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Functional Structure/Activity Relationships

Synthesis and Biological Activity of Novel Succinate Dehydrogenase Inhibitor Derivatives as Potent Fungicide Candidates

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1	Synthesis and Biological Activity of Novel Succinate Dehydrogenase Inhibitor
2	Derivatives as Potent Fungicide Candidates
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13	

14 Abstract

In searching for novel fungicidal leads, the novel bioactive succinate dehydrogenase 15 inhibitor (SDHI) derivatives were designed and synthesized by the inversion of carbonyl and 16 17 amide groups. Bioassay indicated that compound **5i** was outstood with broad-spectrum of in vitro activity against five fungi, its EC₅₀ value (0.73 μ g/mL) was comparable to boscalid (EC₅₀ of 0.51 18 $\mu g/mL$) and fluxapyroxad (EC₅₀ of 0.19 $\mu g/mL$) against Sclerotinia sclerotiorum. For 19 20 *Rhizoctonia cerealis*, **5i** and **5p** with the EC₅₀ of 4.61 μ g/mL and 6.48 μ g/mL respectively, showed significant higher activity than fluxapyroxad with the EC₅₀ of 16.99 µg/mL. In vivo 21 fungicidal activity of 5i exhibited excellent inhibitory rate (100%) against Puccinia sorghi at 50 22 µg/mL, while the positive control boscalid showed only 70% of inhibitory rate. Moreover, 5i 23 showed promising fungicidal activity with 60% inhibitory rate against Rhizoctonia solani at 1 24 $\mu g/mL$, which was better than boscalid (30%). Compound **5i** possessed better in vivo efficacy 25 against P. sorghi and R. solani than boscalid. Molecular docking showed, even the carbonyl 26 oxygen atom of 5i was far from pyrazole ring, it could also form hydrogen bonds towards the 27 hydroxyl hydrogen, amino hydrogen of TYR58 and TRP173 on SDH, respectively, which 28 consisted with the positive control fluxapyroxad. Fluorescence quenching analysis and SDH 29 enzymatic inhibition studies also validated its mode of action. Our studies showed that 5i was 30 31 worthy for further investigation as a promising fungicide candidate.

32 Key words : SDHIs, amide inversion, molecular docking, fungicidal activity, rice disease

34 Introduction

Plant diseases cause a dramatic decline in crop quality and yield, farmers suffer huge economic losses.^{1, 2} Treating crops by fungicides with different mode of action to against plant disease damage is always necessary.³ Succinate dehydrogenase (SDH) inhibitors are widely used to protect crops from all kinds of plant pathogens attack.⁴⁻⁷ As an important membrane complex in the tricarboxylic acid cycle in mitochondrial respiratory chain, SDH is an ideal fungicide target.^{8, 9} SDHIs disrupt the respiration of the pathogen and inhibit its growth by inhibiting the oxidation of succinate to fumarate, and thus cause the pathogen death.^{10, 11}

Among the SDHIs, pyrazole amide derivatives have a central place in fungicidal 42 chemistry because of their broad spectrum of high efficiency, low toxicity and structure diversity. 43 As shown in Figure 1, furametpyr was invented by Sumitomo Chemical in 1986,¹² and then, 44 penflufen was developed by Bayer Cropscience in 2006.¹³ In recent decades, several pyrazole 45 contained fungicides, such as sedaxane,¹⁴ bixafen,¹⁵ fluxapyroxad,¹⁶ benzovindiflupyr,¹⁷ 46 pydiflumetofen¹⁸ and so on, have made a significant contribution to the crop protection against 47 plant diseases. All these representative compounds containing pyrazole moiety show outstanding 48 fungicidal activity. The wide practical interest in this important pyrazole moiety motivates 49 continuous efforts of the exploitation of efficient compounds.¹⁹⁻²¹ 50

In research for some commercially available aromatic amide fungicides, we found mebenil and benodanil, which could be seen as a couple of mirror compounds.²² Many documents reported the lead modification of SDHIs, but the design strategy by inversion of carbonyl and amide groups was rarely used in the studies of novel pyrazole fungicide development. On the basis of the development of fungicides targeted at SDH and prompted by the excellent biological activity of pyrazole carboxamide fungicides, we attempted to investigate the structure of pyrazole-contained derivatives, and target compounds were designed and synthesized by inversion of carbonyl and amide groups of fluxapyroxad (Figure 2). All of the target compounds were identified by ¹H NMR, elemental analysis and HRMS. The in vitro fungicidal activities of these compounds were evaluated against eight common plant pathogenic fungi and in vivo fungicidal activities of these compounds were tested against three different crop diseases. Furthermore, the enzymatic activity and fluorescence quenching analysis of **5**i to SDH were investigated to validate its mode of action.

64

65 MATERIALS AND METHODS

66 Equipment and Materials

67 Melting points were determined using an X-4 melting point apparatus and were uncorrected (Beijing Tech Instruments Co., Beijing, China). ¹H NMR spectra was obtained using 68 a Bruker Avance 400 MHZ spectrometer at 400 MHz in deuterochloroform (CDCl₃) and 69 70 tetramethylsilane (TMS) as internal standards (Bruker, Switzerland). Elemental analyses were performed on a Vario EL III elemental analysis instrument (Elementar, Germany). High 71 resolution mass spectrometry data were obtained with an Agilent 6520 Q-TOF LC/MS 72 instrument equipped with electrospray ionization (ESI) source (Agilent, America). Crystal 73 structure was recorded by a Rigaku 007 Saturn 70 diffraction meter (Rigaku MSC, Japan). All 74 75 solvents were of analytical grade. All of the yields were not optimized. The synthesis procedure of intermediate 4 and all the corresponding data could be found in the Supplementary 76 Information. 77

Molecular Docking. The molecular structures were drawn by ChemBioDraw Ultra 14.0
 and energetically minimized by using Chemdraw 3D Pro MM2 minimized procedure, then saved

as mol2 file. The structure of SDH from *Gallus gallus* (pdb code: 2FBW)²³ was downloaded
from RCSB PDB database and the docking program was performed with SYBYL-X 6.91. The
detail docking procedure was conducted according to the reported literature.²⁴

83 Synthetic Procedures

General Synthesis Procedure for Intermediate (1). The Methyl 2-iodobenzoate (3.6mmol), 84 substituted phenylboronic acid (4.5mmol) and dichloro-bis(triphenyl-phosphine)palladium(II) 85 (5%mmol) were added to a three-neck round-bottom flask equipped with a magnetic stir bar and 86 a reflux condenser. Under an atmosphere of argon, 15 mL of THF was injected to the mixture by 87 syringe, and then 1mol/L Na₂CO₃ was added in one portion. The flask was put into an oil bath 88 89 and heated to 65 °C for 12h. After completion, the flask was cooled to room temperature; 30ml 90 water was added and then extracted with CH_2Cl_2 (3×15 mL). The organic layer was combined and dried over anhydrous Na₂SO₄. Solvent was removed under the reduced pressure. The residue 91 92 was subjected to silica gel chromatography using PE and EtOAc (50:1, v/v) as an eluent to afford 93 intermediate (1) and used for the next step without further purification.

94 *General Synthesis Procedure for Intermediate* (2).

A solution of **1** (2.8mmol) dissolving in 10 mL THF was added in NaOH (28mmol, in 10mL H₂O). The mixture was heated to 80°C and monitored by TLC until the reaction was complete. Solvent was removed under the reduced pressure. The residue was acidified by 2mol/L HCl and precipitation was appeared. The precipitated solid was filtered out and recrystallized in CH₂Cl₂ and hexane to give pure products in an 88-99% yield.

