

## Synthesis of a Tetrasaccharide Building Block of the O-Specific Polysaccharide of *Shigella dysenteriae* Type 1\*

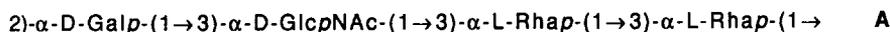
Vince Pozsgay<sup>a,b\*\*</sup>, Cornelis P.J. Glaudemans<sup>b</sup>, John B. Robbins<sup>a</sup>, and Rachel Schneerson<sup>a</sup>

<sup>a</sup>Laboratory of Developmental and Molecular Immunity, National Institute of Child Health and Human Development, and  
<sup>b</sup>Laboratory of Medicinal Chemistry, National Institute of Diabetes, and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, MD 20892 U.S.A.

(Received in USA 20 July 1992)

**Abstract:** A glycosyl trichloroacetimidate derivative (**1**) of the tetrasaccharide  $\alpha$ -D-Galp-(1 $\rightarrow$ 3)- $\alpha$ -D-GlcpNAc-(1 $\rightarrow$ 3)- $\alpha$ -L-Rhap-(1 $\rightarrow$ 3)- $\alpha$ -L-Rhap was synthesized in a highly stereoselective, step-wise manner, using methyl 1-thioglycosides of L-rhamnose, 2-azido-2-deoxy-D-glucose and D-galactose, as major intermediates. The protecting group scenario in compound **1** permits regio-selective deblocking at its "non-reducing end" unit. Therefore **1** is a suitable intermediate for the preparation of extended fragments of the title polysaccharide.

*Shigella dysenteriae* type 1 is a Gram-negative pathogen causing dysentery in humans only, with a high incidence of mortality.<sup>2</sup> The O-specific polysaccharide (O-SP) of its lipopolysaccharide (LPS) envelope is a regular heteropolysaccharide composed<sup>3</sup> of the tetrasaccharide repeating unit **A**. Serum antibodies to the O-

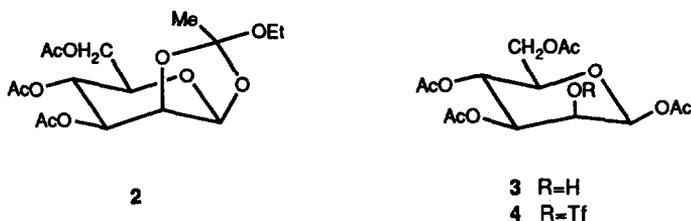


SP may provide the basis for the protection against such infections through vaccination.<sup>2</sup> Host-protective antibodies can be elicited by synthetic antigens in which the native O-SP is covalently linked to a carrier protein.<sup>2,4,5</sup> It is generally agreed upon, that the immunogenicity of the carbohydrate-protein conjugates depends on the carrier protein, the type of attachment, which influences the net charge, the degree of crosslinking, and the carbohydrate-protein ratio.<sup>4</sup> However, there is a considerable degree of controversy concerning the minimal length of the carbohydrate part required for eliciting *protective* antibodies, directed to the carbohydrates of the bacterial cell-surface. There is evidence that the immunogenic properties of the bacterial, cell-surface polysaccharides can also be expressed by carbohydrate structures smaller than the native polysaccharides.<sup>6</sup> Recently, protein-conjugates of synthetic oligosaccharides, corresponding to the inner core regions of *meningococcal* LPSs were shown to induce immunotype-specific antibodies in rabbits.<sup>7</sup> On the other hand, the failure of complete, tetra- and pentasaccharide repeating unit fragments of the capsular polysaccharides of *Pneumococcus* Type 14<sup>8</sup> and Type III Group B *Streptococcus*<sup>9</sup>, respectively, to be antigenic indicates, that the recognition by antibodies generated against native polysaccharides requires extended carbohydrate structures.

\*For a preliminary publication see Ref. 1.

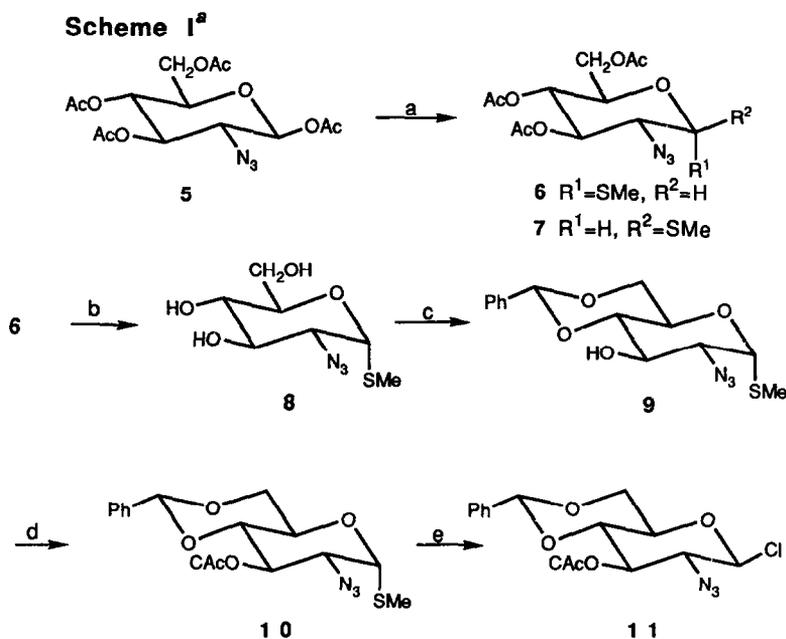
\*\*Mailing address: NIH, Building 8, Rm. B1A-25, Bethesda, MD 20982, U.S.A.





the use of only 1 instead of 2 molar equivalents of the expensive reagent trifluoromethanesulfonic anhydride and keeping the temperature during all operations of the 4→5 conversion below 30°.

The thioglycoside **6** was chosen as the major intermediate. (Scheme I) Compound **6** is a useful  $\alpha$ -D-glucosaminyl donor, since it allows a wide range of protecting group manipulations.<sup>24</sup> The reaction<sup>25</sup> of acetate **5** with (methylthio)trimethylsilane in the presence of TMSOTf gave the crystalline thioglycosides **6** and



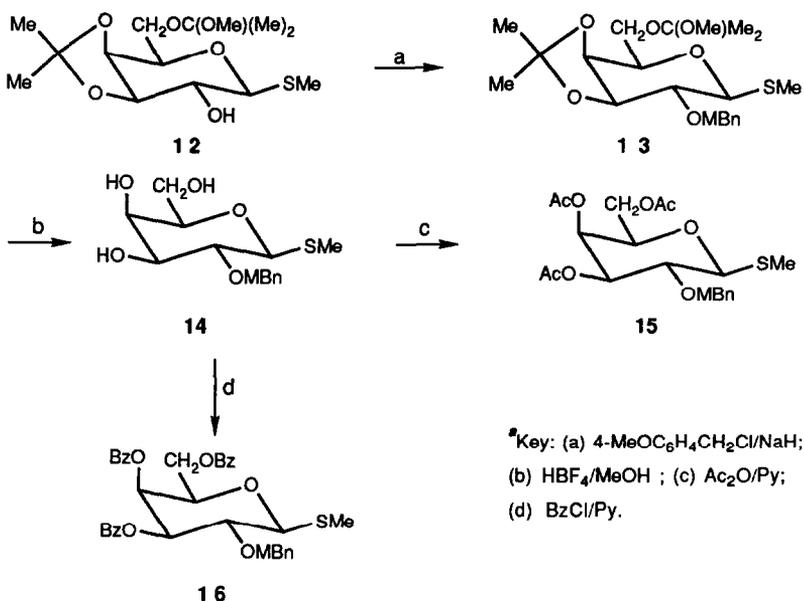
<sup>a</sup>Key: (a) CH<sub>3</sub>SSi(CH<sub>3</sub>)<sub>3</sub>, TMSOTf, (CH<sub>2</sub>Cl)<sub>2</sub>; (b) NaOMe/MeOH; (c) C<sub>6</sub>H<sub>5</sub>CH(OMe)<sub>2</sub>, CSA; (d) CAcCl/Py; (e) Cl<sub>2</sub>.

**7** in a ratio of 4:1, in 93 % yield. The anomeric configuration of the thioglycosides **6** and **7** was ascertained by their <sup>1</sup>J<sub>C-1,H-1</sub> coupling constants (167 Hz and 156 Hz, respectively), which are characteristically lower than those of the *O*-glycosides.<sup>25b</sup> The major,  $\alpha$ -isomer **6** was obtained in pure form by crystallization, without the need of chromatography, and was exposed to NaOMe in methanol to provide the triol **8**. Treatment of **8** with benzaldehyde dimethylacetal in the presence of 10-camphorsulfonic acid gave **9** which was chloroacetylated to

give **10** (88 % yield). Reaction of the thioglycoside **10** with chlorine gave the crystalline chloride **11** in 96 % yield. The 1,2-*trans* configuration in **11** was confirmed by the  $^3J_{H-1,H-2}$  coupling constant being 7.8 Hz.

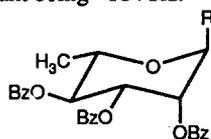
**D-Galactose Units.** Thiogalactoside **16** was selected as the galactosyl donor. In **16** the 4-methoxybenzyl group at *O*-2 serves a dual function. As a non-participating group<sup>16</sup> it allows the stereoselective formation of a 1,2-*cis* interglycosidic linkage upon activation of the anomeric center by thiophilic reagents. The sensitivity of the 4-methoxybenzyl group to oxidation<sup>26</sup> permits selective unmasking of the *HO*-2 group for subsequent chain-elongation. Treatment of the known alcohol<sup>25c</sup> **12** with 4-methoxybenzyl chloride/NaH (Scheme II) gave the crystalline **13** (73 %) which was de-acetalated<sup>27</sup> (HBF<sub>4</sub>/MeOH, 92 %) to give **14**. The position of the 4-methoxybenzyl group in **14** was further confirmed by the <sup>1</sup>H and <sup>13</sup>C NMR data of its triacetate **15** (Experimental). Conventional benzoylation (BzCl/Py) of the triol **14** gave the fully protected, crystalline **16**.

### Scheme II<sup>a</sup>

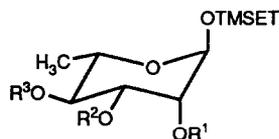


**L-Rhamnose Units.** Acetolysis of the methyl glycoside<sup>28</sup> **17** followed by treatment with  $\alpha,\alpha$ -dichloromethyl methyl ether gave the crystalline chloride<sup>28</sup> **18** (96 %) which was coupled with 2-(trimethylsilyl)ethanol (AgOTf, DTBMP) to provide the glycoside **19** (86 %). We note that attempted coupling of 2,3,4-tri-*O*-acetyl- $\alpha$ -L-rhamnopyranosyl chloride or bromide with 2-(trimethylsilyl)ethanol under Koenigs Knorr-type conditions (AgOTf/base, or Hg(CN)<sub>2</sub>) failed to afford the required glycoside in an acceptable yield. Transesterification (NaOMe/MeOH) of compound **19** provided the triol **20** (92 %). Treatment<sup>29</sup> of the triol **20** with trimethyl orthobenzoate *under vacuum* followed in succession by *in situ* benzoylation and acid

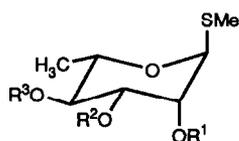
hydrolysis<sup>30,31b</sup> afforded the dibenzoate **21** in 69 % yield. The donor **25** was prepared as follows. The thiorhamnoside<sup>25b</sup> **22** was regioselectively di-*O*-benzoylated [ $\rightarrow$  **23** (79 %)] in analogy to the preparation of compound **21**. Subsequent chloroacetylation at *HO*-3 provided compound **24**. Treatment of **24** with chlorine afforded the rhamnosyl chloride **25** (84 %). The  $\alpha$  anomeric configuration in **25** was ascertained by its  $^1J_{C-1,H-1}$  coupling constant being 184 Hz.



