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Chemoenzymatic Synthesis of (-)-Ribisins A and B from Dibenzo[*b*,*d*]furan

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ABSTRACT: *cis*-Dihydrodiols, derived from monocyclic aromatic compounds, are valuable chiral pool intermediates for the synthesis of cyclic natural products. A drawback of this approach, to the synthesis of polycyclic secondary metabolites, is that additional rings must be annulated. To date, relatively few chiral natural products have been synthesized from polycyclic arene *cis*-dihydrodiols. Fungal metabolites, (–)-ribisins A and B, have now been obtained by functional group manipulation of a tricyclic arene metabolite, obtained from toluene dioxygenase-catalyzed regioselective and stereoselective *cis*-dihydroxylation of dibenzo[b,d]furan. The synthetic sequences were marginally shorter than the alternative routes, using monocyclic arene *cis*-dihydrodiols and required no carbon-carbon bond-forming reactions.



INTRODUCTION

The dibenzo[*b*,*d*]furan (dibenzofuran) skeleton is present in a number of biologically interesting chiral natural products, including (+)-galanthamine, opiates *e.g.* (-)-morphine, and the (-)-ribisins (Figure 1). Ribisins are a family of polyoxygenated dibenzofuran derivatives, which were isolated by Fukuyama and co-workers, from the fruiting bodies of the white rot fungus *Phellinus ribis*.¹ This microbial species has a history of use in traditional medicines where it was used to enhance immunity, as well as to treat gastrointestinal cancer, liver disease, cardiovascular disease and diabetes.²⁻⁴ Ribisins A-D, in particular, enhance the ability of nerve growth factor to promote neurite outgrowth in PC12 cells and may have potential in the treatment of stroke, brain trauma and Alzheimer's disease.¹

Two approaches to the synthesis of ribisins have, to date, been reported.⁵⁻⁷ The key step in Du's synthetic approach was to use a Ferrier rearrangement of an enol ether, derived from *α*-D-glucopyranose, generating all three stereocentres of (-)-ribisin A with the correct absolute configurations.⁵ Banwell's seminal syntheses were chemoenzymatic and used a chiral *cis*-dihydrodiol, derived from bromobenzene, as starting material.^{6, 7} This approach not only gave additional material for further biological evaluation but also led to a correction of the published absolute configuration for (-)ribisin C and the relative stereochemistry for (-)-ribisin B, where two out of the three stereocentres were reassigned.



Figure 1. Examples of natural products containing dibenzo[b,d] furan subunits.

The structures of (-)-ribisins B and C shown in Figure 1 are thus the corrected versions determined by Banwell *et al.* ^{6,7} A possible biosynthetic pathway, to (-)-ribisins A-D from 2hydroxycinnamic acid in *P. ribis, via* oxidoreductasecatalyzed reactions, was reported by Fukuyama *et al.*¹

This proposed route involved the formation of an intermediate dibenzofuran-1,3-(2H,4H)-dione tautomer of dibenzofuran-1,3-diol, and enzyme-catalyzed oxidations to

yield (-)-ribisins A and B. Monooxygenase-catalyzed epoxidation of benzoate esters to form arene oxides, oxepines and phenols,⁸ and laccase- catalyzed oxidation of phenols were reported using *P. ribis* cultures.⁹ The postulated biosynthetic sequence, through a substituted dibenzofuran precursor prompted the current chemoenzymatic approach to (-)-ribisins A and B from dibenzofuran (Scheme 1).



Scheme 1. Proposed biosynthetic route to (-)-ribisins A and B from 2-hydroxycinnamic acid and a dibenzofuran-1,3-diol intermediate



Scheme 2. Synthesis of (-)-ribisin A



Scheme 4. Attempted synthesis of (-)-ribisin C

Biotransformations of monocyclic aromatic compounds are routinely utilized, to produce enantiopure cis-dihydrodiol metabolites for application in the chemoenzymatic synthesis of natural products.^{6, 7, 10-19} Conversely, *cis*-dihydrodiol metabolites, from polycyclic arenes or heteroarenes, are rarely used in chemoenzymatic synthesis.²⁰ Thus, although the isolation and stereochemical assignment of cis-dihydrodiols from tricyclic aromatic substrates was reported, using naphthalene dioxygenase (NDO)²¹ or biphenyl dioxygenase (BPDO)²⁰⁻²³ as biocatalysts, few examples of their employment as homochiral synthons have been documented. Furthermore, biotransformations of polycyclic arene substrates, with the latter dioxygenase types, often resulted in the formation of mixtures of cis-dihydrodiol isomers in relatively low yields. Biotransformations, using the UV4 mutant strain of the soil bacterium P. putida and E. coli recombinant strains, both expressing toluene dioxygenase (TDO), have been widely used in arene *cis*-dihydroxylations. In our laboratories, anilines, phenols, phenolic ethers, dibenzothiophene and dibenzofuran are among the more unexpected substrates recently found to undergo TDOcatalyzed *cis*-dihydroxylations.²⁴⁻²⁷

One possible chemoenzymatic approach to the synthesis of (-)ribisins, was to start with dibenzofuran as substrate, and then to add the two oxygen atoms to the *cis*-dihydrodiol metabolite in a stereo-controlled manner. This approach had the advantage that no carbon-carbon bond-forming reactions occurred in the entire synthetic sequence and the number of steps in the synthesis could, in principle, be reduced.

RESULTS AND DISCUSSION

Based on predictions, from molecular docking studies, and the use of *P. putida* UV4 whole cells (as TDO source), biotransformation of dibenzofuran resulted in exclusive *cis*dihydroxylation at the pseudo-bay region leading to the isolation of metabolite **1** (84% yield), with no evidence of other *cis*-dihydrodiol isomers being formed.²⁷ *cis*-Diol **1**, with dearomatization in one of the rings, was ideally functionalized for conversion to (-)-ribisins A and B as it was enantiopure (>98% *ee*) and had the required (1*S*,2*S*) absolute configuration, based on X-ray crystallographic analysis of a *cis*-diol epoxide derivative (Scheme 2-4).²⁷

Starting from dibenzofuran *cis*-dihydrodiol **1**, a nine-step synthesis of (-)-ribisin A is outlined (Scheme 2). *cis*-Diol **1** was found to be acid-sensitive and readily dehydrated regioselectively, to give dibenzofuran-2-ol. Using the weak acid catalyst PPTS, *cis*-diol **1** was converted to acetonide **2** in 73 % yield along with the dehydration product dibenzofuran-2-ol (11 % yield). During the TDO-catalyzed biotransformation of dibenzofuran, traces of a further *cis*dihydrodiol metabolite, derived from *cis*-dihydroxylation of dibenzofuran-2-ol, was also detected.²⁷

cis-Dihydroxylation of alkene **2**, employing Upjohn osmylation conditions,²⁸ proceeded as expected, exclusively, from the face *anti* to the bulky acetonide protecting group, to give the *cis*-diol acetonide diastereoisomer **3** (91% yield).

Due to the steric bulk of the acetonide group, attempted Mitsunobu inversion of configuration, at the C-4 chiral alcohol center of compound 3 was unsuccessful in our hands. The stereoinversion was later achieved in two steps, by a chemoselective oxidation of the more activated alcohol group of diol 3 to form α -hydroxyketone 4, followed by a stereoselective reduction to give trans-diol 5. Although a number of methods was available, for the chemoselective oxidation of activated alcohols, a problem arose in this particular case; the oxidation product, α -hydroxyketone 4, was found to be very sensitive to further oxidation, leading to carbon-carbon bond cleavage. After screening oxidation protocols, it was found that chemoselective oxidation with OxoneTM in the presence of TEMPO,²⁹ yielded α hydroxyketone 4 in 86% yield. Sodium borohydride reduction of ketone 4 gave exclusively trans-diol 5 in 89% yield, which was protected as *bis-p*-methoxybenzyl ether 6 (54% yield). The acetonide group of ketal 6 was removed under acidic conditions, to furnish the resulting cis-diol 7 (75% yield). Chemoselective oxidation (TEMPO, OxoneTM) of the C-1 hydroxyl group located within the hindered pseudo-bay region of *cis*-diol 7 gave α -hydroxyketone 8 (55% yield). The alcohol group of ketone 8 was converted (Ag₂O/MeI) to methyl ether 9 (66% yield). In the final step, oxidative removal of the PMB protecting groups using DDQ gave (-)-ribisin A in 63% yield. The spectroscopic properties of the synthetic sample were in good agreement with the natural product obtained from P. ribis. The synthesis of (-)ribisin A, from dibenzofuran cis-dihydrodiol 1, was accomplished in nine steps (9% overall vield), a modest improvement over fourteen and eleven steps from the previous two routes.5-7

