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A Divergent Approach to Indoles and Oxazoles from Enamides by Directing-Group-Controlled Cu-Catalyzed Intramolecular C-H Amination and Alkoxylation

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A directing-group-controlled, copper-catalyzed divergent approach to indoles and oxazoles from enamides has been developed. The picolinamide-derived enamides undergo the intramolecular aromatic C–H amination in the presence of a Cu(OPiv)₂ catalyst and an MnO₂ oxidant to form the corresponding indoles in good yields. On the other hand, simpler aryl- or alkyl-substituted enamides are converted to the 2,4,5-trisubstituted oxazole frameworks via vinylic C–H alkoxylation under

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identical conditions. The copper catalysis can provide uniquely divergent access to indole and oxazole

heteroaromatic cores of great importance in medicinal and material chemistry.

Introduction

Nitrogen-containing heteroaromatic rings are prevalent substructures in bioactive molecules, pharmaceutical targets, and functional materials. Synthetic chemists thus have developed many methodologies for the construction of above heterocycles. Among numerous reported procedures, the copper-mediated oxidative cyclization via C-H cleavage now receives significant attention because of its higher synthetic efficiency associated with atom and step economies.¹ Our research group also focused on the unique activity of less toxic, stable, and abundant copper salts and developed several Cu(II)-catalyzed intramolecular C-H amination reactions for the synthesis of carbazoles,^{2a} indolines,^{2b} As the next reaction design, we envisioned the C-H amination of and isoindolinones.^{2c} aryl-substituted enamides: the expected Cu(II)-catalyzed aromatic C–H activation occurs with the aid of picolinamide directing group to form the desired indoles in good yields (Scheme 1, right).^{3,4} Additionally, we have serendipitously found that similar aryl- and alkyl-substituted enamides undergo the vinylic C–H alkoxylation under identical conditions, delivering the corresponding 2.4.5-trisubstituted oxazoles selectively (Scheme 1, left).⁵ Thus, by the judicious choice of directing group, the single enamide skeleton is divergently converted under the same copper-catalyzed conditions to indole⁶ and oxazole⁷ of great interest in medicinal chemistry. Such a directing-group-controlled regiodivergent C–H activation was observed in some palladium- or rhodium-based catalysis⁸ but still remains underdeveloped under cooper-catalyzed conditions.^{2c} The detailed optimization studies and substrate scope are reported herein.

Scheme 1. Directing-Group-Controlled Divergent Approach to Indoles and Oxazoles via Cu-Catalyzed C–H Cleavage



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Results and Discussion

On the basis of our previous success of carbazole synthesis,^{2a} optimization studies commenced with N-picolinoyl enamide 1a-Py (0.10 mmol; Table 1), which was readily prepared by the CsOH-mediated hydroamidation of phenylacetylene with picolinamide.⁹ In an initial experiment, heating a DMF suspension of **1a-Py**, 30 mol % Cu(OAc)₂, and 2.0 equiv of MnO₂ under microwave irradiation (180 °C, 1 h) afforded the desired 2-phenylindole (2a) in 37% GC yield (entry 1). Same as shown in the previous work,^{2a} the corresponding *N*-picolinovl indole was not detected at all, and *NH*-indole **2a** was exclusively formed. After the aqueous workup, we successfully detected the picolinic acid in the aqueous phase by TOF-MS. Thus, the trace water in DMF solvent can promote the spontaneous hydrolysis in situ (vide infra). The addition of AcOH increased the yield to 63% (entry 2). We then tested several acetate-type Cu(II) salts, and bulkier Cu(OPiv)₂, Cu(OCOAd)₂, and Cu(eh)₂ showed better reactivity (entries 3–5). Subsequent screening of acidic additives with Cu(eh)₂ identified PivOH to be optimal (entries 6-8). Additional fine tuning revealed that a combination of Cu(OPiv)₂ and PivOH was best (entries 9 and 10), and finally 91% isolated yield was obtained at slightly higher temperature (entry 11). No reaction occurred in the absence of Cu(OPiv)₂, confirming the copper catalysis in the present transformation (entry 12). Additionally, we also tested other oxidants including AgOAc, K₂S₂O₈, and Mn(OAc)₃ under otherwise optimal conditions, but the indole 2a was observed in only <9% GC yield (data not shown).

Table 1. Optimization for Copper-Catalyzed Intramolecular C–H Amination of Enamide 1a-Py for Synthesis of Indole 2a^a



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| - | entry | Cu(II) | additive (equiv) | temp (°C) | yield $(\%)^b$ |
|-----------------|-------|------------------------|------------------|-----------|----------------|
| 2 <u>-</u> 3 | 1 | Cu(OAc) ₂ | none | 180 | 37 |
| 4 5 6 | 2 | Cu(OAc) ₂ | AcOH (1.0) | 180 | 63 |
| 7 8 9 | 3 | Cu(OPiv) ₂ | AcOH (1.0) | 180 | 69 |
| 10 11 | 4 | Cu(OCOAd) ₂ | AcOH (1.0) | 180 | 72 |
| 12 13 14 | 5 | Cu(eh) ₂ | AcOH (1.0) | 180 | 72 |
| 15 16 17 | 6 | Cu(eh) ₂ | EtCOOH (1.0) | 180 | 71 |
| 18 19 20 | 7 | Cu(eh) ₂ | PivOH (1.0) | 180 | 77 |
| 20 21 22 | 8 | Cu(eh) ₂ | AdCOOH (1.0) | 180 | 52 |
| 23 24 25 | 9 | Cu(eh) ₂ | PivOH (1.5) | 180 | 75 |
| 26 27 28 | 10 | Cu(OPiv) ₂ | PivOH (1.5) | 180 | 84 (84) |
| 29 30 | 11 | Cu(OPiv) ₂ | PivOH (1.5) | 200 | (91) |
| 31 32 33 | 12 | none | PivOH (1.5) | 200 | 0 |
| 34 | | | | | |

^a Reaction conditions: 1a-Pv (0.10 mmol), Cu(II) (0.030 mmol), MnO₂ (0.20 mmol), additive, DMF (0.60 mL), 1 h, N₂, microwave irradiation. ^b Estimated by GC method. Isolated yield is in parentheses. Ad = 1-adamanthyl, eh = 2-ethyl-1-hexanoate, 2-Py = 2-pyridyl.

