.56 (ddd, J = 14.4, 4.8, 3.6 Hz, 1H), 1.69 (dt, J = 13.2, 8.0 Hz, 1H), 1.56 (ddd, J = 14.4, 4.8, 3.6 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  138.0, 128.6, 128.0, 127.9, 81.5, 72.0, 71.7, 66.6, 36.2, 34.1, 30.4. HRMS (ESI) calcd for C<sub>13</sub>H<sub>22</sub>NO<sub>3</sub> [M + NH<sub>4</sub>]<sup>+</sup>, 240.1594; found, 240.1597.

(1S,2R,4R)-2-Benzyloxy-4-tert-butyldiphenylsilyloxymethyl-1-cyclopentanol (29). To a solution of 28 (89.6 mg, 0.40 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (7 mL) were added DMAP (9.8 mg, 0.08 mmmol), TBDPSCl (164.9 mg, 155 µL, 0.60 mmmol), and triethylamine (81.0 mg, 111  $\mu$ L, 0.80 mmol) at 0 °C under argon atmosphere. After stirring at room temperature for 36 h, the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O, and then the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 12:1) to give 29 (164.4 mg, 88% yield) as colorless oil.  $R_f = 0.33$  (petroleum ether/ethyl acetate 4:1);  $[\alpha]_{D}^{25} = -4.2$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.67– 7.64 (m, 4H), 7.43–7.29 (m, 11H), 4.58 (d, J = 11.6 Hz, 1H), 4.51 (d, J = 11.6 Hz, 1H), 4.08 (dt, J = 10.4, 5.2 Hz, 1H), 3.79 (ddd, J = 7.2, 6.8, 4.4 Hz, 1H), 3.64 (dd, J = 9.6, 6.4 Hz, 1H), 3.59 (dd, J = 10.0, 7.2 Hz, 1H), 2.55 (d, J = 5.6 Hz, 1H), 2.19–2.07 (m, 1H), 2.03–1.93 (m, 2H), 1.63 (dt, J = 13.2, 7.6 Hz, 1H), 1.50 (ddd, J = 13.6, 6.8, 4.8 Hz, 1H), 1.05 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 138.3, 135.7, 134.04, 134.02, 129.7, 128.6, 127.9, 127.85, 127.75, 81.0, 72.2, 71.4, 68.6, 36.6, 34.6, 31.5, 27.0, 19.4. Anal. Calcd for C<sub>29</sub>H<sub>36</sub>O<sub>3</sub>Si: C, 75.61; H, 7.88. Found: C, 75.56; H, 8.01. ESI-MS m/z: 461 [M + H]<sup>+</sup>.

(1S,2R,4R)-2-Benzyloxy-4-tert-butyldiphenylsilyloxymethyl-1-cyclopentyl Tosylate (30). To a solution of 29 (1.19 g, 2.58 mmol) in pyridine (32 mL) were added DMAP (0.63 g, 5.16 mmmol) and a solution of TsCl (2.46 g, 12.90 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) at 0 °C under argon atmosphere. After stirring at room temperature for 24 h, the solvent was evaporated and the residue was partitioned between ethyl acetate and H<sub>2</sub>O, then the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 15:1) to give 30 (1.42 g, 89% yield) as colorless oil.  $R_f = 0.44$  (petroleum ether/ethyl acetate 4:1);  $\left[\alpha\right]_D^{25} = +15.0$  (c = 1.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.78 (d, J = 8.4 Hz, 2H), 7.61 (d, I = 7.8 Hz, 4H), 7.42–7.22 (m, 13H), 4.88 (dd, I = 10.0, 5.6Hz, 1H), 4.46 (d, J = 12.0 Hz, 1H), 4.38 (d, J = 12.0 Hz, 1H), 3.78 (td, J = 6.8, 4.4 Hz, 1H), 3.55 (d, J = 7.2 Hz, 2H), 2.39 (s, 3H), 2.15-2.04 (m, 1H), 1.98–1.90 (m, 2H), 1.76 (dt, J = 14.4, 6.0 Hz, 1H), 1.63 (dt, J = 13.6, 7.6 Hz, 1H), 1.01 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 144.5, 138.3, 135.7, 134.6, 133.92, 133.88, 129.8, 129.7, 128.4, 127.9, 127.8, 127.64, 127.61, 81.4, 79.1, 71.5, 68.1, 35.7, 32.1, 31.6, 26.9, 21.8, 19.4. Anal. Calcd for C<sub>36</sub>H<sub>42</sub>O<sub>5</sub>SSi: C, 70.32; H, 6.88; Found: C, 70.25; H, 7.04. HRMS (ESI) calcd for C<sub>36</sub>H<sub>46</sub>NO<sub>5</sub>SSi [M + NH<sub>4</sub>]<sup>+</sup>, 632.2866; found, 632.2860.

(15,2P,4R)-2-Benzyloxy-4-tert-butyldiphenylsilyloxymethyl-1-cyclopentyl Carbonitrile (**31**). To a solution of **30** (256.4 mg, 0.42 mmol) in toluene (25 mL) was added Bu<sub>4</sub>NCN (225.5 mg, 0.84 mmol) under argon atmosphere. After stirring at 85 °C for 24 h, the reaction mixture was concentrated and the residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 25:1) to give **31** (93.0 mg, 48% yield) as colorless oil.  $R_f = 0.47$  (petroleum ether/ethyl acetate 4:1);  $[\alpha]_{D}^{25} = -13.9$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.64–7.62 (m, 4H), 7.44–7.29 (m, 11H), 4.58 (d, J = 12.0 Hz, 1H), 4.55 (d, J = 12.0 Hz, 1H), 4.17 (dd, J = 12.4, 6.4 Hz, 1H), 3.60 (dd, J = 10.4, 6.0 Hz, 1H), 3.54 (dd, J = 10.0, 6.4 Hz, 1H), 2.83 (td, *J* = 8.0, 6.0 Hz, 1H), 2.40–2.29 (m, 1H), 2.18 (dt, *J* = 14.0, 7.6 Hz, 1H), 2.06–1.95 (m, 2H), 1.58–1.51 (m, 1H), 1.05 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  137.7, 135.7, 133.6, 129.9, 128.6, 128.0, 127.9, 127.8, 122.0, 83.3, 72.1, 66.5, 38.7, 34.7, 34.4, 31.5, 27.0, 19.4. HRMS (ESI) calcd for C<sub>30</sub>H<sub>36</sub>NO<sub>2</sub>Si [M + H]<sup>+</sup>, 470.2515; found, 470.2512.

(1S,2R,4R)-2-Benzyloxy-4-tert-butyldiphenylsilyloxymethyl-1-cyclopentylmethylacetamide (32). To a solution of 31 (69.3 mg, 0.15 mmol) in methanol (4.5 mL) were added CoCl<sub>2</sub> (58.4 mg, 0.45 mmol) and then NaBH<sub>4</sub> (85.1 mg, 2.25 mmol) in portions at 0 °C under argon atmosphere. After stirring at the same temperature for 3 h, the reaction was quenched with 2 M HCl (0.9 mL). While the black precipitates disappeared, 2 M NH<sub>3</sub>·H<sub>2</sub>O (1.4 mL) was added. After 5 min, the reaction mixture was concentrated and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) under argon atmosphere. To this solution were added triethylamine (3.4 mL) and acetic anhydride (1.8 mL). After stirring overnight at room temperature, the reaction mixture was concentrated and the residue was partitioned between ethyl acetate and H<sub>2</sub>O, then the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 3:2) to give 32 (45.6 mg, 60% yield) as colorless oil.  $R_{\rm f} = 0.41$  (petroleum ether/ethyl acetate 1:2);  $[\alpha]_{\rm D}^{25} = -16.6$  (c = 0.3, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.65 (d, J = 6.8 Hz, 4H, Ar), 7.44-7.30 (m, 11H, Ar), 6.05 (brs, 1H, NH), 4.58 (d, J = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.39 (d, *J* = 11.6 Hz, 1H, PhCH<sub>2</sub>), 3.66 (dd, *J* = 14.0, 7.6 Hz, 1H, H2), 3.57 (d, J = 6.0 Hz, 2H, H7 $\alpha$ , H7 $\beta$ ), 3.45 (dt, J =12.8, 6.0 Hz, 1H, H6 $\alpha$ ), 3.03 (ddd, J = 13.2, 9.6, 3.6 Hz, 1H, H6 $\beta$ ), 2.24-2.15 (m, 2H, H4, H3α), 2.09-1.98 (m, 1H, H1), 1.85 (s, 3H, CH<sub>3</sub>), 1.71 (ddd, J = 13.6, 8.8, 4.8 Hz, 1H, H5 $\alpha$ ), 1.43–1.32 (m, 2H, H5 $\beta$ , H3 $\beta$ ), 1.06 (s, 9H, tBu). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.1, 138.5, 135.7, 134.0, 129.7, 128.7, 128.0, 127.9, 127.8, 85.4, 71.8, 67.8, 44.0, 43.2, 37.5, 34.7, 29.7, 27.0, 23.3, 19.4. HRMS (ESI) calcd for C<sub>32</sub>H<sub>42</sub>NO<sub>3</sub>Si [M + H]<sup>+</sup>, 516.2934; found, 516.2927.

(1S,2R,4R)-2-Benzyloxy-4-hydroxymethyl-1-cyclopentylmethylacetamide (33). To a solution of 32 (126.1 mg, 0.24 mmol) in THF (14 mL) was added TBAF (1 M in THF, 0.98 mL, 0.98 mmol) under argon atmosphere. After stirring overnight at room temperature, the reaction mixture was concentrated and the residue was purified by column chromatography on silica gel (petroleum ether/acetone = 2:1) to give 33 (68.1 mg, 100% yield) as colorless oil.  $R_f = 0.32$  (petroleum ether/acetone 1:2);  $[\alpha]_D^{25} = -31.0$  (c = 0.6, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.38–7.30 (m, 5H, Ar), 6.05 (s, 1H, NH), 4.59 (d, J = 11.2 Hz, 1H, PhCH<sub>2</sub>), 4.42 (d, J = 11.6 Hz, 1H, PhCH<sub>2</sub>), 3.70 (dd, J = 13.6, 6.8 Hz, 1H, H2), 3.56 (d, J = 6.0 Hz, 2H, H7 $\alpha$ , H7 $\beta$ ), 3.40 (dt, J= 13.2, 5.6 Hz, 1H, H6 $\alpha$ ), 3.07 (ddd, J = 13.2, 9.6, 4.0 Hz, 1H, H6 $\beta$ ), 2.29-2.09 (m, 3H, H1, H4, H3a), 1.96 (brs, 1H, OH), 1.87 (s, 3H,  $CH_3$ ), 1.72 (ddd, J = 13.6, 8.4, 5.2 Hz, 1H, H5 $\alpha$ ), 1.48–1.40 (m, 2H, H5 $\beta$ , H3 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 170.3 (CH<sub>3</sub>CO), 138.2 (Ph), 128.7(Ph), 128.0 (Ph), 127.9 (Ph), 84.8 (C2), 71.7 (PhCH<sub>2</sub>), 67.1 (C7), 43.7 (C6), 43.6 (C1), 37.7 (C4), 34.9 (C3), 29.9 (C5), 23.3 (CH<sub>3</sub>). HRMS (ESI) calcd for C<sub>16</sub>H<sub>24</sub>NO<sub>3</sub> [M + H]<sup>+</sup>, 278.1751; found, 278.1750.

Dibenzyl [ $\alpha$ -Hydroxy-(1R,3R,4S)-3-benzyloxy-4-acetamidomethyl-1-cyclopentyl-methyl]-phosphonate (34). To a solution of DMSO (93.8 mg, 85  $\mu L$ , 1.20 mmol) in  $CH_2Cl_2$  (3.0 mL) was added oxalyl chloride (135.0 mg, 93  $\mu$ L, 1.06 mmol) dropwise at -78°C under argon atmosphere. After stirring at the same temperature for 30 min, a solution of 33 (37.0 mg, 0.13 mmol) in THF (2.0 mL) was slowly added. After 2 h, triethylamine (333 µL, 2.39 mmol) was added and the reaction mixture was stirred for another 1 h. Then the mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O, and the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. To a solution of dibenzyl phosphite (614.9 mg, 518  $\mu$ L, 2.00 mmol, 85%) in THF (2.0 mL) was added NaH (53.2 mg, 1.33 mmol, 60% in mineral oil) at 0 °C under argon atmosphere. After 30 min, a solution of the above residue in THF (2.0 mL) was added to the reaction bottle. The mixture was stirred overnight and quenched with saturated NH<sub>4</sub>Cl solution. Then the mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O and the combined organic layer was washed

with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/acetone = 1:2) to give 34 (48.2 mg, 67% yield) as colorless oil.  $R_{\rm f}$  = 0.23 (petroleum ether/acetone 1:3). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.34–7.28 (m, 15H), 5.83–5.78 (m, 1H), 5.10–4.99 (m, 4H), 4.56–4.49 (m, 1H), 4.39–4.34 (m, 1H), 3.84–3.83 (m, 1H), 3.67–3.62 (m, 1H), 3.27–3.00 (m, 3H), 2.51–2.42 (m, 1H), 2.22–2.01 (m, 2H), 1.97–1.88 (m, 1H), 1.86 (s, 3H), 1.63–1.60 (m, 1H), 1.49–1.41 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  25.03, 24.66. HRMS (ESI) calcd for C<sub>30</sub>H<sub>37</sub>NO<sub>6</sub>P [M + H]<sup>+</sup>, 538.2358; found, 538.2360.