cis-Diol **3** also proved to be a versatile intermediate for the synthesis of (-)-ribisin B (Scheme 3) as all stereocentres already had the correct absolute configurations. To complete its synthesis, regisoselective O-methylation and alcohol oxidation to form a ketone were required. Reaction of diol 3 with anisaldehyde dimethyl acetal formed an exo:endo mixture (57:43) of anisylidene acetal diastereoisomers **10a:10b.** Due to the strong relaxation of the acetal protons $(R_2 \text{ or } R_1 = H)$ by the *ortho* proton on the PMP group, distant nOe values were not observed and, therefore, the exo isomer 10a and endo isomer 10b could not be assigned by this method. However, the chemical shift for C-10c proton in diol **3** was 5.42 ppm while in acetals **10a** and **10b** the chemical shifts of the corresponding pseudo-bay region protons were 5.44 and 5.62 ppm respectively. This indicates that the pmethoxyphenyl group of the minor *endo* acetal isomer **10b** is pseudo-axial and proximate to the C-10c proton. This premise was tentatively used to differentiate between the exo and endo isomers. The difference in chemical shift (0.32 ppm) of the acetal protons 10a (R_2 6.02 ppm) and 10b (R_1 5.70 ppm) can be rationalised by the proximity of proton R_2 to the benzofuran ring, lending further tentative support to the stereochemical assignment. Attempts to regioselectively alkylate diol 3 using sodium hydride and PMB-Cl were unsuccessful.

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After careful multiple elution PLC, only the minor endo diastereoisomer 10b could be fully separated from a small sample of the mixture. DIBAL reduction of acetal **10b** gave. exclusively the unwanted PMB-ether 12. Reduction (DIBAL) of the original mixture of acetal diastereoisomers 10a/10b (57:43) yielded a further mixture (58:42) of readily separable alcohols 11 (30% yield) and 12 (16% yield). The selectivity for the desired exo-acetal 10a may be improved by synthesis of the more sterically demanding 2,4dimethoxyphenyl acetal. Methylation (MeI/NaH) of alcohol 11 yielded methyl ether 13 (90% yield). The ketal group in 10 methyl ether 13 was cleaved, under acidic conditions (TFA, 11 THF, H_2O), in a similar manner to that used for acetonide 6 12 (Scheme 2), to form cis-diol 14 (97% yield). The unwanted regioisomer 12 could, in principle also be converted to 13 intermediate 13 through protecting group manipulation, but 14 this route was not investigated. The C-1 hydroxyl group of 15 diol 14 was chemoselectively oxidized to ketone 15 (catalytic 16 TEMPO with OxoneTM, Scheme 3) (86% yield). Methylation 17 (Ag₂O/MeI) of α -hydroxy ketone 15 gave dimethoxy ketone 18 16 (76% yield). Finally, the PMB protecting group was 19 oxidatively removed (DDQ, H₂O, CH₂Cl₂), from PMB-ether 20 16, to furnish (-)-ribisin B (77% yield) in10% overall yield 21 from cis-dihydrodiol 1 over nine steps. 22

As part of an earlier study of biotransformations (P. putida UV4) of tricylic heteroarenes, the isolation of (1S, 2S)dihydrodiol metabolite 1, and its potential as a chiral synthon, was demonstrated by the synthesis and absolute configuration assignment of the diol epoxide derivative 18 by X-ray crystallography, via treatment of bromohydrin intermediate 17 with sodium methoxide.27 The possibility of epoxide 18 undergoing nucleophilic attack by methoxide anion, using different solvents and temperatures, to yield triols 19 and 20, was examined (Scheme 4). Despite these changes in reaction conditions neither of the triols was obtained. Reactions of unactivated diol epoxides toward nucleophilic attack had been reported by using Al₂O₃ in MeOH solution.³⁰ However, under similar reaction conditions, the required triol 19 was not observed among the range of unidentified products. An alternative synthetic approach to (-)-ribisin C, using chiral intermediate 4, is currently under investigation.

CONCLUSION

The chemoenzymatic syntheses of natural products, from cisdihydrodiol metabolites, have generally been restricted to those obtained from monocyclic arene substrates.6-8, 10-19 cis-Dihydrodiols of acridine and dictamnine, obtained by BPDO-catalyzed cis-dihydroxylation, were among the few examples of tricyclic arenes utilized in the chemoenzymatic synthesis of secondary metabolites.²⁰ Following on from the innovative use of a monocyclic arene cis-dihydrodiol of bromobenzene in chemoenzymatic syntheses,^{6, 7} this study has demonstrated that a tricyclic *cis*-dihydrodiol, formed by TDO-catalyzed *cis*-dihydroxylation of dibenzofuran,²⁷ can also be used as a precursor for the asymmetric synthesis of (-)-ribisins A and B. This alternative approach of using a polycyclic cis-dihydrodiol precursor can now be added to the available methods for the synthesis of chiral natural products containing benzofuran rings.31

EXPERIMENTAL SECTION

¹H and ¹³C{1H} NMR spectra were recorded on Bruker AV-400 and AV-600 instruments using specified solvents. Chemical shifts (δ) are reported in ppm relative to SiMe₄ and coupling constants (J) are given in Hertz (Hz). Optical rotations ($[\alpha]_D$) measurements (10⁻¹ deg cm² g⁻¹) were carried out at ambient temperatures (20-25°C) on a Perkin-Elmer 214 polarimeter and specified solvent concentration (g/100 mL) at the sodium D-line (589 nm). IR spectra were recorded in KBr disc or in thin film. Mass spectra (ES) were recorded on an LCT Premier Mass Spectrometer using an ESI-TOF analyser. Accurate molecular weights were obtained by the peak matching method, using heptacosafluorotributylamine as the standard reference, and were accurate to within $\pm 5 \times 10^{-6}$ ppm. Melting points were recorded in degrees Celsius using a Stuart SMP10 melting point apparatus. Merck Kieselgel 60F₂₅₄ analytical plates were used for TLC analyses. Thermostatically controlled oil baths were used for reactions at elevated temperatures. Preparative layer chromatography (PLC) separations were carried out on glass plates (20 cm x 20 cm) coated with Merck Keiselgel PF_{254/366} silica gel (21 g silica gel in 62 mL water). Flash column chromatography was performed on Fluorochem silicagel 60A 40-63u.

(1R,2S)-1,2-Dihydrodibenzo[b,d]furan-1,2-diol (1).

Dibenzofuran was biotransformed using (P. putida UV4) according to the literature procedure, to give diol 1 in excellent yield (84%).²⁷ This product was used in the current study.

(3aR.10cS)-2.2-Dimethyl-3a.10c-

dihydrobenzo[b][1,3]dioxolo[4,5-e]benzofuran (2).22 To a stirred solution of *cis*-dihydrodiol 1 (270 mg, 1.34 mmol) in a mixture of acetone (6 mL) and 2,2-DMP (6 mL), maintained at 0°C, was added pyridinium *p*-toluenesulfonate (PPTS, 234 mg, 0.93 mmol). The reaction mixture was stirred (2 h) at 0°C and then at room temperature for 12 h. After the addition of a saturated aqueous solution of NaHCO₃ (5 mL), the mixture was concentrated under reduced pressure and extracted with EtOAc (2 x 15 mL). The organic phase was dried (Na₂SO₄), concentrated and the crude product purified by flash column chromatography (4% EtOAc in hexane), to afford acetonide 2, a white solid, (237 mg, 73%); $R_{\rm f} = 0.4$ (10% EtOAc in hexane); $[\alpha]_{\rm D}^{22}$ +130.8 (c 0.66, CHCl₃); HRMS (ES) calc. for $C_{15}H_{14}O_3Na$ (M + Na)⁺ 265.0836 found 265.0841; ¹H-NMR (400 MHz, CDCl₃): δ 7.65 (m, 1H), 7.45 (m, 1H), 7.26 (m, 2H), 6.57 (dd, J = 10.0, 1.3 Hz, 1H), 6.08 (dd, J = 10.0, 3.5 Hz, 1H), 5.43 (d, J = 8.3Hz, 1H), 5.00 (ddd, J = 8.3, 3.5, 1.3 Hz, 1H), 1.52 (s, 3H), 1.38 (s, 3H); ¹³C{¹H}-NMR (100 MHz, CDCl₃): δ 155.1, 151.3, 129.2, 127.1, 124.5, 123.5, 119.5, 118.0, 111.6, 111.0, 106.6, 73.0, 69.6, 27.0, 25.1; IR v_{max}/cm⁻¹ 2068, 1935, 1446. ¹H NMR data was in good agreement with literature values.