Under conditions of entry 11 in Table 1, we performed the aromatic C-H amination of various enamides. The representative indole products are illustrated in Scheme 2. The copper catalysis was compatible with electronically diverse functions including methyl, tert-butyl, trifluoromethyl, and methoxy groups, and the corresponding substituted indoles 2b-d and 2g were obtained in synthetically useful yields. The aromatic C-Cl and C-Br moieties were also tolerated (2e and 2f), which can provide a useful synthetic handle for further manipulations based on the palladium-catalyzed

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cross-coupling chemistry.¹⁰ In the cases of *meta*-substituted enamides, the more sterically accessible C-H was preferably aminated (2h and 2i). The same regioselectivity trend was observed in the reaction of 2-naphtyl-substituted enamide (2). On the other hand, thienyl and benzothienyl substrates were somewhat unstable under the standard conditions with the MnO₂ terminal oxidant, but the corresponding thiophene-fused pyrrole rings 2k and 2l were obtained by using stoichiometric Cu(OPiv)₂ and conventional heating method with an oil bath. The stoichiometric conditions also increased the yield of electron-rich **2g**. In the stoichiometric reactions of theses electron-rich substrates, a small but significant amount of the corresponding N-picolinoylindoles was detected by GC and GCMS analysis. However, upon workup with ethylenediamine, the N-picolinoyl group was smoothly removed to from the NH indoles exclusively (see the Experimental Section for details). The copper-catalyzed C-H amination also accommodated the primary and secondary alkyl substituents to furnish the 2-alkylindoles in good yields (2m and 2n). The reaction could also be conducted on a preparative scale (1.0 mmol) (2a). Although similar intramolecular C–H aminations directed toward indole were already reported,⁴ they still relied on precious Pd catalysts or stoichiometric hypervalent I(III) oxidants. The present protocol is the first successful example of indole synthesis via copper-catalyzed C-H amination, to the best of our knowledge.

Scheme 2. Copper-Catalyzed Intramolecular C–H Amination of Various Enamides 1-Py for Synthesis of Indoles 2^{*a*}

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^a Reaction conditions: 1-Py (0.10 mmol), Cu(OPiv) (0.030 mmol), MnO₂ (0.20 mmol), PivOH (0.15 mmol), DMF (0.60 mL), 1 h, N₂, microwave irradiation. Isolated yields are shown. ^b 1.0 mmol scale.
^c With 2.0 equiv Cu(OPiv)₂ in the absence of MnO₂. At 150 °C with an oil bath for 4 h. ^d Regioisomeric ratio. The major isomer 2h was isolated in 58% yield. ^e Combined isolated yield of regioisomers. Regioisomeric ratio is in parenthesis. ^f Regioisomeric ratio. The major isomer 2j was isolated in 40% yield.

We next investigated the effect of substituent on enamide nitrogen and serendipitously found that the corresponding *N*-benzoyl enamide **1a-Ph** completely changed the reaction course even under otherwise identical conditions: the intramolecular vinylic C–H alkoxylation proceeded selectively to afford the 2,4,6-triphenyloxazole **3a-Ph** in 89% isolated yield (Scheme 3). Both an electron-donating methoxy and an electron-withdrawing trifluoromethyl groups were well tolerated (**3a-OMe** and **3a-CF**₃), and even *tert*-butyl-substituted enamide was converted to the trisubstituted oxazole in 85% yield (**3a-t-Bu**).

Also in this case, the reaction could be easily scaled up to 1.0 mmol (**3a-Ph**). On the other hand, even in the absence of $Cu(OPiv)_2$, a small but significant amount of **3a-Ph** (24% GC yield) was observed.

Prompted by the above intriguing product selectivity switching, we subsequently implemented the reaction of various benzoyl-substituted enamides **1-Ph**. The corresponding triaryloxazoles were uniformly formed with good to excellent yields from a wide range of *para-* and *meta-*substituted enamides **(3b-Ph-3i-Ph)**. The substrates bearing the sterically demanding naphthyl and heteroaromatic thienyl substituents also underwent the cyclization smoothly **(3j-Ph-3l-Ph)**. Additionally, the 4-alkyloxazoles were also readily constructed under identical conditions **(3m-Ph** and **3n-Ph)**.

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Scheme 3. Copper-Catalyzed Intramolecular C-H Alkoxylation of Enamides 1 for Synthesis of

Oxazoles 3^{*a*} 30 mol % Cu(OPiv)₂ R^2 MnO₂, PivOH DMF, 200 °C, 1 h, N₂ С HN \mathbb{R}^2 microwave irradiation N R³ Ο 1 3 R = H: 3a-Ph 89% yield (86% yield)^b (24% yield)^c റ R = OMe: 3a-OMe 87% yield R = CF₃: **3a-CF₃** 89% yield 3a-t-Bu 85% yield R R = Me: 3b-Ph 93% yield R = t-Bu: 3c-Ph 89% yield R = CF₃: **3d-Ph** 92% yield Ο R = Cl: 3e-Ph 78% yield OMe \sim R = Br: **3f-Ph** 53% yield 3g-Ph 93% yield R \cap \cap N 3k-Ph 89% yield R = Me: **3h-Ph** 90% yield 3j-Ph 94% yield R = OMe: 3i-Ph 96% yield \cap ·Bu 3I-Ph 96% yield 3n-Ph 80% yield 3m-Ph 49% yield

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^a Reaction conditions: 1 (0.10 mmol), Cu(OPiv) (0.030 mmol), MnO₂ (0.20 mmol), PivOH (0.15 mmol), DMF (0.60 mL), 1 h, N₂, microwave irradiation. Isolated yields are given. ^b 1.0 mmol scale.
^c GC yield without Cu(OPiv)₂ catalyst.