(3*R*,4*S*)-3,4-*Isopropylidenedioxy-1-cyclopentene-1-methanol* (**35**). To a solution of 17 (100.5 mg, 0.51 mmol) in toluene (10 mL) was slowly added DIBAL-H (1.3 mL, 1.53 mmol, 1.2 M in toluene) at -78 °C under argon atmosphere. After stirring at the same temperature for 4 h, the reaction was quenched with H<sub>2</sub>O and filtered through a pad of Celite. The filtrate was concentrated and purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 3:1) to give **35** (84.5 mg, 98% yield) as colorless oil.  $R_f = 0.15$  (petroleum ether/ethyl acetate 2:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  5.70 (td, *J* = 3.6, 2.0 Hz, 1H), 5.13–5.11 (m, 1H), 4.82 (td, *J* = 6.0, 1.2 Hz, 1H), 4.26–4.15 (m, 2H), 2.62–2.55 (m, 1H), 2.50–2.44 (m, 1H), 1.56 (t, *J* = 6.0 Hz, 1H), 1.43 (s, 3H), 1.35 (s, 3H). The spectroscopic data coincide with the previous report.<sup>80</sup>

(3R,4S)-3,4-Isopropylidenedioxy-1-cyclopentenemethyl Tosylate (36). To a solution of 35 (386.3 mg, 2.27 mmol) in ether (10 mL) were added TsCl (1298.3 mg, 6.81 mmol) and NaOH (408.8 mg, 10.22 mmol) at 0 °C under argon atmosphere. After stirring at room temperature for 24 h, the reaction mixture was partitioned between ethyl acetate and H<sub>2</sub>O, then the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 5:1) to give 36 (712.2 mg, 97% yield) as colorless oil.  $R_{\rm f} = 0.33$  (petroleum ether/ethyl acetate 2:1);  $[\alpha]_{\rm D}^{28} =$ -17.8 (c = 0.9, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.79 (d, J = 8.4 Hz, 1H), 7.35 (d, J = 8.0 Hz, 1H), 5.72 (brs, 1H), 5.04 (d, J = 5.6 Hz, 1H), 4.74 (t, J = 5.6 Hz, 1H), 4.61 (d, J = 12.4 Hz, 1H), 4.55 (d, J = 12.8 Hz, 1H), 2.56-2.50 (m, 1H), 2.45 (s, 3H), 2.39 (d, J = 17.6 Hz, 1H), 1.36 (s, 3H), 1.32 (s, 3H).  $^{13}\mathrm{C}$  NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 145.1, 138.8, 133.1, 130.0, 129.4, 128.1, 110.1, 84.9, 78.0, 68.2, 38.7, 27.5, 25.6, 21.8. HRMS (ESI) calcd for  $C_{16}H_{20}O_5SNa [M + Na]^+$ , 347.0929; found, 347.0928.

(1S,2R,3S,5S)-2,3-O-Isopropylidene-5-azidomethyl-1,2,3-cyclopentanetriol (37). To a solution of 36 (353.3 mg, 1.09 mmol) in DMF (2 mL) was added NaN<sub>3</sub> (354.4 mg, 5.45 mmol) under argon atmosphere. After stirring overnight at room temperature, the reaction mixture was partitioned between ethyl acetate and H2O, and the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was dissolved in THF (25 mL) under argon atmosphere, and then B2H6·THF (3.3 mL, 3.27 mmol, 1 M) was added at 0 °C. After stirring at the same temperature for 5 h, the reaction was quenched with methanol (8.2 mL). After 10 min, 3 N NaOH (2 mL, 6.00 mmol) and H<sub>2</sub>O<sub>2</sub> (aq) (0.69 mL, 6.00 mmol, 27%) were added at 0 °C. After stirring overnight at 40 °C, the solvent was evaporated and the residue was partitioned between CH2Cl2 and H<sub>2</sub>O, and the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 4:1) to give 37 (179.6 mg, 77% yield) as colorless oil.  $R_{\rm f} = 0.27$ (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25} = +6.5$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz,  $CDCl_3$ )  $\delta$  4.71 (td, J = 6.4, 3.2 Hz, 1H, H3), 4.40 (dd, J = 6.8, 2.8 Hz, 1H, H2), 4.03 (dt, J = 4.8, 3.2 Hz, 1H, H1), 3.51  $(dd, J = 12.4, 7.6 Hz, 1H, H6\alpha), 3.43 (dd, J = 12.4, 6.8 Hz, 1H, H6\beta),$ 2.51 (d, J = 3.2 Hz, 1H, OH), 2.24 (dt, J = 14.0, 7.6 Hz, 1H, H4 $\alpha$ ), 2.18 (dt, J = 12.8, 6.8 Hz, 1H, H5), 1.72 (ddd, J = 13.6, 6.0, 3.2 Hz, 1H, H4 $\beta$ ), 1.49 (s, 3H, CH<sub>3</sub>), 1.30 (s, 3H, CH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  111.8, 87.3, 79.2, 79.1, 53.3, 46.5, 33.4, 26.7, 24.2. HRMS (ESI) calcd for C<sub>9</sub>H<sub>19</sub>N<sub>4</sub>O<sub>3</sub> [M + NH<sub>4</sub>]<sup>+</sup>, 231.1452; found, 231.1457.

(1S,2R,3S,5S)-1-Benzyl-2,3-O-isopropylidene-5-azidomethyl-1,2,3-cyclopentane-triol (38). To a solution of 37 (197.0 mg, 0.92 mmol) in DMF (3 mL) were added NaH (110.8 mg, 2.77 mmol, 60% in mineral oil) and BnBr (236.0 mg, 164 µL, 1.38 mmol) at 0 °C under argon atmosphere. After stirring overnight at room temperature, the reaction was guenched with H<sub>2</sub>O and extracted with ethyl acetate. Then the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 20:1) to give 38 (243.0 mg, 87% yield) as colorless oil.  $R_f = 0.49$ (petroleum ether/ethyl acetate 4:1);  $[\alpha]_D^{25} = -19.1$  (c = 0.5, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.37–7.28 (m, 5H, Ar), 4.71 (td, J = 6.4, 3.2 Hz, 1H, H3), 4.66 (d, J = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.54-4.52 (m, 2H, H2, PhCH<sub>2</sub>), 3.76 (dd, J = 4.8, 2.0 Hz, 1H, H1), 3.47 (dd, J =12.2, 6.8 Hz, 1H, H6 $\alpha$ ), 3.37 (dd, J = 12.4, 7.2 Hz, 1H, H6 $\beta$ ), 2.32 (td, J = 14.0, 7.2 Hz, 1H, H5), 2.21 (dt, J = 14.4, 6.4 Hz, 1H, H4 $\alpha$ ), 1.74  $(ddd, J = 14.0, 6.0, 2.8 Hz, 1H, H4\beta), 1.48 (s, 3H, CH<sub>3</sub>), 1.30 (s, 3H, CH<sub>3</sub>)$ CH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 138.0 (Ph), 128.6 (Ph), 127.9 (Ph), 111.7 (C(CH<sub>3</sub>)<sub>2</sub>), 85.6 (C1, C2), 79.5 (C3), 71.7 (PhCH<sub>2</sub>), 53.3 (C6), 44.7 (C5), 33.6 (C4), 26.8 (CH<sub>3</sub>), 24.2 (CH<sub>3</sub>). HRMS (ESI) calcd for C<sub>16</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub>[M + NH<sub>4</sub>]<sup>+</sup>, 321.1927; found, 321.1927.

(15,2*R*,35,55)-1-Benzyl-5-azidomethyl-1,2,3-cyclopentanetriol (**39**). Compound **38** (324.6 mg, 1.07 mmol) was dissolved in 90% (v/v) acetic acid solution (15 mL). After stirring at 70 °C for 5 h, the reaction mixture was concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 1:1) to give **39** (275.2 mg, 98% yield) as colorless oil.  $R_f = 0.26$  (petroleum ether/ethyl acetate 1:2);  $[\alpha]_D^{25} = -14.1$  (c = 0.3, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.37–7.28 (m, 5H), 4.71 (d, J = 12.0 Hz, 1H), 4.63 (d, J = 11.6 Hz, 1H), 4.15 (dt, J = 10.4, 4.8 Hz, 1H), 4.00 (dd, J = 10.4, 5.2 Hz, 1H), 3.66 (t, J = 5.6 Hz, 1H), 3.46 (dd, J = 12.4, 6.0 Hz, 1H), 3.38 (dd, J = 12.0, 6.0 Hz, 1H), 2.38 (d, J = 5.6 Hz, 1H), 2.24 (d, J = 4.8 Hz, 1H), 2.23–2.10 (m, 2H), 1.55–1.50 (m, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  138.3, 128.6, 128.0, 86.4, 78.3, 72.4, 71.6, 54.8, 41.0, 33.4. HRMS (ESI) calcd for C<sub>13</sub>H<sub>21</sub>N<sub>4</sub>O<sub>3</sub> [M + NH<sub>4</sub>]<sup>+</sup>, 281.1608; found, 281.1610.

(1S,2R,3S,4S)-3-Benzyl-4-azidomethyl-1,2,3-cyclopentanetriol 1,2-Cyclicsulfate (40). To a solution of 39 (275.2 mg, 1.04 mmol) in THF (14 mL) was added NaH (166.4 mg, 4.16 mmol, 60% in mineral oil) at -20 °C under argon atmosphere. After 15 min, Im<sub>2</sub>SO<sub>2</sub> (247.8 mg, 1.25 mmol) was added. After stirring at the same temperature for 30 min, the reaction was quenched with saturated NH<sub>4</sub>Cl solution. The mixture was extracted with ethyl acetate, and the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 6:1) to give 40 (270.8 mg, 80% yield) as colorless oil.  $R_f = 0.56$  (petroleum ether/ethyl acetate 2:1);  $[\alpha]_D^{25} = -34.7$  (*c* = 0.3, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 7.40–7.32 (m, 5H), 5.22 (td, J = 7.2, 6.0 Hz, 1H), 5.04 (dd, J = 7.6, 4.0 Hz, 1H), 4.71 (d, J = 11.6 Hz, 1H), 4.55 (d, J = 11.6 Hz, 1H), 4.02 (dd, J = 8.4, 4.4 Hz, 1H), 3.51 (dd, J = 12.4, 4.8 Hz, 1H), 3.40 (dd, J = 12.4, 6.4 Hz, 1H), 2.48 (dt, J = 14.0, 7.2 Hz, 1H), 2.25-2.15 (m, 1H), 2.00 (ddd, J = 14.4, 10.8, 6.0 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 136.6, 128.8, 128.6, 128.2, 87.5, 83.0, 81.3, 72.8, 51.4, 41.2, 32.1. HRMS (ESI) calcd for  $C_{13}H_{19}N_4O_5S$  [M + NH<sub>4</sub>]<sup>+</sup>, 343.1071; found, 343.1075.

(25,35)-2-Benzyloxy-3-azidomethyl-cyclopentanone (41). To a solution of 40 (13.3 mg, 0.04 mmol) in THF (0.9 mL) was added tBuOK (18.0 mg, 0.16 mmol) under argon atmosphere. After stirring at room temperature for 40 min, 5% H<sub>2</sub>SO<sub>4</sub> (0.43 mL) was added and the reaction was stirred overnight at room temperature. The mixture was partitioned between ethyl acetate and H<sub>2</sub>O, then the combined organic layer was washed with saturated NaHCO<sub>3</sub> solution and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ ethyl acetate = 10:1) to give 41 (9.0 mg, 90% yield) as colorless oil. R<sub>f</sub> = 0.56 (petroleum ether/ethyl acetate 2:1);  $[\alpha]_{D}^{25} = -41.9$  (c = 0.4, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.38–7.30 (m, SH), 5.05 (d, J = 11.6 Hz, 1H), 4.69 (d, J = 11.6 Hz, 1H), 3.66 (d, J = 11.2 Hz, 1H), 3.55 (dd, J = 12.4, 3.6 Hz, 1H), 3.40 (dd, J = 12.4, 6.0 Hz, 1H), 2.38

(dd, *J* = 19.6, 9.2 Hz, 1H), 2.33–2.19 (m, 2H), 2.10–2.02 (m,1H), 1.70–1.58 (m,1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  215.9, 137.6, 128.6, 128.4, 128.2, 81.7, 72.9, 52.7, 42.1, 34.9, 20.8. HRMS (ESI) calcd for C<sub>13</sub>H<sub>19</sub>N<sub>4</sub>O<sub>2</sub> [M + NH<sub>4</sub>]<sup>+</sup>, 263.1503; found, 263.1509.