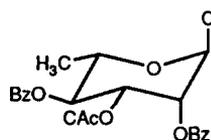
|           |            |
|-----------|------------|
| <b>17</b> | R = OMe    |
| <b>18</b> | R = Cl     |
| <b>19</b> | R = OTMSET |



|           | R <sup>1</sup> | R <sup>2</sup> | R <sup>3</sup> |
|-----------|----------------|----------------|----------------|
| <b>20</b> | H              | H              | H              |
| <b>21</b> | Bz             | H              | Bz             |



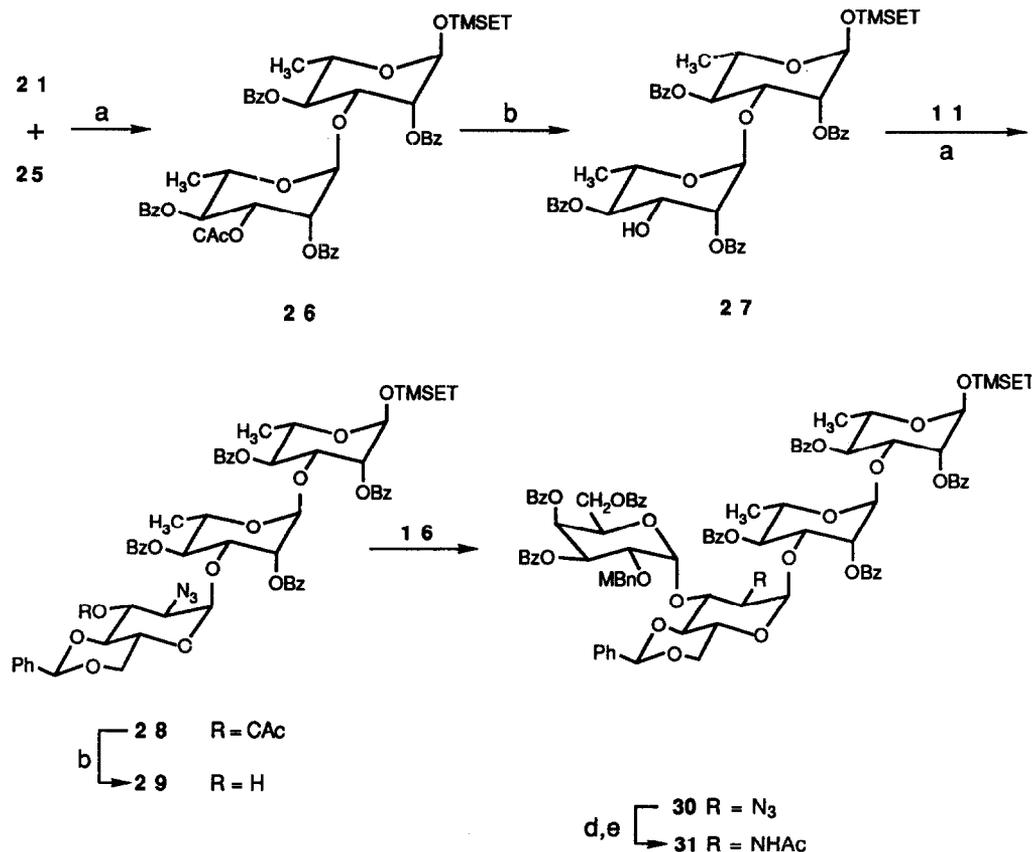
|           | R <sup>1</sup> | R <sup>2</sup> | R <sup>3</sup> |
|-----------|----------------|----------------|----------------|
| <b>22</b> | H              | H              | H              |
| <b>23</b> | Bz             | H              | Bz             |
| <b>24</b> | Bz             | CAC            | Bz             |

**25**

Bz = benzoyl  
 CAC = chloroacetyl  
 TMSET = 2-(trimethylsilyl)ethyl

**Assembly of the tetrasaccharide.** Rhamnosyl chloride **25** was coupled with the acceptor **21** using AgOTf as the promoter and the base 2,6-di-*tert*-butyl-4-methylpyridine. (Scheme III) The disaccharide **26** was obtained in 64 % yield. The  $\alpha$  interglycosidic linkage in **26** was ascertained by the  $^1J_{C-1,H-1}$  coupling constant (172 Hz). Removal of the chloroacetyl group (thiourea in EtOH<sup>32</sup>) afforded the disaccharide alcohol **27**. Our preliminary studies indicated, that the direct activation of the azido-thioglycoside **10** with MeOTf<sup>33</sup> in diethyl ether, in the presence of glycosyl acceptors of moderate nucleophilicity leads to 1,2-*cis*-linked disaccharides in a highly stereocontrolled manner. However, the reactions were sluggish and the yields unsatisfactory. On the other hand, the high reactivity of 2-azido-2-deoxy-glycosyl halides in Koenigs-Knorr-type reactions is amply documented.<sup>16,17</sup> Therefore, in these studies the heterofunctional azido-glycosyl chloride **11**, instead of its precursor thioglycoside **10** was used, as the glucosamine unit. We used glycosyl halides, which were isolated and characterized prior to the glycosylation reactions. This is, however, not always necessary since thioglycosides can be converted *in situ* into the corresponding glycosyl halides. This technology was first reported by Weygand and Ziemann<sup>34</sup> in 1962 and adopted by Bundle's group<sup>35</sup> in 1990 for the synthesis of

1,2-*trans*-linked oligosaccharides. We note, that thioglycoside conversion into a 1,2-*cis*-linked disaccharide



<sup>a</sup>Key: (a) AgOTf, DTBMP, CH<sub>2</sub>Cl<sub>2</sub>; (b) CS(NH<sub>2</sub>)<sub>2</sub>, EtOH; (c) MeOTf, ether; (d) NiCl<sub>2</sub>-H<sub>3</sub>BO<sub>3</sub>-NaBH<sub>4</sub>, EtOH; (e) Ac<sub>2</sub>O.

through the intermediacy of an *in situ*, reagent-generated, glycosyl bromide was also described.<sup>36</sup> Combination of the azido-chloride 11 with the disaccharide nucleophile 27 under AgOTf promotion afforded the trisaccharide 28 (69 %). In this reaction formation of the 1,2-*trans* linked isomer was not observed. The moderate yield of this reaction was due to partial decomposition of the donor 11. Dechloroacetylation of 28 (thiourea<sup>32</sup>) gave the trisaccharide alcohol 29 (90 %). Compound 29 was galactosylated using the thiogalactoside donor 16 under activation by MeOTf<sup>33</sup> to provide the protected tetrasaccharide 30. The proper choice of the solvent was critical to the stereochemical outcome of this reaction. No product containing a 1,2-*trans* interglycosidic linkage could be isolated when diethyl ether was used as the solvent whereas dichloromethane promoted the formation of the



NH<sub>3</sub> as the ionizing gas and by the positive-ion fast atom-bombardment technique performed in the low (FABMS) and high resolution mode (HRFAB), using 3-nitrobenzyl alcohol as the matrix.

**3,4,6-Tri-*O*-acetyl-1,2-*O*-(1-ethoxyethylidene)- $\beta$ -D-mannopyranose (2).** A solution of 1,2,3,4,6-penta-*O*-acetyl- $\alpha,\beta$ -D-mannopyranose (135 g) in a mixture of glacial AcOH (100 mL) and Ac<sub>2</sub>O (10 mL) was treated with 30 % HBr in AcOH (140 mL) at 0°C. The solution was allowed to reach room temperature. After 4 h TLC (1:1 EtOAc-hexane) indicated complete conversion to a faster-moving product. The solution was diluted with CHCl<sub>3</sub> (500 mL) and the mixture was successively extracted with ice-water (5 x 800 mL), 1 % aq. NaHCO<sub>3</sub>, ice-water (500 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. A solution of the syrupy product in MeCN (50 mL) was treated with anhydrous EtOH (80 mL) and 2,4,6-trimethylpyridine (70 mL). After 14 h at room temperature the mixture was diluted with CHCl<sub>3</sub> (500 mL). The mixture was extracted with water (3 x 400 mL). The organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The semisolid obtained was crystallized from EtOH to give **2** (82 g, 63 %): mp 101-103°C, lit.<sup>21</sup> mp 102.5-104°C; [ $\alpha$ ]<sub>D</sub> -19° (c 0.8), lit.<sup>21</sup> [ $\alpha$ ]<sub>D</sub> -15°.

**1,3,4,6-Tetra-*O*-acetyl- $\beta$ -D-mannopyranose (3).** A solution of **2** (50 g) in acetone (300 mL) was treated with 1 N aq. HCl (30 mL) for 5 min at 20°C then was concentrated at 20°C at 20 mmHg. The residue was dissolved in CHCl<sub>3</sub> (300 mL) and the solution was washed with H<sub>2</sub>O (2 x 100 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated to a *thin* syrup. The residue crystallized upon addition of diethyl ether to give **3** (15 g, 32.4 %). A portion was recrystallized from EtOH. Mp 164-166°C, lit.<sup>22</sup> mp 164-165°C; [ $\alpha$ ]<sub>D</sub> -19° (c 0.8), lit.<sup>22</sup> [ $\alpha$ ]<sub>D</sub> -24°.

**1,3,4,6-Tetra-*O*-acetyl-2-*O*-trifluoromethanesulfonyl- $\beta$ -D-mannopyranose (4).** To a stirred solution of **3** (55 g) in CH<sub>2</sub>Cl<sub>2</sub> (600 mL) and pyridine (28 mL) was added trifluoromethanesulfonic anhydride (27 mL) dropwise, at -15°C, during 1 h. The mixture was allowed to reach 5°C then was successively extracted with ice-cold, aq. NaHCO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Crystallization from EtOH gave **4** (71.5 g, 97.5 %), mp 118-120°C, lit.<sup>39</sup> mp 120°C.

**1,3,4,6-Tetra-*O*-acetyl-2-azido-2-deoxy- $\beta$ -D-glucopyranose (5).** A solution of **4** (40.5 g) and sodium azide (11 g) in DMF (250 mL) was stirred for 40 min. Concentration of the solution *below* 30°C gave a syrup which was dissolved in CHCl<sub>3</sub> (200 mL). Extraction with water (3 x 50 mL) followed by drying (Na<sub>2</sub>SO<sub>4</sub>) and concentration gave a syrup which was crystallized from isopropyl ether to give **5** (25 g, 77 %), mp 94-95°C, lit.<sup>19c</sup> mp 97°C; [ $\alpha$ ]<sub>D</sub> +3° (c 1.2), lit.<sup>19c</sup> [ $\alpha$ ]<sub>D</sub> +8°.

**Methyl 2-azido-2-deoxy-3,4,6-tri-*O*-acetyl-1-thio- $\alpha$ - (6) and  $\beta$ -D-glucopyranoside (7).** A solution of **5** (45 g), (methylthio)trimethylsilane (45 mL), and trimethylsilyl trifluoromethanesulfonate (4 mL) in anhydrous (CH<sub>2</sub>Cl)<sub>2</sub> (180 mL) was stirred under reflux for 12 h. The solution was concentrated to a syrup. A solution of the residue in CHCl<sub>3</sub> (200 mL) was extracted with cold, 5 % aq. NaHCO<sub>3</sub>, H<sub>2</sub>O, dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated. Crystallization of the residue from EtOH gave **6** (29.1 g, 66.7 %), mp 96-98°C, [ $\alpha$ ]<sub>D</sub> +167° (c 0.91). Column chromatography of the mother liquor (5:1 hexane-EtOAc) afforded **7** (4.46 g, 10.2 %), mp 73-74°C, [ $\alpha$ ]<sub>D</sub> -41° (c 1.4) and a mixture of **6** and **7** (7.2 g, containing *ca.* 3 % **5**, 16 %). Total yield of **6** and **7**: 93 %. NMR data (CDCl<sub>3</sub>): **6** <sup>1</sup>H,  $\delta$  5.341 (d, 1H, *J*<sub>1,2</sub> 5.5 Hz, H-1), 5.305 (dd, 1H, *J*<sub>2,3</sub> 10.0 Hz, H-3), 5.017 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=10.1 Hz, H-4), 4.402 (ddd, 1H, H-5), 4.315 (dd, 1H *J*<sub>5,6</sub> 4.8 Hz, *J*<sub>6,6'</sub> 12.2 Hz, H-6), 4.090 (dd, 1H *J*<sub>5,6'</sub> 2.0 Hz, H-6'); <sup>13</sup>C,  $\delta$  170.4 and 169.7 (C=O), 84.1 (C-1, *J*<sub>C-1,H-1</sub> 167 Hz), 72.1 (C-4), 68.8 (C-3), 67.9 (C-5), 61.9 (C-6), 61.6 (C-2), 20.6 (CH<sub>3</sub>CO), 12.8 (CH<sub>3</sub>S); **7** <sup>1</sup>H,  $\delta$  5.113 (t, 1H *J*<sub>2,3</sub>=*J*<sub>3,4</sub>=9.4 Hz, H-3), 5.043 (t, 1H, H-4), 4.348 (d, 1H, *J*<sub>1,2</sub> 10.2 Hz, H-1), 4.288 (dd, 1H, *J*<sub>5,6</sub> 4.8 Hz, *J*<sub>6,6'</sub> 12.3 Hz, H-6), 4.163 (dd, 1H *J*<sub>5,6'</sub> 2.2 Hz, H-6'); <sup>13</sup>C  $\delta$  170.4, 169.7, 169.5 (C=O), 84.4 *J*<sub>C-1,H-2</sub> 156 Hz, C-1), 75.8 (C-5), 74.3 (C-3), 68.2 (C-4), 63.1 (C-2), 62.0 (C-6), 20.6, 20.4 (CH<sub>3</sub>CO), 12.4 (CH<sub>3</sub>S). CIMS: **6** and **7** *m/z* 379 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>13</sub>H<sub>19</sub>N<sub>3</sub>O<sub>7</sub>S: C, 43.20; H, 5.30; N, 11.63; S, 8.87. Found: **6** C, 43.75; H, 5.38; N, 11.74; S, 8.98; **7** C, 43.86; H, 5.42; N, 11.68; S, 9.06.

**Methyl 2-azido-2-deoxy-1-thio- $\alpha$ -D-glucopyranoside (8).** A solution of **6** (5.0 g) in anhydrous MeOH (40 mL) was treated with a catalytic amount of MeONa at room temperature for 3 h. The solution was neutralized (Dowex 50X2-200, H<sup>+</sup>), and concentrated. Trituration of the residue in ether gave **8** (2.68 g, 82 %), mp 123-124°C, [ $\alpha$ ]<sub>D</sub> +221° (c 1.2, MeOH). NMR data (D<sub>2</sub>O): <sup>1</sup>H,  $\delta$  5.436 (d, 1H, *J*<sub>1,2</sub> 5.5 Hz, H-1), 4.015 (ddd, 1H, *J*<sub>4,5</sub> 9.7 Hz, H-5), 3.895 (dd, 1H, H-2), 3.874 (dd, 1H, *J*<sub>5,6</sub> 2.3 Hz, H-6), 3.780 (dd, 1H, *J*<sub>5,6'</sub> 5.6 Hz, *J*<sub>6,6'</sub> 12.3 Hz, H-6'), 3.686 (dd, 1H, *J*<sub>2,3</sub> 10.1, H-3), 3.463 (t, 1H, *J*<sub>3,4</sub> 9.7 Hz, H-4), 2.130 (CH<sub>3</sub>); <sup>13</sup>C,  $\delta$  84.9 (C-1), 73.3 (C-3), 73.1 (C-5), 70.7 (C-4), 64.4 (C-2), 61.3 (C-6), 13.0 (CH<sub>3</sub>). CIMS: *m/z* 253 [M+18]<sup>+</sup>, 236 [M+1]<sup>+</sup>. HRFAB: Found *m/z* 236.0706, C<sub>7</sub>H<sub>14</sub>N<sub>3</sub>O<sub>4</sub>S requires 236.0705.