The related copper-mediated and -catalyzed cyclization of enamides were already developed by Stahl^{5a} and Buchwald.^{5b} However, only attempts to construct 2,5-disubstituted oxazoles were reported. Thus, we tested the reaction of **1a-Ph** under literature conditions (Scheme 4): the trisubstituted oxazole **3a-Ph** was formed in 76% GC yield under CuCl₂-mediated conditions developed by Stahl whereas Buchwald's procedure using a CuBr₂ catalyst gave **3a-Ph** in only <2% yield. Under microwave-modified conditions, the CuCl₂/NMI system provided **3a-Ph** in 90% GC yield, but a stoichiometric amount of CuCl₂ was still necessary. These outcomes demonstrate the synthetic advantage of our newly developed Cu(OPiv)₂/MnO₂ catalyst system.

Scheme 4. Attempts to Apply Reported Cu-Promoted Conditions for Synthesis of Oxazole 3a-Ph



To gain some mechanistic insight, several deuterium-labeling experiments with $1a-Py-d_{10}$ and $1a-Ph-d_1$ were carried out (Scheme 5, see the Supporting Information for more detail). All deuterium-labeling

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experiments were performed under the conventional heating conditions with an oil bath, because under microwave-assisted conditions the reaction proceeded in the course of the preheating time up to 200 °C, and the conversion at an early stage was difficult to monitor. We initially checked the irreversibility of C–H cleavage. At an early stage of reaction, no H/D exchange occurred in both indole and oxazole formations (eqs 1 and 2). Thus, the kinetic isotope effect (KIE) value of each reaction was investigated: 4.2 and 1.1 were obtained for **1a-Py** and **1a-Ph**, respectively (eqs 3 and 4).

Scheme 5. Deuterium-Labeling Experiments



On the basis of the above results and literature information, we are tempted to assume the reaction mechanism of **1a-Py** and **1a-Ph** as follows (Scheme 6). For the formation of indole **2a** (cycle A), the starting $Cu(OPiv)_2$ initially undergoes the ligand exchange with **1a-Py** generates the *N*,*N*-bidentately

coordinated Cu complex **4**.¹¹ Subsequent rate-limiting C–H cleavage¹² (**4** \rightarrow **5**) is followed by disproportionation with additional Cu(OPiv)₂ to afford a Cu(III) species **6**.¹³ The indole framework **7** is then constructed by reductive elimination from **6**. Spontaneous hydrolysis of **7** in situ gives the observed *NH*-indole **2a**. On the other hand, the formed CuOPiv is reoxidized by MnO₂¹⁴ into Cu(OPiv)₂ to complete the catalytic cycle A. On the other hand, the oxazole **3a-Ph** formation process (cycle B) includes (1) ligand exchange with **1a-Ph** (Cu(OPiv)₂ \rightarrow **8**), (2) disproportionation-induced oxidation into Cu(III) species **9**, and (3) homolysis of the resulting N–Cu(III) bond along with the regeneration of the staring Cu(OPiv)₂.^{2c,15} The formed amidyl radical **10** undergoes cyclization (**10** \rightarrow **11**) followed by the single electron transfer (SET) oxidation (**11** \rightarrow **12**) and deprotonation to furnish the oxazole **3a-Ph**. The minor background reaction observed in Scheme 3 suggests less effective but significant oxidation aptitude of MnO₂ for the generation of radical species **11**. However, radical inhibitors, TEMPO and galvinoxyl, did not completely shut down oxazole formation as well as indole formation (Scheme 7). Thus, the initial single electron transfer (SET) oxidation, which was originally proposed by Stahl^{5a} is also plausible, and details still remain to be elucidated.¹⁶

Scheme 6. Plausible Mechanism



Scheme 7. Effects of Radical Inhibitors



Conclusion

We have developed a copper-catalyzed oxidative cyclization approach to indoles and oxazoles via C– H amination and alkoxylation, respectively. The directing-group-controlled divergent mechanisms, namely Cu(I)/(II)/(III) organometallic pathway and amidyl radical pathway, are operated, and indoles and oxazoles are selectively obtained from enamides of same skeleton. Such a directing-group-controllable strategy enables diversity-oriented synthesis of *N*-heterocycles through multisite-selective C–H functionalization. Further development of related regioselective and/or -divergent Cu catalysts is ongoing and will be reported in due course.

Experimental Section

Instrumentation and Chemicals ¹H, ¹³C{¹H}, and ¹⁹F{¹H} NMR spectra were recorded at 400 MHz, 100 MHz, and 376 MHz, respectively, for CDCl₃ or DMSO-*d*₆ solutions. HRMS data were obtained by APCI using TOF. GC analysis was carried out using a silicon OV-17 column (2.6 mm i.d. x 1.5 m) or a CBP-1 capillary column (0.5 mm i.d. x 25 m). TLC analyses were performed on commercial glass plates bearing a 0.25 mm layer of Merck silica gel $60F_{254}$. Silica gel (Wakosil C-200) was used for column chromatography. Gel permeation chromatography (GPC) was performed by LC-6AD (pump, SHIMADZU, 3.5 mL/min CHCl₃) and SPD-20A (UV detector, SHIMADZU, 254

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nm) with two in-line GPC H-2001 (20 x 500 mm, particle size: 15 µm) and H-2002 columns (20 x 500 mm, particle size: 15 µm) (preparative columns, Shodex, CHCl₃ eluent) or by LC-20AR (pump, SHIMADZU, 7.5 mL/min EtOAc) and SPD-20A (UV detector, SHIMADZU, 254 nm) with two in-line YMC-GPC T2000 (20 x 600 mm, particle size: 10 µm) (preparative columns, YMC, EtOAc eluent). Microwave irradiation was conducted with Initiator⁺ (Biotage), and the reaction temperature was measured by an internal probe. Unless otherwise noted, materials obtained from commercial suppliers were used as received. Cu(OPiv)₂ was prepared according to the literature.¹⁷ DMF was dried on a Glass Contour Solvent dispensing system (Nikko Hansen & Co., Ltd.) prior to use. Enamides 1, except for 1g-Py and 1g-Ph, were synthesized by the CsOH-mediated hydroamidation of corresponding internal alkynes and benzamides.⁹ Conventional condensation was conducted for the preparation of 1g-Py and 1g-Ph.¹⁸ All reactions were carried out under nitrogen atmosphere unless otherwise noted.