(2S,3S)-2-Benzyloxy-3-acetamidomethyl-cyclopentanone (42). To a solution of 41 (45.5 mg, 0.18 mmol) in pyridine (3 mL) was added thioacetic acid (342.5 mg, 0.32 mL, 4.50 mmol) under argon atmosphere. After stirring at room temperature for 4 h, the reaction mixture was concentrated and the residue was purified by column chromatography on silica gel (petroleum ether/acetone = 2:1) to give 42 (35.9 mg, 74% yield) as white solid.  $R_f = 0.58$  (petroleum ether/ acetone 1:2);  $[\alpha]_{D}^{25} = -24.2$  (c = 0.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.39–7.34 (m, 5H, Ar), 5.62 (s, 1H, NH), 5.12 (d, J = 11.2 Hz, 1H, PhC $H_2$ ), 4.67 (d, J = 11.2 Hz, 1H, PhC $H_2$ ), 3.62 (d, J = 11.6Hz, 1H, H2), 3.52 (dt, J = 13.6, 6.0 Hz, 1H, H6α), 3.24 (ddd, J = 12.4, 7.6, 4.4 Hz, 1H, H6 $\beta$ ), 2.38 (dd, J = 19.6, 9.2 Hz, 1H, H5 $\alpha$ ), 2.24 (dd,  $J = 20.0, 10.4 \text{ Hz}, 1\text{H}, \text{H5}\beta), 2.21-2.13 \text{ (m, 1H, H3)}, 2.09-2.02 \text{ (m, 1H, H3)}$ 1H, H4 $\alpha$ ), 1.82 (s, 3H, CH<sub>3</sub>), 1.51 (dt, J = 22.0, 11.6, 1H, H4 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 216.1 (C1), 170.3 (CH<sub>3</sub>CO), 137.7 (Ar), 128.8 (Ar), 128.7 (Ar), 128.4 (Ar), 84.5 (C2), 72.9 (PhCH<sub>2</sub>), 42.8 (C6), 41.7 (C3), 35.1 (C5), 23.2 (CH<sub>3</sub>CO), 21.2 (C4). HRMS (ESI) calcd for  $C_{15}H_{19}NO_3Na [M + Na]^+$ , 284.1257; found, 284.1257.

Dibenzyl [ $\alpha$ -Hydroxy-(2S,3S)-2-benzyloxy-3-acetamidomethyl-1cyclopentyl]-phosphonate (43). To a solution of dibenzyl phosphite (712.2 mg, 600 µL, 2.31 mmol, 85%) in THF (4.0 mL) was added NaH (61.6 mg, 1.54 mmol, 60% in mineral oil) at 0 °C under argon atmosphere. After 30 min, a solution of 42 (40.2 mg, 0.15 mmol) in THF (4.0 mL) was added slowly to the reaction bottle. The reaction was stirred overnight at room temperature and quenched with saturated NH<sub>4</sub>Cl solution. Then the mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O, and the combined organic layer was washed with brine, dried over Na2SO4, filtered, and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ acetone = 1:2) to give 43 (70.6 mg, 88% yield, R:S = 1:1.6) as colorless oil.  $R_{\rm f} = 0.19$  (petroleum ether/acetone 1:2); 43R:  $[\alpha]_{\rm D}^{20} =$ -13.3 (c = 0.03, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.36-7.20 (m, 15H, Ar), 5.93 (t, J = 4.8 Hz, 1H, NH), 5.06 (s, 1H, PhCH<sub>2</sub>), 5.04 (s, 1H, PhCH<sub>2</sub>), 4.99 (dd, J = 11.6, 7.2 Hz, 1H, PhCH<sub>2</sub>), 4.94 (dd, J = 12.0, 8.4 Hz, 1H, PhCH<sub>2</sub>), 4.69 (d, J = 11.6 Hz, 1H, PhCH<sub>2</sub>), 4.57 (d, J = 11.2 Hz, 1H, PhCH<sub>2</sub>), 3.84 (dd, J = 7.6, 4.8 Hz, 1H, H2), 3.54 (s, 1H, OH), 3.32-3.21 (m, 2H, H6), 2.39-2.29 (m, 2H, H3, H4 $\alpha$ ), 1.96-1.84 (m, 2H, H5 $\alpha$ , H4 $\beta$ ), 1.82 (s, 3H, CH<sub>3</sub>), 1.60-1.50 (m, 1H, H5β). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.5 (CH<sub>3</sub>CO), 137.9 (Ar), 136.62 (d, J = 3.0 Hz Ar), 136.56 (d, J = 3.0 Hz Ar), 128.7 (Ar), 128.63 (Ar), 128.59 (Ar), 128.53 (Ar), 128.46 (Ar), 128.4 (Ar), 128.1 (Ar), 128.0 (Ar), 90.2 (d, J = 2.0 Hz C2), 82.8 (d, J = 165.0 Hz C1), 72.6 (PhCH<sub>2</sub>), 68.4 (d, J = 7.0 Hz PhCH<sub>2</sub>), 68.1 (d, J = 8.0 Hz  $PhCH_2$ , 44.1 (d, J = 7.0 Hz C3), 42.8 (C6), 34.7 (d, J = 6.0 Hz C4), 25.2 ( $\tilde{d}$ , J = 12.0 Hz C5), 23.3 (CH<sub>3</sub>). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$ 24.87. HRMS (ESI) calcd for  $C_{29}H_{35}NO_6P$  [M + H]<sup>+</sup>, 524.2202; found, 524.2200. 43S:  $[\alpha]_{\rm D}^{20} = -38.0$  (c = 0.05, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.37–7.27 (m, 15H, Ar), 5.18 (t, J = 5.4 Hz, 1H, NH), 5.15-5.09 (m, 3H, PhCH<sub>2</sub>), 5.04 (dd, J = 12.0, 7.2 Hz, 1H, PhCH<sub>2</sub>), 4.80 (d, J = 11.4 Hz, 1H, PhCH<sub>2</sub>), 4.54 (d, J = 10.8 Hz, 1H, PhCH<sub>2</sub>), 3.79 (t, J = 8.4 Hz, 1H, H2), 3.26 (s, 1H, OH), 3.20-3.11 (m, 2H, H6), 2.56–2.16 (m, 2H, H4 $\beta$ , H3), 1.91–1.80 (m, 2H, H4 $\alpha$ , H5 $\alpha$ ), 1.73 (s, 3H, CH<sub>3</sub>), 1.24–1.18 (m, 1H, H5 $\beta$ ). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.7 (CH<sub>3</sub>CO), 137.6 (Ar), 136.4 (d, J = 6.0 Hz Ar), 136.3 (d, J = 5.0 Hz Ar), 128.85 (Ar), 128.81 (Ar), 128.78 (Ar), 128.7 (Ar), 128.4 (Ar), 128.32 (Ar), 128.28 (Ar), 84.5 (d, J = 8.0 Hz C2), 77.2 (d, J = 170.0 Hz C1), 73.7 (PhCH<sub>2</sub>), 68.8 (d, J = 7.0 Hz PhCH<sub>2</sub>), 68.3 (d, J = 7.0 Hz PhCH<sub>2</sub>), 43.3 (d, J = 12.0 Hz C3), 41.4 (C6), 33.2 (d, J = 7.0 Hz C4), 23.6 (d, J = 11.0 Hz C5), 23.2 (CH<sub>3</sub>). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  26.36.

Dibenzyl  $\alpha$ -Hydroxy-(3-phenoxy)phenylmethyl-phosphonate (44). To a solution of 3-phenoxybenzaldehyde (219.0 mg, 1.10 mmol) in THF (4.0 mL) were added dibenzyl phosphite (681.3 mg, 574  $\mu$ L, 2.21 mmol, 85%) and triethylamine (223.6 mg, 308  $\mu$ L, 2.21 mmol) under argon atmosphere. The reaction mixture was stirred overnight at room temperature and concentrated. The residue was purified by column chromatography on silica gel (petroleum ether/ ethyl acetate = 2:1) to give 44 (429.0 mg, 84% yield) as white solid.  $R_f$ = 0.20 (petroleum ether/ethyl acetate 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.30–7.18 (m, 14H), 7.12 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.08 (t, *J* = 7.4 Hz, 1H), 6.96–6.94 (m, 3H), 5.03–4.88 (m, SH), 3.56 (brs, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  157.24 (d, *J* = 3.0 Hz), 157.20, 138.7 (d, *J* = 1.0 Hz), 136.2 (d, *J* = 4.0 Hz), 136.1 (d, *J* = 3.0 Hz), 129.8, 129.7 (d, *J* = 2.0 Hz), 128.6 (d, *J* = 4.0 Hz), 128.4 (d, *J* = 5.0 Hz), 128.0 (d, *J* = 5.0 Hz), 123.3, 122.2 (d, *J* = 5.0 Hz), 118.9, 118.6 (d, *J* = 3.0 Hz), 117.9 (d, *J* = 6.0 Hz), 70.9 (d, *J* = 158.0 Hz), 68.8 (d, *J* = 7.0 Hz), 68.5 (d, *J* = 7.0 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  21.66. HRMS (ESI) calcd for C<sub>27</sub>H<sub>25</sub>O<sub>5</sub>NaP [M + Na]<sup>+</sup>, 483.1337; found, 483.1338.

5'-[Benzyl-N,N-bis(1-methylethyl)phosphoramidite]-N<sup>4</sup>-2',3'-Otris-benzyloxy-carbonyl Cytidine (46). To a solution of bis(N,N)diisopropylamino)chlorophosphine (1.35 g, 5.06 mmol) in ether (10 mL) was added a mixture of benzyl alcohol (0.55 g, 0.52 mL, 5.06 mmol) and triethylamine (0.51 g, 0.70 mL, 5.06 mmol) in ether (2.5 mL) at 0 °C under argon atmosphere. After stirring for 30 min at the same temperature, the reaction mixture was stirred for another 30 min at room temperature and then cold hexane (15 mL) was added at 0 °C. After 10 min, the mixture was filtered through a pad of Celite and concentrated under argon atmosphere to afford the benzyloxybis(N,Ndiisopropylamino)phosphine. To a solution of the residue in CH<sub>2</sub>Cl<sub>2</sub> (8.5 mL) was added a mixture of 45 (1.63 g, 2.53 mmol) and Htetrazole (194.9 mg, 2.78 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (8.5 mL) under argon atmosphere. After stirring at room temperature for 4 h, the reaction mixture was concentrated under argon atmosphere and the residue was purified by column chromatography on silica gel (petroleum ether/acetone = 7:2, containing 1% triethylamine) to give 46 (2.06 g, 92% yield) as colorless foam.  $R_f = 0.52$  (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.24 (brs, 1H), 7.38–7.20 (m, 21H), 7.12 (brs, 1H), 6.31-6.26 (m, 1H), 5.38-5.31 (m, 2H), 5.23-5.17 (m, 2H), 5.14-5.04 (m, 4H), 4.80-4.75 (m, 1H), 4.71-4.66 (m, 1H), 4.39 (brs, 1H), 4.06-3.81 (m, 2H), 3.69-3.59 (m, 2H), 1.25-1.16 (m, 12H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  149.13, 148.80. HRMS (ESI) calcd for  $C_{46}H_{52}N_4O_{12}P [M + H]^+$ , 883.3314; found, 883.3343.

Benzyl (N<sup>4</sup>-2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(dibenzylphosphonato)-cyclopentylmethyl]-phosphate (47a). To a solution of 8 (58.2 mg, 0.16 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6.0 mL) were added 46 (282.5 mg, 0.32 mmol) and H-tetrazole (22.4 mg, 0.32 mmol) under argon atmosphere. After stirring at room temperature for 2 h, TBHP (116  $\mu$ L, 0.64 mmol, 5.5 M in decane) was added and the reaction mixture was stirred for another 3 h. Then the solvent was evaporated and the crude product was purified by column chromatography on silica gel (petroleum ether/acetone = 2:1 to 3:2) to give 47a (135.2) mg, 72% yield) as colorless oil.  $R_f = 0.45$  (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.87-7.85 (m, 1H), 7.36-7.23 (m, 31H), 7.13-7.11 (m, 1H), 6.05-6.02 (m, 1H), 5.37-5.04 (m, 14H), 4.78-4.76 (m, 1H), 4.33-4.13 (m, 3H), 2.40-2.38 (m, 1H), 1.82 (brs, 2H), 1.58–1.49 (m, 6H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$ 20.50–20.23 (m), -0.13 - -1.16 (m). HRMS (ESI) calcd for  $C_{60}H_{61}N_3O_{17}P_2Na [M + Na]^+$ , 1180.3368; found, 1180.3330.

Benzyl ( $N^4$ -2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(dibenzylphosphonato)-c-3,c-4-isopropylidenedioxy-r-1-cyclopentylmethyl]-phosphate (47b). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 20 (42.8 mg, 0.10 mmol), 46 (180.1 mg, 0.20 mmol), H-tetrazole (14.0 mg, 0.20 mmol), CH<sub>2</sub>Cl<sub>2</sub> (5.0 mL), 3 h, and TBHP (73  $\mu$ L, 0.40 mmol, 5.5 M in decane), 6 h, eluent (petroleum ether/acetone = 3:2), affording 47b (109.2 mg, 90% yield) as colorless oil.  $R_f = 0.29$  (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.89–7.84 (m, 1H), 7.10 (brs, 1H), 7.38–7.22 (m, 30H), 7.16–7.03 (m, 1H), 6.13–6.02 (m, 1H), 5.36–4.90 (m, 15H), 4.63–4.09 (m, 5H), 2.57–2.44 (m, 1H), 2.26–1.78 (m, 4H), 1.54–1.51 (m, 3H), 1.29–1.22 (m, 3H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  20.52–20.09 (m), 0.12 – -0.76 (m). HRMS (ESI) calcd for C<sub>63</sub>H<sub>65</sub>N<sub>3</sub>O<sub>19</sub>P<sub>2</sub> [M + Na]<sup>+</sup>, 1252.3580; found, 1252.3522.