100	Data for 3',4',5'-trifluoro-[1,1'-biphenyl]-2-carboxylic acid (2a). Yield 95%; white solid;
101	mp 155-156 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 10.80 (s, 1H), 8.03 (dd, <i>J</i> =7.8, 1.4 Hz, 1H), 7.60
102	(td, J=7.5, 1.4 Hz, 1H), 7.49 (td, J=7.7, 1.3 Hz, 1H), 7.30 (dd, J=7.6, 1.3 Hz, 1H), 6.99 – 6.87
103	(m, 2H).
104	Data for 3',4'-dichloro-[1,1'-biphenyl]-2-carboxylic acid (2b). Yield 96%; white solid; mp
105	173-174 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 9.59 (s, 1H), 7.97 (d, <i>J</i> =7.8 Hz, 1H), 7.60 (t, <i>J</i> =7.5
106	Hz, 1H), 7.47 (t, J = 7.6 Hz, 1H), 7.41 (d, J = 8.4 Hz, 2H), 7.32 (d, J = 7.6 Hz, 1H), 7.13 (dd, J
107	=8.2, 2.0 Hz, 1H).
108	Data for 4'-chloro-[1,1'-biphenyl]-2-carboxylic acid (2c). Yield 99%; white solid; mp 170-
109	171°C. ¹ H NMR (400 MHz, CDCl ₃) δ 10.92 (s, 1H), 7.98 (dd, J =7.8, 1.4 Hz, 1H), 7.57 (td, J
110	=7.6, 1.4 Hz, 1H), 7.44 (td, <i>J</i> =7.6, 1.3 Hz, 1H), 7.39 – 7.30 (m, 3H), 7.29 – 7.22 (m, 2H).
111	Data for 4'-(tert-butyl)-5-fluoro-[1,1'-biphenyl]-2-carboxylic acid (2d). Yield 88%; white
112	solid; mp 149-150 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 7.97 (dd, J =8.4, 5.9 Hz, 1H), 7.40 (d, J
113	=8.1 Hz, 2H), 7.27 (s, 1H), 7.25 (d, <i>J</i> =1.8 Hz, 1H), 7.12 – 7.03 (m, 2H), 1.35 (s, 9H).
114	Data for 3',4',5,5'-tetrafluoro-[1,1'-biphenyl]-2-carboxylic acid (2e). Yield 95%; white
115	solid; mp 186-187 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 8.11 (dd, <i>J</i> =8.8, 5.7 Hz, 1H), 7.20 (ddd, <i>J</i>
116	=8.8, 7.7, 2.6 Hz, 1H), 7.03 (dd, <i>J</i> =8.9, 2.6 Hz, 1H), 6.99 – 6.90 (m, 2H).
117	Data for 3',4'-dichloro-5-fluoro-[1,1'-biphenyl]-2-carboxylic acid (2f). Yield 94%; white
118	solid; mp 182-183°C. ¹ H NMR (400 MHz, CDCl ₃) δ 8.08 (td, <i>J</i> =8.3, 6.9, 3.5 Hz, 1H), 7.46 (dd,
119	<i>J</i> =8.4, 3.5 Hz, 1H), 7.41 (q, <i>J</i> =2.2 Hz, 1H), 7.15 (dddd, <i>J</i> =10.5, 6.8, 4.8, 2.0 Hz, 2H), 7.02 (dq,
120	<i>J</i> =9.2, 2.6 Hz, 1H).

121	Data for 5-fluoro-[1,1'-biphenyl]-2-carboxylic acid (2g). Yield 98%; white solid; mp 111-
122	112°C. ¹ H NMR (400 MHz, CDCl ₃) δ11.34 (s, 1H), 7.99 (dd, J =8.6, 5.8 Hz, 1H), 7.38 (dd, J
123	=5.5, 1.8 Hz, 3H), 7.30 (dt, <i>J</i> =6.7, 1.6 Hz, 2H), 7.14 – 7.02 (m, 2H).
124	Data for 4'-chloro-5-fluoro-[1,1'-biphenyl]-2-carboxylic acid (2h). Yield 93%; white solid;
125	mp 150-151 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 10.87 (s, 1H), 8.06 (dd, <i>J</i> =8.7, 5.8 Hz, 1H), 7.42
126	- 7.36 (m, 2H), 7.28 - 7.22 (m, 2H), 7.15 (ddd, <i>J</i> = 8.8, 7.8, 2.6 Hz, 1H), 7.04 (dd, <i>J</i> = 9.2, 2.6 Hz,
127	1H).
128	General Synthesis Procedure for Intermediate (3).

Compound 2 (3 mmol) and CH_2Cl_2 (30 mL) was added to a 100 mL single-neck round bottom flask and stirred in an ice bath. To the mixture, 2 drops of DMF was added and 6 mmol of oxalyl chloride solution was added dropwise. The mixture was slowly rise to room temperature, after stirring at room temperature for 2 h, the mixture was concentrated under the reduced pressure to remove excess oxalyl chloride solution and solvent to afford intermediate (3) and used for the next step without further purification.

135 *General Synthesis Procedure for target compound* (5).

In a 100 mL one-neck round bottom flask, 3 mL of compound **3** was added and dissolved in 30 mL of dry CH_2Cl_2 . After the addition of 6 mmol of triethylamine, the mixture was stirred for 10 min. in an ice bath. Compound **4** (3 mmol) was dissolved in 5 mL of dry CH_2Cl_2 and added dropwise to the reaction system, then the mixture was stirred for 30 min in an ice bath, and stirred at room temperature for 4 h. After the reaction was completed, 15 mL water was added to the reaction solution and extracted with CH_2Cl_2 (3×15 mL). The organic layer was combined and washed with brine, dried over anhydrous sodium sulfate, filtered. The solvent was removed under reduced pressure. The residue was subjected to silica gel chromatography using PE and
EtOAc (10:1-2:1, v/v) as an eluent to afford the target compounds 5a-5u (46-90%).
Data for N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-4'-(trifluoromethyl)-[1,1'biphenyl]-2-carboxamide (5a). Yield 49%; white solid; mp 153-154 °C. ¹H NMR (400 MHz,
CDCl₃) δ 8.25 (s, 1H), 7.81 (dd, *J*=7.6, 1.5 Hz, 1H), 7.67 (d, *J*=8.1 Hz, 2H), 7.60 (td, *J*=7.5, 1.5
Hz, 1H), 7.56 – 7.49 (m, 3H), 7.43 (dd, *J*=7.6, 1.4 Hz, 1H), 7.11 (s, 1H), 3.90 (s, 3H). Elemental

- anal. calcd for $C_{19}H_{13}F_6N_3O$: C, 55.21; H, 3.17; N, 10.17. Found: C, 54.98; H, 3.52; N, 9.92.
- 150 HRMS (ESI) m/z calcd for $C_{19}H_{14}F_6N_3O (M+H)^+ 414.1036$, found 414.1035.

151 Data for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-4'-(trifluoromethyl)-[1,1'biphenyl]-2-carboxamide (5b). Yield 61%; white solid; mp 132-133°C. ¹H NMR (400 MHz, 152 CDCl₃) § 8.19 (s, 1H), 7.80 (dd, J=7.6, 1.5 Hz, 1H), 7.66 (d, J=8.1 Hz, 2H), 7.59 (td, J=7.5, 1.5 153 Hz, 1H), 7.55 – 7.47 (m, 3H), 7.42 (dd, *J*=7.5, 1.4 Hz, 1H), 7.30 (s, 1H), 6.48 (t, *J*=54.6 Hz, 1H), 154 3.86 (s, 3H). Elemental anal. calcd for C₁₉H₁₄F₅N₃O: C, 57.73; H, 3.57; N, 10.63. Found: C, 155 57.09; H, 3.26; N, 10.63. HRMS (ESI) m/z calcd for $C_{19}H_{15}F_5N_3O+(M+H)^+$ 396.1130, found 156 396.1131. 157

Data for 4'-methyl-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-biphenyl]-2carboxamide (**5c**). Yield 68%; white solid; mp 115-116°C. ¹H NMR (400 MHz, CDCl₃) δ 8.26 (s, 1H), 7.82 (dd, *J*=7.7, 1.5 Hz, 1H), 7.51 (td, *J*=7.5, 1.5 Hz, 1H), 7.43 (td, *J*=7.5, 1.4 Hz, 1H), 7.38 (dd, *J*=7.6, 1.4 Hz, 1H), 7.28 – 7.22 (m, 2H), 7.18 (d, *J*=7.9 Hz, 2H), 7.08 (s, 1H), 3.86 (s, 3H), 2.33 (s, 3H). Elemental anal. calcd for C₁₉H₁₆F₃N₃O: C, 63.51; H, 4.49; N, 11.69. Found: C, 63.06; H, 4.64; N, 11.56. HRMS (ESI) m/z calcd for C₁₉H₁₇F₃N₃O (M+ H)⁺ 360.1318, found 360.1322.

165	Data for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-4'-methyl-[1,1'-biphenyl]-2-
166	carboxamide (5d). Yield 79%; white solid; mp 140-141 °C. ¹ H NMR (400 MHz, CDCl ₃) δ 8.21
167	(s, 1H), 7.82 (dd, J=7.7, 1.5 Hz, 1H), 7.53 (td, J=7.5, 1.5 Hz, 1H), 7.49 - 7.36 (m, 2H), 7.34 -
168	7.25 (m, 3H), 7.21 (d, J=7.9 Hz, 2H), 6.39 (t, J=54.7 Hz, 1H), 3.85 (s, 3H), 2.36 (s, 3H).
169	Elemental anal. calcd for C ₁₉ H ₁₇ F ₂ N ₃ O: C, 66.85; H, 5.02; N, 12.31. Found: C, 66.93; H, 5.43;
170	N, 12.23. HRMS (ESI) m/z calcd for $C_{19}H_{18}F_2N_3O$ (M+ H) ⁺ 342.1412, found 342.1414.