Preparation of 1a-Ph-*d*₁**.** In a glovebox filled with nitrogen, diphenylacetylene (67 mg, 0.38 mmol), benzamide (45 mg, 0.38 mmol), and CsOH•OH₂ (63 mg, 0.38 mmol) were placed in a 2.0 mL microwave vessel. The vessel was sealed with a cap and then taken out of the glovebox. Dimethyl sulfoxide-*d*₆ (DMSO-*d*₆, 1.5 mL) was sequentially injected via a syringe. The mixture was stirred for 24 h at 120 °C. The resulting mixture was then quenched with water. The mixture was extracted with ethyl acetate three times, and the combined organic layer was dried over anhydrous Na₂SO₄. After concentration under reduced pressure, silica gel column purification with hexane/ethyl acetate (3/1, v/v) afforded (*Z*)-*N*-(2-deuterio-1,2-diphenylvinyl)benzamide (**1a-Ph-***d*₁; 77 mg, 0.26 mmol, 98% D) in 68% yield.

Typical Procedure for Synthesis of Indoles 2 under Conditions Catalytic in Copper. Thesynthesis of 2a is representative (Scheme 2). $Cu(OPiv)_2$ (8.0 mg, 0.030 mmol),(Z)-N-(1,2-diphenylvinyl)picolinamide (1a-Py; 30 mg, 0.10 mmol), pivalic acid (15 mg, 0.15 mmol),

and MnO_2 (17 mg, 0.20 mmol) were placed in a microwave vessel. The vessel was flushed with nitrogen and then sealed with a cap. DMF (0.60 mL) was sequentially injected via a syringe. The mixture was irradiated under microwave reactor conditions at 200 °C for 1 h. The resulting mixture was then quenched with water, and a small amount of ethylenediamine was added to dissolve the residual copper salts in the aqueous phase. The mixture was extracted with ethyl acetate three times, and the combined organic layer was dried over anhydrous Na₂SO₄. After concentration under reduced pressure, silica gel column purification with hexane/ethyl acetate (10/1, v/v) afforded 2-phenylindole (**2a**; 18 mg, 0.091 mmol) in 91% yield.

2-Phenyl-1*H***-indole** $(2a)^{19}$ white solid; m.p. 187.6-189.3 °C; 18 mg (91%); ¹H NMR (400 MHz, CDCl₃) δ 6.85 (dd, J = 1.1, 1.9 Hz, 1H), 7.15 (ddd, J = 1.1, 7.0, 8.0 Hz, 1H), 7.22 (ddd, J = 1.1, 7.1, 8.2 Hz, 1H), 7.34 (tt, J = 1.1, 7.4 Hz, 1H), 7.40-7.48 (m, 3H), 7.65-7.69 (m, 3H), 8.33 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 100.0, 110.9, 120.3, 120.7, 122.4, 125.2, 127.7, 129.1, 129.3, 132.4, 136.8, 137.9; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₄H₁₂N: 194.0964, found: 194.0960.

6-Methyl-2-(*p*-tolyl)-1*H*-indole (2b) white solid; m.p. 187.6-189.3 °C; 13 mg (59%); m.p. 206.8-208.2 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.39 (s, 3H), 2.47 (s, 3H), 6.73 (dd, J = 0.8, 2.1 Hz, 1H), 6.94 (dd, J = 0.8, 8.0 Hz, 1H), 7.18 (d, J = 0.6 Hz, 1H), 7.22-7.25 (m, 2H), 7.49 (d, J = 8.0 Hz, 1H), 7.50-7.55 (m, 2H), 8.17 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.2, 21.8, 99.1, 110.8, 120.1, 122.0, 124.9, 127.1, 129.7, 129.8, 132.0, 137.1, 137.38, 137.40; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₆H₁₆N: 222.1277, found: 222.1278.

 1H); ¹³C NMR (100 MHz, CDCl₃) δ 31.3, 31.8, 34.7, 34.8, 99.2, 107.1, 118.5, 120.0, 124.8, 126.0, 127.0, 129.9, 136.9, 137.7, 145.7, 150.6; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₂H₂₈N: 306.2216, found: 306.2218.

6-(Trifluoromethyl)-2-(4-(trifluoromethyl)phenyl)-1*H*-indole (2d) white solid; m.p. 115.9-116.8 °C; 30 mg (91%); ¹H NMR (400 MHz, CDCl₃) δ 6.95 (d, J = 1.4 Hz, 1H), 7.38 (dd, J = 1.0, 8.3 Hz, 1H), 7.69-7.77 (m, 6H), 8.58 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 101.6, 108.7 (q, J = 4.3 Hz), 117.3 (q, J = 3.7 Hz), 121.4, 124.0 (q, J = 273.8 Hz), 125.0 (q, J = 271.1 Hz), 125.1 (q, J = 31.7 Hz), 125.5, 126.2 (q, J = 3.4 Hz), 130.2 (q, J = 32.7 Hz), 131.3, 134.9, 135.9, 138.8; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.6, -60.8; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₆H₁₀F₆N: 330.0712, found: 330.0709.

6-Chloro-2-(4-chlorophenyl)-1*H***-indole (2e)** white solid; m.p. 166.2-167.2 °C; 19 mg (74%); ¹H NMR (400 MHz, CDCl₃) δ 6.77 (dd, J = 0.8, 2.2 Hz, 1H), 7.10 (dd, J = 1.8, 8.4 Hz, 1H), 7.38 (t, J = 0.8 Hz, 1H), 7.38-7.43 (m, 2H), 7.52 (d, J = 8.4 Hz, 1H), 7.55-7.58 (m, 2H), 8.26 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 100.4, 110.9, 121.3, 121.6, 126.3, 127.7, 128.4, 129.3, 130.4, 133.8, 137.2, 137.4; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₄H₁₀Cl₂N: 262.0185, found: 262.0184.

