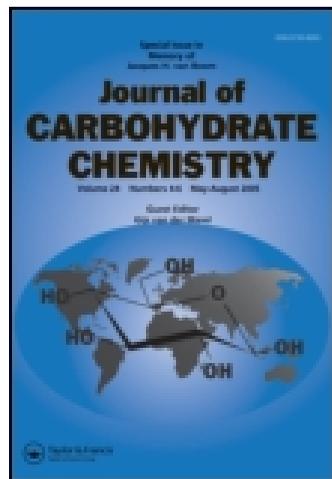


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FeCl₃·6H₂O/C: An Efficient and Recyclable Catalyst for the Synthesis of 2,3-Unsaturated O- and S-Glycosides

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A novel method for synthesizing 2,3-unsaturated glycosides has been developed using a handy and eco-friendly immobilized catalyst, FeCl₃·6H₂O/C. A series of 2,3-unsaturated O- and S-glycosides were obtained for bioassay from corresponding 3,4,6-tri-O-acetyl-D-glucal and D-galactal in good to excellent yields (56%–99%) and high anomeric selectivity ($\alpha/\beta = 7:1$ to $>19:1$). Furthermore, the catalyst was efficient on gram-scale reactions and recyclable for at least three times.

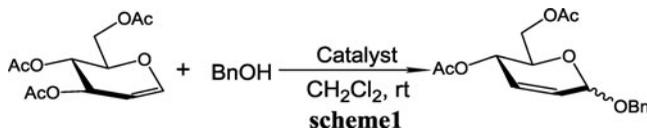
Keywords FeCl₃·6H₂O/C; Tri-O-acetyl-D-glycals; 2,3-Unsaturated glycosides; Ferrier-rearrangement; Recyclable

INTRODUCTION

2,3-Unsaturated glycosides as chiral intermediates have played an important role in the synthesis of bioactive compounds, such as oligosaccharides,^[1] glycopeptide building blocks,^[2] modified carbohydrates,^[3] nucleosides,^[4] and antibiotics.^[5] Besides, the unsaturated part of the sugar ring allows many straightforward modifications such as hydroxylation, oxidation, epoxidation, and hydrogenation, which contributes to their diversity and complexity.^[6–11] As we know, the most valuable method for preparing 2,3-unsaturated glycosides is Lewis acid-catalyzed allelic rearrangement of glycals, which is well known as the Ferrier rearrangement reaction.^[12,13] Up to now, a wide range of Lewis acids and oxidizing agents, such as BF₃·OEt₂,^[12] IDCP,^[14]

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Table 1: Optimization of reaction conditions for the synthesis of 2,3-unsaturated glucosides

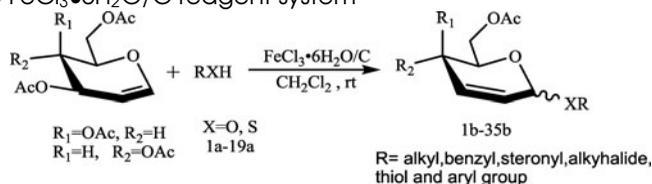
Entry	Conditions	Yield (%)
1	FeCl ₃ ·6H ₂ O/C (0.01eq), 240min	45
2	FeCl ₃ ·6H ₂ O/C (0.02eq), 40min	95
3	FeCl ₃ ·6H ₂ O/C (0.05eq), 10min	99
4	FeCl ₃ ·6H ₂ O/C (0.05eq), 10min	98
5	FeCl ₃ ·6H ₂ O/SiO ₂ (0.05eq), 120min	80
6	FeCl ₃ ·6H ₂ O/4 ÅMS (0.05eq), 120min	traces

TMSOTf,^[15] I₂,^[16] Sc(OTf)₃,^[17] InCl₃,^[18] Yb(OTf)₃,^[19] BiCl₃,^[20] Dy(OTf)₃ immobilized in ionic liquids,^[21] ZnCl₂,^[22] HClO₄-SiO₂,^[23] NiCl₅,^[224] Er(OTf)₃,^[25] ZnCl₂/Al₂O₃,^[26] H₂SO₄-SiO₂,^[27] NaHSO₄-SiO₂,^[28] and CF₃SO₃H-SiO₂,^[29] have been employed for this transformation. However, the present methods for the synthesis of 2,3-unsaturated glycosides possess some drawbacks, such as use of expensive and toxic reagents and/or stoichiometric amount of catalyst, low efficiency, and complicated operation.