**Methyl 2-azido-4,6-*O*-benzylidene-2-deoxy-1-thio- $\alpha$ -D-glucopyranoside (9).** A solution of compound **8** (2.3 g) in MeCN (20 mL) was treated with benzaldehyde dimethylacetal (5 mL) and a catalytic

amount of 10-camphorsulfonic acid at room temperature for 3 h. The mixture was treated with triethylamine and concentrated to a syrup. Trituration in hexane afforded crystalline **9** (2.65g, 84 %), mp 160-161°C,  $[\alpha]_D +160^\circ$  (c 0.58). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 7.3-7.55 (m, 5H, aromatic protons), 5.525 (s, 1H, HCPH), 5.250 (d, 1H, J<sub>1,2</sub> 5.3 Hz, H-1), 4.15-4.27 (m, 2H, H-6,6'), 3.976 (t, 1H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.3 Hz, H-3), 3.828 (dd, 1H, H-2), 3.73-3.82 (m, 1H, H-5), 3.509 (t, 1H, H-4), 2.11 (s, 3H, CH<sub>3</sub>S); <sup>13</sup>C, δ 136.8, 129.4, 128.3, 126.2 (aromatic carbons), 102.1 (CHPh), 85.1 (C-1), 81.7 (C-4), 70.6 (C-3), 68.6 (C-6), 63.7 (C-2), 62.7 (C-5), 13.2 (CH<sub>3</sub>S). CIMS: m/z 341 [M+18]<sup>+</sup>, 324 [M+1]<sup>+</sup>. Anal. Calcd for C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>4</sub>S: C, 52.00; H, 5.30; N, 13.00; S, 9.91. Found: C, 52.57; H, 5.44; N, 12.65; S, 9.76.

**Methyl 2-azido-4,6-O-benzylidene-3-O-chloroacetyl-2-deoxy-1-thio-α-D-glucopyranoside (10).** To a stirred solution of **9** (11.6 g) in a mixture of pyridine (50 mL) and DMF (50 mL) at -50°C was added chloroacetyl chloride (8.3 mL) during 15 min. The mixture was allowed to reach ca 0°C in 20 min then was concentrated under vacuum. A solution of the residue in CHCl<sub>3</sub> (200 mL) was extracted with ice-cold aq. NaHCO<sub>3</sub>, ice-water, was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was crystallized from EtOAc to give **10** (6.6 g). Column chromatography (4:1 hexane-EtOAc) of the mother liquor afforded an additional amount of **10** (6.0 g, total yield 88 %), mp 160-162°C,  $[\alpha]_D +86^\circ$  (c 1.03). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 7.35-7.45 (m, 5H, aromatic resonances), 5.499 (s, 1H, HCPH), 5.447 (t, 1H, J<sub>2,3</sub>=J<sub>3,4</sub>=9.8 Hz, H-3), 5.365 (d, 1H, J<sub>1,2</sub> 5.7 Hz, H-1), 4.25-4.35 (m, 2H, H-5,6), 4.108 (s, 2H, CH<sub>2</sub>Cl), 4.035 (dd, 1H, H-2), 3.800 (m, 1H, H-6'), 3.670 (t, 1H, H-4), 2.351 (s, 3H, CH<sub>3</sub>S); <sup>13</sup>C, δ 166.0 (C=O), 126.1, 128.2, 129.1, 136.6 (aromatic carbons), 101.6 (CHPh), 85.0 (C-1), 79.2 (C-4), 72.3 (C-3), 68.5 (C-6), 63.0 (C-2), 62.2 (C-5), 40.5 (CH<sub>2</sub>Cl), 13.1 (CH<sub>3</sub>S). CIMS: m/z 417 [M+18]<sup>+</sup>. HRFAB: Found m/z 400.0716, C<sub>16</sub>H<sub>19</sub>ClN<sub>3</sub>O<sub>5</sub>S requires 400.0734.

**2-Azido-4,6-O-benzylidene-3-O-chloroacetyl-2-deoxy-β-D-glucopyranosyl chloride (11).** A solution of **10** (2.1 g) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was treated at 0°C with chlorine dissolved in CCl<sub>4</sub>. After 10 min 1-hexene was added until the yellow color disappeared. Concentration of the solution afforded a syrupy residue which crystallized from hexane to give **11** (1.95 g, 96 %), mp 81-83°C,  $[\alpha]_D -80^\circ$  (c 0.82). <sup>1</sup>H NMR data (CDCl<sub>3</sub>): δ 7.9-7.3 (aromatic protons), 5.51 (s, 1H, CHPh), 5.220 (d, 1H, J<sub>1,2</sub> 7.8 Hz, H-1), 4.395 (dd, 1H, J<sub>5,6</sub> 4.9 Hz, J<sub>6,6'</sub> 10.4 Hz, H-6), 4.138 (s, 2H, CH<sub>2</sub>Cl), 3.616 (dd, 1H, J<sub>5,6'</sub> 4.9 Hz). CIMS: m/z 405 [M+18]<sup>+</sup>.

**Methyl 2,3-O-isopropylidene-2-O-(4-methoxybenzyl)-6-O-(2-methoxy-2-methylethyl)-β-D-galactopyranoside<sup>25c</sup> (13).** A solution of methyl 2,3-O-isopropylidene-6-O-(2-methoxy-2-methylethyl)-β-D-galactopyranoside<sup>25c</sup> (33 g) in DMF (100 mL) was treated with NaH (4.1 g, 60 % dispersion in oil) at 0°C, under stirring. 4-Methoxybenzyl chloride (15 mL) was added dropwise. Stirring was continued for 3 h at 0°C. MeOH (5 mL) was added dropwise, followed by H<sub>2</sub>O (500 mL). The mixture was extracted with CHCl<sub>3</sub> (3x100 mL). The combined CHCl<sub>3</sub> extracts were washed with H<sub>2</sub>O (5x100 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>). Crystallization from hexane afforded **13** (21.5 g). Column chromatography of the mother liquor (4:1 hexane-ethyl acetate containing 1 % Et<sub>3</sub>N) gave an additional amount of **13** (10.5 g, total yield 73 %), mp 79-81°C,  $[\alpha]_D -9^\circ$  (c 0.9). NMR data (C<sub>6</sub>D<sub>6</sub>-CHCl<sub>3</sub>, 100:5): <sup>1</sup>H, δ 6.8-7.5 (aromatic protons), 4.870 and 4.760 [2d, 2H, J 11.2 Hz, CH<sub>2</sub> (benzyl)], 4.226 (d, 1H, J<sub>1,2</sub> 10.5 Hz, H-1), 4.056 (dd, 1H, H-3), 3.911 (dd, 1H, J<sub>3,4</sub> 5.7 Hz, J<sub>4,5</sub> 2.2 Hz, H-4), 3.858 (dd, 1H, J<sub>5,6</sub> 6.8 Hz, J<sub>6,6'</sub> 9.7 Hz, H-6), 3.727 (dd, 1H, J<sub>5,6'</sub> 5.3 Hz, H-6'), 3.633 (ddd, 1H, H-5), 3.607 (dd, 1H, J<sub>2,3</sub> 6.4 Hz, H-2), 3.348 [s, 3H, CH<sub>3</sub>O (MBn)], 3.167 (CH<sub>3</sub>OC(CH<sub>3</sub>)<sub>2</sub>), 2.031 (CH<sub>3</sub>S), 1.373 and 1.234 (2s, 2x3H, (CH<sub>3</sub>)<sub>2</sub>CO<sub>2</sub>), 1.283 (s, 6H, (CH<sub>3</sub>)<sub>2</sub>COCH<sub>3</sub>); <sup>13</sup>C, δ 159.3, 130.3-127.2, 113.5 (aromatic carbons), 109.3 [C(CH<sub>3</sub>)<sub>2</sub>], 99.68 (CH<sub>3</sub>OC(CH<sub>3</sub>)<sub>2</sub>), 84.1 (C-1), 79.7 (C-3), 78.4 (C-2), 75.5 (C-5), 73.9 (C-4), 72.8 [CH<sub>2</sub> (MBn)], 60.5 (C-6), 54.4 CH<sub>3</sub>O [(MBn)], 47.9 (CH<sub>3</sub>OC(CH<sub>3</sub>)<sub>2</sub>), 27.7 and 26.0 (CH<sub>3</sub>)<sub>2</sub>CO<sub>2</sub>), 24.1 [(CH<sub>3</sub>)<sub>2</sub>COCH<sub>3</sub>], 11.8 (CH<sub>3</sub>S). CIMS: m/z 460 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>22</sub>H<sub>34</sub>O<sub>7</sub>S: C, 59.71; H, 7.74; S, 7.24. Found: C, 59.90; H, 7.77; S, 7.29.

**Methyl 2-O-(4-methoxybenzyl)-1-thio-β-D-galactopyranoside (14).** A solution of **13** (32 g) in MeOH (220 mL) was treated with 50 % aq. HBF<sub>4</sub> (3 mL) at 0°C for 1 h. The resulting crystalline mass was treated with ether (50 mL). Filtration afforded **14** (19.3 g). The filtrate was treated with triethylamine and was concentrated. The residue was triturated with ether (100 mL). Filtration afforded a second crop of **14** (3.5 g, total yield 92 %), mp 173-175°C,  $[\alpha]_D -3^\circ$  (c 0.34, MeOH). NMR data (DMSO-d<sub>6</sub>): <sup>1</sup>H NMR, δ 6.85-7.33 (m, 4H, aromatic protons), 4.700, 4.610 [2d, 2H, J 10 Hz, CH<sub>2</sub> (MBn)], 4.238 (d, 1H, J<sub>1,2</sub> 9.3 Hz, H-1), 3.736 (s, 3H, CH<sub>3</sub>O), 3.71 (m, 1H, H-4), 3.46-3.54 (m, 3H, H-3,6,6'), 3.36-3.43 (m, 2H, H-2,5), 2.108 (s, 3H, CH<sub>3</sub>S); <sup>13</sup>C, δ 158.4, 130.8, 129.1, 113.2 (aromatic carbons), 84.2 (C-1), 78.9 (C-5), 77.8 (C-2), 74.4 (C-3), 73.5 [CH<sub>2</sub> (MBn)], 68.7 (C-4), 60.4 (C-6), 54.9 (CH<sub>3</sub>O), 11.8 (CH<sub>3</sub>S). CIMS: m/z 348 [M+18]<sup>+</sup>. Anal. Calcd for C, 59.71; H, 7.74; S, 7.24. Found: C, 59.81, H, 7.73; S, 7.33.

**Methyl 3,4,6-tri-*O*-acetyl-2-*O*-(4-methoxybenzyl)-1-thio- $\beta$ -D-galactopyranoside (15).** A solution of **14** (500 mg) in anhydrous pyridine (5 mL) at 0°C was treated with Ac<sub>2</sub>O (4 mL). The solution was allowed to reach room temperature in 2 h. Concentration to a syrup under vacuum followed by crystallization from hexane afforded **15** (635 mg, 92 %), mp 96-98°C, [ $\alpha$ ]<sub>D</sub> +10° (c 1.1). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  6.83-6.9 and 7.22-7.28 (m, 4H, aromatic protons), 5.405 (dd, 1H, *J*<sub>3,4</sub> 4 Hz, *J*<sub>4,5</sub> 1 Hz, H-4), 4.996 (dd, 1H *J*<sub>2,3</sub> 9.7 Hz, H-3), 4.76 and 4.56 [2d, 2H, *J* 10.4 Hz, CH<sub>2</sub> (MBn)], 4.441 (d, 1H, *J*<sub>1,2</sub> 10.5 Hz, H-1), 4.161 (dd, 1H, *J*<sub>5,6</sub> 6.6 Hz, *J*<sub>6,6'</sub> 11.2 Hz, H-6), 4.087 (dd, 1H, *J*<sub>5,6'</sub> 6.6 Hz, H-6'), 3.864 (dt, 1H, H-5), 3.793 (s, 1H, CH<sub>3</sub>O), 3.659 (t, 1H, H-2); <sup>13</sup>C,  $\delta$  170.2, 170.0, 169.7 (C=O), 158.3, 129.7, 129.6, 113.7 (aromatic carbons), 85.9 (C-1), 75.3 (C-2), 75.1 [CH<sub>2</sub> (MBn)], 74.0 (C-3,5), 67.7 (C-4), 61.5 (C-6), 55.3 (CH<sub>3</sub>O), 20.7 (CH<sub>3</sub>CO), 13.2 (CH<sub>3</sub>S). CIMS: *m/z* 474 [M+18]<sup>+</sup>, 457 [M+1]<sup>+</sup>. Anal. Calcd for C<sub>21</sub>H<sub>28</sub>O<sub>9</sub>S: C, 55.25; H, 6.18; S, 7.02. Found: C, 55.34; H, 6.22; S, 7.08.