6-Bromo-2-(4-bromophenyl)-1*H***-indole (2f)** white solid; m.p. 187.6-188.3 °C; 18 mg (51%); ¹H NMR (400 MHz, CDCl₃) δ 6.78 (d, *J* = 1.4 Hz, 1H), 7.23 (dd, *J* = 1.4, 8.4 Hz, 1H), 7.49 (t, *J* = 8.7 Hz, 3H), 7.54-7.58 (m, 3H), 8.27 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 100.5, 113.9, 116.0, 121.9, 123.9, 126.4, 126.6, 128.0, 130.8, 132.3, 137.3, 137.6; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₄H₁₀Br₂N: 349.9175, found: 349.9179.

2-(4-Methoxyphenyl)-1*H***-indole (2g)** white solid; m.p. 228.5-229.7 °C; 14 mg (62%); ¹H NMR (400 MHz, CDCl₃) δ 3.86 (s, 3H), 6.72 (dd, *J* = 0.8, 2.0 Hz, 1H), 6.96-7.00 (m, 2H), 7.09-7.19 (m, 2H),

7.37-7.39 (m, 1H), 7.58-7.61 (m, 3H), 8.25 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 55.4, 98.9, 110.7, 114.5, 120.2, 120.4, 121.9, 125.2, 126.5, 129.4, 136.7, 138.0, 159.4; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₅H₁₄NO: 224.1070, found: 224.1072.

5-Methyl-2-(*m***-tolyl)-1***H***-indole (2h)** white solid; m.p. 160.9-161.6 °C; 19 mg (58%); ¹H NMR (400 MHz, CDCl₃) δ 2.40 (s, 3H), 2.44 (s, 3H), 6.72 (dd, *J* = 0.6, 2.0 Hz, 1H), 7.00 (dd, *J* = 1.3, 8.2 Hz, 1H), 7.11 (d, *J* = 7.5 Hz, 1H), 7. 24-7.32 (m, 2H), 7.40 (d, *J* = 0.6 Hz, 1H), 7.43 (d, *J* = 8.0 Hz, 1H), 7.45 (s, 1H), 8.20 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.5, 21.6, 99.4, 110.5, 120.3, 122.2, 123.9, 125.8, 128.4, 128.9, 129.4, 129.6, 132.4, 135.1, 138.1, 138.6; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₆H₁₆N: 222.1277, found: 222.1275.

Α 10:1 mixture of 5-methoxy-2-(3-methoxyphenyl)-1*H*-indole (2i) and 7-methoxy-2-(3-methoxyphenyl)-1*H*-indole (2i') white solid; m.p. 132.2-133.9 °C; 19 mg (75%); ¹H NMR (400 MHz, CDCl₃) for mixture δ 3.86 (s, 0.91 × 3H for 2i), 3.87 (s, 0.91 × 3H for 2i), 3.88 (s, $0.090 \times 3H$ for **2i'**), 3.98 (s. $0.090 \times 3H$ for **2i'**), 6.75 (dd. J = 0.7, 2.0 Hz, $0.91 \times 1H$ for **2i**), 6.80 (d. J =2.3 Hz, $0.090 \times 1H$ for **2i'**), 6.84-6.88 (m, 0.91 × 2H for **2i**, 0.090 × 2H for **2i'**), 7.03 (t, J = 7.9 Hz, $0.090 \times 1H$ for **2i'**), 7.07-7.08 (m, 0.91 × 1H for **2i**, 0.090 × 1H for **2i'**), 7.17 (t, J = 2.0 Hz, 0.91 × 1H for **2i**), 7.21-7.24 (m, 0.91×1 H for **2i**, 0.090×1 H for **2i**'), 7.26-7.28 (m, 0.91×1 H for **2i**, 0.090×1 H for 2i'), 7.32-7.36 (m, 0.91 × 1H for 2i, 0.090 × 1H for 2i'), 8.23 (s, 0.91 × 1H for 2i), 8.54 (s, 0.090 × 1H for 2i'); ¹³C NMR (100 MHz, CDCl₃) for 2i δ 55.4, 55.8, 100.1, 102.2, 110.9, 111.7, 112.7, 113.0, 117.6, 129.6, 130.1, 132.0, 133.8, 138.5, 154.5, 160.1; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₆H₁₆NO₂: 254.1176, found: 254.1174.

2-(Naphthalen-2-yl)-1*H***-benzo[***f***]indole (2j) white solid; m.p. 251.3-253.0 °C; 12 mg (40%); ¹H NMR (400 MHz, DMSO-***d***₆) δ 7.25-7.35 (m, 3H), 7.54-7.62 (m, 2H), 7.90-8.05 (m, 6H), 8.13-8.15 (m,**

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2H), 8.52 (s, 1H), 11.73 (s, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 99.2, 106.5, 117.5, 122.7, 123.7, 124.2, 124.5, 126.9, 127.3, 127.7, 128.2, 128.3, 128.5, 128.9, 129.0, 129.7, 130.6, 131.08, 131.14, 133.6, 138.6, 142.3; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₂H₁₆N: 294.1277; found: 294.1275.

2-Butyl-1*H***-indole (2m)** yellow oil; 13 mg (66%); ¹H NMR (400 MHz, CDCl₃) δ 0.95 (t, *J* = 7.3 Hz, 3H), 1.41 (sext, *J* = 7.5 Hz, 2H), 1.66-1.73 (m, 2H), 2.74 (t, *J* = 7.6 Hz, 2H), 6.23 (dd, *J* = 0.9, 2.0 Hz, 1H), 7.04-7.12 (m, 2H), 7.27-7.29 (m, 1H), 7.51-7.53 (m, 1H), 7.82 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.9, 22.4, 28.0, 31.3, 99.5, 110.3, 119.6, 119.7, 120.9, 128.9, 135.8, 140.0; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₂H₁₆N: 174.1277, found: 174.1275.