Benzyl ( $N^4$ -2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(diben-zylphosphonato)-(15,3R,4S)-3-benzyloxy-4-acetamido-1-cyclopen-

*tylmethyl]-phosphate* (47c). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 27S (69.3 mg, 0.13 mmol), 46 (233.1 mg, 0.26 mmol), and *H*-tetrazole (18.5 mg, 0.26 mmol), 1 h; TBHP (96 μL, 0.53 mmol, 5.5 M in decane), eluent (petroleum ether/acetone = 1:1), affording 47c (132.6 mg, 76% yield) as colorless oil.  $R_f$  = 0.17 (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.82–7.66 (m, 2H), 7.37–7.16 (m, 35H), 7.14–6.80 (m, 1H), 6.01–5.68 (m, 1H), 5.50–5.27 (m, 2H), 5.18–4.99 (m, 12H), 4.83–4.69 (m, 1H), 4.61–4.49 (m, 2H), 4.37–4.03 (m, 5H), 3.84–3.76 (m, 1H), 2.83–2.75 (m, 1H), 2.39–2.00 (m, 2H), 1.93–1.89 (m, 3H), 1.73–1.60 (m, 1H), 1.49–1.38 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 19.73–19.35 (m), -0.27 – -1.56 (m). HRMS (ESI) calcd for C<sub>69</sub>H<sub>70</sub>N<sub>4</sub>O<sub>19</sub>P<sub>2</sub>Na [M + Na]<sup>+</sup>, 1343.4002; found, 1343.4027.

Benzyl  $(N^4-2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(diben$ zylphosphonato)-(1R,3R,4R)-3-benzyloxy-4-acetamido-1-cyclopentylmethyl]-phosphate (47d). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 27R (67.8 mg, 0.13 mmol), 46 (229.6 mg, 0.26 mmol), H-tetrazole (18.2 mg, 0.26 mmol), CH<sub>2</sub>Cl<sub>2</sub> (5.0 mL), TBHP (95 μL, 0.52 mmol, 5.5 M in decane), and eluent (petroleum ether/acetone = 1:1), affording 47d (134.9 mg, 79% yield) as colorless oil.  $R_f = 0.10$  (petroleum ether/ acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.98 (brs, 1H), 7.82– 7.76 (m, 1H), 7.36-7.20 (m, 35H), 7.13-7.02 (m, 1H), 6.03-6.00 (m, 1H), 5.90-5.66 (m, 1H), 5.35-5.21 (m, 2H), 5.18-4.89 (m, 12H), 4.85-4.71 (m, 1H), 4.61-4.48 (m, 2H), 4.34-4.02 (m, 4H), 3.83-3.74 (m, 1H), 2.72-2.60 (m, 1H), 2.31-2.06 (m, 2H), 1.99-1.89 (m, 3H), 1.84–1.66 (m, 2H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$ 19.73-19.54 (m), -0.32 - -1.16 (m). HRMS (ESI) calcd for  $C_{69}H_{70}N_4O_{19}P_2Na [M + Na]^+$ , 1343.4002; found, 1343.4050.

Benzyl (N<sup>4</sup>-2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(dibenzylphosphonato)-(1R,3R,4S)-3-benzyloxy-4-acetamidomethyl-1-cyclopentylmethyl]-phosphate (47e). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 34 (43.1 mg, 0.08 mmol), 46 (141.3 mg, 0.16 mmol), H-tetrazole (11.2 mg, 0.16 mmol), CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL), TBHP (58 μL, 0.32 mmol, 5.5 M), and eluent (petroleum ether/acetone = 1:2), affording 47e (70.8 mg, 66% yield) as colorless oil.  $R_{\rm f} = 0.30$  (petroleum ether/acetone 1:3). <sup>1</sup>H NMR (400 MHz, CDCl<sub>2</sub>)  $\delta$  8.00–7.74 (m, 1H), 7.38–7.25 (m, 36H), 7.14-7.07 (m, 1H), 6.21-5.92 (m, 2H), 5.40-4.96 (m, 14H), 4.79-4.71 (m, 1H), 4.52-4.10 (m, 5H), 3.58-3.49 (m, 1H), 3.34-3.24 (m, 1H), 3.12-3.96 (m, 1H), 2.53-2.42 (m, 1H), 2.29-2.20 (m, 1H), 2.05–1.95 (m, 3H), 1.84–1.82 (m, 3H), 1.70–1.58 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  19.83–19.36 (m), -0.10 – -1.06 (m). HRMS (ESI) calcd for  $C_{70}H_{72}N_4O_{19}P_2Na [M + Na]^+$ , 1357.4158; found, 1357.4200.

Benzyl  $(N^4-2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(diben$ zylphosphonato)-1-((2S,3S)-2-benzyloxy-3-acetamidomethyl-cyclopentyl)]-phosphate (47f). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 43 (62.9 mg, 0.12 mmol), 46 (211.9 mg, 0.24 mmol), H-tetrazole (16.8 mg, 0.24 mmol), CH2Cl2 (3.0 mL), TBHP (87 µL, 0.48 mmol, 5.5 M in decane), and eluent (petroleum ether/acetone = 1:2), affording 47f(121.4 mg, 76% yield) as colorless oil.  $R_f = 0.24$  (petroleum ether/ acetone 1:3). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.86–7.74 (m, 1H), 7.56 (brs, 1H), 7.38–7.20 (m, 35H), 7.14–7.11 (m, 1H), 6.07–5.79 (m, 2H), 5.52-5.22 (m, 2H), 5.20-4.92 (m, 12H), 4.80-4.44 (m, 2H), 4.33-4.06 (m, 3H), 3.90-3.73 (m, 1H), 3.38-3.07 (m, 2H), 2.51-2.27 (m, 3H), 1.98–1.82 (m, 1H), 1.82–1.71 (m, 3H), 1.24–1.11 (m, 1H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 22.08–19.25 (m), -5.94 – -6.69 (m). HRMS (ESI) calcd for  $C_{69}H_{71}N_4O_{19}P_2$  [M + H]<sup>+</sup>, 1321.4182; found, 1321.4223.

Benzyl ( $N^4$ -2',3'-O-Tris-benzyloxycarbonylcytidin-5'-yl)-[(dibenzylphosphonato)-(3-phenoxy)phenylmethyl]-phosphate (47g). This compound was prepared in the same manner as described in the preparation of 47a. Quantities: 44 (52.5 mg, 0.11 mmol), 46 (201.3 mg, 0.23 mmol), H-tetrazole (16.0 mg, 0.23 mmol), CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL), TBHP (83  $\mu$ L, 0.46 mmol, 5.5 M in decane), and eluent (petroleum ether/acetone = 2:1), affording 47g (116.3 mg, 81% yield) as colorless oil.  $R_f$  = 0.31 (petroleum ether/acetone 1:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.82–7.75 (m, 1H), 7.49 (brs, 1H), 7.38–6.91 (m, 40H), 6.04–5.99 (m, 1H), 5.69–5.62 (m, 1H), 5.32–5.28 (m, 1H), 5.20–4.76 (m, 13H), 4.19–4.04 (m, 3H). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>)  $\delta$  16.96–16.37 (m), –0.44 – –1.14 (m). HRMS (ESI) calcd for C<sub>67</sub>H<sub>61</sub>N<sub>3</sub>O<sub>18</sub>P<sub>2</sub>Na [M + Na]<sup>+</sup>, 1280.3323; found, 1280.3300.

Trisodium Cytidin-5'-yl-(cyclopentylphosphonatomethyl)-phosphate (2). To a solution of 47a (135.2 mg, 0.117 mmol) in DMF (2.0 mL) were added Pd/C (135.2 mg, 10%) and 1,4-cyclohexadiene (593.8 mg, 0.68 mL, 7.188 mmol, 97%) under argon atmosphere. The reaction mixture was stirred at room temperature until TLC showed complete consumption of the starting material. Then the mixture was filtered and concentrated. The residue was purified by column chromatography on RP-18 silica gel ( $H_2O \rightarrow H_2O/MeOH = 5:1$ ) to give crude product 2 (42.4 mg, 75% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ion-exchange (IR 120 Na<sup>+</sup>) and lyophilized from water to give 2s and 2l as white powder. 2s: RP-HPLC, column A, 3.5% CH<sub>3</sub>CN, 0.05 M TEAB (triethylammonium bicarbonate (pH 7.2-7.5)) buffer, 15 mL/min flow;  $t_{\rm R} = 7.27$  min;  $[\alpha]_{\rm D}^{30} = +17.0$  (c = 1.5, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.09 (d, J = 7.8 Hz, 1H), 6.22 (d, I = 7.8 Hz, 1H), 5.94 (d, I = 4.2 Hz, 1H), 4.35–4.31 (m, 2H), 4.28– 4.26 (m, 2H), 4.24-4.17 (m, 2H), 2.31-2.24 (m, 1H), 1.82-1.73 (m, 2H), 1.62–1.42 (m, 6H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 163.5, 154.0, 143.5, 96.64, 90.3, 83.8 (d, J = 9.1 Hz), 77.8 (dd, J = 155.5, 7.6 Hz), 75.0, 69.8, 65.0 (d, J = 4.5 Hz), 42.0, 30.1 (d, J = 7.6 Hz), 28.9 (d, J = 7.6 Hz), 25.7, 25.5. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O) δ 16.94, 0.61. HRMS (ESI) calcd for  $C_{15}H_{24}N_3O_{11}P_2 [M - H]^-$ , 484.0886; found, 484.0895. 21: RP-HPLC, column A, 3.5% CH<sub>3</sub>CN, 0.05 M TEAB (triethylammonium bicarbonate (pH 7.2–7.5)) buffer, 15 mL/min flow;  $t_{\rm R}$  = 9.03 min;  $[\alpha]_{D}^{30} = +0.9 \ (c = 1.3, H_2O)$ . <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.08 (d, J = 7.8 Hz, 1H), 6.21(d, J = 7.8 Hz, 1H), 5.95 (d, J = 3.6 Hz, 1H), 4.36 (t, J = 5.4 Hz, 1H), 4.33 (t, J = 4.2 Hz, 1H), 4.28–4.15 (m, 4H), 2.30-2.25 (m, 1H), 1.81-1.73 (m, 2H), 1.59-1.45 (m, 6H). <sup>13</sup>C NMR (151 MHz,  $D_2O$ )  $\delta$  163.6, 154.1, 143.4, 96.6, 90.2, 83.8 (d, J = 9.1 Hz), 77.9 (dd, J = 155.5, 7.6 Hz), 74.9, 69.8, 64.7 (d, J = 4.5 Hz), 42.0, 30.1 (d, J = 7.6 Hz), 28.9 (d, J = 6.0 Hz), 25.6, 25.5. <sup>31</sup>P NMR (162 MHz,  $D_2O$ )  $\delta$  16.54 (d, J = 13.0 Hz), 0.47 (d, J = 13.0 Hz). HRMS (ESI) calcd for  $C_{15}H_{24}N_3O_{11}P_2\ [M-H]^-\!\!,$  484.0886; found, 484 0882