**Methyl 3,4,6-tri-*O*-benzoyl-2-*O*-(4-methoxybenzyl)-1-thio- $\beta$ -D-galactopyranoside (16).** A solution of **14** (3 g) in pyridine (20 mL) at 0°C was treated with benzoyl chloride (5 mL). The mixture was kept at 25°C for 2 h. Methanol (10 mL) was added under cooling with ice. The solution was concentrated to a syrup which was dissolved in CHCl<sub>3</sub> (50 mL). The CHCl<sub>3</sub> solution was extracted with H<sub>2</sub>O (5x20 mL), dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography of the residue (8:1 → 4:1 hexane-EtOAc) gave **16** (5.29 g, 90 %), which was crystallized from diisopropyl ether, mp 150-151°C, [ $\alpha$ ]<sub>D</sub> +71° (c 1.45). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  6.62-8.06 (m, 19H, aromatic protons), 5.924 (bd, 1H, H-4), 5.458 (dd, 1H, *J*<sub>2,3</sub> 9.7 Hz, *J*<sub>3,4</sub> 3.4 Hz, H-3), 4.59 and 4.77 [2d, 2H, CH<sub>2</sub> (MBn)], 4.638 (d, 1H, *J*<sub>1,2</sub> 9.8 Hz, H-1), 4.625 (dd, 1H, H-6), 4.341 (dd, 1H, *J*<sub>5,6'</sub> 6.6 Hz, *J*<sub>6,6'</sub> 11.2 Hz, H-6'), 4.225 (bt, 1H, H-5), 3.963 (t, 1H H-2), 3.689 (s, 3H, CH<sub>3</sub>O), 2.340 (s, 3H, CH<sub>3</sub>S); <sup>13</sup>C,  $\delta$  165.8, 165.2 (C=O), 159.2, 133.0-133.4, 129.3-129.9, 128.1-128.5-113.6 (aromatic carbons), 86.1 (C-1), 75.3 (C-2), 75.1 [CH<sub>2</sub> (MBn)], 74.6 (C-3,5), 68.8 (C-4), 62.2 (C-6), 55.2 (CH<sub>3</sub>O), 13.3 (CH<sub>3</sub>S). CIMS: *m/z* 660 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>36</sub>H<sub>34</sub>O<sub>9</sub>S: C, 67.27; H, 5.33; S, 4.99. Found: C, 66.90; H, 5.47; S, 4.81.

**2,3,4-Tri-*O*-benzoyl- $\alpha$ -L-rhamnopyranosyl chloride (18).** A solution of methyl 2,3,4-tri-*O*-benzoyl- $\alpha$ -L-rhamnopyranoside<sup>28</sup> (20 g) in Ac<sub>2</sub>O (80 mL) was treated with 100  $\mu$ L of cc. H<sub>2</sub>SO<sub>4</sub> for 4 h at 25°C. NaHCO<sub>3</sub> (1 g) was added and the solution was concentrated. The residual syrup was dissolved in CHCl<sub>3</sub>. The CHCl<sub>3</sub> solution was extracted with aq. NaHCO<sub>3</sub>, water, was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. A solution of the residue in CH<sub>2</sub>Cl<sub>2</sub> (150 mL) was treated with  $\alpha,\alpha$ -dichloromethyl methyl ether (10 mL), and freshly fused ZnCl<sub>2</sub> (ca. 0.2 g) for 3 h at 25°C. The mixture was cooled to 0°C then was poured into ice-cold, aq. NaHCO<sub>3</sub>. The aqueous solution was extracted with CHCl<sub>3</sub>. The organic phase was washed with H<sub>2</sub>O, was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Crystallization of the residue from EtOAc afforded **18** (19.5 g, 96 %), mp 165-166°C, lit.<sup>28</sup> mp 165-166°C, [ $\alpha$ ]<sub>D</sub> +129° (c 0.77), lit.<sup>28</sup> [ $\alpha$ ]<sub>D</sub> +136°. NMR data (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  7.22-8.12 (aromatic protons), 6.231 (d, 1H, *J*<sub>1,2</sub> 1.4 Hz, H-1), 6.096 (dd, 1H, *J*<sub>2,3</sub> 3.3 Hz, *J*<sub>3,4</sub> 10.3 Hz, H-3), 5.815 (dd, 1H, H-2), 5.752 (t, 1H, H-4), 4.491 (dq, 1H, H-5), 1.417 (d, 3H, *J*<sub>5,6</sub> 6.3 Hz, H-6); <sup>13</sup>C,  $\delta$  165.6 (C=O), 128.3-133.7 (aromatic carbons), 89.2 (C-1), 72.8 (C-2), 71.0 (C-4), 69.8 (C-5), 68.7 (C-3), 17.3 (C-6). CIMS: *m/z* 512 [M+18]<sup>+</sup>, 459 [M-35]<sup>+</sup>. Anal. Calcd for C<sub>27</sub>H<sub>23</sub>O<sub>7</sub>Cl: C, 65.52; H, 4.68; Cl, 7.16. Found: C, 65.27; H, 4.69; Cl, 7.32.

**2-(Trimethylsilyl)ethyl 2,3,4-tri-*O*-benzoyl- $\alpha$ -L-rhamnopyranoside (19).** A mixture of **18** (19 g), 2,6-di-*tert*-butyl-4-methylpyridine (7 g), 2-(trimethylsilyl)ethanol (8 mL), powdered, 4A molecular sieves (10 g), and anhydrous CH<sub>2</sub>Cl<sub>2</sub> (100 mL) was stirred for 1 h then cooled to -20°C. AgOTf (10 g) was added in one portion. The reaction mixture was stirred for 2 h at 0°C. More AgOTf (8 g) was added and stirring was continued at 0°C for 3 h. The mixture was treated with ice cold, aq. NaHCO<sub>3</sub> (5 %) and was then filtered. The filtrate was extracted with H<sub>2</sub>O, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. The residue was purified by column chromatography (4:1 hexane-EtOAc) to give **19** as a syrup (19 g, 86 %), [ $\alpha$ ]<sub>D</sub> +122° (c 0.8). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  7.22-8.14 (aromatic protons), 5.851 (dd, 1H, *J*<sub>2,3</sub> 3.5 Hz, *J*<sub>3,4</sub> 10.0 Hz, H-3), 5.668 (t, 1H, *J*<sub>4,5</sub> 10.0 Hz, H-4), 5.627 (dd, 1H, H-2), 5.032 (d, 1H, *J*<sub>1,2</sub> 1.6 Hz, H-1), 4.209 (dq, 1H, H-5), 3.907, 3.646 (2x1H, OCH<sub>2</sub>CH<sub>2</sub>), 1.366 (d, 3H, H-6), 1.00-1.12 (2H, CH<sub>2</sub>Si), 0.093 (s, 9H, SiMe<sub>3</sub>); <sup>13</sup>C,  $\delta$  165.7, 165.6, 165.4 (C=O), 128.2-133.3 (aromatic carbons), 97.0 (C-1), 71.9 (C-4), 71.2 (C-2), 70.1 (C-3), 66.6 (C-5), 65.8 (OCH<sub>2</sub>), 18.8 (SiCH<sub>2</sub>), 17.7 (C-6), -1.3 (CH<sub>3</sub>)<sub>3</sub>Si. CIMS: *m/z* 594 [M+18]<sup>+</sup>, 459 [M-C<sub>5</sub>H<sub>13</sub>OSi]<sup>+</sup>. Anal. Calcd for C<sub>32</sub>H<sub>36</sub>O<sub>8</sub>Si: C, 66.64; H, 6.29. Found: C, 66.68; H, 6.34.

**2-(Trimethylsilyl)ethyl  $\alpha$ -L-rhamnopyranoside (20).** A solution of **19** (15 g) in anhydrous MeOH (100 mL) was treated with a catalytic amount of MeONa for 24 h. The solution was neutralized [Dowex 50x2 (H<sup>+</sup>)] and concentrated. Column chromatography of the residue (EtOAc) afforded **20** as a syrup (6.3 g, 92

%),  $[\alpha]_D -75^\circ$  (c 1.5). NMR data (MeOH- $d_4$ ):  $^1\text{H}$ ,  $\delta$  4.680 (d, 1H,  $J_{1,2}$  1.6 Hz, H-1), 3.775 (dd, 1H, H-2), 3.655 (dd, 1H,  $J_{2,3}$  3.3 Hz,  $J_{3,4}$  9.4 Hz, H-3), 3.588 (dq, 1H, H-5), 3.371 (t, 1H,  $J_{4,5}$  9.4 Hz, H-4), 1.261 (d, 3H,  $J_{5,6}$  6.3 Hz, H-6);  $^{13}\text{C}$ ,  $\delta$  101.2 (C-1), 73.9 (C-4), 72.4 (C-3), 72.3 (C-2), 69.6 (C-5), 65.8 (OCH<sub>2</sub>), 18.6 (CH<sub>2</sub>Si), 17.9 (C-6), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. CIMS:  $m/z$  282 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>11</sub>H<sub>24</sub>O<sub>5</sub>Si: C, 49.97; H, 9.15. Found: C, 49.36; H, 8.91.

**2-(Trimethylsilyl)ethyl 2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (21).** A solution of compound 20 (3.5 g) in EtOAc (30 mL) was mixed with trimethyl orthobenzoate (4.5 mL) at 25°C, then most of EtOAc was removed under vacuum. The mixture was treated with 10-camphorsulfonic acid (40 mg) followed by evacuation of the reaction flask. After 10 min pyridine (5 mL) was added, the solution was cooled to 0°C and treated with benzoyl chloride (2.5 mL) for 1 h at 0°C. MeOH (5 mL) was added under stirring, then the mixture was concentrated and treated with aq. 80 % AcOH (30 mL). After 5 min the solution was concentrated and the residue was partitioned between CHCl<sub>3</sub> and H<sub>2</sub>O. The organic phase was concentrated. Column chromatography (8:1 hexane-EtOAc) of the residue afforded 21 (4.3 g, 69 %) as a syrup which crystallized on standing, mp 133-134°C,  $[\alpha]_D +31^\circ$  (c 0.9). NMR data (CDCl<sub>3</sub>):  $^1\text{H}$ ,  $\delta$  8.05-8.15, 7.56-7.64, 7.43-7.52 (aromatic protons), 5.358 (dd, 1H,  $J_{1,2}$  1.7 Hz,  $J_{2,3}$  3.5 Hz, H-2), 5.263 (t, 1H,  $J_{3,4}=J_{4,5}=9.7$  Hz, H-4), 4.985 (d, 1H, H-1), 4.320 (ddd, 1H, H-3), 4.078 (dq, 1H, H-5), 2.424 (d, 1H,  $J_{\text{H-3,OH}}$  8.2 Hz, HO-3), 1.320 (d, 1H, H-6);  $^{13}\text{C}$ ,  $\delta$  167.0, 166.0 (C=O), 128.4-133.4 (aromatic carbons), 96.9 (C-1), 75.7 (C-4), 73.6 (C-2), 69.1 (C-3), 66.2 (C-5), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>Si), 18.0 (OCH<sub>2</sub>CH<sub>2</sub>Si), 17.7 (C-6), -1.2 [Si(CH<sub>3</sub>)<sub>3</sub>]. Anal. Calcd for C<sub>25</sub>H<sub>32</sub>O<sub>7</sub>Si: C, 63.53; H, 6.83. Found: C, 63.44; H, 6.85.

**Methyl 2,4-di-O-benzoyl-1-thio- $\alpha$ -L-rhamnopyranoside (23).** 10-Camphorsulfonic acid (40 mg) was added to a stirred mixture of methyl 1-thio- $\alpha$ -L-rhamnopyranoside<sup>25b</sup> (1 g), DMF (0.5 mL) and trimethyl orthobenzoate (3 mL) followed, *immediately*, by exposure of the reaction flask to vacuum. After 5 min pyridine (2 mL) was added and the mixture was cooled to 0°C. Benzoyl chloride (2 mL) was added under stirring and the reaction mixture was allowed to reach 25°C in ca. 30 min. The mixture was then cooled to 0°C and was treated with MeOH (5 mL) followed by removal of the volatiles in vacuum. The residue was treated with aq. 80 % AcOH (10 mL). After 10 min the mixture was concentrated. Column chromatography of the residue (7:1 hexane-EtOAc) afforded 23 as an amorphous solid (1.64 g, 79 %),  $[\alpha]_D -21^\circ$  (c 1.2), lit.<sup>29</sup>  $[\alpha]_D -21^\circ$ . For NMR data see Ref. 29. CIMS:  $m/z$  420 [M+18]<sup>+</sup>, 403 [M+1]<sup>+</sup>.

**Methyl 2,4-di-O-benzoyl-3-O-chloroacetyl-1-thio- $\alpha$ -L-rhamnopyranoside (24).** A solution of 23 (15 g) in a 1:1 mixture of pyridine and DMF (100 mL) was treated at -45°C with chloroacetyl chloride (4.5 mL) under stirring. After 5 min the reaction mixture was concentrated under reduced pressure at 30°C. Column chromatography (7:1 hexane-EtOAc) of the residue afforded 24 as an amorphous solid (16.2 g, 91%),  $[\alpha]_D +23^\circ$  (c 1.3). NMR data (CDCl<sub>3</sub>):  $^1\text{H}$ ,  $\delta$  8.1-7.3 (aromatic protons), 5.681 (dd, 1H,  $J_{1,2}$  1.4 Hz,  $J_{2,3}$  3.2 Hz, H-2), 5.630 (dd, 1H,  $J_{3,4}$  9.9 Hz, H-3), 5.527 (t, 1H,  $J_{4,5}$  9.9 Hz, H-4), 5.316 (d, 1H, H-1), 4.446 (dq, 1H, H-5), 3.887 and 3.817 (2d 2x1H,  $J$  for each 15 Hz, CH<sub>2</sub>Cl), 2.219 (s, 3H, CH<sub>3</sub>S), 1.363 (d, 3H, H-6);  $^{13}\text{C}$ ,  $\delta$  166.5, 165.6 (C=O), 128.5-133.6 (aromatic carbons), 83.4 (C-1), 71.6 (C-2,3,4), 67.2 (C-5), 40.4 (CH<sub>2</sub>Cl), 17.6 (C-6), 13.8 (CH<sub>3</sub>S). CIMS:  $m/z$  496 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>23</sub>H<sub>23</sub>ClO<sub>7</sub>S: C, 57.68; H, 4.84; Cl, 7.40; S, 6.69. Found: C, 57.78; H, 4.92; Cl, 7.31; S, 6.63.