2-Cyclohexyl-1*H***-indole (2n)** white solid; m.p. 104.2-105.6 °C; 16 mg (79%); ¹H NMR (400 MHz, CDCl₃) δ 1.23-1.51 (m, 5H), 1.74-1.77 (m, 1H), 1.83-1,87 (m, 2H), 2.07-2.10 (m, 2H), 2.68-2.75 (m, 1H), 6.23 (d, *J* = 1.0 Hz, 1H), 7.04-7.13 (m, 2H), 7.30 (d, *J* = 8.0 Hz, 1H), 7.53 (d, *J* = 7.6 Hz, 1H), 7.90 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 26.1, 26.3, 33.0, 37.3, 97.5, 110.3, 119.6, 119.9, 120.9, 128.6, 135.5, 145.1; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₄H₁₈N: 200.1434, found: 200.1434.

Typical Procedure for Synthesis of Indoles 2 under Conditions Stoichiometric in Copper. The synthesis of **2k** is representative (Scheme 2). $Cu(OPiv)_2$ (53 mg, 0.20 mmol), (*Z*)-*N*-(1,2-di(thiophen-2-yl)vinyl)picolinamide (**1k-Py**; 31 mg, 0.10 mmol), and pivalic acid (15 mg, 0.15 mmol) were placed in a 2.0 mL microwave vessel. The vessel was flushed with nitrogen and then sealed with a cap. DMF (0.60 mL) was sequentially injected via a syringe. The mixture was heated at 150 °C for 4 h with an oil bath. The resulting mixture was then quenched with water, and a small amount of ethylenediamine was added to dissolve the residual copper salts in the aqueous phase. The mixture was extracted with ethyl acetate three times, and the combined organic layer was dried over anhydrous Na₂SO₄. After concentration under reduced pressure, silica gel column purification with

hexane/ethyl acetate (10/1, v/v) afforded 5-(thiophen-2-yl)-4*H*-thieno[3,2-*b*]pyrrole (**2k**; 8.2 mg, 0.41 mmol) in 41% yield.

5-(Thiophen-2-yl)-4*H***-thieno[3,2-***b***]pyrrole (2k) white solid; m.p. 110.6-111.9 °C; 8.4 mg (41%); ¹H NMR (400 MHz, CDCl₃) \delta 6.66 (d,** *J* **= 1.8 Hz, 1H), 6.95 (dd,** *J* **= 0.6, 5.2 Hz, 1H), 7.05 (dd,** *J* **= 3.6, 5.0 Hz, 1H), 7.09 (d,** *J* **= 5.2 Hz, 1H), 7.12 (dd,** *J* **= 1.1, 3.6 Hz, 1H), 7.20 (dd,** *J* **= 1.1, 5.1 Hz, 1H), 8.32 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) \delta 99.7, 110.9, 121.6, 123.4, 124.2, 125.5, 127.8, 131.6, 136.1, 139.0; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₀H₈NS₂: 206.0093, found: 206.0095.**

2-(Benzo[*b***]thiophen-2-yl)-1***H***-benzo[4,5]thieno[3,2-***b***]pyrrole (2l) brown solid; m.p. 196.2-197.5 ^oC; 5.8 mg (19%); ¹H NMR (400 MHz, CDCl₃) δ 6.84 (d,** *J* **= 1.9 Hz, 1H), 7.26-7.40 (m, 5H), 7.74-7.82 (m, 4H), 8.84 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 102.1, 117.4, 118.4, 122.2, 123.3, 123.4, 124.2, 124.30, 124.33, 124.6, 124.8, 126.4, 131.5, 133.6, 135.4, 138.5, 140.4, 142.3; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₈H₁₂NS₂: 306.0406, found: 306.0394.**

Typical Procedure for Synthesis of Oxazoles 3. The synthesis of **3a-Ph** is representative (Scheme 3). $Cu(OPiv)_2$ (8.0 mg, 0.030 mmol), (*Z*)-*N*-(1,2-diphenylvinyl)benzamide (**1a-Ph**; 30 mg, 0.10 mmol), pivalic acid (15 mg, 0.15 mmol), and MnO₂ (17 mg, 0.2 mmol) were placed in a microwave vessel. The vessel was flushed with nitrogen and then sealed with a cap. DMF (0.60 mL) was sequentially injected via a syringe. The mixture was irradiated under microwave reactor conditions at 200 °C for 1 h. The resulting mixture was then quenched with water, and a small amount of ethylenediamine was added to dissolve the residual copper salts in the aqueous phase. The mixture was extracted with ethyl acetate three times, and the combined organic layer was dried over anhydrous Na₂SO₄. After concentration under reduced pressure, silica gel column purification with hexane/ethyl acetate (20/1, v/v) afforded 2,4,5-triphenyloxazole (**3a-Ph**; 27 mg, 0.089 mmol) in 89% yield.

2,4,5-Triphenyloxazole (**3a-Ph**)²⁰ white solid; m.p. 112.9-113.5 °C; 260 mg (86%); 27 mg (89%) on a 0.10 mmol scale; ¹H NMR (400 MHz, CDCl₃) δ 7.32-7.42 (m, 6H), 7.45-7.51 (m, 3H), 7.66-7.69 (m, 2H), 7.71-7.74 (m, 2H), 8.15-8.17 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 126.5, 126.6, 127.4, 128.2, 128.3, 128.57, 128.65, 128.7, 128.8, 129.0, 130.4, 132.6, 136.8, 145.6, 160.2; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₁H₁₆NO: 298.1226, found: 298.1228.

2-(4-Methoxyphenyl)-4,5-diphenyloxazole (**3a-OMe**) white solid; m.p. 110.9-111.9 °C; 29 mg (87%); ¹H NMR (400 MHz, CDCl₃) δ 3.86 (s, 3H), 6.98-7.00 (m, 2H), 7.32-7.42 (m, 6H), 7.65-7.67 (m, 2H), 7.71-7.73 (m, 2H), 8.07-8.10 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 55.4, 114.2, 120.2, 126.5, 128.1, 128.17, 128.19, 128.4, 128.6, 128.7, 129.2, 132.7, 136.6, 145.0, 160.3, 161.4; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₂H₁₈NO₂: 328.1332, found: 328.1331.