Trisodium Cytidin-5'-yl-(c-3,c-4-dihydroxy-r-1-cyclopentylphosphonatomethyl)-phosphate (3). To a solution of 47b (11.0 mg, 0.009 mmol) in  $CH_2Cl_2$  (1.0 mL) was added  $BCl_3$  (108  $\mu$ L, 0.108 mmol, 1 M in heptane) at 0 °C under argon atmosphere. After stirring at room temperature for 30 min, the reaction was quenched with methanol and the mixture was concentrated. The residue was purified by column chromatography on RP-18 silica gel (H<sub>2</sub>O) to afford the crude product 3 (4.2 mg, 91% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ion-exchange (IR 120 Na<sup>+</sup>), and lyophilized from water to give 3s and 3l as white powder. 3s: RP-HPLC, column A, 0-5 min linear gradient 0-1.0% CH<sub>3</sub>CN, H<sub>2</sub>O (0.1% TFA), 15 mL/min flow,  $t_{\rm R} = 4.20 \text{ min}; [\alpha]_{\rm D}^{30} = +6.7 (c = 1.1, H_2 \text{O}).$  <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.98 (d, J = 7.8 Hz, 1H), 6.14 (d, J = 7.2 Hz, 1H), 5.97 (d, J = 4.2 Hz, 1H), 4.35 (t, J = 5.4 Hz, 1H), 4.31 (d, J = 4.2 Hz, 1H), 4.29–4.19 (m, 4H), 4.00-3.96 (m, 2H), 2.41-2.33 (m, 1H), 2.17-2.08 (m, 2H), 1.77–1.69 (m, 2H). <sup>13</sup>C NMR (151 MHz,  $D_2O$ )  $\delta$  166.2, 157.5, 142.5, 97.2, 90.2, 83.5 (d, J = 9.1 Hz), 77.0 (dd, J = 157.0, 9.1 Hz), 75.0, 73.4, 73.3, 69.9, 65.1 (d, J = 4.5 Hz), 35.6, 34.6 (d, J = 7.6 Hz), 33.7 (d, J = 6.0 Hz).  $^{31}\mathrm{P}$  NMR (162 MHz, D2O)  $\delta$  15.85, 0.41. HRMS (ESI) calcd for  $C_{15}H_{24}N_3O_{13}P_2$  [M - H]<sup>-</sup>, 516.0784; found, 516.0777. 3*l*: RP-HPLC, column A, 0-5 min linear gradient 0-1.0% CH<sub>3</sub>CN, H<sub>2</sub>O (0.1% TFA), 15 mL/min flow,  $t_{\rm R} = 4.43$  min;  $[\alpha]_{\rm D}^{30} = +2.9$  (c = 0.9, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.96 (d, J = 7.2 Hz, 1H), 6.12 (d, *J* = 7.8 Hz, 1H), 5.98 (d, *J* = 4.2 Hz, 1H), 4.37 (t, *J* = 4.8 Hz, 1H), 4.30 (t, J = 4.8 Hz, 1H), 4.27-4.19 (m, 4H), 3.99-3.95 (m, 2H), 2.41-2.33 (m, 1H), 2.15-2.07 (m, 2H), 1.78-1.70 (m, 2H). <sup>13</sup>C NMR  $(151 \text{ MHz}, D_2 \text{O}) \delta 166.9, 158.5, 142.2, 97.3, 90.0, 83.4 \text{ (d, } J = 9.1 \text{ Hz}\text{)},$ 77.1 (dd, J = 155.5, 7.6 Hz), 75.0, 73.4, 73.3, 69.8, 64.9 (d, J = 4.5 Hz), 35.6, 34.5 (d, J = 7.6 Hz), 33.7 (d, J = 6.0 Hz). <sup>31</sup>P NMR (162 MHz,

D<sub>2</sub>O)  $\delta$  15.6, 0.31. HRMS (ESI) calcd for C<sub>15</sub>H<sub>24</sub>N<sub>3</sub>O<sub>13</sub>P<sub>2</sub> [M – H]<sup>-</sup>, 516.0784; found, 516.0779.

Trisodium Cytidin-5'-yl-[(1S,3R,4R)-3-hydroxy-4-acetamido-1-cyclopentyl-phosphonato-methyl]-phosphate (4S). This compound was prepared in the same manner as described in the preparation of 2. Quantities: 47c (53.3 mg, 0.04 mmol), Pd/C (53.3 mg, 10%), 1,4cyclohexadiene (198.2 mg, 234 µL, 2.40 mmol, 97%), DMF (1.0 mL), and eluent (H<sub>2</sub>O), affording crude product 4S (17.7 mg, 79% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to their sodium salt form by ion-exchange (IR 120 Na<sup>+</sup>) and lyophilized from water to give 4Ss and 4Sl as white powder. 4Ss: RP-HPLC, column A, 0-5 min linear gradient 0-1.0% CH<sub>3</sub>CN, H<sub>2</sub>O (0.1% TFA), 15 mL/min flow,  $t_{\rm R} = 4.28$  min;  $[\alpha]_{\rm D}^{30} = +4.0$  (c = 1.9, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.07 (d, J = 7.8 Hz, 1H), 6.20 (d, J = 7.8 Hz, 1H), 5.94 (d, J = 3.0 Hz, 1H), 4.32–4.18 (m, 6H), 4.05– 4.00 (m, 1H), 3.97-3.94 (m, 1H), 2.69-2.67 (m, 1H), 2.24 (dt, J = 13.8, 7.2 Hz, 1H), 2.06 (dt, J = 13.8, 7.8 Hz, 1H), 1.96 (s, 3H), 1.80 (ddd, J = 13.8, 9.0, 4.8 Hz, 1H), 1.61 (dt, J = 13.2, 9.6 Hz, 1H).<sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.6, 163.6, 154.0, 143.5, 96.6, 90.3, 83.7 (d, J = 9.1 Hz), 77.0, 76.6 (dd, J = 158.6, 9.1 Hz), 75.0, 69.8, 65.1 (d, J = 4.5 Hz), 58.2, 37.0, 35.7 (d, J = 9.1 Hz), 31.9 (d, J = 4.5 Hz), 22.7 ppm.  $^{31}\text{P}$  NMR (162 MHz, D2O)  $\delta$  15.66, 0.56. HRMS (ESI) calcd for  $C_{17}H_{27}N_4O_{13}P_2$  [M - H]<sup>-</sup>, 557.1050; found, 557.1055. 4*Sl*: RP-HPLC, column A, 0-5 min linear gradient 0-1.0% CH<sub>3</sub>CN, H<sub>2</sub>O (0.1% TFA), 15 mL/min flow,  $t_{\rm R} = 4.80$  min;  $[\alpha]_{\rm D}^{30} = -3.9$  (c = 1.0, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.97 (d, *J* = 7.8 Hz, 1H), 6.12 (d, J = 7.2 Hz, 1H), 5.99 (d, J = 4.2 Hz, 1H), 4.36 (t, J = 5.4 Hz, 1H), 4.30 (t, J = 4.2 Hz, 1H), 4.27-4.23 (m, 3H), 4.19 (ddd, J = 11.4, 4.2, 2.4Hz, 1H), 4.03 (dt, J = 7.8, 5.4 Hz, 1H), 3.92 (dd, J = 15.0, 7.2 Hz, 1H), 2.71–2.65 (m, 1H), 2.20 (dt, J = 13.2, 7.8 Hz, 1H), 2.16 (dt, J = 14.4, 8.4 Hz, 1H), 1.93 (s, 3H), 1.75 (ddd, J = 13.8, 9.0, 4.8 Hz, 1H), 1.54 (dt, J = 12.6, 9.6 Hz, 1H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O)  $\delta$  174.6, 166.2, 157.6, 142.4, 97.2, 89.9, 83.5 (d, J = 9.1 Hz), 77.1, 76.4 (dd, J = 158.6, 9.1 Hz), 74.9, 69.9, 64.9 (d, J = 6.0 Hz), 58.1, 36.9, 33.8 (d, J = 10.6 Hz), 33.3 (d, J = 3.0 Hz), 22.6. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  15.65, 0.20. HRMS (ESI) calcd for  $C_{17}H_{27}N_4O_{13}P_2$  [M - H]<sup>-</sup>, 557.1050; found, 557.1069.

Trisodium Cytidin-5'-yl-[(1S,3R,4R)-3-benzyloxy-4-acetamido-1cyclopentyl-phosphonatomethyl]-phosphate (5S). To a solution of 47c (117.7 mg, 0.089 mmol) in EtOH (2.7 mL) were added Pd/C (235.4 mg, 10%) and 1,4-cyclohexadiene (847.0 mg, 1.0 mL, 10.68 mmol, 97%) under argon atmosphere. The reaction mixture was stirred at room temperature until TLC showed complete consumption of the starting material. Then the mixture was filtered and concentrated. The residue was purified by column chromatography on RP-18 silica gel (H<sub>2</sub>O  $\rightarrow$  MeOH/H<sub>2</sub>O = 1:1) to give crude product 5S (25.2 mg, 44% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ionexchange (IR 120 Na<sup>+</sup>) and lyophilized from water to give 5Ss and 5Sl as white powder. 5Ss: RP-HPLC, column B, 0-18 min linear gradient 7.0-8.8% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/ min flow;  $t_{\rm R} = 15.72$  min;  $[\alpha]_{\rm D}^{30} = -1.7$  (c = 0.7,  $H_2$ O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.94 (d, J = 7.2 Hz, 1H), 7.42–7.35 (m, 5H), 6.08 (d, J = 7.8 Hz, 1H), 5.97 (d, J = 4.2 Hz, 1H), 4.59 (d, J = 11.4 Hz, 1H), 4.51 (d, J = 12.0 Hz, 1H), 4.32 (t, J = 5.4 Hz, 1H), 4.27–4.07 (m, 6H), 3.87 (dt, J = 7.2, 4.8 Hz, 1H), 2.68–2.60 (m, 1H), 2.20 (dt, J = 12.6, 7.8 Hz, 1H), 2.12 (dt, J = 13.8, 9.0 Hz, 1H), 1.94–1.90 (m, 1H), 1.89 (s, 3H), 1.50 (dt, J = 12.6, 10.2 Hz, 1H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O)  $\delta$  174.0, 166.5, 157.9, 142.2, 138.1, 129.4, 129.1, 128.9, 97.1, 89.9, 84.2, 83.3 (d, J = 9.1 Hz), 76.7 (dd, J = 157.0, 7.6 Hz), 75.0, 71.8, 69.7, 64.7 (d, J = 4.5 Hz), 56.4, 37.6, 34.3 (d, J = 9.1 Hz), 32.3 (d, J = 4.5 Hz), 22.6 ppm. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  15.55, 0.11. HRMS (ESI) calcd for  $C_{24}H_{33}N_4O_{13}P_2$  [M - H]<sup>-</sup>, 647.1519; found, 647.1513. **5***Sl*: RP-HPLC, column B, 0-18 min linear gradient 7.0-8.8% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_{\rm R}$  = 16.70 min;  $[\alpha]_{D}^{30} = +0.5 \ (c = 0.4, H_2O).$ <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.97 (d, J = 7.8 Hz, 1H), 7.42–7.36 (m, 5H), 6.11 (d, J = 7.2 Hz, 1H), 5.94 (d, J = 3.6 Hz, 1H), 4.61 (d, J = 12.0 Hz, 1H), 4.55 (d, J = 12.0 Hz, 1H), 4.29-4.24 (m, 5H), 4.19-4.16 (m, 1H), 4.14-4.10 (m, 1H), 3.91 (dt, J = 6.6, 4.8 Hz, 1H), 2.70–2.62 (m, 1H), 2.25 (dt, J = 13.8, 7.2 Hz,

1H), 2.08–2.02 (m, 1H), 1.92 (s, 3H), 1.92–1.89 (m, 1H), 1.58 (dt, *J* = 13.2, 9.6 Hz, 1H). <sup>13</sup>C NMR (151 MHz,  $D_2O$ )  $\delta$  174.1, 165.7, 156.9, 142.6, 138.2, 129.4, 129.1, 128.9, 97.0, 90.3, 84.5, 83.4 (d, *J* = 9.1 Hz), 76.4 (dd, *J* = 155.5, 7.6 Hz), 75.0, 71.8, 69.8, 65.2 (d, *J* = 4.5 Hz), 56.4, 37.5, 34.0 (d, *J* = 10.6 Hz), 32.3 (d, *J* = 4.5 Hz), 22.7. <sup>31</sup>P NMR (162 MHz,  $D_2O$ )  $\delta$  15.52, 0.48. HRMS (ESI) calcd for  $C_{24}H_{33}N_4O_{13}P_2$  [M – H]<sup>-</sup>, 647.1519; found, 647.1525.