**2,4-Di-O-benzoyl-3-O-chloroacetyl- $\alpha$ -L-rhamnopyranosyl chloride (25).** A solution of 24 (10 g) in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was treated with a solution of Cl<sub>2</sub> in CCl<sub>4</sub> at 0°C until the mixture remained yellow. After 10 min the solution was concentrated. Column chromatography (7:1 hexane-EtOAc) of the residue afforded 25 as a syrup (8.2 g, 84 %),  $[\alpha]_D +52^\circ$  (c 1). NMR data (CDCl<sub>3</sub>):  $^1\text{H}$ ,  $\delta$  8.1-7.4 (aromatic protons), 6.160 (d, 1H, H-1), 5.941 (dd, 1H,  $J_{2,3}$  3.5 Hz,  $J_{3,4}$  10.2 Hz, H-3), 5.710 (dd, 1H,  $J_{1,2}$  1.7 Hz, H-2), 5.569 (t, 1H,  $J_{4,5}$  10.2 Hz, H-4), 4.413 (dq, 1H, H-5), 3.896 and 3.832 (2d, 2x1H,  $J$  for each 15 Hz, CH<sub>2</sub>Cl), 1.39 (d, 1H,  $J_{5,6}$  6.3 Hz, H-6);  $^{13}\text{C}$ ,  $\delta$  166.5, 165.5, 165.3 (C=O), 128.6-133.9 (aromatic carbons), 89.0 (C-1,  $^1J_{\text{C-1,H-1}}$  184 Hz), 72.3 (C-2), 70.7 (C-4), 69.8 (C-3,5), 40.3 (CH<sub>2</sub>Cl), 17.3 (C-6). CIMS:  $m/z$  484 [M+18]<sup>+</sup>.

**2-(Trimethylsilyl)ethyl 2,4-di-O-benzoyl-3-O-(2,4-di-O-benzoyl-3-O-chloroacetyl- $\alpha$ -L-rhamnopyranosyl)- $\alpha$ -L-rhamnopyranoside (26).** Silver trifluoromethanesulfonate (3.9 g) was added to a stirred mixture of 21 (2.2 g), 25 (3.8 g), 2,6-di-*tert*-butyl-4-methylpyridine (1.6 g) and powdered 4A molecular sieves (5 g) in CH<sub>2</sub>Cl<sub>2</sub> (45 mL) at -40°C. The stirring was continued for 1 h during which the reaction mixture reached room temperature. Ice-cold, 5 % aq. NaHCO<sub>3</sub> was added and the mixture was filtered. The mixture was diluted with CHCl<sub>3</sub> (50 mL) and was extracted with water. The organic layer was concentrated. Column chromatography (7:1 hexane-EtOAc) of the residue afforded 26 as an amorphous solid (2.7 g, 64 %),  $[\alpha]_D$

+99° (c 0.8). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.26-7.33 (aromatics), 5.560 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>A</sub>), 5.467 (dd, 1H, *J*<sub>1,2</sub> 1.6 Hz, *J*<sub>2,3</sub> 3.3 Hz, H-2<sub>A</sub>), 5.428 (dd, 1H, *J*<sub>2,3</sub> 3.3 Hz, H-3<sub>B</sub>), 5.305 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.7 Hz, H-4<sub>B</sub>), 5.164 (dd, 1H, *J*<sub>1,2</sub> 1.7 Hz, *J*<sub>2,3</sub> 3.4 Hz, H-2<sub>B</sub>), 5.135 (d, 1H, H-1<sub>B</sub>), 5.013 (d, 1H, H-1<sub>A</sub>), 4.470 (dd, 1H, H-3<sub>A</sub>), 4.105 (dq, 1H, H-5<sub>B</sub>), 4.062 (dq, 1H, H-5<sub>A</sub>), 3.776 and 3.672 (2d, 2x1H, *J* 15.6 Hz, CH<sub>2</sub>Cl), 1.340 (d, 3H, H-6<sub>A</sub>), 1.160 (d, 3H, H-6<sub>B</sub>), 0.07 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>Si); <sup>13</sup>C, δ 166.2-194.9 (5 C=O), 133.5-128.4 (aromatic carbons), 99.2 (C-1<sub>B</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 172 Hz), 96.9 (C-1<sub>A</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 171 Hz), 76.0 (C-3<sub>A</sub>), 73.4 (C-4<sub>A</sub>), 72.5 (C-2<sub>A</sub>), 71.3, 70.5, 70.1 (C-2<sub>B</sub>, 3<sub>B</sub>, 4<sub>B</sub>) 67.4 (C-5<sub>B</sub>), 66.6 (C-5<sub>A</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 40.3 (CH<sub>2</sub>Cl), 17.9 (CH<sub>2</sub>Si), 17.7 (C-6<sub>A</sub>), 17.3 (C-6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. CIMS: *m/z* 921 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>47</sub>H<sub>57</sub>ClO<sub>14</sub>Si: C, 62.48; H, 5.69, Cl, 3.92. Found: C, 63.31; H, 5.98; Cl, 3.79.

**2-(Trimethylsilyl)ethyl 2,4-di-*O*-benzoyl-3-*O*-(2,4-di-*O*-benzoyl-α-*L*-rhamnopyranosyl)-α-*L*-rhamnopyranoside (27).** A solution of 26 (2.5 g) and thiourea (1.5 g) in EtOH (50 mL) was stirred at 25°C for 36 h. The solution was concentrated and the residue was triturated with CHCl<sub>3</sub> (50 mL). The mixture was filtered, and the filtrate was extracted with ice-cold, 5% aq. HCl, H<sub>2</sub>O, 5% aq. NaHCO<sub>3</sub>, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated. Column chromatography (4:1 hexane-EtOAc) of the residue afforded 27 as an amorphous solid (2.25 g, 90%), [α]<sub>D</sub> +83° (c 1.1). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.24-7.33 (aromatic protons), 5.553 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>A</sub>), 5.476 (dd, 1H, *J*<sub>1,2</sub> 1.7 Hz, *J*<sub>2,3</sub> 3.2 Hz, H-2<sub>A</sub>), 5.176 (d, 1H, *J*<sub>1,2</sub> 1.4 Hz, H-1<sub>B</sub>), 5.087 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>B</sub>), 5.020 (dd, 1H, H-2<sub>B</sub>), 5.012 (dd, 1H, H-1<sub>A</sub>), 4.450 (dd, 1H, H-3<sub>A</sub>), 4.076 (ddd, 1H, H-3<sub>B</sub>), 4.062 (dq, 1H, H-5<sub>A</sub>), 4.006 (dq, 1H, H-5<sub>B</sub>), 2.221 (d, 1H, *J*<sub>3,H<sub>O</sub></sub> 7.7 Hz, HO), 1.335 (d, 3H, H-6<sub>A</sub>), 1.138 (d, 3H, H-6<sub>B</sub>), 0.070 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>Si); <sup>13</sup>C, δ 166.7, 166.0, 165.3 (C=O), 133.5-128.3 (aromatic carbons), 99.2 (C-1<sub>B</sub>), 96.8 (C-1<sub>A</sub>), 76.3 (C-3<sub>A</sub>), 75.1 (C-4<sub>B</sub>), 73.2 (C-4<sub>A</sub>), 72.9 (C-2<sub>B</sub>), 72.7 (C-2<sub>A</sub>), 68.5 (C-3<sub>B</sub>), 66.9 (C-5<sub>B</sub>), 66.6 (C-5<sub>A</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 17.9 (CH<sub>2</sub>Si), 17.7, 17.3 (C-6<sub>A</sub>, 6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. CIMS: *m/z* 844 [M+18]<sup>+</sup>. Anal. Calcd for C<sub>45</sub>H<sub>50</sub>O<sub>13</sub>Si: C, 65.36; H, 6.09. Found: C, 65.78; H, 6.50.

**2-(Trimethylsilyl)ethyl 3-*O*-[3-*O*-(2-azido-4,6-*O*-benzylidene-3-*O*-chloroacetyl)-2-deoxy-α-*D*-glucopyranosyl]-2,4-di-*O*-benzoyl-α-*L*-rhamnopyranosyl]-2,4-di-*O*-benzoyl-α-*L*-rhamnopyranoside (28).** To a stirred solution of 11 (2.1 g), 27 (2.2 g) and 2,6-di-*tert*-butyl-4-methylpyridine (1.1 g) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) containing powdered, 4A molecular sieves (3 g) at -50°C was added AgOTf (1.5 g) and the mixture was stirred for 2 h during which time the temperature of the mixture was allowed to reach 0°C. Processing as described for 27, using 4:1 hexane-EtOAc as chromatographic eluent, afforded 28 as an amorphous solid (1.95 g, 69%), [α]<sub>D</sub> +141° (c 0.5). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.25-6.87 (aromatic protons), 5.549 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=10 Hz, H-4<sub>A</sub>), 5.506 (dd, 1H, *J*<sub>1,2</sub> 1.6 Hz, *J*<sub>2,3</sub> 3.4 Hz, H-2<sub>A</sub>), 5.402 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.7 Hz, H-4<sub>B</sub>), 5.225 (dd, 1H, H-2<sub>B</sub>), 5.181 (s, 1H, HCPH), 5.150 (d, 1H, *J*<sub>1,2</sub> 1.6 Hz, H-1<sub>B</sub>), 5.087 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-3<sub>C</sub>), 4.985 (d, 1H, H-1<sub>A</sub>), 4.769 (d, 1H, *J*<sub>1,2</sub> 3.8 Hz, H-1<sub>C</sub>), 4.472 (dd, 1H, *J*<sub>2,3</sub> 9.8 Hz, *J*<sub>3,4</sub> 3.4 Hz, H-3<sub>A</sub>), 4.143 (dd, 1H, *J*<sub>2,3</sub> 3.3 Hz, H-3<sub>B</sub>), 4.7 (2dq, 2H, H-5<sub>A</sub>, 5<sub>B</sub>), 3.92 (s, 2H, CH<sub>2</sub>Cl), 3.722 (dd, 1H, *J*<sub>5,6</sub> 4.7 Hz, *J*<sub>6,6'</sub> 9.8 Hz, H-6<sub>C</sub>), 3.484 (ddd, 1H, H-5<sub>C</sub>), 3.39-3.30 (m, 2H, H-4<sub>C</sub>, 6'<sub>C</sub>), 2.948 (dd, 1H, H-2<sub>C</sub>), 1.346 (d, 3H, *J*<sub>5,6</sub> 6.3 Hz, H-6<sub>A</sub>), 1.171 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>B</sub>), 0.06 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>Si); <sup>13</sup>C, δ 165.9, 165.6, 165.3, 165.1 (C=O), 136.6, 133.5-133.1, 129.8-127.8, 126.2 (aromatic carbons), 101.2 (CHPh), 99.1 (C-1<sub>B</sub>), 96.9 (C-1<sub>A</sub>), 94.7 (C-1<sub>C</sub>), 78.5 (C-4<sub>C</sub>), 75.6 (C-3<sub>A</sub>), 73.6 (C-4<sub>A</sub>), 72.3 (C-2<sub>A</sub>), 71.6 (C-4<sub>B</sub>), 71.5 (C-3<sub>B</sub>), 70.6 (C-3<sub>C</sub>), 68.0 (C-6<sub>C</sub>), 67.7 (C-5<sub>B</sub>), 67.5 (C-2<sub>B</sub>), 66.5 (C-5<sub>A</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 62.5 (C-5<sub>C</sub>), 60.9 (C-2<sub>C</sub>), 40.4 (CH<sub>2</sub>Cl), 17.9 (CH<sub>2</sub>Si), 17.7 (C-6<sub>A</sub>), 17.4 (C-6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. HRFAB: Found *m/z* 1176.3572, C<sub>60</sub>H<sub>63</sub><sup>35</sup>CN<sub>3</sub>O<sub>18</sub>Si (M+H<sup>+</sup>+H<sub>2</sub>) requires 1176.3564.