The side product, dibenzo[b,d]furan-2-ol (27 mg, 11%), was also isolated from the crude product. NMR data in good agreement with literature values.³² ¹H-NMR (400 MHz, $CDCl_3$) δ_H 7.86 (d, J = 7.8 Hz, 1H), 7.53 (d, J = 8.2 Hz, 1H), 7.44 (td, J = 7.1, 1.3 Hz, 1H), 7.42 (d, J = 8.8 Hz, 1H), 7.36 (d, J = 2.7 Hz, 1H), 7.30 (1H, td, J = 7.5, 0.9 Hz, H), 6.95 $(1H, dd, J = 8.8, 2.7 Hz, 1H), 4.98 (1H, br s, OH); {}^{13}C{}^{1}H{}$ -NMR (100 MHz, CDCl₃) $\delta_{\rm C}$ 157.0, 151.5, 151.0, 127.3, 125.1, 124.2, 122.5, 120.7, 115.3, 112.1, 111.8, 106.3.

(3aS,4R,5S,10cR)-2,2-Dimethyl-3a,4,5,10c-

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tetrahydrobenzo[b][1,3]dioxolo[4,5-e]benzofuran-4,5-diol (3). To a stirred solution of acetonide 2 (17.5 mg, 0.072 mmol) and N-methylmorpholine-N-oxide (10.0 mg), in a mixture of acetone (4.5 mL) and water (0.5 mL), maintained at 0°C, was added a solution of osmium tetroxide in BuOH (2.5 wt %, 70 µl). After stirring the reaction mixture overnight at room temperature, a saturated aqueous solution of Na₂S₂O₅ (1 mL) was added to it and the stirring continued for another 10 mins. The reaction mixture was concentrated under reduced pressure, water (2 mL) added to it, the mixture thoroughly extracted with EtOAc (2 x 5 mL) and the organic extract was dried (Na₂SO₄) and concentrated. The crude product obtained was purified by flash column chromatography (60% EtOAc in hexane) to afford cis-diol 3 (18.2 mg, 91%), a white solid, $R_{\rm f} = 0.2$ (40% EtOAc in hexane): $[\alpha]_{D}^{22}$ +30.6 (c 0.36, MeOH); HRMS (ES) calc. for C₁₅H₁₆O₅Na (M + Na)⁺ 299.0887 found 299.0895; ¹H-NMR (400 MHz, CDCl₃): δ_H 7.67 (m, 1H), 7.50 (m, 1H), 7.33 (dt, J = 7.3, 1.3 Hz, 1H), 7.28 (dt, J = 7.5, 1.3 Hz, 1H), 5.42 (d, J = 5.5 Hz, 1H), 5.02 (d, J = 3.9 Hz, 1H), 4.60 (dd, J = 7.3, 5.5 Hz, 1H), 4.26 (dd, J = 7.3, 3.9 Hz, 1H), 2.88 (br s, 1H), 2.76 (br s, 1H), 1.52 (s, 3H,), 1.41 (s, 3H,). ¹³C{¹H}-NMR (100 MHz, CDCl₃): δ_C 155.9, 151.9, 126.4, 125.3, 123.4, 120.2, 113.0, 111.8, 111.0, 71.8, 69.5, 64.0, 29.7, 28.21, 26.3; IR v_{max}/cm⁻¹ 3454, 2988, 2935, 1958, 1630, 1245.

(3aS,4S,10cR)-4-Hydroxy-2,2-dimethyl-3a,4-

25 dihvdrobenzo[b][1,3]dioxolo[4,5-e]benzofuran-5(10cH)-26 one (4). A stirred solution of acetonide diol 3 (20 mg, 0.072 27 mmol) containing tetrabutylammonium bromide (1 mg) in 28 dry CH₂Cl₂ (2 mL) was treated with TEMPO (1 mg) and 29 Oxone (55 mg, 0.18 mmol). The reaction mixture was stirred 30 overnight at room temperature, concentration under reduced 31 pressure and the crude material purified by flash column 32 chromatography (20% EtOAc in hexane), to give hydroxy 33 ketone 4 (17.2 mg, 86%), a white solid; $R_{\rm f} = 0.2$ (30% EtOAc 34 in hexane); $[\alpha]_{D}^{22}$ +35.0 (c 0.8, CHCl₃); HRMS (ES) calc. 35 for C₁₅H₁₅O₅ (M + H)⁺ 275.0919 found 275.0922; ¹H-NMR 36 (600 MHz, CDCl₃): δ_H 7.85 (d, J = 7.8 Hz, 1H), 7.63 (d, J =37 8.6 Hz, 1H), 7.58 (dt, J = 7.2, 1.2 Hz, 1H), 7.43 (dt, J = 7.2, 38 1.0 Hz, 2H), 5.58 (d, J = 5.0 Hz, 1H), 4.73-4.68 (m, 1H), 3.58 39 (br s, 1H), 1.59 (s, 3H), 1.57 (s, 3H); ¹³C{¹H}-NMR (150 40 MHz, CDCl₃): δ_{C} 185.4, 157.4, 146.0, 130.1, 128.1, 125.5, 124.8. 122.3. 113.1. 112.9. 81.5. 76.4. 68.4. 28.1. 26.5: IR 41 $v_{\rm max}/{\rm cm}^{-1}$ 3430, 3089, 3066, 2990, 2810, 1711, 1597, 1542, 42 1216, 1065, 1001, 900. 43

(3aS,4R,5R,10cR)-2,2-Dimethyl-3a,4,5,10c-

45 tetrahydrobenzo[b][1,3]dioxolo[4,5-e]benzofuran-4,5-diol 46 (5). An ice-cooled solution of hydroxyketone 4 (205 mg, 0.75 47 mmol) in MeOH (10 mL) was treated with sodium 48 borohydride (57 mg, 1.5 mmol) and the mixture stirred (0.5 49 h) at room temperature and concentrated under reduced 50 pressure. The residue was dissolved in CH₂Cl₂ (20 mL), 51 washed with 1 M HCl (5 mL) and the aqueous washing 52 extracted with CH₂Cl₂ (2 x 5 mL). The combined organic 53 phase was dried (MgSO₄) and concentrated to afford a pure sample of *trans*-diol 5 (184 mg, 89%), a white solid; $R_f = 0.2$ 54 (50% EtOAc in hexane); $[\alpha]_D^{22}$ +35.9 (c 1.0, CHCl₃); HRMS 55 (ES) calc. for $C_{15}H_{20}NO_5$ (M + NH₄)⁺ 294.1341 found 56 294.1355; ¹H-NMR (600 MHz, CDCl₃): δ_H 7.65 (d, J = 7.657 Hz, 1H), 7.50 (d, J = 8.1 Hz, 1H), 7.32 (dt, J = 7.5, 1.4 Hz, 58

1H), 7.27 (dt, J = 7.6, 1.0 Hz, 1H), 5.39 (d, J = 5.4 Hz, 1H), 4.79 (d, J = 5.9 Hz, 1H), 4.49 (dd, J = 6.9, 5.5 Hz, 1H), 4.31(dd, J = 6.9, 5.9 Hz, 1H), 3.33 (br s, 1H), 2.95 (br s, 1H), 1.50 (s, 3H), 1.41 (s, 3H); ${}^{13}C{}^{1}H$ -NMR (150 MHz, CDCl₃): δ_C 155.9, 152.5, 126.4, 125.1, 123.4, 120.1, 111.8, 111.7, 111.6, 78.3, 74.3, 69.5, 67.6, 28.3, 26.5; IR v_{max} /cm⁻¹ 3498, 2998, 2902, 1967, 1629, 1220.