Trisodium Cytidin-5'-yl-[(1R,3R,4R)-3-benzyloxy-4-acetamido-1cyclopentyl-phosphonatomethyl]-phosphate (5R). This compound was prepared in the same manner as described in the preparation of 5S. Quantities: 47d (210.7 mg, 0.16 mmol), Pd/C (421.4 mg, 10%), 1,4-cyclohexadiene (1.61 g, 1.9 mL, 19.08 mmol, 97%), EtOH (4.8 mL), and eluent (H<sub>2</sub>O  $\rightarrow$  MeOH/H<sub>2</sub>O = 1:1), affording crude product 5R (51.8 mg, 50% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ion-exchange (IR 120 Na<sup>+</sup>), and lyophilized from water to give 5Rs and 5Rl as white powder. 5Rs: RP-HPLC, column B, 0-16 min linear gradient 8-9.5% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_{\rm R} = 11.95$  min;  $[\alpha]_{\rm D}^{30} = +0.7$  (c = 0.3, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.96 (d, J = 7.8 Hz, 1H), 7.41–7.35 (m, 5H), 6.09 (d, J = 7.8 Hz, 1H), 5.93 (d, J = 3.6 Hz, 1H), 4.61 (d, J = 12.0 Hz, 1H), 4.55 (d, J = 11.4 Hz, 1H), 4.31 (t, J = 5.4 Hz, 1H), 4.28-4.17 (m, 5H), 4.10-4.07 (m, 1H), 3.91 (dt, J = 7.2, 6.6 Hz, 1H), 2.52–2.48 (m, 1H), 2.38 (dt, J = 12.6, 6.6 Hz, 1H), 2.12 (dt, J = 13.8, 9.6 Hz, 1H), 1.92 (s, 3H), 1.68-1.62 (m, 2H). <sup>13</sup>C NMR (151 MHz,  $D_2O$ )  $\delta$  174.0, 166.9, 158.3, 142.1, 138.2, 129.4, 129.2, 128.9, 97.1, 90.4, 85.3, 83.3 (d, J = 9.1 Hz), 76.4 (dd, J = 152.5, 15.1 Hz), 75.2, 72.1, 69.4, 64.8, 55.4, 37.6, 34.2 (d, J = 7.6 Hz), 33.8 (d, J = 4.5 Hz), 22.7. <sup>31</sup>P NMR (162 MHz,  $D_2O$ )  $\delta$  14.68, 0.56. HRMS (ESI) calcd for  $C_{24}H_{33}N_4O_{13}P_2$  [M - H]<sup>-</sup>, 647.1519; found, 647.1537. SRI: RP-HPLC, column B, 0-16 min linear gradient 8-9.5% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_{\rm R}$  = 12.58 min;  $[\alpha]_{D}^{30} = -3.7 \ (c = 0.4, H_2O).$ <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  7.98 (d, J = 7.8 Hz, 1H), 7.41–7.34 (m, 5H), 6.09 (d, J = 7.8 Hz, 1H), 5.90 (d, J = 3.0 Hz, 1H), 4.60 (d, J = 12.0 Hz, 1H), 4.53 (d, J = 11.4 Hz, 1H), 4.32 (t, J = 5.4 Hz, 1H), 4.28 - 4.19 (m, 5H), 4.05 (dt, J = 9.6, 4.8 Hz, 1H),3.91 (dt, J = 8.4, 6.6 Hz, 1H), 2.50 (brs, 1H), 2.27 (dt, J = 12.6, 6.0 Hz, 1H), 2.17 (dt, J = 13.8, 10.2 Hz, 1H), 1.89 (s, 3H), 1.68–1.62 (m, 2H).  $^{13}\mathrm{C}$  NMR (151 MHz, D2O)  $\delta$  173.9, 166.9, 158.3, 142.0, 138.2, 129.3, 129.1, 128.9, 97.0, 90.3, 85.0, 83.0 (d, J = 9.1 Hz), 77.2 (d, J = 157.0 Hz), 75.2, 72.1, 69.2 (d, J = 7.6 Hz), 64.3, 55.3, 37.3, 35.0 (d, J = 9.1 Hz), 32.6 (d, J = 6.0 Hz), 22.6. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$ 14.95, 0.36. HRMS (ESI) calcd for  $C_{24}H_{33}N_4O_{13}P_2$  [M - H]<sup>-</sup>, 647.1519; found, 647.1525.

Trisodium Cytidin-5'-yl-[(1R,3R,4S)-3-benzyloxy-4-acetamidomethyl-1-cyclopentyl-phosphonatomethyl]-phosphate (6). This compound was prepared in the same manner as described in the preparation of 5S. Quantities: 47e (68.4 mg, 0.051 mmol), Pd/C (136.8 mg, 10%), 1,4-cyclohexadiene (508.2 mg, 0.60 mL, 6.120 mmol, 97%), EtOH (2.2 mL), and eluent (H<sub>2</sub>O  $\rightarrow$  MeOH/H<sub>2</sub>O = 1:1), affording crude product 6 (17.6 mg, 52% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ion-exchange (IR 120 Na<sup>+</sup>) and lyophilized from water to give 6s and 6l as white powder. 6s: RP-HPLC, column B, 0-16 min linear gradient 8-11.5% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_{\rm R}$  = 12.65 min;  $[\alpha]_{D}^{30} = -3.6 \ (c = 0.2, H_2O).$ <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)  $\delta$  8.01 (d, J = 7.8 Hz, 1H), 7.41–7.37 (m, 5H), 6.08 (d, J = 7.8 Hz, 1H), 5.93 (d, J = 3.6 Hz, 1H, 4.59 (d, J = 11.4 Hz, 1H), 4.46 (d, J = 11.4 Hz, 1H), 4.33 $(t, J = 6.0 \text{ Hz}_{1}, 1\text{H}), 4.26-4.17 \text{ (m, 5H)}, 3.75 \text{ (dt, } J = 8.4, 6.6 \text{ Hz}, 1\text{H}),$ 3.20 (dd, J = 13.8, 7.2 Hz, 5H), 3.12 (dd, J = 13.8, 7.2 Hz, 1H), 2.47 (dt, J = 12.6, 6.6 Hz, 1H), 2.41 (brs, 1H), 2.18–2.12 (m, 1H), 1.99– 1.93 (m, 1H), 1.92 (s, 3H), 1.64 (td, J = 11.4, 9.0 Hz, 1H), 1.54 (ddd, J = 12.6, 7.8, 4.8 Hz, 1H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O)  $\delta$  174.7, 166.9, 158.3, 142.2, 138.2, 129.4, 129.3, 128.9, 97.1, 90.5, 84.1, 83.4 (d, J = 9.1 Hz), 79.0 (d, J = 149.5 Hz), 75.3, 71.9, 69.2, 64.5 (d, J = 4.5 Hz), 43.6, 43.1, 38.4, 35.1 (d, J = 4.5 Hz), 31.5 (d, J = 7.6 Hz), 22.6. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O) δ 14.38, 0.70. HRMS (ESI) calcd for  $C_{25}H_{35}N_4O_{13}P_2 [M - H]^-$ , 661.1676; found, 661.1675. 6l: RP-HPLC, column B, 0-16 min linear gradient 8-11.5% CH<sub>3</sub>CN, 100 mM

NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_R$  = 13.77 min; [α]<sub>D</sub><sup>30</sup> = -7.1 (*c* = 0.3, H<sub>2</sub>O). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 7.98 (d, *J* = 7.6 Hz, 1H), 7.45-7.35 (m, 5H), 6.11 (d, *J* = 7.6 Hz, 1H), 5.94 (d, *J* = 3.6 Hz, 1H), 4.56 (d, *J* = 11.2 Hz, 1H), 4.45 (d, *J* = 11.2 Hz, 1H), 4.33 (t, *J* = 5.6 Hz, 1H), 4.30-4.22 (m, 5H), 3.76 (dt, *J* = 8.4, 6.4 Hz, 1H), 3.21 (dd, *J* = 13.2, 6.8 Hz, 1H), 3.10 (dd, *J* = 13.6, 6.8 Hz, 1H), 2.43 (brs, 1H), 2.32 (dt, *J* = 12.4, 6.4 Hz, 1H), 2.10-2.07 (m, 1H), 2.04-1.96 (m, 1H), 1.91 (s, 3H), 1.68-1.62 (m, 1H), 1.59-1.55 (m, 1H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.6, 166.9, 158.3, 142.2, 138.3, 129.3, 129.1, 128.9, 97.1, 90.3, 83.8, 83.1 (d, *J* = 9.1 Hz), 78.1 (d, *J* = 154.0 Hz), 75.2, 72.0, 69.0, 64.0, 43.7, 43.2, 37.9, 36.5 (d, *J* = 7.6 Hz), 29.1, 22.6. <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O) δ 14.36, 0.42. HRMS (ESI) calcd for C<sub>25</sub>H<sub>35</sub>N<sub>4</sub>O<sub>13</sub>P<sub>2</sub> [M - H]<sup>-</sup>, 661.1676; found, 661.1702.

Trisodium Cytidin-5'-yl-[phosphonato-1-((2S,3S)-2-benzyloxy-3acetamido-methyl-cyclopentyl)]-phosphate (48). This compound was prepared in the same manner as described in the preparation of 5S. Quantities: 47f (16.1 mg, 0.012 mmol), Pd/C (16.1 mg, 10%), 1,4cyclohexadiene (59.3 mg, 70 µL, 0.720 mmol, 97%), EtOH (0.5 mL), and eluent (H<sub>2</sub>O  $\rightarrow$  MeOH/H<sub>2</sub>O = 1:1), affording crude product 48 (5.8 mg, 73% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ionexchange (IR 120 Na<sup>+</sup>), and lyophilized from water to give 48s and 48l as white powder. 48s: RP-HPLC, column A, 6.0% CH<sub>3</sub>CN, 0.05 M TEAB (triethylammonium bicarbonate (pH 7.2-7.5)) buffer, 15 mL/ min flow;  $t_{\rm R} = 6.13$  min;  $[\alpha]_{\rm D}^{30} = -22.2$  (c = 0.9, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz,  $D_2O$ )  $\delta$  7.98 (d, J = 7.8 Hz, 1H, H6'), 7.37–7.33 (m, 5H, Ar), 6.11 (d, J = 7.8 Hz, 1H, H7'), 5.77 (d, J = 1.8 Hz, 1H, H1'), 4.92 (d, J = 11.4 Hz, 1H, PhCH<sub>2</sub>), 4.54 (d, J = 11.4 Hz, 1H, PhCH<sub>2</sub>), 4.41 (dd, J = 7.2, 4.8 Hz, 1H, H3'), 4.34–4.28 (m, 2H, H5' $\alpha$ , H5' $\beta$ ), 4.12–4.10 (m, 1H, H4'), 3.91 (ddd, J = 9.6, 5.4, 4.2 Hz, 1H, H2), 3.50-3.49 (m, 1H, H2'), 3.28 (dd, J = 13.2, 5.4 Hz, 1H, H6 $\alpha$ ), 3.13 (dd, J = 13.8, 6.6 Hz, 1H, H6 $\beta$ ), 2.43–2.33 (m, 2H, H3, H4 $\alpha$ ), 2.27 (td, J = 15.6, 9.0 Hz, 1H, H4 $\beta$ ), 2.00–1.94 (m, 1H, H5 $\alpha$ ), 1.87 (s, 3H, CH<sub>3</sub>), 1.37– 1.31 (m, 1H, H5β). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.7 (CH<sub>3</sub>CO), 164.7 (C8'), 155.2 (C9'), 142.8 (C6'), 138.9 (Ph), 129.2 (Ph), 129.1 (Ph), 128.8 (Ph), 96.4 (C7'), 90.7 (C1'), 86.0 (dd, J = 157.0, 7.6 Hz C1), 85.9 (dd, J = 6.0, 3.0 Hz C2), 82.7 (d, J = 6.0 Hz C4'), 75.1 (C2'), 74.0 (PhCH<sub>2</sub>), 68.2 (C3'), 64.2 (d, J = 6.0 Hz C5'), 43.1 (d, J = 9.1 Hz C3), 42.1 (C6), 32.2 (d, J = 6.0 Hz C4), 23.3 (d, J = 9.1 Hz C5), 22.6 (CH<sub>2</sub>). <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  17.21, -2.48. HRMS (ESI) calcd for C<sub>24</sub>H<sub>33</sub>N<sub>4</sub>O<sub>13</sub>P<sub>2</sub> [M – H]<sup>-</sup>, 647.1519; found, 647.1522. 481: RP-HPLC, column A, 6.0% CH<sub>3</sub>CN, 0.05 M TEAB (triethylammonium bicarbonate (pH 7.2-7.5)) buffer, 15 mL/min flow;  $t_{\rm R} = 10.55$  min;  $[\alpha]_{\rm D}^{30} = -11.5$  (c = 0.5, H<sub>2</sub>O). <sup>1</sup>H NMR (600 MHz,  $D_2O$ )  $\delta$  7.94 (d, J = 7.8 Hz, 1H, H6'), 7.44–7.35 (m, 5H, Ar), 6.02 (d, J = 7.8 Hz, 1H, H7'), 5.92 (d, J = 3.6 Hz, 1H, H1'), 4.99 (d, J = 12.0 Hz, 1H, PhCH<sub>2</sub>), 4.61 (d, J = 12.0 Hz, 1H, PhCH<sub>2</sub>), 4.35 (t, J = 5.4 Hz, 1H, H3'), 4.31–4.29 (m, 1H, H5' $\alpha$ ), 4.26 (t, J = 4.2 Hz, 1H, H2'), 4.24–4.19 (m, 3H, H4', H2, H5' $\beta$ ), 3.16 (dd, J = 13.8, 6.0 Hz, 1H, H6 $\alpha$ ), 3.08 (dd, J = 13.8, 7.2 Hz, 1H, H6 $\beta$ ), 2.44–2.34 (m, 2H,  $H4\alpha$ ,  $H4\beta$ ), 2.33–2.28 (m, 1H, H3), 1.84 (s, 3H, CH<sub>3</sub>), 1.83–1.77 (m, 1H, H5 $\alpha$ ), 1.55–1.48 (m, 1H, H5 $\beta$ ). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O)  $\delta$ 174.6 (CH<sub>3</sub>CO), 166.8 (C8'), 158.3 (C9'), 142.1 (C6'), 139.0 (Ph), 129.6 (Ph), 129.2 (Ph), 128.8 (Ph), 97.2 (C7'), 90.4 (C2), 90.4 (d, J = 160.1, 9.1 Hz C1), 90.2 (C1'), 83.1 (d, J = 9.1 Hz C4'), 75.0 (C2'), 73.9 (PhCH<sub>2</sub>), 69.4 (C3'), 64.5 (d, J = 6.0 Hz C5'), 42.7 (C3), 42.4 (C6), 32.0 (C4), 24.6 (d, J = 6.0 Hz C5), 22.5 (CH<sub>3</sub>). <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  15.60 (d, J = 25.9 Hz), -3.45 (d, J = 25.9 Hz). HRMS (ESI) calcd for  $C_{24}H_{33}N_4O_{13}P_2 [M - H]^-$ , 647.1519; found, 647.1511.