**2-(Trimethylsilyl)ethyl 3-*O*-[3-*O*-(2-azido-4,6-*O*-benzylidene-2-deoxy-α-*D*-glucopyranosyl)-2,4-di-*O*-benzoyl-α-*L*-rhamnopyranosyl]-2,4-di-*O*-benzoyl-α-*L*-rhamnopyranoside (29).** A solution of 28 (1.7 g) and thiourea (2.0 g) in EtOH (50 mL) was stirred at 25°C for 36 h. Work-up as described for 27 afforded 29 as an amorphous solid (1.46 g, 90%), [α]<sub>D</sub> +117° (c 0.6). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.22-6.99 (aromatics), 5.546 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.9 Hz, H-4<sub>A</sub>), 5.499 (dd, 1H, *J*<sub>1,2</sub> 1.6 Hz, *J*<sub>2,3</sub> 3.4 Hz, H-2<sub>A</sub>), 5.369 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>B</sub>), 5.215 (s, 1H, HCPH), 5.195 (dd, 1H, H-2<sub>B</sub>), 5.146 (d, 1H, *J*<sub>1,2</sub> 1.5 Hz, H-1<sub>B</sub>), 4.983 (d, 1H, H-1<sub>A</sub>), 4.586 (d, 1H, *J*<sub>1,2</sub> 4.6 Hz, H-1<sub>C</sub>), 4.463 (dd, 1H, *J*<sub>3,4</sub> 9.8 Hz, H-3<sub>A</sub>), 4.110 (dd, 1H, *J*<sub>2,3</sub> 3.2 Hz, *J*<sub>3,4</sub> 9.9 Hz, H-3<sub>B</sub>), 4.073 (dq, 1H, H-5<sub>B</sub>), 4.030 (dq, 1H, H-5<sub>A</sub>), 3.414 (ddd, 1H, H-5<sub>C</sub>), 3.754 (dt, *J*<sub>2,3</sub>=*J*<sub>3,4</sub>=10 Hz, *J*<sub>3,OH</sub> 3.4 Hz, H-3<sub>C</sub>), 3.679 (dd, 1H, *J*<sub>5,6</sub> 4.0 Hz, *J*<sub>6,6'</sub> 9.4 Hz, H-6<sub>C</sub>), 3.332 (t, *J*<sub>5,6'</sub> 9.4 Hz, H-6<sub>C</sub>), 3.175 (t, 1H, H-4<sub>C</sub>), 2.916 (dd, 1H, *J*<sub>2,3</sub> 10 Hz, H-2<sub>C</sub>), 1.348, 1.135 (2d, 2x3H, H-6<sub>A</sub>, H-6<sub>B</sub>), 0.070 [(CH<sub>3</sub>)<sub>3</sub>Si]. <sup>13</sup>C, δ 166.0-165.2 (C=O), 136.7-126.4 (aromatic carbons), 101.7 (CHPh), 99.1 (C-1<sub>B</sub>), 96.9 (C-1<sub>A</sub>), 94.9 (C-1<sub>C</sub>), 81.2 (C-4<sub>C</sub>), 75.8 (C-3<sub>A</sub>), 73.5 (C-4<sub>A</sub>), 72.3 (C-2<sub>A</sub>), 72.0 (C-4<sub>B</sub>), 71.7 (C-3<sub>B</sub>), 68.4 (C-3<sub>C</sub>), 68.2 (CH<sub>2</sub>CH<sub>2</sub>Si), 68.0 (C-2<sub>B</sub>), 67.4 (C-5<sub>A</sub>), 66.6 (C-5<sub>B</sub>), 65.7 (C-6<sub>C</sub>), 62.6 (C-2<sub>C</sub>), 62.4 (C-5<sub>C</sub>), 17.9 (CH<sub>2</sub>Si), 17.7, 17.35 (C-6<sub>A</sub>, 6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. FAB/MS: *m/z* 1074

[M-N<sub>2</sub>+1]<sup>+</sup>. Anal. Calcd for C<sub>58</sub>H<sub>63</sub>N<sub>3</sub>O<sub>17</sub>Si: C, 63.20; H, 5.76; N, 3.81. Found: C, 63.28; H, 5.80, N, 3.80.

**2-(Trimethylsilyl)ethyl 3-O-{3-O-[2-azido-4,6-O-benzylidene-3-O-(3,4,6-tri-O-benzoyl-2-O-/4-methoxybenzyl/-α-D-galactopyranosyl)-2-deoxy-α-D-glucopyranosyl]-2,4-di-O-benzoyl-α-L-rhamnopyranosyl}-2,4-di-O-benzoyl-α-L-rhamnopyranoside (30).** To a solution of **16** (1.7 g), **29** (1.39 g), 2,6-di-*tert*-butyl-4-methylpyridine (1.5 g) in ether (40 mL), containing powdered, 4A molecular sieves (4 g) was added methyl triflate<sup>33</sup> (MeOTf) (300 μL). Stirring was continued for 24 h. Triethylamine (1 mL) was added and the mixture was filtered. The solids were washed with CHCl<sub>3</sub>. The filtrate was concentrated. Column chromatography (3:1 hexane-EtOAc) of the residue afforded **30** as an amorphous solid (1.9 g, 89 %), [α]<sub>D</sub> +136° (c 0.7). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.22-6.41 (aromatic protons), 5.752 (dd, *J*<sub>3,4</sub> 3.2 Hz, H-4<sub>D</sub>), 5.607 (dd, 1H, *J*<sub>2,3</sub> 10.7 Hz, H-3<sub>D</sub>), 5.562 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>A</sub>), 5.514 (dd, 1H, *J*<sub>1,2</sub> 1.6 Hz, *J*<sub>2,3</sub> 3.4 Hz, H-2<sub>A</sub>), 5.417 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.9 Hz, H-4<sub>B</sub>), 5.383 (d, 1H, *J*<sub>1,2</sub> 3.4 Hz, H-1<sub>D</sub>), 5.248 (dd, 1H, H-2<sub>B</sub>), 5.219 (s, 1H *H*CPh), 5.160 (d, 1H, H-1<sub>B</sub>), 4.990 (d, 1H, H-1<sub>A</sub>), 4.652 (d, 1H, *J*<sub>1,2</sub> 3.9 Hz, H-1<sub>C</sub>), 4.468 (dd, 1H, H-3<sub>A</sub>), 4.270 and 3.944 [2d, 2H, CH<sub>2</sub> (MBn)], 3.651 (s, 3H, CH<sub>3</sub>O), 3.032 (dd, 1H, *J*<sub>2,3</sub> 10.4 Hz, H-3), 1.358 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>A</sub>), 1.139 (d, 3H, *J*<sub>5,6</sub> 6.1 Hz, H-6<sub>B</sub>), 0.07 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>Si); <sup>13</sup>C, δ 165.9, 165.5, 165.3, 165.1 (C=O), 102.1 (CHPh), 99.1 (C-1<sub>B</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 168 Hz), 97.0 (C-1<sub>A</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 171 Hz), 96.9 (C-1<sub>D</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 171 Hz), 94.8 (C-1<sub>C</sub>, <sup>1</sup>*J*<sub>C-1,H-1</sub> 170 Hz), 82.0 (C-4<sub>C</sub>), 76.1 (C-3<sub>A</sub>), 73.4 (C-4<sub>B</sub>), 72.4, 72.3, 71.9, 71.5, 70.9 (C-2<sub>A</sub>, 2<sub>B</sub>, 2<sub>D</sub>, 3<sub>C</sub>, 4<sub>A</sub>), 70.5 (CH<sub>2</sub> (MBn)), 69.0, 68.9, 67.8, 67.5, 66.6(2C) (C-3<sub>B</sub>, 3<sub>D</sub>, 4<sub>D</sub>, 5<sub>A</sub>, 5<sub>B</sub>, 5<sub>D</sub>), 68.4 (C-6<sub>C</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 62.1, 61.4 (C-2<sub>C</sub>, 5<sub>C</sub>), 61.1 (C-6<sub>D</sub>), 17.9 (CH<sub>2</sub>Si), 17.7, 17.4 (C-6<sub>A</sub>, 6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. FABMS: *m/z* 1668 [M-N<sub>2</sub>+1]<sup>+</sup>. Anal. Calc for C<sub>93</sub>H<sub>93</sub>N<sub>3</sub>O<sub>26</sub>Si: 65.83; H, 5.52; N, 2.48. Found: C, 66.30; H, 6.02; N, 2.34.

**2-(Trimethylsilyl)ethyl 3-O-{3-O-[2-acetamido-4,6-O-benzylidene-3-O-(3,4,6-tri-O-benzoyl-2-O-/4-methoxybenzyl/-α-D-galactopyranosyl)-2-deoxy-α-D-glucopyranosyl]-2,4-di-O-benzoyl-α-L-rhamnopyranosyl}-2,4-di-O-benzoyl-α-L-rhamnopyranoside (31).** To a solution containing **30** (1.8 g), NiCl<sub>2</sub> (0.8 g), and H<sub>3</sub>BO<sub>3</sub> (0.5 g) in EtOH was added a 1 % solution of NaBH<sub>4</sub> in EtOH dropwise until the black color persisted. More NiCl<sub>2</sub> (1.6 g), H<sub>3</sub>BO<sub>3</sub> (1.0 g) was added followed by NaBH<sub>4</sub> in EtOH until the starting material disappeared. The mixture was concentrated to *ca* half its original volume then cooled to 0°C. Acetic anhydride (10 mL) was added. After 15 min the solution was concentrated. The residue was equilibrated between CHCl<sub>3</sub> and H<sub>2</sub>O, the organic phase was dried and concentrated. Column chromatography (3:1 hexane-EtOAc) of the residue afforded **31** as an amorphous solid (780 mg, 43 %), [α]<sub>D</sub> +128° (c 0.5). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.22-7.08, 6.59-6.43 (aromatic resonances), 6.65 (d, 1H, *H*NAc), 5.84 (d, 1H, *J*<sub>3,4</sub> 3.4 Hz, H-4<sub>D</sub>), 5.60 (dd, 1H, *J*<sub>2,3</sub> 10.5 Hz, H-3<sub>D</sub>), 5.52 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=10.0 Hz, H-4<sub>A</sub>), 5.48 (dd, 1H, *J*<sub>1,2</sub> 1.5 Hz, *J*<sub>2,3</sub> 3.5 Hz, H-2<sub>A</sub>), 5.28 (d, 1H, *J*<sub>1,2</sub> 4.0 Hz, H-1<sub>D</sub>), 5.23 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.9 Hz, H-4<sub>B</sub>), 5.17 (s, 1H, *H*CPh), 5.16 (dd, 1H, H-2<sub>B</sub>), 5.09 (d, 1H, H-1<sub>B</sub>), 4.99 (d, 1H, H-1<sub>A</sub>), 3.65 (s, 3H, CH<sub>3</sub>O), 1.68 (s, 3H, CH<sub>3</sub>CON), 1.32 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>A</sub>), 1.25 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>B</sub>), 0.07 (s, 9H, [(CH<sub>3</sub>)<sub>3</sub>Si]). <sup>13</sup>C, δ 169.9 [C=O (AcNH)], 102.0 (CHPh), 100.2, 99.5, 96.9(2C) (C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 82.1 (C-4<sub>C</sub>), 70.3 [CH<sub>2</sub> (MBn)], 68.4 (C-6<sub>C</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 61.1 (C-5<sub>C</sub>), 60.4 (C-6<sub>D</sub>), 55.0 (OCH<sub>3</sub>), 51.4 (C-2<sub>C</sub>), 22.3 (CH<sub>3</sub>CON), 17.9 (CH<sub>2</sub>Si), 17.6 (C-6<sub>A</sub>), 17.4 (C-6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. FABMS: *m/z* 1712 [M+1]<sup>+</sup>. Anal. Calcd for C<sub>95</sub>H<sub>97</sub>NO<sub>27</sub>Si: 66.62; H, 5.71; N, 0.82. Found: C, 67.01; H, 6.10; N, 0.82.

**2-(Trimethylsilyl)ethyl 3-O-{3-O-[2-acetamido-4,6-O-benzylidene-3-O-(3,4,6-tri-O-benzoyl-α-D-galactopyranosyl)-2-deoxy-α-D-glucopyranosyl]-2,4-di-O-benzoyl-α-L-rhamnopyranosyl}-2,4-di-O-benzoyl-α-L-rhamnopyranoside (32).** A mixture of **31** (700 mg), 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (200 mg), CH<sub>2</sub>Cl<sub>2</sub> (10 mL), and H<sub>2</sub>O (2 mL) was stirred at 25°C for 12 h. The mixture was extracted with aq. 5 % NaHCO<sub>3</sub>. The organic phase was concentrated. Column chromatography (2:1 hexane-EtOAc) of the residue afforded **32** as an amorphous solid (640 mg, 98 %), [α]<sub>D</sub> +107 (c 0.8). NMR data (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.22-7.2 (aromatic protons), 6.49 (d, 1H, *J*<sub>H-2,NH</sub> 9.9 Hz, *H*N), 5.875 (bd, 1H, H-4<sub>D</sub>), 5.544 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.8 Hz, H-4<sub>A</sub>), 5.515 (dd, 1H, H-2<sub>A</sub>), 5.508 (dd, 1H, *J*<sub>3,4</sub> 3.3 Hz, H-3<sub>D</sub>), 5.294 (s, 1H, *H*CPh), 5.239 (t, 1H, *J*<sub>3,4</sub>=*J*<sub>4,5</sub>=9.6 Hz, H-4<sub>B</sub>), 5.156 (dd, 1H, *J*<sub>1,2</sub> 1.7 Hz, *J*<sub>2,3</sub> 3.3 Hz, H-2<sub>B</sub>), 5.093 (d, 1H, H-1<sub>B</sub>), 5.072 (d, 1H *J*<sub>1,2</sub> 3.6 Hz, H-1<sub>D</sub>), 4.996 (d, 1H *J*<sub>1,2</sub> 1.5 Hz, H-1<sub>A</sub>), 4.462 (dd, 1H, H-3<sub>A</sub>), 3.960 (dd, 1H, H-3<sub>B</sub>), 2.66 (d, 1H, *H*O), 1.66 (s, 3H, CH<sub>3</sub>CON), 1.339 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>A</sub>), 1.220 (d, 3H, *J*<sub>5,6</sub> 6.2 Hz, H-6<sub>B</sub>), 0.070 (s, 9H, (CH<sub>3</sub>)<sub>3</sub>Si); <sup>13</sup>C, δ 170.2 (CH<sub>3</sub>CON), 166.1-165.4 [C=O (Bz)], 136.4, 133.8-132.8, 129.8-127.8, 126.0 (aromatic carbons), 101.1 (CHPh), 100.8, 99.2 (2C), 96.9 (C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>), 81.0 (C-4<sub>C</sub>), 75.8 (C-3<sub>A</sub>), 73.6 (C-4<sub>B</sub>), 68.0 (C-6<sub>C</sub>), 65.7 (OCH<sub>2</sub>CH<sub>2</sub>), 63.2 (C-5<sub>C</sub>), 61.2 (C-6<sub>D</sub>), 51.7 (C-2<sub>C</sub>), 22.4 (CH<sub>3</sub>O), 17.9 (CH<sub>2</sub>Si), 17.6 (C-6<sub>A</sub>), 17.4 (C-6<sub>B</sub>), -1.3 [(CH<sub>3</sub>)<sub>3</sub>Si]. FABMS: *m/z* 1592 [M+1]<sup>+</sup>. Anal. Calcd for C<sub>87</sub>H<sub>89</sub>NO<sub>26</sub>Si: 65.61; H, 5.63; N, 0.88. Found: C, 65.88; H, 5.87; N, 0.88.