(3aR,4S,5R,10cR)-4,5-bis((4-Methoxybenzyl)oxy)-2,2-

dimethyl-3a,4,5,10c-tetrahydrobenzo[b][1,3]dioxolo[4,5elbenzofuran (6). To a solution of trans-diol 5 (18 mg, 0.065 mmol) and 4-methoxybenzyl-2,2,2-trichloroacetimidate (92 mg) in toluene (4 mL) was added lanthanum (III) triflate (7.6 mg). After stirring (5 mins) the reaction mixture at room the solvent was removed under reduced temperature, pressure and the crude product purified by PLC (10% EtOAc in hexane), to furnish methoxybenzyl ether 6 (18 mg, 54%), a pale-yellow oil; $R_f = 0.2$ (20% EtOAc in hexane); $\left[\alpha\right]_{\mathrm{D}}^{22}$ +23.6 (c 1.2, CHCl₃); HRMS (ES) calc. for C₃₁H₃₂O₇Na (M + Na)⁺ 539.2046 found 539.2048; ¹H-NMR (600 MHz, CDCl₃): δ_H 7.65 (d, J = 7.6 Hz, 1H), 7.50 (d, J = 8.2 Hz, 1H), 7.39 (d, J = 8.5 Hz, 2H), 7.32 (td, J = 7.6 1.2 Hz, 1H) 7.29-7.25 (m, 3H), 6.90 (d, J = 8.5 Hz, 2H), 6.86 (d, J = 8.5 Hz, 1H), 5.35 (d, J = 5.7 Hz, 1H), 4.96 (d, J = 11.2 Hz, 1H), 4.80 (d, J = 11.2 Hz, 1H), 4.76 (d, J = 11.1 Hz, 1H), 4.69 (d, J = 11.1 Hz, 1H)11.1 Hz, 1H), 4.67 (1H, d, J = 5.8 Hz, 1H), 4.51 (1H, dd, J =6.9, 5.8 Hz, 1H), 4.13 (dd, J = 6.9, 5.8 Hz, H-4), 3.82 (s, 3H), 3.80 (s, 3H), 1.50 (s, 3H), 1.40 (s, 1H); ${}^{13}C{}^{1}H$ -NMR (150) MHz, CDCl₃): δ_{C} 159.4 (4), 155.9, 153.0, 130.2 (2), 129.8, 126.5, 124.9, 123.2, 120.0, 113.8 (4), 112.2, 111.7, 111.2, 80.2, 78.2, 73.9, 72.9, 72.7, 69.3, 55.3 (2), 28.2, 26.2; IR $v_{\rm max}/{\rm cm}^{-1}$ 2968, 2931, 2844, 1520, 1261, 1107, 871.

(1R,2R,3S,4R)-3,4-bis((4-Methoxybenzyl)oxy)-1,2,3,4-

tetrahydrodibenzo[b,d]furan-1,2-diol (7). A solution of methoxybenzyl ether 6 (18 mg, 0.035 mmol) in THF-H₂O (4:1, 3 mL) was treated with TFA (70 µL). The reaction mixture was stirred (40 h) at 60°C, concentrated under reduced pressure, and the crude product purified by flash column chromatography (40% EtOAc in hexane), to give cisdiol 7 (12.5 mg, 75%), a colorless oil; $R_f = 0.2$ (50% EtOAc in hexane): $[\alpha]_D^{22}$ +19.0 (c 1.3, CHCl₃); HRMS (ES) calc. for C₂₈H₂₈O₇Na

(M + Na)⁺ 499.1733 found 499.1732; ¹H-NMR (400 MHz, CDCl₃): δ_H 7.77 (d, J = 7.6 Hz, 1H), 7.50 (d, J = 8.2 Hz, 1H), 7.35-7.30 (m, 3H), 7.27 (td, J = 7.7, 0.9 Hz, 1H) 7.21 (d, J =8.6 Hz, 2H), 6.90 (d, J = 8.6 Hz, 2H), 6.86 (d, J = 8.7 Hz, 2H), 5.08 (dd, J=6.8, 4.2 Hz, 1H), 4.91 (d, J=11.2 Hz, 1H), 4.74 (d, J = 11.2 Hz, 1H), 4.73 (d, J = 11.4 Hz, 1H), 4.70 (d, J = 4.7 Hz, 1H), 4.59 (d, J = 11.4 Hz, 1H), 4.22 (1H, dd, J =7.0, 4.2 Hz, 1H), 4.03 (ddd, J = 10.5, 6.4, 4.2 Hz, 1H), 3.82 (s, 3H), 3.80 (s, 3H); ${}^{13}C{}^{1}H$ -NMR (150 MHz, CDCl₃): δ_C 159.6 (2), 155.9, 151.3, 129.8 (2), 129.7, 129.6 (2), 129.4, 126.8, 125.0, 123.2, 120.9, 115.9, 114.0 (4), 111.6, 77.9, 73.5, 73.2, 72.4, 70.4, 64.3, 55.4 (2); IR v_{max}/cm⁻¹ 3996, 2954, 2917, 2849, 1575, 1542, 1249, 1089, 1011.

(2S,3S,4R)-2-Hydroxy-3,4-bis((4-methoxybenzyl)oxy)-3,4-

dihydrodibenzo[b,d]furan-1(2H)-one (8). To a stirred solution of cis-diol 7 (12 mg, 0.025 mmol), containing tetrabutylammonium bromide (1 mg) in dry CH₂Cl₂ (2 mL), was added TEMPO (1 mg) and Oxone[™] (18 mg, 0.06 mmol). After stirring the reaction mixture, overnight at room

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temperature, the solvent was removed under reduced pressure and the crude product purified by flash column 2 chromatography (30% EtOAc in hexane), to afford α hydroxyketone 8 (6.6 mg, 55%), a white solid; $R_{\rm f} = 0.3$ (30%) EtOAc in hexane); $[\alpha]_D^{22}$ +34.0 (c 0.7, CHCl₃); HRMS (ES) calc. for $C_{28}H_{26}O_7Na$ (M + Na)⁺ 497.1576 found 6 497.1506; ¹H-NMR (400 MHz, CDCl₃): δ_H 8.03 (m, 1H), 7.57 (m, 1H), 7.44-7.34 (m, 6H), 6.91 (d, J = 8.6 Hz, 2H), 6.88 (d, J = 8.6 Hz, 2H), 5.10 (d, J = 10.9 Hz, 1H), 5.01 (d, J)8 = 10.7 Hz, 1H), 4.98 (d, J = 7.9 Hz, 1H), 4.94 (d, J = 10.9 9 Hz, 1H), 4.83 (d, J = 10.7 Hz, 1H), 4.45 (dd, J = 9.8, 1.9 Hz, 10 1H), 4.06 (dd, J = 9.8, 7.9 Hz, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 11 3.80 (d, J = 2.0 Hz, 1H); ¹³C{¹H}-NMR (150 MHz, CDCl₃): 12 δ_{C} 191.0, 167.4, 159.4 (2), 156.2, 130.0 (4), 129.6, 126.3, 13 125.1, 122.3, 122.0, 114.9, 113.9 (4), 111.9, 85.8, 77.8, 75.2, 14 75.1, 75.1, 74.7, 55.3 (2); IR v_{max}/cm^{-1} 3453, 3070, 2927, 15 2852, 2778, 1771, 1707, 1625, 1515, 1366, 1169, 1014. 16