171 Data for 3',4',5'-trifluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-172 biphenyl]-2-carboxamide (**5e**). Yield 73%; white solid; mp 150-151 °C. ¹H NMR (400 MHz, 173 CDCl₃) δ 8.23 (s, 1H), 7.73 (dd, *J*=7.6, 1.5 Hz, 1H), 7.55 (dtd, *J*=22.9, 7.5, 1.5 Hz, 2H), 7.37 (dd, 174 *J*=7.5, 1.4 Hz, 1H), 7.22 (s, 1H), 7.09 – 6.96 (m, 2H), 3.92 (s, 3H). Elemental anal. calcd for 175 C₁₈H₁₁F₆N₃O: C, 54.14; H, 2.78; N, 10.52. Found: C, 54.20; H, 2.96; N, 10.35. HRMS (ESI) m/z 176 calcd for C₁₈H₁₂F₆N₃O (M+ H)⁺ 400.0879, found 400.0882.

Data for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-3',4',5'-trifluoro-[1,1'-177 biphenyl]-2-carboxamide (5f). Yield 88%; white solid; mp 142-143°C. ¹H NMR (400 MHz, 178 CDCl₃) δ 8.21 (s, 1H), 7.76 (dd, J=7.5, 1.6 Hz, 1H), 7.62 – 7.48 (m, 2H), 7.42 (s, 1H), 7.38 (dd, 179 J = 7.5, 1.4 Hz, 1H, 7.11 - 6.99 (m, 2H), 6.61 (t, J = 54.5 Hz, 1H), 3.89 (s, 3H). Elemental anal. 180 calcd for C₁₈H₁₂F₅N₃O: C, 56.70; H, 3.17; N, 11.02. Found: C, 56.52; H, 3.27; N, 11.02. HRMS 181 (ESI) m/z calcd for $C_{18}H_{13}F_5N_3O (M+H)^+ 382.0973$, found 382.0975. 182 Data for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-[1,1'-biphenyl]-2-carboxamide 183

185 J=7.7, 1.4 Hz, 1H), 7.57 (td, J=7.5, 1.5 Hz, 1H), 7.49 (td, J=7.5, 1.4 Hz, 1H), 7.46 – 7.34 (m,

(5g). Yield 90%; white solid; mp 87-88 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.22 (s, 1H), 7.84 (dd,

186 6H), 7.27 (s, 1H), 6.41 (t, J=54.8 Hz, 1H), 3.86 (s, 3H). Elemental anal. calcd for $C_{18}H_{15}F_2N_3O$:

187 C, 66.05; H, 4.62; N, 12.84. Found: C, 65.68; H, 4.63; N, 12.86. HRMS (ESI) m/z calcd for 188 $C_{18}H_{16}F_2N_3O (M+H)^+$ 328.1256, found 328.1258.

Data for 3',4'-dichloro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-biphenyl]-189 2-carboxamide (5h). Yield 66%; white solid; mp 117-118 °C. ¹H NMR (400 MHz, CDCl₃) δ 190 8.25 (s, 1H), 7.77 (dd, J=7.7, 1.4 Hz, 1H), 7.65 – 7.37 (m, 5H), 7.28 – 7.14 (m, 2H), 3.93 (s, 3H). 191 192 Elemental anal. calcd for $C_{18}H_{12}Cl_2F_3N_3O$: C, 52.20; H, 2.92; N, 10.14.Found: C,51.90; H,2.78; N,10.21. HRMS (ESI) m/z calcd for $C_{18}H_{13}Cl_2F_3N_3O$ (M+H)⁺ 414.0382, found 414.0381. 193 Data for 3',4'-dichloro-N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-[1,1'-biphenyl]-194 2-carboxamide (5i). Yield 72%; white solid; mp 100-101 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.18 195 (s, 1H), 7.76 (dd, J=7.6, 1.4 Hz, 1H), 7.60 – 7.47 (m, 3H), 7.44 (d, J=8.3 Hz, 1H), 7.41 – 7.34 196 (m, 2H), 7.21 (dd, J=8.3, 2.1 Hz, 1H), 6.56 (t, J=54.6 Hz, 1H), 3.86 (s, 3H). Elemental anal. 197 calcd for C₁₈H₁₃Cl₂F₂N₃O: C, 54.57; H, 3.31; N, 10.61. Found: C, 54.16; H, 3.50; N, 10.45. 198 HRMS (ESI) m/z calcd for $C_{18}H_{14}C_{12}F_2N_3O$ (M+ H)⁺ 396.0477, found 396.0470. 199 Data for 4'-chloro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-biphenyl]-2-200 carboxamide (5j). Yield 46%; white solid; mp 139-140 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.26 201 (s, 1H), 7.79 (dd, J = 7.6, 1.4 Hz, 1H), 7.52 (dtd, J = 30.2, 7.6, 1.4 Hz, 2H), 7.42 – 7.35 (m, 3H), 202 203 7.35 - 7.29 (m, 2H), 7.14 (s, 1H), 3.90 (s, 3H). Elemental anal. calcd for C₁₈H₁₃ClF₃N₃O: C, 56.93; H, 3.45; N, 11.06. Found: C, 56.83; H, 3.95; N, 10.82. HRMS (ESI) m/z calcd for 204 C₁₈H₁₄ClF₃N₃O (M+ H)⁺ 380.0772, found 380.0772. 205

206 Data for 4'-chloro-N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-[1,1'-biphenyl]-2-207 carboxamide (**5k**). Yield 82%; white solid; mp 128-129°C. ¹H NMR (400 MHz, CDCl₃) δ 8.19 208 (s, 1H), 7.77 (dd, *J*=7.6, 1.5 Hz, 1H), 7.51 (dtd, *J*=28.9, 7.5, 1.4 Hz, 2H), 7.41 – 7.29 (m, 6H), 209 6.50 (t, *J*=54.6 Hz, 1H), 3.85 (s, 3H). Elemental anal. calcd for C₁₈H₁₄ClF₂N₃O: C, 59.76; H, 3.90; N, 11.62. Found: C, 59.28; H, 4.02; N, 11.48. HRMS (ESI) m/z calcd for C₁₈H₁₅ClF₂N₃O
(M+ H)⁺ 362.0866, found 362.0872.

for 5-fluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-biphenyl]-2-212 Data carboxamide (51). Yield 61%; white solid; mp 82-83 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.25 (s, 213 1H), 7.86 (dd, J = 8.6, 5.8 Hz, 1H), 7.40 (gg, J = 7.1, 2.6, 1.8 Hz, 5H), 7.20 - 7.00 (m, 3H), 3.88 214 215 (s, 3H). Elemental anal. calcd for C₁₈H₁₃F₄N₃O: C, 59.51; H, 3.61; N, 11.57. Found: C, 59.47; H, 4.03; N, 11.60. HRMS (ESI) m/z calcd for $C_{18}H_{14}F_4N_3O$ (M+ H)⁺ 364.1068, found 364.1070. 216 for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-5-fluoro-[1,1'-biphenyl]-2-217 Data carboxamide (5m). Yield 66%; white solid; mp 95-96°C. ¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 218 1H), 7.86 (dd, J = 8.6, 5.7 Hz, 1H), 7.52 – 7.33 (m, 5H), 7.25 (s, 1H), 7.22 – 7.10 (m, 2H), 6.41 219 (t, J = 54.8 Hz, 1H), 3.86 (s, 3H). Elemental anal. calcd for C₁₈H₁₄F₃N₃O: C, 62.61; H, 4.09; N, 220 12.17. Found: C, 62.03; H, 3.93; N, 12.27. HRMS (ESI) m/z calcd for $C_{18}H_{15}F_{3}N_{3}O$ (M+ H)⁺ 221 346.1162, found 346.1164. 222

223 Data for 3',4',5,5'-tetrafluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-224 biphenyl]-2-carboxamide (**5n**). Yield 61%; white solid; mp 155-156 °C. ¹H NMR (400 MHz, 225 CDCl₃) δ 8.22 (s, 1H), 7.76 (dd, *J*=8.6, 5.6 Hz, 1H), 7.22 (td, *J*=8.2, 2.6 Hz, 1H), 7.16 (s, 1H), 226 7.08 (dd, *J*=9.0, 2.6 Hz, 1H), 7.03 (dd, *J*=7.6, 6.2 Hz, 2H), 3.92 (s, 3H). Elemental anal. calcd for 227 C₁₈H₁₀F₇N₃O: C, 51.81; H, 2.42; N, 10.07. Found: C, 51.86; H, 2.60; N, 9.98. HRMS (ESI) m/z 228 calcd for C₁₈H₁₁F₇N₃O (M+ H)⁺ 418.0785, found 418.0778.