Trisodium Cytidin-5'-yl-[(3-phenoxy)phenylphosphonatomethyl]-phosphate (49). This compound was prepared in the same manner as described in the preparation of 2. Quantities: 47g (61.2 mg, 0.05 mmol), Pd/C (61.2 mg, 10%), 1,4-cyclohexadiene (485.3 mg, 573  $\mu$ L, 5.88 mmol, 97%), DMF (1.0 mL), and eluent (H<sub>2</sub>O  $\rightarrow$  MeOH/ H<sub>2</sub>O = 1:1), affording crude product 49 (17.0 mg, 60% yield). The diastereoisomers were further separated by preparative RP-HPLC, converted to the form of sodium salt by ion-exchange (IR 120 Na<sup>+</sup>), and lyophilized from water to give 49s and 49l as white powder. 49s: RP-HPLC, column B,  $\lambda$  = 260 nm, 0–12 min linear gradient 4–8% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow; t<sub>R</sub> = 10.10 min. **491**: RP-HPLC, column B,  $\lambda$  = 260 nm, 0–12 min linear gradient 4–8% CH<sub>3</sub>CN, 100 mM NH<sub>4</sub>HCO<sub>3</sub> buffer (pH = 8.4), 15 mL/min flow;  $t_{\rm R}$  = 10.80 min. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  = 7.66 (d, J = 7.6 Hz, 1H), 7.34–7.29 (m, 3H), 7.26 (t, J = 7.6 Hz, 1H), 7.15 (brs, 1H), 7.11 (t, J = 7.6 Hz, 1H), 6.98–6.96 (m, 2H), 6.84–6.82 (m, 1H), 6.03 (d, J = 7.6 Hz, 1H), 5.80 (d, J = 3.6 Hz, 1H), 5.02 (dd, J = 13.2, 10.4 Hz, 1H), 3.97–3.95 (m, 2H), 3.90–3.86 (m, 2H), 3.70 ppm (d, J = 12.0 Hz, 1H). The spectroscopic data coincide with the previous report.<sup>36</sup>

**RP-HPLC Based Sialyltransferase Assay.** The assay was performed according to the UV/RP-HPLC method developed by Schmidt group.<sup>30</sup> For details of the procedures, see Supporting Information.

## ASSOCIATED CONTENT

#### **Supporting Information**

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jmed-chem.5b01181.

Procedures for preparing compounds 9, 10, and 11, HPLC purity of the target compounds, details of biological assay, and NMR spectral data (PDF) Molecular formula strings (CSV)

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#### Notes

The authors declare no competing financial interest.

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# REFERENCES

(1) Angata, T.; Varki, A. Chemical diversity in the sialic acids and related alpha-keto acids: an evolutionary perspective. *Chem. Rev.* 2002, 102, 439–469.

(2) Varki, A. Glycan-based interactions involving vertebrate sialicacid-recognizing proteins. *Nature* **200**7, 446, 1023–1029.

(3) Severi, E.; Hood, D. W.; Thomas, G. H. Sialic acid utilization by bacterial pathogens. *Microbiology* **2007**, *153*, 2817–2822.

(4) Varki, N. M.; Varki, A. Diversity in cell surface sialic acid presentations: implications for biology and disease. *Lab. Invest.* 2007, *87*, 851–857.

(5) Harduin-Lepers, A.; Krzewinski-Recchi, M. A.; Colomb, F.; Foulquier, F.; Groux-Degroote, S.; Delannoy, P. Sialyltransferases functions in cancers. *Front. Biosci., Elite Ed.* **2012**, *4*, 499–515.

(6) Dall Olio, F.; Malagolini, N.; di Stefano, G.; Minni, F.; Marrano, D.; Serafini-Cessi, F. Increased CMP-NeuAc:Gal beta 1,4GlcNAc-R alpha 2,6 sialyltransferase activity in human colorectal cancer tissues. *Int. J. Cancer* **1989**, *44*, 434–439.

(7) Yamamoto, H.; Saito, T.; Kaneko, Y.; Kersey, D.; Yong, V. W.; Bremer, E. G.; Mkrdichian, E.; Cerullo, L.; Leestma, J.; Moskal, J. R. alpha2,3-sialyltransferase mRNA and alpha2,3-linked glycoprotein sialylation are increased in malignant gliomas. *Brain Res.* **1997**, *755*, 175–179.

(8) Burchell, J.; Poulsom, R.; Hanby, A.; Whitehouse, C.; Cooper, L.; Clausen, H.; Miles, D.; Taylor-Papadimitriou, J. An alpha2,3 sialyltransferase (ST3Gal I) is elevated in primary breast carcinomas. *Glycobiology* **1999**, *9*, 1307–1311.

(9) Wang, P. H.; Li, Y. F.; Juang, C. M.; Lee, Y. R.; Chao, H. T.; Tsai, Y. C.; Yuan, C. C. Altered mRNA expression of sialyltransferase in squamous cell carcinomas of the cervix. *Gynecol. Oncol.* **2001**, *83*, 121–127.

(10) Thampoe, I. J.; Furukawa, K.; Vellve, E.; Lloyd, K. O. Sialyltransferase levels and ganglioside expression in melanoma and other cultured human cancer cells. *Cancer Res.* **1989**, *49*, 6258–6264.

(11) Kanani, A.; Sutherland, D. R.; Fibach, E.; Matta, K. L.; Hindenburg, A.; Brockhausen, I.; Kuhns, W.; Taub, R. N.; van den Eijnden, D. H.; Baker, M. A. Human leukemic myeloblasts and myeloblastoid cells contain the enzyme cytidine S'-monophosphate-Nacetylneuraminic acid:Gal beta 1–3GalNAc alpha (2–3)-sialyltransferase. *Cancer Res.* **1990**, *50*, 5003–5007.

(12) Wang, X.; Niu, Y.; Cao, X.; Zhang, L.; Zhang, L.-H.; Ye, X.-S. 3D-QSAR analysis of sialyltransferase inhibitors. *Bioorg. Med. Chem.* 2003, 11, 4217–4224.

(13) Wang, X.; Zhang, L.-H.; Ye, X.-S. Recent development in the design of sialyltransferase inhibitors. *Med. Res. Rev.* **2003**, *23*, 32–47.

(14) Drinnan, N. B.; Halliday, J.; Ramsdale, T. Inhibitors of sialyltransferases: potential roles in tumor growth and metastasis. *Mini-Rev. Med. Chem.* **2003**, *3*, 501–517.

(15) Jung, K.-H.; Schwörer, R.; Schmidt, R. R. Sialyltransferase Inhibitors. *Trends Glycosci. Glycotechnol.* **2003**, *15*, 275–289.

(16) Kijima-Suda, I.; Miyamoto, Y.; Toyoshima, S.; Itoh, M.; Osawa, T. Inhibition of experimental pulmonary metastasis of mouse colon adenocarcinoma 26 sublines by a sialic acid:nucleoside conjugate having sialyltransferase inhibiting activity. *Cancer Res.* **1986**, *46*, 858–862.

(17) Müller, B.; Martin, T. J.; Schaub, C.; Schmidt, R. R. Synthesis of phosphonate analogues of CMP-Neu5Ac determination of (2–6)-sialyltransferase inhibition. *Tetrahedron Lett.* **1998**, *39*, 509–512.

(18) Cohen, S. B.; Halcomb, R. L. Synthesis and characterization of an anomeric sulfur analogue of CMP-sialic acid. *J. Org. Chem.* 2000, 65, 6145–6152.

(19) Tanaka, T.; Ozawa, M.; Miura, T.; Inazu, T.; Tsuji, S.; Kajimoto, T. Synthesis of novel mimetics of CMP-sialic acid as the inhibitors of sialyltransferases. *Synlett* **2002**, *2002*, 1487–1490.

(20) Whalen, L. J.; McEvoy, K. A.; Halcomb, R. L. Synthesis and evaluation of phosphoramidate amino acid-based inhibitors of sialyltransferases. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 301–304.

(21) Kumar, R.; Nasi, R.; Bhasin, M.; Huan Khieu, N.; Hsieh, M.; Gilbert, M.; Jarrell, H.; Zou, W.; Jennings, H. J. Sialyltransferase inhibitors: consideration of molecular shape and charge/hydrophobic interactions. *Carbohydr. Res.* **2013**, *378*, 45–55.

(22) Hindsgaul, O.; Kaur, K. J.; Srivastava, G.; Blaszczyk-Thurin, M.; Crawley, S. C.; Heerze, L. D.; Palcic, M. M. Evaluation of deoxygenated oligosaccharide acceptor analogs as specific inhibitors of glycosyltransferases. *J. Biol. Chem.* **1991**, *266*, 17858–17862.

(23) Kajihara, Y.; Kodama, H.; Wakabayashi, T.; Sato, K.; Hashimoto, H. Characterization of inhibitory activities and binding mode of synthetic 6'-modified methyl N-acetyl-beta-lactosaminide toward rat liver CMP-D-NeuSAc: D-galactoside- $(2\rightarrow 6)$ -alpha-Dsialyltransferase. *Carbohydr. Res.* **1993**, 247, 179–193.

(24) Huet, G.; Hennebicq-Reig, S.; de Bolos, C.; Ulloa, F.; Lesuffleur, T.; Barbat, A.; Carriere, V.; Kim, I.; Real, F. X.; Delannoy, P.; Zweibaum, A. GalNAc-alpha-O-benzyl inhibits NeuAcalpha2–3 glycosylation and blocks the intracellular transport of apical glycoproteins and mucus in differentiated HT-29 cells. *J. Cell Biol.* **1998**, *141*, 1311–1322.

(25) Okazaki, K.; Nishigaki, S.; Ishizuka, F.; Kajihara, Y.; Ogawa, S. Potent and specific sialyltransferase inhibitors: imino-linked 5a'-carbadisaccharides. *Org. Biomol. Chem.* **2003**, *1*, 2229–2230.

(26) Xia, J.; Xue, J.; Locke, R. D.; Chandrasekaran, E. V.; Srikrishnan, T.; Matta, K. L. Synthesis of fluorinated mucin core 2 branched oligosaccharides with the potential of novel substrates and enzyme inhibitors for glycosyltransferases and sulfotransferases. *J. Org. Chem.* **2006**, *71*, 3696–3706.

(27) Hinou, H.; Sun, X.; Ito, Y. Bisubstrate-type inhibitor of sialyltransferases. *Tetrahedron Lett.* **2002**, 43, 9147–9150.

(28) Hinou, H.; Sun, X. L.; Ito, Y. Systematic syntheses and inhibitory activities of bisubstrate-type inhibitors of sialyltransferases. *J. Org. Chem.* **2003**, *68*, 5602–5613.

(29) Izumi, M.; Wada, K.; Yuasa, H.; Hashimoto, H. Synthesis of bisubstrate and donor analogues of sialyltransferase and their inhibitory activities. *J. Org. Chem.* **2005**, *70*, 8817–8824.

(30) Schaub, C.; Müller, B.; Schmidt, R. R. New sialyltransferase inhibitors based on CMP-quinic acid: development of a new sialyltransferase assay. *Glycoconjugate J.* **1998**, *15*, 345–354.

(31) Amann, F.; Schaub, C.; Müller, B.; Schmidt, R. R. New potent sialyltransferase inhibitors—synthesis of donor and of transition-state analogues of sialyl donor CMP-NeuSAc. *Chem. - Eur. J.* **1998**, *4*, 1106–1115.

(32) Müller, B.; Schaub, C.; Schmidt, R. R. Efficient sialyltransferase inhibitors based on transition-state analogues of the sialyl donor. *Angew. Chem., Int. Ed.* **1998**, 37, 2893–2897.

(33) Schaub, C.; Müller, B.; Schmidt, R. R. Sialyltransferase inhibitors based on CMP-quinic acid. *Eur. J. Org. Chem.* **2000**, 2000, 1745–1758.

(34) Sun, H.; Yang, J.; Amaral, K. E.; Horenstein, B. A. Synthesis of a new transition-state analog of the sialyl donor. Inhibition of sialyltransferases. *Tetrahedron Lett.* **2001**, *42*, 2451–2453.

(35) Schwörer, R.; Schmidt, R. R. Efficient sialyltransferase inhibitors based on glycosides of N-acetylglucosamine. J. Am. Chem. Soc. 2002, 124, 1632–1637.

(36) Skropeta, D.; Schwörer, R.; Haag, T.; Schmidt, R. R. Asymmetric synthesis and affinity of potent sialyltransferase inhibitors based on transition-state analogues. *Glycoconjugate J.* **2004**, *21*, 205–219.

(37) Wu, C. Y.; Hsu, C. C.; Chen, S. T.; Tsai, Y. C. Soyasaponin I, a potent and specific sialyltransferase inhibitor. *Biochem. Biophys. Res. Commun.* 2001, 284, 466–469.

(38) Lee, K. Y.; Kim, H. G.; Hwang, M. R.; Chae, J. I.; Yang, J. M.; Lee, Y. C.; Choo, Y. K.; Lee, Y. I.; Lee, S. S.; Do, S. I. The Hexapeptide inhibitor of Galbeta 1,3GalNAc-specific alpha 2,3-sialyltransferase as a generic inhibitor of sialyltransferases. *J. Biol. Chem.* **2002**, 277, 49341– 49351.