17 (2S, 3R, 4R)-2-Methoxy-3, 4-bis((4-methoxybenzyl)oxy)-3, 4-18 dihydrodibenzo[b,d]furan-1(2H)-one (9). A glass tube, containing a mixture of α -hydroxyketone 8 (6.5 mg, 0.014 19 mmol), Ag₂O (12 mg) and MeI (10 µL) in dry THF (2 mL), 20 was sealed under argon and the mixture stirred (24 h) at 60°C. 21 The solvent was removed under reduced pressure and the 22 crude product purified by flash column chromatography 23 (15% EtOAc in hexane) to yield methyl ether 9 (4.4 mg, 24 66%), a colorless oil; $R_f = 0.2$ (20% EtOAc in hexane); $[\alpha]_D^{22}$ 25 +47.7 (c 0.4, CHCl₃); HRMS (ES) calc. for C₂₉H₃₂NO₇ (M + 26 NH₄)⁺ 506.2179 found 506.2193; ¹H-NMR (400 MHz, 27 CDCl₃): $\delta_H 8.06$ (d, J = 7.2 Hz, 1H), 7.55 (d, J = 7.8 Hz, 1H), 28 7.43-7.28 (m, 6H), 6.91 (d, J = 8.8 Hz, 2H), 6.89 (d, J = 8.8Hz, 2H), 5.05 (d, J = 11.1 Hz, 1H), 4.95-4.88 (m, 2H), 4.86 29 (d, J = 10.7 Hz, 1H), 4.75 (d, J = 10.7 Hz, 1H), 4.14 (dd, J =30 9.3, 7.3 Hz, 1H), 3.99 (d, J = 9.3 Hz, 1H), 3.83 (s, 3H), 3.82 31 (s, 3H), 3.77 (3H, s, 3H); ¹³C{¹H}-NMR (150 MHz, CDCl₃): 32 $\delta_{\rm C}$ 190.6, 165.9, 159.6, 159.5, 156.1, 130.2, 129.9, 129.5, 33 126.2, 124.9, 122.8, 122.3, 115.9, 113.9, 111.7, 86.2, 83.7, 34 75.3, 74.8, 74.3, 61.0, 55.3; IR v_{max} /cm⁻¹ 2969, 2882, 1735, 35 1679, 1280, 1021, 976 36

37 (2S,3R,4R)-3,4-Dihydroxy-2-methoxy-3,4-38 dihydrodibenzo[b,d]furan-1(2H)-one (-)- (Ribisin A).^{1,7} To a 39 solution of methyl ether 9 (9.0 mg, 0.018 mmol) in a mixture 40 of CH₂Cl₂ and H₂O (4:1, 2 mL) was added DDO (12.5 mg) and the mixture stirred (18 h) at room temperature. Water (2 41 mL) and CH₂Cl₂ (5 mL) were added to the stirring mixture, 42 the organic phase separated, and the remaining aqueous 43 phase extracted with CH₂Cl₂ (5 mL). The combined organic 44 extract was dried (MgSO₄), concentrated, and the residue 45 purified by flash column chromatography (50% EtOAc in 46 hexane) to afford (-)-ribisin A (2.9 mg, 63%), a pale-yellow 47 solid; $R_{\rm f} = 0.2$ (60% EtOAc in hexane); $[\alpha]_{\rm D}^{22}$ 48 -16.0 (c 0.2, MeOH); HRMS (ES) calc. for C₂₆H₂₄O₁₀Na (2M 49 + Na)+ 519.1267 found 519.1259; 1H-NMR (600 MHz, 50 CDCl₃): δ_H 7.93 (dm, J = 7.6 Hz, 1H), 7.56 (dm, J = 8.1 Hz, 51 1H), 7.36 (dt, J = 7.4, 1.4 Hz, 1H), 7.32 (dt, J = 7.6, 1.0 Hz, 52 1H), 4.93 (d, J = 7.6 Hz, 1H), 3.97 (d, J = 9.9 Hz, 1H), 3.92 (dd, J = 9.9, 7.6 Hz, 1H), 3.66 (s, 3H); ¹³C{¹H}-NMR (150 53 MHz, CDCl₃): δ_C 193.1, 169.4, 157.6, 127.2, 126.0, 124.3, 54 122.9, 116.1, 112.8, 87.2, 78.9, 70.4, 60.8. 55 56

(3aR, 3bS, 5S, 6aS, 11cR)-5-(4-Methoxyphenyl)-2, 2-imethyl-3a, 3b, 6a, 11c-tetrahydrobenzo[4', 5']furo[2', 3':5, 6]benzo[1, 2d:3,4-d']bis([1,3]dioxole) and (3aR,3bS,5R,6aS,11cR)-5-(4methoxyphenyl)-2,2-Dimethyl-3a,3b,6a,11c-

tetrahydrobenzo[4',5']furo[2',3':5,6]benzo[1,2-d:3,4-

d']bis([1,3]dioxole) (10a and 10b). To an ice-cooled solution of cis-diol 3 (40 mg, 0.145 mmol) and anisaldehyde dimethyl acetal (0.10 mL) in dry CH₂Cl₂ (3 mL) was added (+)-10camphorsulfonic acid (14 mg). The reaction mixture was stirred (3 h) at 0°C, warmed to room temperature and treated with saturated aqueous solution of NaHCO₃ (2 mL). The organic phase was separated, the remaining aqueous solution extracted with CH₂Cl₂ (2 x 3 mL), and the combined organic solution dried (MgSO₄) and concentrated. The resulting yellow oil was purified by flash column chromatography (8% EtOAc in hexane) to give a mixture of acetals exo 10a and endo 10b (57:43), a pale-yellow oil (50 mg, 87%); $R_f = 0.2$ (5% EtOAc in hexane); HRMS (ES) calc. for $C_{23}H_{22}O_6Na$ (M + Na)⁺ 417.1314 found 417.1309; Minor endo-diastereoisomer 10b: ¹H-NMR (400 MHz, CDCl₃): $\delta_{\rm H}$ 7.74 (dm, J = 6.9 Hz, 1H), 7.51 (dm, J = 7.1 Hz, 1H), 7.40-7.27 (m, 9H,), 7.14 (d, J = 8.7Hz, 2 H), 6.76 (d, J = 8.7 Hz, 2H), 6.02 (s, 1H), 5.44 (d, J =5.1 Hz, 1H), 5.39 (d, J = 6.5 Hz, 1H), 5.01 (dd, J = 6.5, 2.8 Hz, 1H), 4.91 (dd, J = 5.1, 2.8 Hz, 1H), 3.72 (s, 3H), 1.46 (s, 3H), 1.12 (s, 3H). Major exo-diastereoisomer 10a: ¹H-NMR (400 MHz, CDCl₃): δ_H 7.76 (dm, J = 7.3 Hz, 1H), 7.54 (dm, J= 7.7 Hz, 1H), 7.39 (d, J = 8.7 Hz, 2 H), 7.37-7.27 (m, 9H), 5.70 (s, 1H), 5.62 (d, J = 5.8 Hz, 1H), 5.51 (d, J = 4.9 Hz, 1H), 5.01 (dd, J = 5.8, 3.0 Hz, 1H), 4.91 (dd, J = 5.0, 3.0 Hz, 1H), 3.81 (s, 3H), 1.45 (s, 3H), 1.09 (s, 3H).

(3aS,4S,5S,10cR)-5-((4-Methoxvbenzvl)oxv)-2,2-dimethyl-3a,4,5,10c-tetrahydrobenzo[b][1,3]dioxolo[4,5-

e]benzofuran-4-ol (11) and (3aR,4R,5S,10cR)-4-((4-Methoxybenzyl)oxy)-2,2-dimethyl-3a,4,5,10c-

tetrahydrobenzo[b][1,3]dioxolo[4,5-e]benzofuran-5-ol (12). To a solution of cis-diol 3 (120 mg, 0.43 mmol) and (+)camphorsulfonic acid (40 mg), in dry CH₂Cl₂ (5 mL) maintained at -15°C, was added 1-(dimethoxymethyl)-4methoxybenzene (0.30 mL, 1.6 mmol). After stirring (4 h) the reaction mixture it was diluted with CH₂Cl₂ (10 mL), allowed to warm up to room temperature and washed with a saturated aqueous solution of NaHCO₃ (10 mL). The organic phase was dried (MgSO₄) and concentrated under reduced pressure to give a mixture of diastereomers 10a and 10b. This was taken up in dry CH₂Cl₂ (10 mL), cooled to -78°C and treated with a solution of DIBAL (1 M, 4.3 mL) in hexane. The mixture was stirred (6 h) at -78°C, allowed to warm to room temperature, and a saturated aqueous solution of potassium sodium tartrate (2 mL) was added. After stirring the reaction mixture overnight, the organic phase was separated and the remaining aqueous phase extracted with CH₂Cl₂ (5 mL). The combined organic extract was dried (MgSO₄) and concentrated. The crude product obtained was purified by flash column chromatography (25% EtOAc in hexane) to give benzylic ether 11, a pale-yellow solid, and homobenzylic ether 12, a pale-yellow oil.