Data for N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-3',4',5,5'-tetrafluoro-[1,1'biphenyl]-2-carboxamide (50). Yield 60%; white solid; mp 146-147 °C. ¹H NMR (400 MHz,
CDCl₃) δ 8.19 (s, 1H), 7.79 (dd, *J*=8.6, 5.6 Hz, 1H), 7.37 (s, 1H), 7.22 (td, *J*=8.3, 2.6 Hz, 1H),
7.07 (ddd, *J*=17.8, 8.3, 4.4 Hz, 3H), 6.61 (t, *J*=54.5 Hz, 1H), 3.89 (s, 3H). Elemental anal. calcd

for C₁₈H₁₁F₆N₃O: C, 54.14; H, 2.78; N, 10.52. Found: C, 54.01; H, 2.85; N, 10.69. HRMS (ESI)
m/z calcd for C₁₈H₁₂F₆N₃O (M+ H)⁺ 400.0879, found 400.0871.

235Data for 3',4'-dichloro-5-fluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-236biphenyl]-2-carboxamide (**5p**). Yield 48%; white solid; mp 117–118 °C. ¹H NMR (400 MHz,237CDCl₃) δ 8.22 (s, 1H), 7.80 (dd, *J*=8.6, 5.6 Hz, 1H), 7.54 – 7.44 (m, 2H), 7.21 (ddd, *J*=8.3, 6.5,2382.3 Hz, 2H), 7.10 (dd, *J*=9.1, 2.6 Hz, 2H), 3.91 (s, 3H). Elemental anal. calcd for239C₁₈H₁₁Cl₂F₄N₃O: C, 50.02; H, 2.57; N, 9.72. Found: C, 49.25; H, 2.59; N, 9.73. HRMS (ESI)240m/z calcd for C₁₈H₁₂Cl₂F₄N₃O (M+ H)⁺ 432.0288, found 432.0285.241Data for 3',4'-dichloro-N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-5-fluoro-[1,1'-

biphenyl]-2-carboxamide (5q). Yield 53%; white solid; mp 111-112 °C. ¹H NMR (400 MHz,
CDCl₃) δ 8.16 (s, 1H), 7.77 (dd, *J*=8.6, 5.6 Hz, 1H), 7.51 (d, *J*=2.1 Hz, 1H), 7.46 (d, *J*=8.2 Hz,
1H), 7.35 (s, 1H), 7.23 – 7.14 (m, 2H), 7.08 (dd, *J*=9.1, 2.6 Hz, 1H), 6.55 (t, *J*=54.5 Hz, 1H),
3.86 (s, 3H).Elemental anal. calcd for C₁₈H₁₂Cl₂F₃N₃O: C, 52.20; H, 2.92; N, 10.14. Found: C,
51.80; H, 4.06; N, 10.17. HRMS (ESI) m/z calcd for C₁₈H₁₃Cl₂F₃N₃O (M+ H)⁺ 414.0382, found
414.0386.

248 Data for 4'-chloro-5-fluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-249 biphenyl]-2-carboxamide (**5r**). Yield 50%; white solid; mp 132-133°C. ¹H NMR (400 MHz, 250 CDCl₃) δ 8.24 (s, 1H), 7.81 (dd, *J*=8.6, 5.7 Hz, 1H), 7.46 – 7.28 (m, 4H), 7.17 (td, *J*=8.3, 2.6 Hz, 251 1H), 7.12 – 7.04 (m, 2H), 3.90 (s, 3H). Elemental anal. calcd for C₁₈H₁₂ClF₄N₃O: C, 54.35; H, 252 3.04; N, 10.56. Found: C, 54.18; H, 2.95; N, 10.59. HRMS (ESI) m/z calcd for C₁₈H₁₃ClF₄N₃O 253 (M+ H)⁺ 398.0678, found 398.0672.

Data for 4'-chloro-N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-5-fluoro-[1,1'biphenyl]-2-carboxamide (**5s**). Yield 61%; white solid; mp 129-130 °C. ¹H NMR (400 MHz, 256 CDCl₃) δ 8.17 (s, 1H), 7.79 (dd, *J*=8.6, 5.7 Hz, 1H), 7.42 – 7.27 (m, 5H), 7.16 (td, *J*=8.2, 2.6 Hz,

257 1H), 7.08 (dd, *J*=9.3, 2.6 Hz, 1H), 6.49 (t, *J*=54.6 Hz, 1H), 3.85 (s, 3H). Elemental anal. calcd for

258 C₁₈H₁₃ClF₃N₃O: C, 56.93; H, 3.45; N, 11.06. Found: C, 57.03; H, 3.55; N, 11.12. HRMS (ESI)

259 m/z calcd for $C_{18}H_{14}ClF_3N_3O (M+H)^+$ 380.0772, found 380.0769.

260 Data for 4'-(tert-butyl)-5-fluoro-N-(1-methyl-3-(trifluoromethyl)-1H-pyrazol-4-yl)-[1,1'-261 biphenyl]-2-carboxamide (**5t**). Yield 73%; white solid; mp 125-126 °C. ¹H NMR (400 MHz, 262 CDCl₃) δ 8.30 (s, 1H), 7.90 (dt, *J*=8.5, 4.2 Hz, 1H), 7.45 (dd, *J*=8.5, 2.2 Hz, 2H), 7.31 (dd, *J*=8.6, 263 2.3 Hz, 2H), 7.19 – 6.99 (m, 3H), 3.89 (s, 3H), 1.32 (s, 9H). Elemental anal. calcd for 264 C₂₂H₂₁F₄N₃O: C, 63.00; H, 5.05; N, 10.02 Found: C, 62.72; H, 5.29; N, 9.95. HRMS (ESI) m/z 265 calcd for C₂₂H₂₂F₄N₃O (M+ H)⁺ 420.1694, found 420.1699.

266 Data for 4'-(tert-butyl)-N-(3-(difluoromethyl)-1-methyl-1H-pyrazol-4-yl)-5-fluoro-[1,1'-267 biphenyl]-2-carboxamide (**5u**). Yield 75%; white solid; mp 115-116 °C. ¹H NMR (400 MHz, 268 CDCl₃) δ 8.24 (s, 1H), 7.90 (dd, *J*=8.6, 5.8 Hz, 1H), 7.47 (d, *J*=8.1 Hz, 2H), 7.35 (d, *J*=8.1 Hz, 269 2H), 7.27 (s, 1H), 7.20 – 7.08 (m, 2H), 6.36 (t, *J*=54.8 Hz, 1H), 3.87 (s, 3H), 1.35 (s, 270 9H).Elemental anal. calcd for C₂₂H₂₂F₃N₃O: C, 65.82; H, 5.52; N, 10.47. Found: C, 65.65; H, 271 5.47; N, 10.55. HRMS (ESI) m/z calcd for C₂₂H₂₃F₃N₃O (M+ H)⁺ 402.1788, found 402.1792.

X-ray Diffraction. Compound **5i** was recrystallized by a slow evaporation from a solution of dichloromethane/n-hexane (v/v=2:5) to afford a single crystal suitable for X-ray crystallography for structure validation. Cell dimensions and intensities were measured using a Rigaku 007 Saturn 70 diffractometer with graphite monochromated Mo K α radiation. Compound **5i**: Monoclinic, a=10.994 (2) Å, b=19.344 (4) Å, c=8.4094 (17) Å, U=1759.4(6) Å³, T=113(2), space group P2(1)/c. A total of 18736 reflections were measured, of which 4184 were unique ($R_{int}=0.0768$) in the range of 1.883 <2 Θ < 27.893°(-14 \leq h \leq 13, -25 \leq k \leq 25, -11 \leq 1 \leq 11), and 2882 observed reflections with I > 2σ (I) were used in the refinement on F². The structure of **5**i was solved by direct method with the SHELXTL-97 program and all of the non-H atoms were refined anisotropically by full matrix least-squares to give the Final R indices R₁=0.0599, wR₂=0.1903. The atomic coordinates of **5**i were deposited at the Cambridge Crystallographic Data Centre (CCDC) and CCDC-1942098 contained the supplementary crystallographic data for this paper. The crystallographic data of **5**i can be download from the CCDC and the Supporting Information here.