Recently, iron (III) catalysts have been widely used in organic synthesis,^[30,31] and some of these types of catalysts, such as FeCl₃,^[32] Fe₂(SO₄)₃·xH₂O,^[33] FeCl₃ immobilized in ionic liquids,^[34] and Fe(OTf)₃,^[35] have been successfully utilized for the Ferrier rearrangement. In our previous research for diverse syntheses of glycosides, we have successfully used immobilized catalysts for Ferrier rearrangement.^[27,36] The present work aimed at developing a new and reusable immobilized iron (III) catalyst for Ferrier rearrangement.

Initially, we screened different carriers, including carbon, SiO₂, and molecular sieves, to immobilize iron (III) chloride hexahydrate (FeCl₃·6H₂O) according to the literature method.^[37,38] We found that FeCl₃·6H₂O/C was the most promising for catalyzing the reaction between 3,4,6-tri-*O*-acetyl-*D*-glucal and benzyl alcohol, because other immobilized iron (III) catalysts, including FeCl₃·6H₂O/SiO₂ and FeCl₃·6H₂O/4 ÅMS, could only give the desired product either in longer time or poorer yield (Table 1). Therefore, FeCl₃·6H₂O/C was explored in detail to establish the proper reaction conditions. We found that 0.05 equiv FeCl₃·6H₂O/C was sufficient to promote the reaction and gave an excellent yield (99%) in 10 min at rt using CH₂Cl₂ as solvent. When a smaller amount of catalyst was used, the reaction proceeded much slower, while the reaction rate and yield were not improved with a larger amount of catalyst.

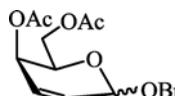
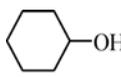
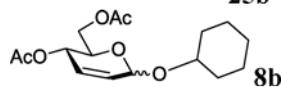
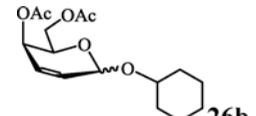
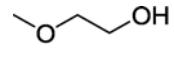
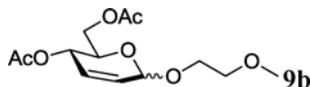
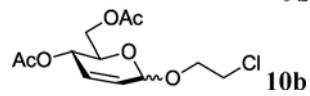
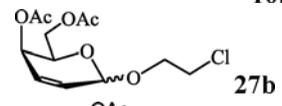
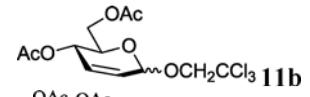
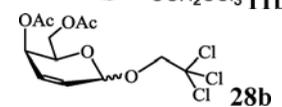
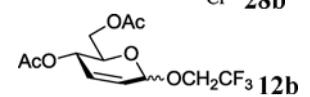
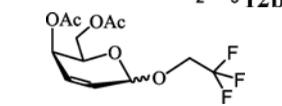
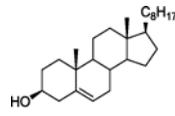
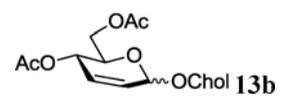
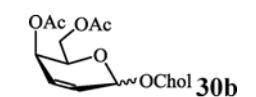
With the optimal conditions established, we next investigated the reaction scope using a variety of acceptors. As shown in Table 2, glycosidation of

Table 2: Ferrier reaction of 3, 4, 6-tri-O-acetyl-D-glycal with acceptors in the presence of the FeCl₃·6H₂O/C reagent system

Entry	Acceptors	Products	Time (h)	Yield (%) ^a	α/β ratios ^b
1	CH ₃ (CH ₂)CH ₂ OH 1a		1.5	93 ⁽³⁶⁾	7.5:1
2			1.5	93 ⁽⁴⁰⁾	>19:1
3	CH ₃ (CH ₂) ₆ CH ₂ OH 2a		1.5	97 ⁽²⁵⁾	7:1
4			2.5	85 ⁽²⁵⁾	>19:1
5	(CH ₃) ₂ CHOH 3a		1	90 ⁽³⁶⁾	9:1
6			2	82 ⁽²⁴⁾	>19:1
7			2	88 ⁽³⁶⁾	15:1
8			1.5	86 ⁽³⁶⁾	7:1
9			2	89 ⁽²⁴⁾	>19:1
10			5.5	75 ⁽³⁶⁾	7:1
11			4	87 ⁽²⁾	>19:1
12	PhCH ₂ OH 7a		10 min	99 ⁽³⁶⁾	8:1