Benzylic ether 11 (52 mg, 30%); $R_f = 0.2$ (30% EtOAc in hexane); $[\alpha]_{D^{22}}$ +8.4 (c 1.5, CHCl₃); HRMS (ES) calc. for C₂₃H₂₈NO₆ (M + NH₄)⁺ 414.1917 found 414.1928; ¹H-NMR (600 MHz, CDCl₃): δ_H 7.65 (d, J = 3.9 Hz, 1H), 7.48 (m, 1H), 7.32 (m, 2H), 7.30 (m, 1H), 7.26 (m, 1H), 6.89 (m, 2H), 5.39 (d, J = 5.6 Hz, 1H), 4.91 (t, J = 4.3 Hz, 1H), 4.82 (d, J = 11.7Hz, 1H), 4.72 (d, *J* = 11.7 Hz, 1H), 4.66 (dd, *J* = 7.3, 5.6 Hz, 1H), 4.02 (dd, J = 7.3, 4.0 Hz, 1H), 3.80 (s, 3H), 2.83 (d, J = 4.8 Hz, 1H), 1.50 (s, 3H), 1.33 (s, 3H); ¹³C{¹H}-NMR (150 MHz, CDCl₃): δ_C 159.7, 155.9, 152.0, 129.8, 129.6, 128.9, 126.4, 125.1, 123.3, 120.1, 114.1, 111.7, 110.8, 78.2, 75.8, 73.0, 69.6, 62.9, 55.3, 28.1, 26.3; IR v_{max}/cm^{-1} 2957, 2924, 2853, 1637, 1080, 1034, 923.

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Homobenzyl ether **12** (27 mg, 16%); $R_f = 0.2$ (30% EtOAc in hexane); $[\alpha]_D^{22}-8.2$ (*c* 0.9, CHCl₃); HRMS (ES) calc. for C₂₃H₂₈NO₆ (M + NH₄)⁺ 414.1917 found 414.1928; ¹H-NMR (600 MHz, CDCl₃): δ_H 7.69 (d, J = 7.5 Hz, 1H), 7.52 (d, J = 8.3 Hz, 1H), 7.35 (dt, J = 7.3, 1.2 Hz, 1H), 7.32 (d, J = 8.6 Hz, 2H), 7.29 (dt, J = 7.5, 1.0 Hz, 1H), 6.89 (d, J = 8.6 Hz, 2H), 5.42 (d, J = 5.7 Hz, 1H), 4.84 (d, J = 11.2 Hz, 1H), 4.74 (d, J = 4.1 Hz, 1H), 4.71 (d, J = 11.2 Hz, 1H), 4.57 (dd, J = 7.5, 5.8 Hz, 1H), 4.19 (td, J = 7.5, 4.0 Hz, 1H), 3.81 (s, 3H), 2.64 (d, J = 7.5 Hz, 1H), 1.51 (s, 3H), 1.43 (s, 3H); ¹³C {¹H}-NMR (150 MHz, CDCl₃): δ_C 159.6, 155.9, 152.2, 129.8, 129.4, 126.5, 125.3, 123.4, 120.3, 114.0, 113.6, 111.7, 110.8, 77.6, 72.3, 71.5, 70.3, 69.5, 55.3, 28.2, 26.2; IR ν_{max}/cm^{-1} 2958, 2925, 2853, 1641, 1080, 1012, 907.

A small portion of the minor diastereoisomer **10b** was separated from the acetal mixture **10a/10b** by multiple elution PLC. This sample on similar DIBAL reduction gave exclusively isomer **12**.

(3aR,4S,5S,10cR)-4-Methoxy-5-((4-methoxybenzyl)oxy)-2,2dimethyl-3a,4,5,10c-tetrahydrobenzo[b][1,3]dioxolo[4,5-

elbenzofuran (13). To a solution of alcohol 11 (16 mg, 0.040 mmol) in dry THF (4 mL) was added sodium hydride (5 mg), under an argon atmosphere, followed by the addition of methyl iodide (12 μ L). The reaction mixture was stirred (4 h) at room temperature, the solvent removed under reduced pressure, and the crude product purified by flash column chromatography (10% EtOAc-hexane), to afford methyl ether 13 (14.9 mg, 90%), colorless oil; $R_f = 0.2$ (20% EtOAc in hexane); $[\alpha]_D^{22}$ -95.6 (*c* 0.5, CHCl₃); HRMS (ES) calc. for C₂₄H₃₀NO₆ (M + NH₄)⁺ 428.2073 found 428.2052; ¹H-NMR (600 MHz, CDCl₃): δ_H 7.68 (d, J = 7.7 Hz, 1H), 7.51 (d, J =8.2 Hz, 1H), 7.37 (d, J = 8.8 Hz, 2H), 7.34 (dt, J = 7.3, 1.3 Hz, 8H), 7.29 (dt, J = 7.61.0 Hz, 9H), 6.89 (d, J = 8.6 Hz, 1H), 5.43 (d, J = 6.2 Hz, 1H), 4.81 (d, J = 3.6 Hz, 1H), 4.75 (d, J = 11.7 Hz, 1H), 4.74 (dd, J = 8.5, 6.2 Hz, 1H), 4.68 (d, J = 11.7 Hz, 1H), 3.80 (s, 3H), 3.67 (dd, J = 8.5, 3.6 Hz, 1H), 3.48 (s, 3H), 1.52 (s, 3H), 1.49 (s, 3H); ¹³C{¹H}-NMR (150 MHz, CDCl₃): δ_{C} 159.4, 155.9, 152.9, 129.9, 129.8, 126.4, 125.2, 123.4, 120.4, 113.8, 113.4, 111.7, 110.7, 81.6, 76.1, 71.7, 69.4, 66.9, 58.1, 55.3, 28.2, 25.8; IR v_{max}/cm⁻¹ 2956, 2924, 2854, 1514, 1249, 1109, 874.

(1R,2R,3S,4S)-3-Methoxy-4-((4-methoxybenzyl)oxy)-

1,2,3,4-tetrahydrodibenzo[b,d]furan-1,2-diol (14). А 46 solution of methyl ether 13 (8.0 mg, 0.020 mmol) in THF-47 $H_2O(9:1, 2 \text{ mL})$ was treated with TFA (10 μ L). The reaction 48 mixture was stirred (50°C) overnight, concentrated under 49 reduced pressure, and the product purified by flash column 50 chromatography (60% EtOAc in hexane), to yield cis-diol 14 51 (7.0 mg, 97%), colorless oil; $R_{\rm f} = 0.2$ (60% EtOAc-hexane); 52 $\left[\alpha\right]_{D}^{22}$ -139.0 (c 0.7, CHCl₃); HRMS (ES) calc. for C₂₁H₂₂O₆Na (M + Na)⁺ 393.1314 found 393.1302; ¹H-NMR 53 (600 MHz, CDCl₃): δ_H 7.68 (d, J = 7.7 Hz, 1H), 7.51 (d, J =54 8.2 Hz, 1H), 7.40 (d, J = 8.6 Hz, 2H), 7.34 (dt, J = 7.3, 1.3 55 Hz, 1H), 7.28 (dt, J = 7.6, 1.0 Hz, 1H), 6.90 (d, J = 8.6 Hz, 56 2H), 5.16 (d, J = 4.2 Hz, 1H), 4.84 (d, J = 3.9 Hz, 1H), 4.80 57 (d, J = 11.7 Hz, 1H), 4.74 (d, J = 11.7 Hz, 1H), 4.37 (dd, J = 11.7 Hz, 1H)58

10.2, 3.9 Hz, 1H), 3.81 (s, 3H), 3.78 (dd, J = 10.1, 3.9 Hz, 1H), 3.39 (s, 3H), 3.22 (br s, 2H); ¹³C {¹H}-NMR (150 MHz, CDCl₃): δ_C 159.5, 156.0, 152.4, 130.2, 129.6, 126.4, 125.2, 123.3, 120.1, 116.0, 113.9, 111.7, 78.1, 71.7, 68.3, 65.9, 62.5, 57.2, 55.3; IR v_{max} /cm⁻¹ 3998, 2955, 2917, 2849, 1576, 1382, 1222, 1033, 751.