286 Bioassays

In vitro fungicidal activity. The fungicidal activity of the target compounds was tested in 287 vitro against Alternaria solani, Botrytis cinerea, Gibberella zeae, Phytophthora infestans (Mont) 288 de Bary, Physalospora piricola, Pellicularia sasakii, Rhizoctonia cerealis and Sclerotinia 289 sclerotiorum by using mycelium growth rate test.²⁵ The commercial SDHIs boscalid and 290 fluxapyroxad were selected as positive control. The synthesized compounds and controls were 291 dissolved in DMSO to prepare the 20 mg/mL stock solution before diluting by PDA. Target 292 compounds at a concentration 50 µg/mL were used for the initial preliminary screening with 293 three replicates. Their relative inhibitory rate (I, %) was calculated according to the following 294 equation: 295

296

$$I(\%) = [(C-T)/(C-4)]*100$$

I, inhibitory rate; C, colony diameter of control (mm); T, colony diameter of treatment(mm).

The compounds with excellent inhibitory rate were selected for the determination of EC_{50} values. The 20 mg/mL stock solutions were diluted by PDA to obtain a series of concentrations for the same procedures as described in the above experiments. The EC_{50} data were calculated according to the concentration and the probit of the corresponding inhibitory rate respectively.
The results were summarized in Table 1 and Table 2.

In vivo fungicidal activity. The in vivo fungicidal activities of the target compounds against *Erysiphe graminis* (wheat white powder), *Puccinia sorghi* Schw (corn rust) and *Rhizoctonia solani* (rice sheath blight) were evaluated with three replicates by using pot bioassay as reported previously.^{26,27} The biological assay was carry out by Shenyang Sinochem Agrochemicals R&D Co. Ltd. and the results were summarized in **Table 3** and **Table 4**.

Fluorescence auenching analysis of succinate dehydrogenase. Succinate 309 dehydrogenase of R. cerealis was extracted according to operation instructions (BC0955, 310 311 solarbio, Beijing) for the assay. Fluorescence quenching analysis was performed with F-4500 fluorescence spectrophotometer (Hitachi, Tokyo, Japan). Fluorescence emission spectra was 312 recorded at excitation wavelength of 280 nm with a wavelength ringing from 300-390 nm at 4°C, 313 314 and the emission band-width was set at 10 nm with medium sensitive. Compound 5i with 0 to 4 mg/L was added into the succinate dehydrogenase for fluorescence determination, with boscalid, 315 fluxapyroxad and tricyclazole as control. The fluorescence intensity under the treatment was 316 equal which in SDH and compound interaction minus fluorescence intensity of compound itself. 317

SDH enzymatic activity. The SDH enzyme activity of **5i** was determined by using a succinate dehydrogenase assay kit (Solabio, BC0950) and assessed as reported previously.²⁸ *R*. *cerealis* was grown in sterilized Fries medium for 5 d and then treated with different concentrations of **5i** and fluxapyroxad, respectively. The SDH enzyme activity was measured after 24 h of treatment with the selected compounds, and the absorbance value was recorded at 600 nm by using a microplate reader. Compounds were tested at 6 different concentrations with three replicates and the IC₅₀ value was calculated by GraphPad Prim version 6.02.

325 **RESULTS AND DISCUSSION**

Molecular Docking Analysis. Novel SDHI derivatives are rationally designed by 326 reversing of -CO-NH- group and we speculate whether these compounds also target at SDH and 327 form hydrogen bonds at the pocket. Molecular docking of selected compounds and fluxapyroxad 328 were carried out for evaluation its potent mode of action of target compounds. All reported SDHs 329 are highly reserved in spatial structure, subunit and electron transport pathway, and even the 330 ubiquinone binding site in the prokaryotes and eukaryotes is also highly reserved.²⁹ For these 331 reasons, avian (Gallus gallus) respiratory complex II with carboxin bound (pdb code: 2FBW) 332 was chose for molecular docking study. In order to illustrate the binding mode between ligand 333 334 and receptor, 5i was selected as an example. As shown in Figure 3, 5i located in the same site and adopted a similar binding conformation as compared to fluxapyroxad in the three-335 dimensional schematic diagrams. Compound **5i** bounded to the receptor protein, the carboxyl 336 oxygen formed two hydrogen bonds with the amino acid residue TRP173 and TYR58, 337 respectively. The binding mode was similar to that of fluxapyroxad, this indicated that, the target 338 compounds also possessed strong interaction with SDH. 339

Chemical Synthesis. The synthetic procedures for the target compounds are described in 340 Scheme 1. The Suzuki-Miyaura cross-coupling of aryl halide and aryl boronic acid was 341 conducted under argon at 80 °C to produce the key intermediate 1.30 Intermediate 1 was then 342 reacted with NaOH via a direct ester hydrolysis under 80 °C to yield the compound 2. Compound 343 2 was reacted with oxalyl chloride in CH₂Cl₂ with DMF to afford intermediate 3. Intermediate 3 344 was then reacted with pyrazole amine 4 in CH₂Cl₂ in the presence of Et₃N to afford target 345 compound 5 with moderate to good yields. To further validate the structure of the title 346 compounds, the structure of 5i was further identified by X-ray diffraction studies (see Figure. 4). 347

Fungicidal Activity. The results of in vitro and in vivo fungicidal activities of the target 348 compounds (5a to 5u) were summarized in Table 1-4. In general, most of the compounds 349 displayed considerable to excellent fungicidal activities against eight phytopathogens in vitro at 350 50 µg/mL. (Table 1). No obvious difference of fungicidal activity between 3-CF₃ substituent 351 prazole compounds and $3-CF_2H$ substituent prazole compounds was observed. Most of these 352 353 compounds were highly active against B. cinerea, R. cerealis and S. sclerotiorum. As shown in Table 1, 5c, 5d and 5j showed over 90% of inhibitory rate against B. cinereal, which were 354 comparable to boscalid and fluxapyroxad. For G. zeae, 5i and 5j showed better activity than 355 positive controls. Compound 5i, 5p, 5q, 5s displayed higher activity against P. infestans than 356 boscalid and fluxapyroxad, 5h, 5i, 5j, 5p, 5q, 5r, 5s exhibited excellent activities against P. 357 sasakii, which were better than boscalid and fluxapyroxad. Eight compounds showed over 86% 358 of inhibitory rate against R. cerealis, boscalid and fluxapyroxad only showed 35% and 86% of 359 inhibitory rate, respectively. All compounds exhibited excellent activities against S. sclerotiorum. 360 361 Several compounds with superior fungicidal activities were selected for the calculation of EC₅₀ value. As shown in Table 2, several compounds showed good activity against Botrytis 362 cinerea, but not as effective as that of the positive controls. Compounds 5i and 5q displayed 363 364 excellent activities against P. infestans and P. sasakii with the EC_{50} values lower than the positive control fluxapyroxad. Nine compounds exhibited better fungicidal activity against R. 365 366 *cerealis* than fluxapyroxad, especially, 5i showed excellent fungicidal activity with the EC_{50} value of 4.61 μ g/mL, which was more active than that of fluxapyroxad with its EC₅₀ value of 367 16.99 µg/mL. The EC₅₀ values of all target compounds were tested against S. sclerotiorum, 5i 368 369 also showed the highest fungicidal activity with its EC_{50} value of 0.73 µg/mL, which was 370 comparable to that of boscalid with its EC_{50} value of 0.51 µg/mL. Therefore, 5i exhibited

promising fungicidal activity against five fungi with broader spectrum of fungicidal activity andcould be an alternative to SDHIs candidate.