(39) Lin, T. W.; Chang, W. W.; Chen, C. C.; Tsai, Y. C. Stachybotrydial, a potent inhibitor of fucosyltransferase and sialyltransferase. *Biochem. Biophys. Res. Commun.* **2005**, *331*, 953–957.

(40) Chang, K. H.; Lee, L.; Chen, J.; Li, W. S. Lithocholic acid analogues, new and potent alpha-2,3-sialyltransferase inhibitors. *Chem. Commun.* 2006, 629–631.

(41) Hidari, K. I.; Oyama, K.; Ito, G.; Nakayama, M.; Inai, M.; Goto, S.; Kanai, Y.; Watanabe, K.; Yoshida, K.; Furuta, T.; Kan, T.; Suzuki, T. Identification and characterization of flavonoids as sialyltransferase inhibitors. *Biochem. Biophys. Res. Commun.* **2009**, *382*, 609–613.

(42) Pauling, L. Molecular architecture and biological reactions. *Chem. Eng. News* **1946**, *24*, 1375–1377.

(43) Chang, W. W.; Yu, C. Y.; Lin, T. W.; Wang, P. H.; Tsai, Y. C. Soyasaponin I decreases the expression of alpha2,3-linked sialic acid on the cell surface and suppresses the metastatic potential of B16F10 melanoma cells. *Biochem. Biophys. Res. Commun.* **2006**, 341, 614–619. (44) Chen, J. Y.; Tang, Y. A.; Huang, S. M.; Juan, H. F.; Wu, L. W.; Sun, Y. C.; Wang, S. C.; Wu, K. W.; Balraj, G.; Chang, T. T.; Li, W. S.; Cheng, H. C.; Wang, Y. C. A novel sialyltransferase inhibitor suppresses FAK/paxillin signaling and cancer angiogenesis and metastasis pathways. *Cancer Res.* **2011**, *71*, 473–483.

(45) Heasley, B. Recent developments in the stereocontrolled synthesis of highly substituted cyclopentane core structures: from drug discovery research to natural product synthesis. *Curr. Org. Chem.* 2014, *18*, 641–686.

(46) Daluge, S. M.; Good, S. S.; Faletto, M. B.; Miller, W. H., St; Clair, M. H.; Boone, L. R.; Tisdale, M.; Parry, N. R.; Reardon, J. E.; Dornsife, R. E.; Averett, D. R.; Krenitsky, T. A. 1592U89, a novel carbocyclic nucleoside analog with potent, selective anti-human immunodeficiency virus activity. *Antimicrob. Agents Chemother.* **1997**, *41*, 1082–1093. (47) Zhu, R.; Snyder, A. H.; Kharel, Y.; Schaffter, L.; Sun, Q.; Kennedy, P. C.; Lynch, K. R.; Macdonald, T. L. VPC01091-Asymmetric synthesis of conformationally constrained fingolimod analogues-discovery of an orally active sphingosine 1-phosphate receptor type-1 agonist and receptor type-3 antagonist. *J. Med. Chem.* **2007**, *50*, 6428–6435.

(48) Shetty, A. K.; Peek, L. A. Peramivir for the treatment of influenza. *Expert Rev. Anti-Infect. Ther.* 2012, 10, 123-143.

(49) Arjona, O.; Gomez, A. M.; Lopez, J. C.; Plumet, J. Synthesis and conformational and biological aspects of carbasugars. *Chem. Rev.* 2007, *107*, 1919–2036.

(50) Horenstein, B. A. Quantum mechanical analysis of an  $\alpha$ -carboxylate-substituted oxocarbenium ion. Isotope effects for formation of the sialyl cation and the origin of an unusually large secondary 14C isotope effect. *J. Am. Chem. Soc.* **1997**, *119*, 1101–1107.

(51) Bruner, M.; Horenstein, B. A. Isotope trapping and kinetic isotope effect studies of rat liver alpha- $(2\rightarrow 6)$ -sialyltransferase. *Biochemistry* **1998**, 37, 289–297.

(52) Bruner, M.; Horenstein, B. A. Use of an altered sugar-nucleotide to unmask the transition state for  $alpha(2\rightarrow 6)$  sialyltransferase. *Biochemistry* **2000**, *39*, 2261–2268.

(53) Dufner, G.; Schwörer, R.; Müller, B.; Schmidt, R. R. Base- and sugar-modified cytidine monophosphate *N*-acetylneuraminic acid (CMP-NeuSAc) analogues - synthesis and studies with  $\alpha$ (2–6)-sialyltransferase from rat liver. *Eur. J. Org. Chem.* **2000**, 2000, 1467–1482.

(54) Palmer, A. M.; Jäger, V. Pyrrolidine *N*-oxides by stereoselective addition of grignard and lithium compounds to 4,5-dideoxy-2,3-O-isopropylidene-D-erythro-4-pentenose *N*-benzyl nitrone and subsequent Cope–House cyclization. *Eur. J. Org. Chem.* **2001**, 2001, 1293–1308.

(55) Ivanova, N. A.; Valiullina, Z. R.; Shitikova, O. V.; Miftakhov, M. S. Reaction of methyl-4-methylene-2,3-O-isopropylidene-β-D-ribofuranoside with N-bromosuccinimide in aqueous tetrahydrofurane. *Russ. Russ. J. Org. Chem.* **2007**, *43*, 742–746.

(56) Jigajinni, V. B.; Wightman, R. H. An enantiospecific synthesis of (+)-disparlure from carbohydrate precursors. *Carbohydr. Res.* **1986**, *147*, 145–148.

(57) Denmark, S. E.; Forbes, D. C.; Hays, D. S.; DePue, J. S.; Wilde, R. G. Catalytic epoxidation of alkenes with oxone. *J. Org. Chem.* **1995**, *60*, 1391–1407.

(58) Soai, K.; Ookawa, A. Mixed solvents containing methanol as useful reaction media for unique chemoselective reductions within lithium borohydride. *J. Org. Chem.* **1986**, *51*, 4000–4005.

(59) Wang, H.; She, J.; Zhang, L.-H.; Ye, X.-S. Silver(I) oxide mediated selective monoprotection of diols in pyranosides. *J. Org. Chem.* **2004**, *69*, 5774–5777.

(60) Dondoni, A.; Perrone, D.; Semola, M. T. Thiazole-based stereoselective routes to leucine and phenylalanine hydroxyethylene dipeptide isostere inhibitors of renin and HIV-1 aspartic protease. *J. Org. Chem.* **1995**, *60*, 7927–7933.

(61) Rosen, T.; Lico, I. M.; Chu, D. T. W. A convenient and highly chemoselective method for the reductive acetylation of azides. *J. Org. Chem.* **1988**, *53*, 1580–1582.

(62) Kolakowski, R. V.; Shangguan, N.; Sauers, R. R.; Williams, L. J. Mechanism of thio acid/azide amidation. *J. Am. Chem. Soc.* **2006**, *128*, 5695–5702.

(63) Alexander, V.; Choi, W. J.; Chun, J.; Kim, H. O.; Jeon, J. H.; Tosh, D. K.; Lee, H. W.; Chandra, G.; Choi, J.; Jeong, L. S. A new DNA building block, 4'-selenothymidine: synthesis and modification to 4'-seleno-AZT as a potential anti-HIV agent. *Org. Lett.* **2010**, *12*, 2242–2245.

(64) Baeckvall, J. E.; Plobeck, N. A. New synthesis of the 6Hpyrido[4,3-b]carbazoles ellipticine and olivacine via cycloaddition of 2phenylsulfonyl 1,3-dienes to indoles. *J. Org. Chem.* **1990**, *55*, 4528– 4531. (65) Yoon, N. M.; Gyoung, Y. S. Reaction of diisobutylaluminum hydride with selected organic compounds containing representative functional groups. *J. Org. Chem.* **1985**, *50*, 2443–2450.

(66) Tsang, D. S.; Yang, S.; Alphonse, F. A.; Yudin, A. K. Stereoselective isomerisation of N-allyl aziridines into geometrically stable Z enamines by using rhodium hydride catalysis. *Chem. - Eur. J.* **2008**, *14*, 886–894.

(67) Burgess, K.; Ohlmeyer, M. J. Manipulation of substratecontrolled diastereoselectivities in hydroborations of acyclic allylamine derivatives. J. Org. Chem. **1991**, *56*, 1027–1036.

(68) Cívicos, J. F.; Alonso, D. A.; Nájera, C. Oxime–Palladacyclecatalyzed Suzuki–Miyaura arylation and alkenylation of aryl imidazolesulfonates under aqueous and phosphane-free conditions. *Eur. J. Org. Chem.* **2012**, 2012, 3670–3676.

(69) Carpintero, M.; Jaramillo, C.; Fernández-Mayoralas, A. Stereoselective synthesis of carba- and C-glycosyl analogs of fucopyranosides. *Eur. J. Org. Chem.* **2000**, 2000, 1285–1296.

(70) Taylor, S. D.; Lunn, F. A.; Bearne, S. L. Ground state, intermediate, and multivalent nucleotide analogue inhibitors of cytidine 5'-triphosphate synthase. *ChemMedChem* **2008**, *3*, 1853–1857.

(71) Johnson, D. C., II; Widlanski, T. S. Facile deprotection of O-Cbz-protected nucleosides by hydrogenolysis: an alternative to Obenzyl ether-protected nucleosides. *Org. Lett.* **2004**, *6*, 4643–4646.

(72) Dumbre, S.; Derouaux, A.; Lescrinier, E.; Piette, A.; Joris, B.; Terrak, M.; Herdewijn, P. Synthesis of modified peptidoglycan precursor analogues for the inhibition of glycosyltransferase. *J. Am. Chem. Soc.* **2012**, *134*, 9343–9351.

(73) Marschner, C.; Penn, G.; Griengl, H. Synthesis of  $\alpha$ - and  $\beta$ -D-carbaribofuranose from (+)-norborn-5-en-2-one. *Tetrahedron Lett.* **1990**, 31, 2873–2874.

(74) Allevi, P.; Cribiù, R.; Anastasia, M. Facile and rapid regeneration of free amino acids from N-benzyloxycarbonyl-5-oxazolidinones and from N-benzyloxycarbonylamino derivatives by treatment with BCl<sub>3</sub> in dichloromethane. *Tetrahedron Lett.* **2004**, *45*, 5841–5843.

(75) Bajwa, J. S. Chemoselective deprotection of benzyl esters in the presence of benzyl ethers, benzyloxymethyl ethers and n-benzyl groups by catalytic transfer hydrogenation. *Tetrahedron Lett.* **1992**, *33*, 2299–2302.

(76) Kuhn, B.; Benz, J.; Greif, M.; Engel, A. M.; Sobek, H.; Rudolph, M. G. The structure of human  $\alpha$ -2,6-sialyltransferase reveals the binding mode of complex glycans. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **2013**, *69*, 1826–1838.

(77) Legaigneur, P.; Breton, C.; El Battari, A.; Guillemot, J. C.; Auge, C.; Malissard, M.; Berger, E. G.; Ronin, C. Exploring the acceptor substrate recognition of the human beta-galactoside alpha 2,6-sialyltransferase. *J. Biol. Chem.* **2001**, *276*, 21608–21617.

(78) Boers, R. B.; Randulfe, Y. P.; van der Haas, H. N. S.; van Rossum-Baan, M.; Lugtenburg, J. Synthesis and spectroscopic characterization of 1–13C- and 4–13C-Plastoquinone-9. *Eur. J. Org. Chem.* 2002, 2094–2108.

(79) Matveeva, E. D.; Podrugina, T. A.; Grishin, Y. K.; Pavlova, A. S.; Zefirov, N. S. Phosphonium-iodonim ylides in nucleophilic substitution reactions. *Russ. J. Org. Chem.* **200**7, *43*, 201–206.

(80) Tadano, K.; Maeda, H.; Hoshino, M.; Iimura, Y.; Suami, T. A novel transformation of four aldoses to some optically pure pseudohexopyranoses and a pseudopentofuranose, carbocyclic analogs of hexopyranoses and pentofuranose. Synthesis of derivatives of (1S,2S,3R,4S,5S)-, (1S,2S,3R,4S,5S)-, (1S,2S,3R,4S,5S)-, (1S,2S,3R,4S,5R)-2,3,4,5-tetrahydroxy-1-(hydroxymethyl)-cyclohexanes and (1S,2S,3S,4S)-2,3,4-trihydroxy-1(hydroxymethyl)-cyclopentane. J. Org. Chem. **1987**, *52*, 1946–1956.

(81) McMurry, J. E.; Dushin, R. G. Total synthesis of  $(\pm)$ -crassin by titanium-induced pinacol coupling. J. Am. Chem. Soc. **1989**, 111, 8928–8929.

(82) Kikuchi, H.; Okazaki, K.; Sekiya, M.; Uryu, Y.; Ueda, K.; Katou, Y.; Kurata, S.; Oshima, Y. Synthesis and innate immunosuppressive effect of 1,2-cyclopentanediol derivatives. *Eur. J. Med. Chem.* **2011**, *46*, 1263–1273.