(2S,3S,4S)-2-Hydroxy-3-methoxy-4-((4-methoxybenzyl)oxy)-3,4dihydrodibenzo[b,d]furan-1(2H)-one (15). TEMPO (1 mg) and OxoneTM (28 mg, 0.09 mmol) were sequentially added to a stirred solution of cis-diol 14 (15 mg, 0.041 mmol) and tetrabutylammonium bromide (2 mg) in dry CH₂Cl₂ (4 mL). After stirring the reaction mixture, overnight at room temperature, it was concentrated under reduced pressure, and the residue purified by flash column chromatography (50% EtOAc in hexane) to afford ketone 15 (12.8 mg, 86%), a pale-yellow oil; $R_{\rm f} = 0.3$ (50% EtOAc $[\alpha]_{D}^{22}$ -118.0 hexane): 1.0. in (cCHCl₃); HRMS (ES) calc. for $C_{21}H_{20}O_6Na (M + Na)^+ 391.1158$ found 391.1155; ¹H-NMR (600 MHz, CDCl₃): δ_H 8.08 (dm, 1H), 7.58 (m, 1H), 7.44 (td, J = 7.3, 1.5 Hz, 1H), 7.42-7.38 (m, 3H), 6.91 (d, J = 8.7 Hz, 2H), 5.04 (d, J = 3.8 Hz, 1H), 4.88 (dd, J =9.9, 1.0 Hz, 1H), 4.83 (d, J = 11.7 Hz, 1H), 4.80 (d, J = 11.7 Hz, 1H), 3.81 (s, 3H), 3.72 (dd, J = 9.9, 3.7 Hz, 1H), 3.59 (d, J = 1.2Hz, 1H), 3.54 (3H, s, 3H); ${}^{13}C{}^{1}H$ -NMR (150 MHz, CDCl₃); δ c191.5, 165.9, 159.4, 156.2, 130.1 (2), 129.1, 126.7, 125.3, 122.5 (2), 116.1, 114.0 (2), 111.9, 83.1, 73.6, 72.5, 67.4, 58.2, 55.4; IR v_{max} /cm⁻¹ 3844, 3681, 2948, 2880, 1652, 1410, 1128, 1044, 852.

(2S,3R,4S)-2,3-Dimethoxy-4-((4-methoxybenzyl)oxy)-3,4-Following the *dihydrodibenzo[b,d]furan-1(2H)-one* (16). procedure described for the synthesis of methyl ether 9, a mixture of ketone 15 (9.2 mg, 0.025 mmol), Ag₂O (8.0 mg) and MeI (10 μ L) in dry THF (2 mL) was heated in a sealed tube under argon atmosphere. The crude product obtained was purified by flash column chromatography (20% EtOAc in hexane) to give dimethoxy ketone **16** (7.3 mg, 76%), colorless oil; $R_{\rm f} = 0.2$ (30%) EtOAc in hexane); $[\alpha]_D^{22}$ -139.0 (*c* 0.7, CHCl₃); HRMS (ES) calc. for C₂₂H₂₃O₆ (M + H)⁺ 383.1495 found 383.1471; ¹H-NMR (600 MHz, CDCl₃): δ_H 8.09 (dm, J = 7.7 Hz, 1H), 7.55 (m, 1H), 7.43-7.35 (m, 4H), 6.92 (d, J = 8.7 Hz, 2H), 5.05 (d, J = 3.8 Hz, 1H), 4.88 (d, J = 11.8 Hz, 1H), 4.80 (d, J = 11.8 Hz, 1H), 4.31 (d, J =8.5 Hz, 1H), 3.98 (dd, J = 8.4, 3.7 Hz, 1H), 3.82 (s, 3H), 3.70 (s, 3H), 3.52 (s, 3H); ${}^{13}C{}^{1}H$ -NMR (150 MHz, CDCl₃): δ_C 190.9, 165.3, 159.6, 155.9, 130.0, 129.3, 126.2, 125.0, 123.1, 122.5, 116.4, 114.0, 111.7, 82.5, 81.9, 72.6, 68.7, 60.5, 59.0, 55.4; IR v max/cm⁻¹ 2947, 2880, 1739, 1684, 1484, 1281, 1041.

(2S,3R,4S)-4-Hydroxy-2,3-dimethoxy-3,4-

dihydrodibenzo[b,d]furan-1(2H)-one [(-)-Ribisin B].^{1,7}

Following the procedure described for the last step in the literature synthesis of (-)-ribisin A, a solution of dimethoxy ketone **16** (7.6 mg, 0.020 mmol) in CH₂Cl₂ and H₂O (4:1, 2 mL) was reacted with DDQ (5 mg). Purification of the crude product by flash column chromatography (25% EtOAc in hexane) furnished (-)-ribisin B (4.0 mg, 77%), as a colorless oil; $R_f = 0.2$ (30% EtOAc in hexane); $[\alpha]_D^{22}$ -25.8 (*c* 0.4, CHCl₃); HRMS (ES) calc. for C₁₄H₁₅O₅ (M + H)⁺ 263.0919 found 263.0912; ¹H-NMR (600 MHz, CDCl₃): δ_H 8.08 (dm, J = 7.6 Hz, 1H), 7.56 (dm, J = 7.7 Hz, 1H), 7.41-7.35 (m, 2H), 5.30 (dd, J = 7.1, 4.2 Hz, 1H), 4.15 (d, J = 6.7 Hz, 1H), 4.01 (dd, J = 6.7, 4.2 Hz, 1H), 3.62 (s, 3H), 3.61 (s, 3H), 3.05 (d, J = 7.1 Hz, 1H); ¹³C{¹H}-NMR (150 MHz, CDCl₃): δ_C 190.2, 165.2, 155.8, 126.1, 124.9, 123.1, 122.3, 115.3, 111.8, 81.9, 81.7, 63.8, 59.8, 59.5.

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Supporting Information

Copies of ¹H and ¹³C NMR spectra are presented for all intermediates.

REFERENCES

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57 58 59

- 1. Liu, Y. H.; Kubo, M.; Fukuyama, Y., Nerve Growth Factor-Potentiating Benzofuran Derivatives from the Medicinal Fungus *Phellinus ribis*. J. Nat. Prod. **2012**, 75, 2152-7.
- Liu, Y. H.; Wang, F. S., Structural characterization of an active polysaccharide from *Phellinus ribis*. *Carbohydr. Polym.* 2007, 70, 386-92.
- 3. Lee, I. K.; Lee, J. H.; Yun, B. S., Polychlorinated compounds with PPARgamma agonistic effect from the medicinal fungus *Phellinus ribis*. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4566-8.
- 4. Kubo, M.; Liu, Y. H.; Ishida, M.; Harada, K.; Fukuyama, Y., A New Spiroindene Pigment from the Medicinal Fungus Phellinus ribis. *Chem. Pharm. Bull.* **2014**, *62*, 122-4.
- Zhang, C. L.; Liu, J.; Du, Y. G., Total synthesis of ribisin A. *Tetrahedron Lett.* 2014, 55, 959-61.
- Lan, P.; Banwell, M. G.; Ward, J. S.; Willis, A. C., Chemoenzymatic Total Synthesis and Reassignment of the Absolute Configuration of Ribisin C. Org. Lett. 2014, 16, 228-31.
- Lan, P.; Banwell, M. G.; Willis, A. C., Chemoenzymatic Total Syntheses of Ribisins A, B, and D, Polyoxygenated Benzofuran Derivatives Displaying NGF-Potentiating Properties. *J. Org. Chem.* 2014, 79, 2829-42.
- Boyd, D. R.; Sharma, N. D.; Harrison, J. S.; Malone, J. F.; McRoberts, W. C.; Hamilton, J. T. G.; Harper, D. B., Enzymecatalysed synthesis and reactions of benzene oxide/oxepine derivatives of methyl benzoates. *Org. Biomol. Chem.* 2008, *6*, 1251-9.
- Min, K. L.; Kim, Y. H.; Kim, Y. W.; Jung, H. S.; Hah, Y. C., Characterization of a novel laccase produced by the wood-rotting fungus Phellinus ribis. *Arch. Biochem. Biophys.* 2001, 392, 279-86.
- Tang, F.; Lan, P.; Bolte, B.; Banwell, M. G.; Ward, J. S.; Willis, A. C., Total Synthesis of (+)-Viridianol, a Marine-Derived Sesquiterpene Embodying the Decahydrocyclobuta d indene Framework. *J. Org. Chem.* 2018, *83*, 14049-56.
- 11. Dlugosch, M.; Banwell, M. G., Synthesis of a Highly Functionalised and Homochiral 2-Iodocyclohexenone Related to the C-Ring of the Polycyclic, Indole Alkaloids Aspidophytine and Haplophytine. *Aust. J. Chem.* **2018**, *71*, 573-9.