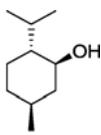
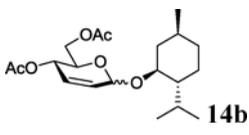
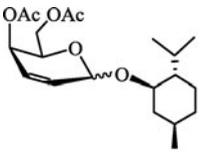
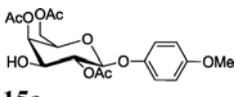
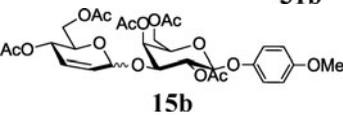
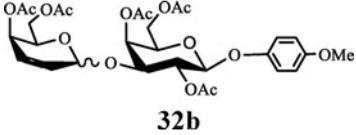
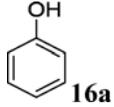
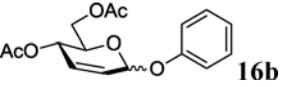
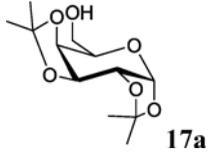
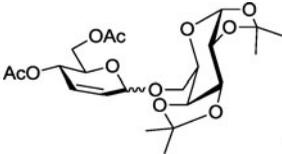
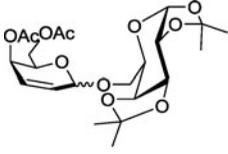
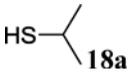
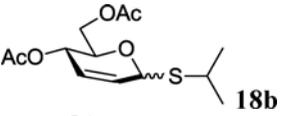
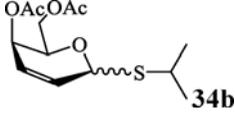
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Table 2: Ferrier reaction of 3, 4, 6-tri-*O*-acetyl-*D*-glyceral with acceptors in the presence of the FeCl₃•6H₂O/C reagent system (Continued)

Entry	Acceptors	Products	Time (h)	Yield (%) ^a	α/β ratios ^b
13		 25b	40 min	92 ⁽²⁾	> 19:1
14	 8a	 8b	5.5	88 ⁽³⁶⁾	10:1
15		 26b	2.5	82 ⁽⁴¹⁾	> 19:1
16	 9a	 9b	4.5	85 ⁽³⁶⁾	8:1
17	HOCH₂CH₂Cl 10a	 10b	1.5	94 ⁽³⁶⁾	7.5:1
18		 27b	0.5	85 ⁽⁴²⁾	> 19:1
19	Cl₃CCH₂OH 11a	 11b	1	76 ⁽³⁶⁾	9:1
20		 28b	1.5	80	> 19:1
21	F₃CCH₂OH 12a	 12b	1	69	7:1
22		 29b	1	61	> 19:1
23	 13a	 13b	2.5	87 ⁽³⁶⁾	18:1
24		 30b	1.5	68	> 19:1

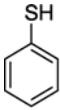
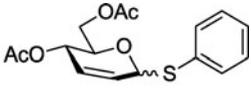
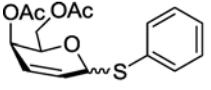
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Table 2: Ferrier reaction of 3, 4, 6-tri-*O*-acetyl-*D*-glyceral with acceptors in the presence of the FeCl₃·6H₂O/C reagent system (Continued)

Entry	Acceptors	Products	Time (h)	Yield (%) ^a	α/β ratios ^b
25			1	84 ⁽⁴⁰⁾	>19:1
26	14a		1.5	67	>19:1
27			1	74	>19:1
28	15a		1.5	57	>19:1
29			0.5	56 ^{c(21)}	7:1
30			3.5	82 ⁽⁴³⁾	7:1
31	17a		3	66 ⁽⁴³⁾	>19:1
32			0.5	91 ⁽⁴⁴⁾	9:1
33	18a		0.5	75 ⁽⁴⁴⁾	>19:1

(Continued on next page)

Table 2: Ferrier reaction of 3, 4, 6-tri-*O*-acetyl-*D*-glyceral with acceptors in the presence of the FeCl₃•6H₂O/C reagent system (Continued)

Entry	Acceptors	Products	Time (h)	Yield (%) ^a	α/β ratios ^b
34	 19a	 19b	40 min	88 ⁽⁴⁴⁾	7:1
35		 35b	0.5	86 ⁽²⁴⁾	> 19:1

^a Isolated yields.^b α/β ratios were determined by ¹H NMR.^c The dosage of FeCl₃•6H₂O/C is 0.001 eq.