**2-(Trimethylsilyl)ethyl 3-O-(3-O-[2-acetamido-4,6-O-benzylidene-3-O-(3,4,6-tri-O-benzoyl-2-O-chloroacetyl- $\alpha$ -D-galactopyranosyl)-2-deoxy- $\alpha$ -D-glucopyranosyl]-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl)-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (33).** To a solution of 32 (610 mg) in pyridine (3 mL) at 0°C was added under stirring chloroacetyl chloride (150  $\mu$ L). After 15 min the mixture was diluted with  $\text{CHCl}_3$ , and the resulting solution was extracted with ice-cold, aq. 5%  $\text{NaHCO}_3$ . The organic phase was concentrated. Column chromatography (2:1 hexane-EtOAc) of the residue afforded 33 as an amorphous solid (600 mg, 92%):  $[\alpha]_D +114^\circ$  (c 0.6). NMR data ( $\text{CDCl}_3$ ):  $^1\text{H}$ ,  $\delta$  8.22-7.08 (aromatic protons), 6.74 (d, 1H,  $J_{\text{H-2,NH}}$  9.8 Hz, HN), 5.978 (bd, 1H  $J_{3,4}$  3.4 Hz, H-4<sub>D</sub>), 5.669 (dd, 1H,  $J_{1,2}$  4.4 Hz,  $J_{2,3}$  10.8 Hz, H-2<sub>D</sub>), 5.529 (t, 1H,  $J_{3,4}=J_{4,5}=9.8$  Hz, H-4<sub>A</sub>), 5.497 (dd, 1H, H-2<sub>A</sub>), 5.496 (dd, 1H,  $J_{2,3}$  10.4 Hz,  $J_{3,4}$  3.8 Hz, H-3<sub>D</sub>), 5.227 (dd, 1H,  $J_{1,2}$  3.8 Hz, H-2<sub>D</sub>), 5.213 (s, 1H, HCPH), 5.204 (t, 1H,  $J_{3,4}=J_{4,5}=9.8$  Hz, H-4<sub>B</sub>), 5.154 (dd, 1H,  $J_{1,2}$  1.7 Hz,  $J_{2,3}$  3.4 Hz, H-2<sub>B</sub>), 5.086 (d, 1H, H-1<sub>B</sub>), 4.991 (d, 1H,  $J_{1,2}$  1.5 Hz, H-1<sub>A</sub>), 3.322, 3.317 (2d, 2H,  $J$  15 Hz,  $\text{H}_2\text{CCl}$ ), 1.711 (s, 3H,  $\text{CH}_3\text{CON}$ ), 1.324 (d, 3H,  $J_{5,6}$  6.2 Hz, H-6<sub>A</sub>), 1.205 (d, 3H,  $J_{5,6}$  6.2 Hz, H-6<sub>B</sub>), 0.07 (s, 9H,  $(\text{CH}_3)_3\text{Si}$ ).  $^{13}\text{C}$ ,  $\delta$  170.1 ( $\text{CH}_3\text{C}=\text{O}$ ), 166.4-164.9 [ $\text{C}=\text{O}$  (Bz)], 101.3 (CHPh), 100.1, 99.4, 97.1, 96.9 (C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 81.6 (C-4<sub>C</sub>), 75.8 (C-3<sub>A</sub>), 73.7 (C-4<sub>B</sub>), 68.1 (C-6<sub>C</sub>), 65.7 ( $\text{OCH}_2\text{CH}_2$ ), 63.0 (C-5<sub>C</sub>), 60.1 (C-6<sub>D</sub>), 51.6 (C-2<sub>C</sub>), 39.9 ( $\text{CH}_2\text{Cl}$ ), 22.4 ( $\text{CH}_3\text{CO}$ ), 17.9 ( $\text{CH}_2\text{Si}$ ), 17.6 (C-6<sub>A</sub>), 17.4 (C-6<sub>B</sub>), -1.3 [ $(\text{CH}_3)_3\text{Si}$ ]. FABMS:  $m/z$  1668  $[\text{M}+1]^+$ . Anal. Calcd for  $\text{C}_{89}\text{H}_{90}\text{ClNO}_{27}\text{Si}$ : C, 64.04; H, 5.43; N, 0.84; Cl, 2.12. Found: C, 63.61; H, 5.66; N, 0.95; Cl, 2.37.

**2-(Trimethylsilyl)ethyl 3-O-(3-O-[2-acetamido-4,6-di-O-acetyl-3-O-(3,4,6-tri-O-benzoyl-2-O-chloroacetyl- $\alpha$ -D-galactopyranosyl)-2-deoxy- $\alpha$ -D-glucopyranosyl]-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl)-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (35).** A solution of 33 (520 mg) in AcOH (5 mL) was warmed to 70°C. Water (1 mL) was added and the solution was stirred at 70°C for 6 h. The solution was concentrated. Residual solvents were removed by addition and distillation of toluene (3 x 5 mL). Column chromatography (2:1 hexane-EtOAc) of the residue afforded the diol 34 (280 mg) (FABMS:  $m/z$  1580  $[\text{M}+1]^+$ ) which was treated with pyridine (1 mL) and  $\text{Ac}_2\text{O}$  (1 mL) at 25°C for 3 h. Removal of the solvents followed by column chromatography (3:1 hexane-EtOAc) afforded 35 (205 mg, 70%),  $[\alpha]_D +127^\circ$ . NMR data ( $\text{CDCl}_3$ ):  $^1\text{H}$ ,  $\delta$  8.22-7.22 (aromatic protons), 5.959 (bd, 1H,  $J_{3,4}$  3.2 Hz, H-4<sub>D</sub>), 5.618 (dd, 1H,  $J_{2,3}$  10.6 Hz, H-3<sub>D</sub>), 5.52 (t, 1H,  $J_{3,4}=J_{4,5}=9.8$  Hz, H-4<sub>A</sub>), 5.500 (dd, 1H, H-2<sub>A</sub>), 5.460 (dd, 1H,  $J_{1,2}$  3.8 Hz, H-2<sub>D</sub>), 5.220 (t, 1H,  $J_{3,4}=J_{4,5}=9.6$  Hz, H-4<sub>B</sub>), 5.140 (dd, 1H,  $J_{1,2}$  1.6 Hz,  $J_{2,3}$  3.2 Hz, H-2<sub>B</sub>), 5.089 (d, 1H, H-1<sub>D</sub>), 4.99 (d, 1H,  $J_{1,2}$  1.5 Hz, H-1<sub>A</sub>), 4.970 (t, 1H,  $J_{3,4}=J_{4,5}=9.3$  Hz, H-4<sub>C</sub>), 4.55, 8.82 (2d, 2H,  $J$  15 Hz,  $\text{H}_2\text{CCl}$ ), 1.936, 1.709, 1.609 (3s, 3H each,  $3\text{CH}_3\text{CO}$ ), 1.327 (d, 3H,  $J_{5,6}$  6.3 Hz, H-6<sub>A</sub>), 1.204 (d, 3H,  $J_{5,6}$  6.1 Hz, H-6<sub>B</sub>), 0.07 (s, 9H,  $(\text{CH}_3)_3\text{Si}$ );  $^{13}\text{C}$ ,  $\delta$  170.4-164.8 (11 lines,  $\text{C}=\text{O}$ ), 99.2, 98.7, 96.9, 96.3 (C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 76.0 (C-3<sub>A</sub>), 73.5 (C-4<sub>A</sub>), 65.7 ( $\text{OC H}_2\text{CH}_2$ ), 60.7, 60.6 (C-6<sub>C</sub>, 6<sub>D</sub>), 51.2 (C-2<sub>C</sub>), 40.7 ( $\text{CH}_2\text{Cl}$ ), 22.3 ( $\text{CH}_3\text{CON}$ ), 20.5 (2C) ( $\text{CH}_3\text{CO}$ ), 17.9 ( $\text{CH}_2\text{Si}$ ), 17.5 (C-6<sub>A</sub>), 17.4 (C-6<sub>B</sub>), -1.3 [ $(\text{CH}_3)_3\text{Si}$ ]. FABMS:  $m/z$  1664  $[\text{M}+1]^+$ . Anal. Calcd for  $\text{C}_{86}\text{H}_{90}\text{ClNO}_{29}\text{Si}$ : C, 62.03; H, 5.45; N, 0.84. Found: C, 61.77; H, 5.55; N, 0.85.

**O-(3,4,6-Tri-O-benzoyl-2-O-chloroacetyl- $\alpha$ -D-galactopyranosyl)-(1 $\rightarrow$ 3)-2-acetamido-4,6-di-O-acetyl-2-deoxy- $\alpha$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl-(1 $\rightarrow$ 3)-2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl trichloroacetimidate (1).** A solution of 35 (160 mg) in a 1:1 mixture of  $\text{CH}_2\text{Cl}_2$  and trifluoroacetic acid (2 mL) was kept at 25°C for 4 h then the volatiles were removed. Toluene (3 x 5 mL) was added and evaporated from the residue. The solid residue was dissolved in  $\text{CH}_2\text{Cl}_2$  (2 mL) and the solution was cooled to -20°C.  $\text{CCl}_3\text{CN}$  (0.4 mL) and 1,8-diazabicyclo[5.4.0]undec-7-ene (10  $\mu$ L) were added. The solution was stirred at -20°C for 1 h then was allowed to reach ca 20°C in 1 h. Removal of the volatiles was followed by column chromatography (2:1 hexane-EtOAc) to give 1 as an amorphous solid (120 mg, 73%),  $[\alpha]_D +124^\circ$  (c 0.4). NMR data ( $\text{CDCl}_3$ ):  $^1\text{H}$ ,  $\delta$  6.48 (d, 1H,  $J_{\text{H-2,NH}}$  8 Hz, NH), 6.467 (d, 1H,  $J_{1,2}$  1.6 Hz, H-1<sub>A</sub>), 5.958 (bd, 1H, H-4<sub>D</sub>), 5.732 (dd, 1H,  $J_{2,3}$  2.9 Hz, H-2<sub>A</sub>), 5.647 (t, 1H,  $J_{3,4}=J_{4,5}=9.9$  Hz, H-4<sub>A</sub>), 5.614 (dd, 1H,  $J_{3,4}$  3.2 Hz, H-3<sub>D</sub>), 5.452 (dd, 1H,  $J_{1,2}$  3.6 Hz,  $J_{2,3}$  10.8 Hz, H-2<sub>D</sub>), 5.258 (t, 1H,  $J_{3,4}=J_{4,5}=9.7$  Hz, H-4<sub>B</sub>), 5.166 (dd, 1H, H-2<sub>B</sub>), 5.122 (d, 1H, H-1<sub>B</sub>), 5.102 (d, 1H,  $J_{1,2}$  3.7 Hz, H-1<sub>D</sub>), 4.985 (t, 1H,  $J_{3,4}=J_{4,5}=9.5$  Hz, H-4<sub>C</sub>), 4.06, 3.82 (2d,  $\text{H}_2\text{CCl}$ ), 1.948, 1.706, 1.590 ( $\text{CH}_3\text{CO}$ ), 1.383 (d, 3H, H-6<sub>A</sub>), 1.210 (d, 1H, H-6<sub>B</sub>).  $^{13}\text{C}$ ,  $\delta$  99.2 ( $^1J_{\text{C-1,H-1}}$  174 Hz), 98.3 ( $^1J_{\text{C-1,H-1}}$  172 Hz), 96.3 ( $^1J_{\text{C-1,H-1}}$  173 Hz), (C-1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>), 94.7 ( $^1J_{\text{C-1,H-1}}$  179 Hz) (C-1<sub>A</sub>), 60.7, 60.6 (C-6<sub>C</sub>, 6<sub>D</sub>), 52.1 (C-2<sub>C</sub>), 40.7 ( $\text{CH}_2\text{Cl}$ ), 22.2 ( $\text{CH}_3\text{CON}$ ), 20.5 ( $\text{CH}_3\text{COO}$ ), 17.6, 17.4 (C-6<sub>A</sub>, 6<sub>B</sub>). FABMS:  $m/z$  1546  $[\text{M}-\text{C}_2\text{HCl}_3\text{NO}]^+$ . Anal. Calcd for  $\text{C}_{83}\text{H}_{78}\text{O}_{29}\text{Cl}_4\text{N}_2$ : C, 58.32; H, 4.60; N, 1.64; Cl, 8.30. Found: C, 58.08; H, 4.69; N, 1.62; Cl, 8.22.