The in vivo fungicidal activity of the target compounds was tested against E. graminis, P. 373 sorghi and R. solani for comparison with the commercial fungicide boscalid and fluxapyroxad 374 (Table 3). Most of the compounds exhibited excellent fungicidal activity against *P. sorghi* and *R.* 375 376 solani at the concentration 200 µg/mL. Compound 5c, 5l, 5o, 5t displayed 100% of inhibitory rate against E. graminis and similar activity as that of boscalid and fluxapyroxad. Compound 5d, 377 5f, 5g, 5h, 5i, 5g exhibited more than 90% of excellent fungicidal activity against P. sorghi, 378 379 which was comparable to those of boscalid and fluxapyroxad. Compound 5g, 5h, 5i, 5t displayed over 80% of inhibitory rate against R. solani. 380

Some compounds with higher inhibitory rate were selected for further greenhouse in vivo 381 fungicidal activity evaluation. As shown in **Table 4**, **5g** and **5h** were not as effective as **5i** against 382 P. sorghi and R. solani. Compound 5i and fluxapyroxad exhibited 100% inhibitory rate against P. 383 sorghi at the concentration of 50 µg/mL, they were better than boscalid (70%). Furthermore, 384 compound 5i showed excellent inhibitory rate (60%) against R. solani at 1µg/mL, which was 385 better than boscalid (30%) at the same concentration. In general, 5i presented excellent activity 386 387 against R. cerealis in vitro and was highly effective against R. solani in vivo. Both in vitro and in vivo activity determination indicated that, 5i could be a promising candidate against R. solani for 388 rice disease control. 389

SDH Enzymatic Inhibition Activity. Compound **5i** with promising fungicidal activity was selected and evaluated for SDH enzymatic inhibition determination to its target site validation. As shown in **Table 5**, **5i** exhibited very good SDH inhibition with an IC₅₀ of 0.60 μ g/mL, while the IC₅₀ of fluxapyroxad was 0.29 μ g/mL, **5i** showed the same level of inhibitory activity as fluxapyroxad. Compound 5i with carbonyl group adjacent to benzene ring contrast to
 fluxapyroxad which carbonyl group was adjacent to pyrazole could also exhibit excellent SDH
 enzymatic inhibition activity and formed strong interaction with SDH.

Fluorescence Quenching Analysis of SDH. The interaction of protein and small 397 molecules could be revealed by fluorescence quenching analysis. Fluorescence quenching of the 398 SDH by **5i**, boscalid and fluxapyroxad were conducted to validate interactions between protein 399 and compounds. As shown in Figure 5C, with the increase of the concentration of 5i, the 400 fluorescence intensity for SDH gradually quenched, it was similar to that of the positive control 401 boscalid and fluxapyroxad (Figure 5A, 5B), there showed no significant fluorescence changes 402 when treated by a negative control tricyclazole, a melanin synthesis inhibitor³¹ (Figure 5D). 403 Boscalid and fluxapyroxad, carboxamide fungicides, were SDHIs.³² Pesticide target discovery 404 and validation is a most important basis for novel pesticide development.³³ Compound 5i 405 exhibited the similar fluorescence quenching rules as that of boscalid and fluxapyroxad, which 406 indicated that they might act at the same target site. 407

In summary, a series of novel SDHIs derivatives were designed and synthesized by 408 inversion of carbonyl and amide groups of fluxapyroxad. Bioassay discovered highly active 5i 409 with broad-spectrum of outstanding in vitro fungicidal activity and better in vivo efficacy against 410 P. sorghi and R. solani. Fluorescence quenching analysis for SDH target validation indicated that, 411 5i exhibited the similar properties as that of the two SDHI positive controls boscalid and 412 fluxapyroxad, these results revealed that, the designed compounds might also act at SDH. Our 413 results provided a new clue for the molecular design and development of the novel highly active 414 415 SDHIs.

416 ASSOCIATED CONTENT

417 Supporting Information

- The Supporting Information is available free of charge on the ACS Publications website at DOI:
- ¹H NMR spectra for all compounds and HRMS spectra for target compounds
- 420 Crystal data of **5i** (CIF)

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Figure 3. Binding modes of 5i (A) and fluxapyroxad (B) with SDH.



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545 Figure 4. Crystal structure for 5i by X-ray diffraction determination. (CCDC number: 1942098)



549 Figure 5. Fluorescence spectra of succinate dehydrogenase: A, boscalid; B, fluxapyroxad; C, compound 5i; D,
550 tricyclazole "a to e" were "0 to 4 mg/L".



Compd	\mathbb{R}^1	\mathbb{R}^2	R ³	\mathbb{R}^4	R ⁵	Compd	\mathbb{R}^1	\mathbb{R}^2	R ³	\mathbb{R}^4	R ⁵	Compd	\mathbb{R}^1	\mathbb{R}^2	R ³	R ⁴	R ⁵
5a	Н	Н	CF ₃	Н	CF ₃	5h	Н	Н	Cl	Cl	CF ₃	50	F	F	F	F	CHF ₂
5b	Н	Н	CF_3	Н	CHF_2	5i	Н	Н	Cl	Cl	CHF_2	5p	F	Н	Cl	Cl	CF_3
5c	Н	Н	CH_3	Н	CF ₃	5j	Н	Н	Cl	Н	CF_3	5q	F	Н	Cl	Cl	CHF_2
5d	Н	Н	CH_3	Н	CHF_2	5k	Н	Н	Cl	Н	CHF_2	5r	F	Н	Cl	Н	CF_3
5e	Н	F	F	F	CF ₃	51	F	Н	Н	Н	CF_3	5 s	F	Н	Cl	Н	CHF_2
5f	Н	F	F	F	CHF_2	5m	F	Н	Н	Н	CHF_2	5t	F	Н	tBu	Н	CF_3
5g	Н	Н	Н	Н	CHF_2	5n	F	F	F	F	CF_3	5u	F	Н	tBu	Н	CHF_2
					Sche	me 1 The	e syn	thetio	c rou	te of	target c	ompound	ds				

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Table 1 In vitro fungicidal activity of target compounds against Phytopathogens at 50 µg/mL

Compd			Myceli	ium growtl	n inhibitory	/ rate (%)		
	AS ^a	BC	GZ	PI	PP	PS	RC	SS
5a	36±2	49 ± 1	35 ±0	21 ±0	13 ±1	22 ±2	52 ±1	100
5b	54 ± 1	76 ± 1	59 ± 1	49 ± 1	30 ± 0	56 ± 1	80 ± 0	91 ± 1
5c	56±3	93 ± 0	61 ± 2	46 ± 2	25 ± 0	54 ± 2	100	96 ± 0
5d	46 ± 1	90 ± 0	44 ± 2	36 ± 2	35 ± 0	46 ± 1	71 ± 0	95 ± 0
5e	44 ± 0	66 ± 1	57 ± 1	26 ± 2	11 ± 2	32 ± 1	65 ± 1	89 ± 2
5f	49 ± 1	68 ± 0	56 ± 1	28 ± 1	32 ± 1	27 ±2	68 ± 0	88 ± 1
5g	41±0	63 ± 1	30 ± 2	33 ± 2	21 ± 1	24 ±2	56 ± 2	91 ±0
5h	59±1	71 ±2	65 ± 1	51 ± 1	35 ± 1	61 ± 0	95 ±0	96 ± 0
5i	62±0	71 ± 0	67 ± 1	62 ± 0	49 ± 0	71 ± 0	92 ±0	100
5j	56±1	90 ± 1	69 ± 1	51 ± 0	38 ± 0	61 ± 1	100	95 ± 0
5k	44±1	83 ± 1	52 ± 1	46 ± 1	25 ± 1	54 ± 0	80 ± 0	91 ± 0
51	46±0	76 ± 0	44 ± 1	28 ± 1	28 ± 0	27 ± 1	83 ± 1	89 ± 3
5m	46 ± 1	56 ± 1	35 ± 2	21 ± 1	21 ± 0	24 ± 1	68 ± 1	86 ± 2
5n	51 ± 1	68 ± 0	59 ± 1	36 ± 1	10 ± 2	46 ± 1	74 ± 1	96 ± 1
50	49 ± 0	59 ± 1	61 ± 0	51 ± 0	17 ± 0	54 ± 1	76 ± 1	100
5p	59±1	68 ± 1	61 ± 0	59 ± 1	13 ± 2	66 ± 0	91 ±0	100
5q	59±0	78 ± 1	59 ± 0	72 ± 0	39 ± 1	76 ± 0	89 ± 0	100
5r	56±3	88 ± 1	59 ± 1	51 ± 1	32 ± 1	66 ± 0	100	96 ± 1
5s	56 ± 1	80 ± 1	59 ± 1	56 ± 1	21 ± 1	68 ± 0	86 ± 1	100
5t	46 ± 3	49 ± 3	44 ± 0	33 ± 3	21 ± 1	44 ± 1	68 ± 1	100
5u	54 ± 2	59 ± 2	57 ± 1	41 ± 1	32 ± 2	54 ± 1	79 ± 2	100
Boscalid	90 ± 0	95 ± 1	57 ± 2	23 ± 2	58 ± 1	22 ±2	35 ± 1	100
Fluxapyroxad	95 ± 1	100	67 ± 2	54 ±0	82 ± 0	54 ± 1	86 ± 1	100

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³: AS : Alternaria solani, BC : Botrytis cinerea, GZ : Gibberella zeae, PI : Phytophthora infestans

560 (Mont.) de Bary, PP: *Physalospora piricola*, PS: *Pellicularia sasakii*, RC: *Rhizoctonia cerealis*, SS

561 : *Sclerotinia sclerotiorum*.