4,5-Diphenyl-2-(4-(trifluoromethyl)phenyl)oxazole (**3a-CF**₃) white solid; m.p. 137.8-139.9 °C; 33 mg (89%); ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.44 (m, 6H), 7.66-7.75 (m, 6H), 8.30 (d, *J* = 8.1 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 123.9 (q, *J* = 272.4 Hz), 125.8 (q, *J* = 3.9 Hz), 126.6, 126.7, 128.1, 128.5, 128.6, 128.7, 128.8, 128.9, 130.5, 131.9 (q, *J* = 32.7 Hz), 132.2, 137.2, 146.4, 158.7; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.8; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₂H₁₅F₃NO: 366.1100, found: 366.1104.

2-(*tert***-Butyl)-4,5-diphenyloxazole (3a-***t***-Bu) white solid; m.p. 58.9-60.3 °C; 24 mg (85%); ¹H NMR (400 MHz, CDCl₃) δ 1.48 (s, 9H), 7.27-7.38 (m, 6H), 7.56-7.59 (m, 2H), 7.63-7.66 (m, 2H); ¹³C NMR**

(100 MHz, CDCl₃) δ 28.7, 33.8, 126.3, 127.9, 128.1, 128.2, 128.55, 128.59, 129.4, 132.9, 134.9, 144.7, 169.9; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₉H₂₀NO: 278.1539, found: 278.1537.

2-Phenyl-4,5-di-*p*-tolyloxazole (**3b-Ph**) white solid; m.p. 131.3-132.0 °C; 30 mg (93%); ¹H NMR (400 MHz, CDCl₃) δ 2.39 (s, 3H), 2.39 (s, 3H), 7.18-7.22 (m, 4H), 7.44-7.50 (m, 3H), 7.57 (d, *J* = 8.2 Hz, 2H), 7.62 (d, *J* = 8.2 Hz, 2H), 8.13-8.16 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.38, 21.42, 126.3, 126.4, 126.5, 127.5, 128.0, 128.7, 129.3, 129.4, 129.8, 130.2, 136.3, 137.9, 138.5, 145.5, 159.8; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₃H₂₀NO: 326.1539, found: 326.1538.

4,5-Bis(4-(*tert***-butyl)phenyl)-2-phenyloxazole (3c-Ph)** white solid; m.p. 154.2-155.6 °C; 37 mg (89%); ¹H NMR (400 MHz, CDCl₃) δ 1.35 (s, 9H), 1.36 (s, 9H), 7.41-7.49 (m, 7H), 7.65-7.70 (m, 4H), 8.13-8.16 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 31.25, 31.34, 34.7, 34.8, 125.5, 125.6, 126.2, 126.3, 126.4, 127.6, 127.7, 128.7, 129.8, 130.2, 136.3, 145.4, 151.1, 151.6, 159.8; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₉H₃₂NO: 410.2478, found: 410.2474.

2-Phenyl-4,5-bis(4-(trifluoromethyl)phenyl)oxazole (**3d-Ph**) white solid; m.p. 165.3-166.0 °C; 40 mg (92%); ¹H NMR (400 MHz, CDCl₃) δ 7.50-7.53 (m, 3H), 7.66-7.70 (m, 4H), 7.77 (d, *J* = 8.0 Hz, 2H), 7.84 (d, *J* = 8.0 Hz, 2H), 8.13-8.18 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 123.8 (q, *J* = 270.5 Hz), 124.0 (q, *J* = 270.5 Hz), 125.8 (q, *J* = 3.6 Hz), 125.9 (q, *J* = 3.9 Hz), 126.66, 126.74, 128.4, 128.9, 130.0, 130.6 (q, *J* = 33.0 Hz), 130.7 (q, *J* = 32.3 Hz), 131.87, 131.89, 135.7, 137.0, 144.9, 161.2; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.8, -62.7; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₃H₁₄F₆NO: 434.0974, found: 434.0976.

4,5-Bis(4-chlorophenyl)-2-phenyloxazole (3e-Ph) white solid; m.p. 145.9-147.0 °C; 29 mg (78%); ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.40 (m, 4H), 7.47-7.50 (m, 3H), 7.57-7.59 (m, 2H), 7.63-7.66 (m, 2H), 8.11-8.14 (m, 2H); ¹³C NMR (100 MHz; CDCl₃) δ 126.5, 127.0, 127.2, 127.8, 128.9, 129.0, 129.1, 129.4, 130.7, 130.8, 134.3, 134.7, 136.1, 144.7, 160.5; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₁H₁₄Cl₂NO: 366.0447, found: 366.0446.

4.5-Bis(4-bromophenyl)-2-phenyloxazole (**3f-Ph**) white solid; m.p. 165.9-167.9 °C; 17 mg (53%); ¹H NMR (400 MHz, CDCl₃) δ 7.48-7.60 (m. 11H), 8.12-8.14 (m. 2H); ¹³C NMR (100 MHz, CDCl₃)) δ 122.5, 122.9, 126.5, 127.0, 127.6, 128.0, 128.9, 129.6, 130.70, 130.71, 131.2, 132.0, 132.1, 136.2, 160.6; HRMS (APCI) m/z ($[M+H]^+$) calcd for C₂₁H₁₄Br₂NO: 453.9437, found: 453.9423.

4-(4-Methoxyphenyl)-2,5-diphenyloxazole (3g-Ph) white solid; m.p. 127.2-127.9 °C; 31 mg (93%); ¹H NMR (400 MHz, CDCl₃) δ 3.85 (s, 3H), 6.93-6.96 (m, 2H), 7.30-7.41 (m, 3H), 7.45-7.51 (m, 3H), 7.63-7.69 (m, 4H), 8.13-8.16 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 55.3, 114.1, 125.0, 126.39, 126.44, 127.5, 128.4, 128.7, 128.8, 129.2, 129.5, 130.3, 136.6, 144.9, 159.6, 160.0; HRMS (APCI) m/z $([M+H]^+)$ calcd for C₂₂H₁₈NO₂: 328.1132, found: 328.1131.