- Dlugosch, M.; Ma, X. H.; Yang, S. X.; Banwell, M. G.; Ma, C. X.; Ward, J. S.; Carr, P., Syntheses of Structurally and Stereochemically Varied Forms of C₇N Aminocyclitol Derivatives from Enzymatically Derived and Homochiral *cis*-1,2-Dihydrocatechols. *Org. Lett.* 2018, 20, 7225-8.
- Baidilov, D.; Rycek, L.; Trant, J. F.; Froese, J.; Murphy, B.; Hudlicky, T., Chemoenzymatic Synthesis of Advanced Intermediates for Formal Total Syntheses of Tetrodotoxin. *Angew. Chem., Int. Ed.* 2018, *57*, 10994-8.
- Borra, S.; Kumar, M.; McNulty, J.; Baidilov, D.; Hudlicky, T., Chemoenzymatic Synthesis of the Antifungal Compound (-)-Pestynol by a Convergent, Sonogashira Construction of the Central Yne-Diene. *Eur. J. Org. Chem.* 2019, 77-9.
- Borra, S.; Lapinskaite, R.; Kempthorne, C.; Liscombe, D.; McNulty, J.; Hudlicky, T., Isolation, Synthesis, and Semisynthesis of Amaryllidaceae Constituents from Narcissus and Galanthus sp.: De Novo Total Synthesis of 2-*epi*-Narciclasine. *J. Nat. Prod.* 2018, *81*, 1451-9.
- Endoma-Arias, M. A. A.; Makarova, M.; Dela Paz, H. E.; Hudlicky, T., Chemoenzymatic Total Synthesis of (+)-Oxycodone from Phenethyl Acetate. *Synthesis-Stuttgart* 2019, *51*, 225-32.
- 17. Hudlicky, T., Benefits of Unconventional Methods in the Total Synthesis of Natural Products. *Acs Omega* **2018**, *3*, 17326-40.
- Lewis, S. E., Applications of biocatalytic arene *ipso*, *ortho cis*-dihydroxylation in synthesis. *Chem. Commun.* 2014, *50*, 2821-30.
- Lewis, S. E., Asymmetric Dearomatisation Under Enzymatic Conditions. In Asymmetric Dearomatisation Reactions, You, S. L., Ed. Wiley-VCH: Weinheim, 2016; pp 279-346.
- Boyd, D. R.; Sharma, N. D.; Carroll, J. G.; Loke, P. L.; O'Dowd, C. R.; Allen, C. C. R., Biphenyl dioxygenase-catalysed *cis*dihydroxylation of tricyclic azaarenes: chemoenzymatic synthesis of arene oxide metabolites and furoquinoline alkaloids. *RSC Adv.* 2013, *3*, 10944-55.
- Resnick, S. M.; Gibson, D. T., Regio- and stereospecific oxidation of fluorene, dibenzofuran, and dibenzothiophene by naphthalene dioxygenase from Pseudomonas sp strain NCIB 9816-4. *Appl. Environ. Microbiol.* 1996, 62, 4073-80.
- 22. Cerniglia, C. E.; Morgan, J. C.; Gibson, D. T., Bacterial and Fungal Oxidation of

		Dibenzofuran. Biochem. J. 1979, 180, 175-
1		85.
2	23.	Bianchi, D.; Bosetti, A.; Cidaria, D.;
3		Bernardi, A.; Gagliardi, I.; Damico, P.,
4		Oxidation of polycyclic aromatic
5		heterocycles by Pseudomonas fluorescens
6		11C1. Appl. Microbiol. Biotechnol. 199 7,
/	24	4/, 390-9. Poud D. P.: Sharma N. D.: Malana, I. E.:
8	24.	Boyu, D. K., Shahina, N. D., Maione, J. F., McInture P. B. Λ : Stevenson P. I : Allen
9		C C R Kwit M Gawronski I
10		Structure stereochemistry and synthesis of
11		enantionure cyclohexenone <i>cis</i> -diol
12		bacterial metabolites derived from phenols.
13		Org. Biomol. Chem. 2012, 10, 6217-29.
14	25.	Boyd, D. R.; Sharma, N. D.; Malone, J. F.;
15		McIntyre, P. B. A.; McRoberts, C.; Floyd,
10		S.; Allen, C. C. R.; Gohil, A.; Coles, S. J.;
17		Horton, P. N.; Stevenson, P. J., Toluene
18		Dioxygenase-Catalyzed Synthesis and
19		Reactions of cis-Diol Metabolites Derived
20		from 2-and 3-Methoxyphenols. J. Org.
21		Chem. 2015, 80, 3429-39.
22	26.	Boyd, D. R.; Sharma, N. D.; McIntyre, P.
23		B. A.; Stevenson, P. J.; McRoberts, W. C.;
24		Gohil, A.; Hoering, P.; Allen, C. C. R.,
25		Enzyme-Catalysed Synthesis of Cyclohex-
20		2-en-1-one <i>cls</i> -Diols from Substituted
27		Hudroxycyclobex 2 on 1 ones 4dy Synth
20		Catal 2017 350 4002 14
29	27	Boyd D R · Sharma N D · Brannigan I
30 21	27.	N · McGivern C I · Nockemann P ·
27		Stevenson P I McRoberts C Hoering
32		P.: Allen, C. C. R., <i>Cis</i> -Dihydroxylation of
34		Tricyclic Arenes and Heteroarenes
35		Catalyzed by Toluene Dioxygenase: A
36		Molecular Docking Study and
37		Experimental Validation. Adv. Synth. Catal.
38		2019 , <i>361</i> , 2526-37.
30	28.	Vanrheenen, V.; Kelly, R. C.; Cha, D. Y.,
40		Improved Catalytic OsO ₄ Oxidation of
40		Olefins to <i>cis</i> -1,2-Glycols using Tertiary
42		Amine Oxides as Oxidant. <i>Tetrahedron</i>
43	20	<i>Lett.</i> 1976 , 1973-6.
44	29.	Bolm, C.; Magnus, A. S.; Hildebrand, J. P.,
45		Catalytic synthesis of aldenydes and
46		TEMPO/Ovene Organia Lattery 2000 2
47		1173_5
48	30	Hudlicky T: Fan R I : Tsunoda T:
49	50.	Luna H · Andersen C · Price I D
50		Biocatalysis as a Rational Approach to
51		Enantiodivergent Synthesis of Highly
52		Oxygenated Compounds - (+)-Pinitol and (-
53)-Pinitol and Other Cyclitols. Isr. J. Chem.
54		1991 , <i>31</i> , 229-38.
55	31.	Heravi, M. M.; Zadsirjan, V.; Hamidi, H.;
56		Amiri, P. H. T., Total synthesis of natural
57		products containing benzofuran rings. RSC
58		Adv. 2017, 7, 24470-521.
59		
60		ACS Pa

32.	Oliveira, A.; Oliveira-Campos, A. M. F.;
	Raposo, M. M. M.; Griffiths, J.; Machado,
	A. E. H., Fries rearrangement of
	dibenzofuran-2-yl ethanoate under
	photochemical and Lewis-acid-catalysed
	conditions. Tetrahedron 2004, 60, 6145-54.