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Table 2 The in vitro EC_{50} value ($\mu g/mL$) of selected compounds

5b $16.11 (11.58-23.34)$ $y=3.9205+0.9050 x$ 0.9649 5c $7.90 (6.65-9.32)$ $y=3.5335+1.6762 x$ 0.9887 5d $14.91 (11.67-19.41)$ $y=3.0082 + 1.7329 x$ 0.9765 5h $19.88 (14.43-24.93)$ $y=3.6833+1.0469 x$ 0.9802 5i $17.22 (14.27-19.81)$ $y=3.0418+1.6371 x$ 0.9919 5j $16.21 (11.52-21.06)$ $y=4.2262+0.6529 x$ 0.9694 B. cinerea5k $24.38 (19.50-27.69)$ $y=2.7646+1.6426 x$ 0.9911 5q $35.34 (26.97-45.26)$ $y=2.2335+1.7245 x$ 0.9636 5q $35.34 (26.97-45.26)$ $y=2.6208+1.5635 x$ 0.9884 5r $25.56 (18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 5s $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9907 Fluxapyroxad $0.10 (0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 $P.$ infestans $5q$ $24.22 (20.09-30.66)$ $y=3.0215+1.4420 x$ 0.9839 $P.$ infestans $5q$ $24.45 (23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976	Fungi	Compd.	EC ₅₀ (95% confidence interval)	Regression equation	R ²
5c7.90 (6.65-9.32) $y=3.5335+1.6762 x$ 0.98875d14.91 (11.67-19.41) $y=3.0082 + 1.7329 x$ 0.97655h19.88 (14.43-24.93) $y=3.6833+1.0469 x$ 0.98025i17.22 (14.27-19.81) $y=3.0418+1.6371 x$ 0.99195j16.21 (11.52-21.06) $y=4.2262+0.6529 x$ 0.9694B. cinerea5k24.38 (19.50-27.69) $y=2.7646+1.6426 x$ 0.99115q35.34 (26.97-45.26) $y=2.6208+1.5635 x$ 0.98845r25.56 (18.78-32.85) $y=4.2162+0.5721 x$ 0.97925s8.19 (6.07-11.51) $y=3.6524+1.4798 x$ 0.9628Boscalid0.45 (0.36-0.53) $y=5.2928+0.7623 x$ 0.9908Fluxapyroxad0.10 (0.08-0.11) $y=3.0215+1.4420 x$ 0.9839P. infestans5q24.22 (20.09-30.66) $y=2.9475+1.4804 x$ 0.9976Fluxapyroxad38.84 (27.58-60.05) $y=3.3449+1.0493 x$ 0.9643		5b	16.11 (11.58-23.34)	y=3.9205+0.9050 x	0.9649
5d $14.91 (11.67-19.41)$ $y=3.0082 + 1.7329 x$ 0.9765 5h $19.88 (14.43-24.93)$ $y=3.6833+1.0469 x$ 0.9802 5i $17.22 (14.27-19.81)$ $y=3.0418+1.6371 x$ 0.9919 5j $16.21 (11.52-21.06)$ $y=4.2262+0.6529 x$ 0.9694 B. cinerea5k $24.38 (19.50-27.69)$ $y=2.7646+1.6426 x$ 0.9911 5q $53.34 (26.97-45.26)$ $y=2.2335+1.7245 x$ 0.9636 5q $55.56 (18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 5s $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9908 Fluxapyroxad $0.10 (0.08-0.11)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans5q $24.22 (20.09-30.66)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5c	7.90 (6.65-9.32)	y=3.5335+1.6762 x	0.9887
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		5d	14.91 (11.67-19.41)	y=3.0082+1.7329 x	0.9765
5i $17.22(14.27-19.81)$ $y=3.0418+1.6371 x$ 0.9919 5j $16.21(11.52-21.06)$ $y=4.2262+0.6529 x$ 0.9694 B. cinerea5k $24.38(19.50-27.69)$ $y=2.7646+1.6426 x$ 0.9911 5l $43.29(24.00-73.56)$ $y=2.2335+1.7245 x$ 0.9636 5q $5s$ $8.19(6.07-45.26)$ $y=2.6208+1.5635 x$ 0.9884 5r $25.56(18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 5s $8.19(6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45(0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9908 Fluxapyroxad $0.10(0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 Fluxapyroxad $0.10(0.08-0.11)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84(27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5h	19.88 (14.43-24.93)	y=3.6833+1.0469 x	0.9802
5j $16.21 (11.52-21.06)$ $y=4.2262+0.6529 x$ 0.9694 B. cinerea5k $24.38 (19.50-27.69)$ $y=2.7646+1.6426 x$ 0.9911 5l $43.29 (24.00-73.56)$ $y=2.2335+1.7245 x$ 0.9636 5q $35.34 (26.97-45.26)$ $y=2.6208+1.5635 x$ 0.9884 5r $25.56 (18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 5s $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9908 Fluxapyroxad $0.10 (0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 P. infestans5q $24.45 (23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5i	17.22 (14.27-19.81)	<i>y</i> =3.0418+1.6371 <i>x</i>	0.9919
B. cinerea $5k$ $24.38 (19.50-27.69)$ $y=2.7646+1.6426 x$ 0.9911 $5l$ $43.29 (24.00-73.56)$ $y=2.2335+1.7245 x$ 0.9636 $5q$ $35.34 (26.97-45.26)$ $y=2.6208+1.5635 x$ 0.9884 $5r$ $25.56 (18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 $5s$ $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9907 Fluxapyroxad $0.10 (0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 P. infestans $5q$ $24.22 (20.09-30.66)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5j	16.21 (11.52-21.06)	y = 4.2262 + 0.6529 x	0.9694
5l $43.29 (24.00-73.56)$ $y=2.2335+1.7245 x$ 0.9636 5q $35.34 (26.97-45.26)$ $y=2.6208+1.5635 x$ 0.9884 5r $25.56 (18.78-32.85)$ $y=4.2162+0.5721 x$ 0.9792 5s $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9907 Fluxapyroxad $0.10 (0.08-0.11)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans5q $24.22 (20.09-30.66)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643	B. cinerea	5k	24.38 (19.50-27.69)	<i>y</i> =2.7646+1.6426 <i>x</i>	0.9911
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		51	43.29 (24.00-73.56)	<i>y</i> =2.2335+1.7245 <i>x</i>	0.9636
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		5q	35.34 (26.97-45.26)	<i>y</i> =2.6208+1.5635 <i>x</i>	0.9884
5s $8.19 (6.07-11.51)$ $y=3.6524+1.4798 x$ 0.9628 Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9907 Fluxapyroxad $0.10 (0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 5i $24.22 (20.09-30.66)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans5q $24.45 (23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5r	25.56 (18.78-32.85)	y=4.2162+0.5721 x	0.9792
Boscalid $0.45 (0.36-0.53)$ $y=5.2928+0.7623 x$ 0.9907 Fluxapyroxad $0.10 (0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 5i $24.22 (20.09-30.66)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans 5q $24.45 (23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5 s	8.19 (6.07-11.51)	y=3.6524+1.4798 x	0.9628
Fluxapyroxad $0.10(0.08-0.11)$ $y=6.0470+1.0169 x$ 0.9908 5i $24.22(20.09-30.66)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans5q $24.45(23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84(27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		Boscalid	0.45 (0.36-0.53)	y=5.2928+0.7623 x	0.9907
5i $24.22 (20.09-30.66)$ $y=3.0215+1.4420 x$ 0.9839 P. infestans5q $24.45 (23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		Fluxapyroxad	0.10 (0.08-0.11)	y=6.0470+1.0169 x	0.9908
P. infestans $5q$ $24.45(23.22-29.23)$ $y=2.9475+1.4804 x$ 0.9976 Fluxapyroxad $38.84(27.58-60.05)$ $y=3.3449+1.0493 x$ 0.9643		5 i	24.22 (20.09-30.66)	y=3.0215+1.4420 x	0.9839
Fluxapyroxad $38.84 (27.58-60.05)$ $y=3.3449+1.0493 \times 0.9643$	P. infestans	; 5q	24.45 (23.22-29.23)	y=2.9475+1.4804 x	0.9976
	Ū.	Fluxapyroxad	38.84 (27.58-60.05)	y=3.3449+1.0493 x	0.9643
5i 19.74 (15.44-27.77) $y=3.2913+1.3284 \times 0.9652$		5 i	19.74 (15.44-27.77)	y=3.2913+1.3284 x	0.9652
<i>P. sasakii</i> 5q $18.79(15.37-24.32)$ <i>y</i> = $3.4240+1.2413 \times 0.9864$	P. sasakii	5q	18.79 (15.37-24.32)	y=3.4240+1.2413 x	0.9864
Fluxapyroxad $38.18(30.54-51.26)$ $y=2.5966+1.5260 \times 0.9881$		Fluxapyroxad	38.18 (30.54-51.26)	y=2.5966+1.5260 x	0.9881
5b 17.75 (15.43-19.82) $y=3.5447+1.1907 \times 0.9926$		5b	17.75 (15.43-19.82)	y=3.5447+1.1907 x	0.9926
5c $14.60(10.38-22.73)$ $y=2.8779+1.8342 \times 0.9408$		5c	14.60 (10.38-22.73)	y=2.8779+1.8342 x	0.9408
5d $25.85(17.71-39.23)$ $y=2.3516+1.8853 \times 0.9566$		5d	25.85 (17.71-39.23)	y=2.3516+1.8853 x	0.9566
5h $15.45(12.14-19.43)$ $y=3.4256+1.3460 \times 0.9820$		5h	15.45 (12.14-19.43)	y=3.4256+1.3460 x	0.9820
5i 4.61 (3.95-5.36) $y=3.8287+1.7728 \times 0.9921$		5 i	4.61 (3.95-5.36)	y=3.8287+1.7728 x	0.9921
5 $y=3.2247+1.8335 \times 0.9410$		5j	9.80 (6.31-14.84)	y=3.2247+1.8335 x	0.9410
5k 20.75 (17.45-23.34) $y=3.0136+1.5349 \times 0.9934$		5k	20.75 (17.45-23.34)	y=3.0136+1.5349 x	0.9934
5 31.52 (18.66-56.03) $y=3.3679+1.0929 \times 0.9475$	D 1:	51	31.52 (18.66-56.03)	v=3.3679+1.0929 x	0.9475
<i>R. cerealis</i> 5n 12.52 (10.89-14.03) $y=3.7241+1.1842 \times 0.9950$	R. cerealis	5n	12.52 (10.89-14.03)	v=3.7241+1.1842 x	0.9950
50 11.03 (9.11-11.61) $y=3.7202+1.2741 \times 0.9937$		50	11.03 (9.11-11.61)	v=3.7202+1.2741 x	0.9937
5p $6.48(5.41-7.76)$ $y=4.2499+0.9598 \times 0.9917$		5p	6.48 (5.41-7.76)	v=4.2499+0.9598 x	0.9917
5q 12.13 (9.96-15.41) $y=3.7561+1.1587 \times 0.9812$		5g	12.13 (9.96-15.41)	v=3.7561+1.1587 x	0.9812
5r 16.96 (14.89-18.83) $y=1.7841+2.6537 \times 0.9964$		5r	16.96 (14.89-18.83)	v=1.7841+2.6537 x	0.9964
5s 19.91 (16.72-23.42) $y=1.8063+2.4936 \times 0.9897$		5 s	19.91 (16.72-23.42)	v=1.8063+2.4936 x	0.9897
5u 17.78 (16.70-19.55) $y=3.0593+1.5796 \times 0.9970$		5u	17.78 (16.70-19.55)	v=3.0593+1.5796 x	0.9970
Fluxapyroxad $16.99(14.06-20.62)$ $y=2.7777+1.8349 \times 0.9881$		Fluxapyroxad	16.99 (14.06-20.62)	y=2.7777+1.8349 x	0.9881
5a 12.15 (10.78-13.01) $y=4.4216+0.5519 \times 0.9966$		5a	12.15 (10.78-13.01)	y = 4.4216 + 0.5519 x	0.9966
5b $20.55(14.67-32.31)$ $y=3.4946+1.1542 \times 0.9643$		5b	20.55 (14.67-32.31)	v=3.4946+1.1542 x	0.9643
5c $8.55(4.99-14.19)$ $y=4.2416+0.8353 \times 0.9184$		5c	8.55 (4.99-14.19)	v = 4.2416 + 0.8353 x	0.9184
5d $34.28(21.89-59.91)$ $y=3.0072+1.3091 \times 0.9535$		5d	34.28 (21.89-59.91)	v=3.0072+1.3091 x	0.9535
5e $15.64 (10.40-24.95)$ $y=1.9826+2.5505 \times 0.9327$		5e	15.64 (10.40-24.95)	v=1.9826+2.5505 x	0.9327
S. scierotiorum 5f $6.93 (4.86-9.20)$ $y=4.3711+0.7671 \times 0.9661$. sclerotiorum	^m 5f	6.93 (4.86-9.20)	y=4.3711+0.7671 x	0.9661
5g 14.26 (13.14-16.86) $v=3.6601+1.1822 \times 0.9936$		5g	14.26 (13.14-16.86)	y=3.6601+1.1822 x	0.9936
5h $1.08 (0.73-1.50)$ $v=4.9935+0.6235 \times 0.9606$		5h	1.08 (0.73-1.50)	v=4.9935+0.6235 x	0.9606
5i $0.73 (0.49-1.04)$ $v=5.1180+0.6857 \times 0.9659$		5i	0.73 (0.49-1.04)	v=5.1180+0.6857 x	0.9659
5j $8.20 (4.69-14.08)$ $y=4.0936+1.0227 \times 0.9248$		5i	8.20 (4.69-14.08)	y = 4.0936 + 1.0227 x	0.9248