## REFERENCES

- (1) Pozsgay, V.; Glaudemans, C. P. J.; Robbins, J. B.; Schneerson, R. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 255.
- (2) (a) Chu, C. Y.; Liu, B.; Watson, D.; Szu, S.; Bryla, D.; Shiloach, J.; Schneerson, R.; Robbins, J. B. *Infect. Immun.* **1991**, *59*, 4450. (b) Chu, C. Y.; Schneerson, R.; Robbins, J. B. *Infect. Immun.* In press.
- (3) Chemical structure: (a) Dmitriev, B. A.; Knirel, Y. A.; Kochetkov, N. K. *Eur. J. Biochem.* **1976**, *66*, 559. Biological repeating unit: (b) Sturm, S.; Jann, B.; Jann, K.; Fortnagel, P.; Timmis, K. N. *Microbial Pathogenesis* **1986**, *1*, 307.
- (4) For a review on the conjugation methods see: Dick, Jr., W. E.; Beurret, M. *Contrib. Microbiol. Immunol.* **1989**, *10*, 48.
- (5) Robbins, J. B.; Schneerson, R. *J. Infect. Dis.* **1990**, *161*, 821.
- (6) Paoletti L. C.; Kasper, D. L.; Michon, F.; DiFabio, J.; Holme, K.; Jennings, H. J.; Wessels, M. R. *J. Biol. Chem.* **1990**, *265*, 18278.
- (7) Verheul, A. F. M.; Boons, G. J. P. H.; van der Marel, G. A.; van Boom, J. H.; Jennings, H. J.; Snippe, H.; Verhoef, J.; Hoogerhout, P.; Poolman, J. T. *Infect. Immun.* **1991**, *59*, 3566.
- (8) Wessels, M. R.; Kasper, D. L. *J. Exp. Med.* **1989**, *169*, 2121.
- (9) Wessels, M. R.; Pozsgay, V.; Kasper, D. L.; Jennings, H. J. *J. Biol. Chem.* **1987**, *262*, 8262.
- (10) By definition (*Eur. J. Biochem.* **1982**, *126*, 433) oligosaccharides consist of two to ten monosaccharide residues.
- (11) Pavliak, V.; Nashed, E.M.; Pozsgay, V.; Kovac, P.; Karpas, A.; Chu, C. Y.; Schneerson, R.; Robbins, J. B.; Glaudemans, C. P. J. *J. Biol. Chem.* Submitted.
- (12) For a recent review on the synthesis of oligosaccharide fragments of bacterial, O-specific polysaccharides see: Bundle, D. R. *Topics Curr. Chem.* **1990**, *154*, 1.
- (13) For syntheses of the rhamnobiase fragment see: (a) Pozsgay, V.; Brisson, J.-R.; Jennings, H. J. *Can. J. Chem.* **1987**, *65*, 2764. (b) Backinowsky, L. V.; Gomtsyan, A. R.; Byramova, N. E.; Kochetkov, N. K.; Yankina, N. F. *Bioorg. Khim.* **1985**, *11*, 1562. (c) Jaworska, A.; Zamojski, A. *Carbohydr. Res.* **1984**, *126*, 191. (d) Wessel, H. P.; Bundle, D. R. *Carbohydr. Res.* **1983**, *124*, 301. (e) Pozsgay, V.; Nanasi, P.; Neszmelyi, A. *Carbohydr. Res.* **1981**, *90*, 215. (f) Josephson, S.; Bundle, D. R. *J. Chem. Soc. Perkin I* **1980**, 297. (g) Schwarzenbach, D.; Jeanloz, R. W. *Carbohydr. Res.* **1980**, *81*, 323. (h) Liptak, A.; Nanasi, P.; Neszmelyi, A.; Wagner, H. *Tetrahedron*, **1980**, *36*, 1261. (i) Lafitte, C.; Du, A. M. N. P.; Winternitz, F.; Wylde, R.; Pratiel-Sosa, F. *Carbohydr. Res.* **1978**, *67*, 91.
- (14) For the synthesis of tri- and tetrasaccharide fragments see: Classon, B.; Garegg, P. J.; Hallgren, C. *Abstracts of the XIVth Int. Carbohydr. Symp.* **1988**, B90, Stockholm; Garegg, P. J.; Hallgren, C. *J. Carbohydr. Chem.* **1992**, *11*, 445.
- (15) Concurrently with these studies, syntheses of two tetrasaccharides related to A were described: (a) Pavliak, V.; Kovac, P.; Glaudemans, C. P. J. *Carbohydr. Res.* **1992**, *229*, 103. (b) Kovac, P.; Edgar, K. *J. Org. Chem.* **1992**, *57*, 2455.
- (16) Paulsen, H. *Angew. Chem.* **1982**, *94*, 184; *Angew. Chem. Int. Ed. Engl.* **1982**, *21*, 155 and references therein.
- (17) More recent applications of Paulsen's azide-technology include: (a) Vos, J. N.; Westerduin, P.; van Boeckel, C. A. A. *Bioorg. Med. Chem. Lett.* **1991**, *1*, 143. (b) Duchaussoy, P.; Lei, P. S.; Petitou, Duchaussoy, P.; Lederman, I.; Choay, J.; Sinay, P. *Carbohydr. Res.* **1988**, *179*, 163. (d) Chiba, T.; Jacquinet, J.-C.; Sinay, P.; Petitou, M.; Choay, J. *Carbohydr. Res.* **1988**, *174*, 253. (e) van Boeckel, C. A. A.; Basten, J. E. M.; Lucas, H.; van Aelst, S. F. *Angew. Chem.* **1988**, *100*, 1217; *Angew. Chem. Int. Ed. Engl.* **1988**, *27*, 1177. (f) Petitou, M.; Duchaussoy, P.; Lederman, I.; Choay, J.; Jacquinet, J.-C.; Sinay, P.; Torri, G. *Carbohydr. Res.* **1987**, *167*, 67. (g) Schmid, R. R. in *Stereochemistry of Organic and Bioorganic Transformations* (Bartmann, W.; Sharpless, B., Eds.) VCH Publishers, **1987**, p. 169. (h) Petitou, M.; Duchaussoy, P.; Lederman, I.; Choay, J.; Sinay, P.; Jacquinet, J.-C.; Torri, G. *Carbohydr. Res.* **1986**, *147*, 221. (i) van Boeckel, C. A. A.; Beetz, T.; Vos, J. N.; de Jong, A. J. M.; van Aelst, S. F.; van den Bosch, R. H.; Mertens, J. M. R.; van der Vlugt, F. A. J. *Carbohydr. Chem.* **1985**, *4*, 293.
- (18) Methods for the conversion of the azido group into an amino group at C-2 of glycosides include: (a) catalytic hydrogenation (e.g. Ref. 17h). (b) reduction with  $\text{NiCl}_2 \cdot \text{H}_3\text{BO}_3 \cdot \text{NaBH}_4$  (Paulsen, H.;

- Sinwell, V. *Chem. Ber.* **1978**, *111*, 869). (c) catalytic transfer hydrogenation with Pd/C-HCO<sub>2</sub>NH<sub>4</sub> (Ref. 17b). (d) reduction with triphenylphosphine (e. g. Classon, B.; Garegg, P. J.; Oscarson, P. J.; Tiden, A.-K. *Carbohydr. Res.* **1991**, *216*, 187). (e) reduction with H<sub>2</sub>S (e. g. Paulsen, H.; Lorentzen, J. P. *Carbohydr. Res.* **1986**, *150*, 63). (f) one-step conversion of the azido group into an acetamido group by AcSH (Rosen, T.; Lico, I. M.; Chu, D. T. W. *J. Org. Chem.* **1988**, *53*, 1580). (g) conversion into acetamido group with trioctylphosphine/AcOH (Ciommer, M.; Kunz, H. *Synlett*, **1991**, 593.)
- (19) (a) Vasella, A.; Witzig, C.; Chiara, J.-L.; Martin-Lomas, M. *Helv. Chim. Acta* **1991**, *74*, 2073. (b) Pavliak, V.; Kovac, P. *Carbohydr. Res.* **1991**, *210*, 333. (c) Bovin, N.V.; Zurabyan, S.E.; Khorlin, A. Y. *Carbohydr. Res.* **1981**, *98*, 25. For the synthesis of the racemate of compound 5, see: Lehmann, J.; Moritz, A. *Liebigs Ann. Chem.* **1991**, 937.
- (20) (a) Vos, J. N.; van Boom, J. H.; van Boeckel, C. A. A.; Beetz, T. J. *Carbohydr. Res.* **1984**, *3*, 117. (b) Kloosterman, M.; de Nijs, M. P.; van Boom, J. H. *J. Carbohydr. Chem.* **1986**, *5*, 215.
- (21) Klyashchitskii, B. A.; Strakhova, G. D.; Shvets, V. I.; Sokolov, S. D.; Preobrazhenskii, N. A. *Zh. Obshch. Khim.* **1970**, *40*, 236.
- (22) (a) Kovac, P. *Carbohydr. Res.* **1986**, *153*, 168. (b) Bovin, N. V.; Zurabyan, S. E.; Khorlin, A. Y. *Izv. Akad. Nauk SSSR, Ser. Khim.* **1981**, 1638. (c) Deferrari, J. O.; Gross, E. G.; Thiel, I. M. E. *Methods Carbohydr. Chem.* **1972**, *6*, 365. (d) Deferrari, J. O.; Gross, E. G.; Mastronardi, I. O. *Carbohydr. Res.* **1967**, *4*, 432.
- (23) The author thanks Professor K. Takeo for this suggestion.
- (24) Starting material for the previously described heterofunctional 2-azido-2-deoxy-D-glucosamine donors was the corresponding, 1,6-anhydro derivative. (e. g. (a) Paulsen, H.; Stenzel, W. *Chem. Ber.* **1978**, *111*, 2334; *ibid.* **1978**, *111*, 2348. (b) Ref. 17a.) The aggressive conditions of the acidolytic opening of the anhydro ring precluded the use of an acid-sensitive protecting-group scenario.
- (25) (a) Pozsgay, V.; Jennings, H. J. *Tetrahedron Lett.* **1987**, *28*, 1375. (b) Pozsgay, V.; Jennings, H. J. *J. Org. Chem.* **1988**, *53*, 4042. (c) Pozsgay, V.; Jennings, H. J. *Carbohydr. Res.* **1988**, *179*, 61. (d) Kameyama, A.; Ishida, H.; Kiso, M.; Hasegawa, A. *Carbohydr. Res.* **1989**, *193*, c1.
- (26) (a) Oikawa, Y.; Yoshioko, T.; Yonemitsu, O. *Tetrahedron Lett.* **1982**, *23*, 885. (b) Johansson R.; Samuelsson, B. *J.C.S. Perkin I.* **1984**, 2371.
- (27) Pozsgay, V.; Jennings, H. J.; Kasper, D. L. *J. Carbohydr. Chem.* **1987**, *6*, 41.
- (28) Ness, R. K.; Fletcher, H. G.; Hudson, C. S. *J. Am. Chem. Soc.* **1951**, *73*, 296.
- (29) Pozsgay, V. *Carbohydr. Res.* In press.
- (30) King, J. F.; Allbutt, A. D. *Can. J. Chem.* **1970**, *48*, 1754.
- (31) Applications of the method of King and Allbutt (Ref. 30) in the carbohydrate field include: (a) Lemieux, R. U.; Driguez, H. *J. Am. Chem. Soc.* **1975**, *97*, 4069. (b) Garegg, P. J.; Hultberg, H. *Carbohydr. Res.* **1979**, *72*, 276. (c) Wessel, H.-P.; Bundle, D. *Carbohydr. Res.* **1983**, *124*, 301. (d) Wessel, H.-P.; Bundle, D. *J. Chem. Soc. Perkin I*, **1985**, 2251. (e) Garegg, P. J.; Oscarson, S. *Carbohydr. Res.* **1895**, *136*, 207. (f) Paulsen, H.; Hasenkamp, T.; Paal, M. *Carbohydr. Res.* **1985**, *144*, 45. (g) Tsvetkov, Y. E.; Backinowsky, L. V.; Kochetkov, N. K. *Carbohydr. Res.* **1989**, *193*, 75. (h) Veeneman, G. H.; van Leeuwen, S. H.; Zuurmond, H. M.; van Boom, J. H. *J. Carbohydr. Chem.* **1990**, *9*, 783. (i) Auzanneau, F.-I.; Bundle, D. R. *Carbohydr. Res.* **1991**, *212*, 13.
- (32) Bertolini, M. J.; Glaudemans, C. P. J. *Carbohydr. Res.*, **1970**, *15*, 263.
- (33) (a) Lonn, H. *Carbohydr. Res.* **1985**, *139*, 105. (b) Lonn, H. *J. Carbohydr. Chem.* **1987**, *6*, 301.
- (34) (a) Weygand, F.; Ziemann, H. *Ann. Chem.* **1962**, *657*, 179. (b) See also: Horton, D.; Hutson, D. H. *Adv. Carbohydr. Chem.* **1963**, *18*, 123.
- (35) (a) Kihlberg, J.; Leigh, D. A.; Bundle, D. R. *J. Org. Chem.* **1990**, *55*, 2860. (b) Kihlberg, J.; Eichler, E.; Bundle, D. R. *Carbohydr. Res.* **1991**, *211*, 59. (c) Kihlberg, J.; Bundle, D. R. *Carbohydr. Res.* **1991**, *216*, 67. (d) Ref. 31 (i).
- (36) Andersson, F.; Fugedi, P.; Garegg, P. J.; Nashed, M. *Tetrahedron Lett.* **1986**, *27*, 3919.
- (37) Jansson, K.; Ahlfors, S.; Frejd, T.; Kihlberg, J.; Magnusson, G. *J. Org. Chem.* **1988**, *53*, 5629.
- (38) Schmidt, R. R. *Angew. Chem.* **1986**, *98*, 213. *Angew. Chem. Int. Ed. Engl.* **1986**, *25*, 212.
- (39) Hamacher, K. *Carbohydr. Res.* **1984**, *128*, 291.

#### Acknowledgements.

We thank Dr. Lewis Pannell and Mr. Noel Whittaker for the mass spectra.