2-Phenyl-4,5-di-*m*-tolyloxazole (3h-Ph) colorless liquid; 29 mg (90%); ¹H NMR (400 MHz, CDCl₃) δ 2.36 (s, 3H), 2.38 (s, 3H), 7.14-7.18 (m, 2H), 7.23-7.29 (m, 2H), 7.45-7.50 (m, 5H), 7.53 (s, 1H), 7.60 (s, 1H), 8.14-8.17 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 21.47, 21.49, 123.8, 125.2, 126.5, 127.1, 127.5, 128.4, 128.6, 128.77, 128.82, 129.96, 129.99, 129.3, 130.3, 132.5, 136.8, 138.3, 138.4, 145.7, 160.0; HRMS (APCI) m/z ($[M+H]^+$) calcd for C₂₃H₂₀NO; 326.1539, found: 326.1537.

4,5-Bis(3-methoxyphenyl)-2-phenyloxazole (**3i-Ph**) white solid; m.p. 73.9-75.2 °C; 34 mg (96%); ¹H NMR (400 MHz, CDCl₃) δ 3.78 (s, 3H), 3.82 (s, 3H), 6.88-6.93 (m, 2H), 7.23-7.24 (m, 1H), 7.27-7.33 (m, 5H), 7.46-7.51 (m, 3H), 8.15-8.17 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 55.29, 55.33, 111.8, 113.3, 114.6, 119.1, 120.7, 126.5, 127.3, 128.8, 129.6, 129.8, 130.1, 130.41, 130.43, 133.8, 136.9, 145.5, 159.7, 159.8, 160.0; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₃H₂₀NO₃: 358.1438, found: 358.1436.

4,5-Di(naphthalen-2-yl)-2-phenyloxazole (**3j-Ph**) white solid; m.p. 180.3-181.3 °C; 37 mg (94%); ¹H NMR (400 MHz, CDCl₃) δ 7.48-7.55 (m, 7H), 7.75 (dd, J = 1.7, 8.6 Hz, 1H), 7.80-7.88 (m, 7H), 8.23-8.26 (m, 3H), 8.33 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 124.2, 125.89, 125.92, 126.31, 126.34, 126.4, 126.6, 126.68, 126.74, 127.4, 127.7, 127.8, 128.2, 128.36, 128.38, 128.39, 128.85, 128.88, 130.0, 130.5, 133.17, 133.21, 133.3, 133.5, 137.2, 146.0, 160.5; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₉H₂₀NO: 398.1539, found: 398.1537.

2-Phenyl-4,5-di(thiophen-2-yl)oxazole (**3k-Ph**) white solid; m.p. 102.9-104.2 °C; 28 mg (89%); ¹H NMR (400 MHz, CDCl₃) δ 7.10 (dd, J = 3.6, 5.2 Hz, 1H), 7.13 (dd, J = 3.6, 5.1 H, 1H), 7.39 (dd, J = 1.0, 5.1 Hz, 1H), 7.42 (dd, J = 1.0, 5.1 Hz, 1H), 7.47-7.52 (m, 3H), 7.55-7.56 (m, 2H), 8.11-8.16 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 126.26, 126.31, 126.6, 126.76, 126.83, 126.9, 127.5, 127.6, 128.8, 129.7, 130.7, 131.4, 134.0, 140.4, 159.9; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₇H₁₂NOS₂: 310.0355, found: 310.0356.

4,5-Bis(benzo[*b***]thiophen-2-yl)-2-phenyloxazole (3l-Ph)** white solid; m.p. 236.9-238.6 °C; 39 mg (96%); ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.54 (m, 4H), 7.50-7.54 (m, 3H), 7.82-7.91 (m, 6H),

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8.18-8.20 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 122.26, 122.29, 123.4, 123.6, 123.9, 124.1, 124.6, 124.86, 124.92, 125.4, 126.6, 126.8, 128.9, 129.3, 131.0, 132.8, 133.9, 139.4, 139.8, 140.0, 140.2, 141.4, 160.6; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₅H₁₆NOS₂: 410.0668, found: 410.0677.

4-Butyl-2,5-diphenyloxazole (**3m-Ph**) colorless liquid; 14 mg (49%); ¹H NMR (400 MHz, CDCl₃) δ 0.97 (t, *J* = 7.3 Hz, 3H), 1.47 (sext, *J* = 7.5 Hz, 2H), 1.74-1.82 (m, 2H), 2.82 (t, *J* = 7.9 Hz, 2H), 7.34 (t, *J* = 7.4 Hz, 1H), 7.42-7.49 (m, 5H), 7.67-7.69 (m, 2H), 8.08-8.10 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 14.0, 22.7, 27.1, 31.1, 125.5, 126.3, 127.6, 127.7, 128.7, 128.8, 129.3, 130.1, 138.2, 145.2, 159.5; HRMS (APCI) m/z ([M+H]⁺) calcd for C₁₉H₂₀NO: 278.1539, found: 278.1540.

4-Cyclohexyl-2,5-diphenyloxazole (**3n-Ph**) white solid; m.p. 133.6-134.0 °C; 24 mg (80%); ¹H NMR (400 MHz, CDCl₃) δ 1.38-1.42 (m, 3H), 1.75-1.89 (m, 7H), 2.86-2.93 (m, 1H), 7.34 (tt, *J* = 1.2, 7.4 Hz, 1H), 7.42-7.49 (m, 5H), 7.63-7.66 (m, 2H), 8.07-8.10 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 25.9, 26.5, 31.0, 36.0, 126.0, 126.4, 127.7, 127.8, 128.7, 128.8, 129.4, 129.9, 142.8, 144.3, 159.7; HRMS (APCI) m/z ([M+H]⁺) calcd for C₂₁H₂₂NO: 304.1696, found: 304.1697.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publication website at DOI: 10.1021/acs.joc.7b00682.

 1 H, 13 C{ 1 H}, and 19 F{ 1 H} NMR spectra for products and kinetic profiles (PDF)

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Notes

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