5k	7.57 (5.24-11.15)	<i>y</i> =3.8165+1.3999 <i>x</i>	0.9552
51	24.20 (15.74-34.15)	y=2.5859+1.7845 x	0.9657
5m	3.98 (2.74-5.87)	y = 4.5779 + 0.7132 x	0.9860
5n	16.50 (13.56-21.11)	<i>y</i> =3.2996+1.4007 <i>x</i>	0.9830
50	13.21 (12.40-15.93)	<i>y</i> =4.0860+0.8218 <i>x</i>	0.9941
5p	8.46 (7.00-10.29)	y=3.6722+1.4763 x	0.9866
5q	5.98 (4.60-7.87)	y=4.0054+1.3166 x	0.9776
5r	8.87 (7.43-10.66)	y=4.0090+1.0814 x	0.9881
55	3.72 (2.35-6.05)	<i>y</i> =4.5508+0.8075 <i>x</i>	0.9575
5t	3.94 (2.48-6.06)	<i>y</i> =4.6745+0.5696 <i>x</i>	0.9621
5u	5.53 (4.18-7.14)	y=4.5031+0.7048 x	0.9769
Boscalid	0.51 (0.36-0.67)	<i>y</i> =5.3117+0.9572 <i>x</i>	0.9760
Fluxapyroxad	0.19 (0.15-0.23)	<i>y</i> =5.6713+0.8878 <i>x</i>	0.9805

Comnd	Fungicidal acti	vity (%) at 2	200 µg/mL
Compa	E. graminis ^a	P. sorghi	R. solani
5a	0	0	0
5b	0	0	0
5c	100	20	50
5d	20	90	10
5e	0	0	0
5f	0	90	60
5g	0	98	90
5h	0	90	90
5i	0	100	100
5j	0	20	40
5k	0	0	0
51	100	20	40
5m	0	40	50
5n	0	0	20
50	100	0	30
5р	0	30	0
5q	0	98+ ^b	0
5r	0	20	50
5 s	0	20	80
5t	100	0	100
5u	90	30	50
Boscalid	100	100	80
Fluxapyroxad	100	100	70

 Table 3 In vivo fungicidal activities of the target compound

^a: *E. graminis*, wheat white powder; *P. sorghi*, corn rust; *R. solani*, Rice Sheath Blight.

572 ^b: Existing phytotoxicity.

Table 4. Greenhouse in vivo fungicidal activity validation

Compd	P. sorghi			R. solani				
-	50µg/mL	10µg/mL	1µg/mL	50µg/mL	10µg/mL	1µg/mL		
5g	30	0	0	30	20	0		
5h	85	0	0	20	10	0		
5i	100	40	0	80	70	60		
Boscalid	70	30	0	90	60	30		
Fluxapyroxad	100	100	85	100	100	100		

⁵⁷³

576Table 5 Inhibitory effect of **5i** against *R. cerealis* SDH

Compd	$IC_{50}(\mu g/mL)$	Regression equation	R ²
 5i	0.60	<i>y</i> =5.2854+1.2844 <i>x</i>	0.9806
Fluxapyroxad	0.29	<i>y</i> =5.7100+1.3059 <i>x</i>	0.9529

Graphic for table of contents

579

578