tri-*O*-acetyl-*D*-glucal with primary, secondary, benzyl, allyl, propargyl, and halogenated alcohols; thioalcohol; and thiophenol proceeded smoothly under the specified conditions. The reaction afforded 2,3-unsaturated glucosides in good to excellent yields (69%–99%) with good anomeric selectivity ($\alpha/\beta > 7:1$). Comparing the results among entry 17, entry 19, and entry 21, it is clear that the reaction rate became slower and the yield was decreased with the reduction in the nucleophilicity of acceptors. Additionally, complex alcohols such as cholesterol also produced a high yield of product (87%, entry 23). Moreover, we could obtain disaccharide smoothly while using sugar-derived alcohols as the acceptor (entries 27 and 30). When the dosage of FeCl₃•6H₂O/C reduced to 0.001 equiv, we successfully obtained the phenol glycoside in 56% yield (entry 29), which is hardly synthesized with other catalyst systems.^[32,33]

However, the reaction between tri-*O*-acetyl-*D*-galactal and benzyl alcohol needed harsher conditions. We found that 0.5 equiv FeCl₃•6H₂O/C was required to promote the glycosylation with 92% yield and only α anomeric selectivity. Likewise, a number of 2,3-unsaturated galactosides were obtained successfully from primary, secondary, benzyl, allyl, propargyl, and halogenated alcohols; thioalcohol; thiophenol; cholesterol; and sugar-derived alcohols, with sole α anomeric selectivity, which probably resulted from the steric and anomeric effects.

The structure and stereochemistry of all glycosidation products were determined through spectroscopic analysis and also by comparison with the reported data.^[39–44] In vitro antitumor activity of some of the compounds was evaluated against the human k562 cancer cell line by the standard MTT assay.^[45] They were proven to possess moderate antitumor activity as shown in Table 3.

More significantly, this synthetic methodology was shown to be reliable on a 2 g scale of tri-*O*-acetyl-*D*-glucal in the presence of benzyl alcohol to give

Table 3: Inhibition of resistant K562 cell proliferation by some glycosidation products

Compound	Sample concentration (μg/mL)	Inhibition rate (%)
DDP	10	60.1
9b	100	23.2
13b	100	11.5
22b	100	24.0
31b	100	23.1

DDP: *cis*-dichlorodiamminoplatinum-II.

an excellent yield (95%), and the catalyst could be easily recovered by simple filtration and drying at 120°C for 3 h. Furthermore, over three cycles, the catalyst did not show any significant loss in activity.^[32,39]

In summary, we have developed a mild and environmentally friendly iron (III)-catalyzed glycosylation method for synthesizing 2,3-unsaturated glucosides and 2,3-unsaturated galactosides in high anomeric selectivity. This method was applied to a wide range of acceptors, including primary, secondary, benzyl, allyl, propargyl, and halogenated alcohols; thioalcohol; thiophenol; cholesterol; and sugar derivatives. Furthermore, the efficient applicability of this methodology to gram scales of synthesis and the recyclability of the catalyst make this approach highly practical.

EXPERIMENTAL

General Experimental Methods

¹H NMR spectra and ¹³C NMR spectra were recorded on a Bruker DRX-500 MHz spectrometer using tetramethylsilane as internal standard and CDCl₃ as solvent. Mass spectra were determined on LTQ-XL (Thermo Scientific, USA) with an (ESI) ion trap mass spectrometer. Silica gel (10–40 μm, Yantai, China) was used for column chromatography. TLC plates (10–40 μm, Yantai, China) were applied to monitor the reactions. All the reagents were purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China) and were used as received without further purification. The supported ferric catalysts were prepared according to the reported methods.^[37,38]

Preparation of FeCl₃·6H₂O/C Catalyst

The preparation of FeCl₃·6H₂O/C followed the literature method.^[38] In brief, 1 g of iron (III) chloride hexahydrate, 1 g of activated carbon, and 4.0 mL of absolute ethanol were added to a round-bottom flask. Then, the mixture was

refluxed for 30 min. Finally, the solvent was evaporated under reduced pressure and the resulting catalyst was baked in an oven at 120°C for 2 h.^[46]

General Experimental Procedure for the Synthesis of 2,3-Unsaturated Glucosides

To a stirred solution of tri-*O*-acetyl-*D*-glucal (100 mg, 0.37 mmol) in DCM (1 mL) were added the corresponding alcohol (0.41 mmol, 1.1 equiv) and FeCl₃·6H₂O/C (5 mol%) at ambient temperature. After the reaction was completed (monitored by TLC), the reaction mixture was filtered and the catalyst was washed with dichloromethane. After evaporation of the solvent under vacuum, the crude products were purified by silica gel column chromatography (petroleum ether/EtOAc = 10/1).

General Experimental Procedure for the Synthesis of 2,3-Unsaturated Galactosides

To a stirred solution of tri-*O*-acetyl-*D*-galactal (50 mg, 0.185 mmol) in DCM (1 mL) were added the corresponding alcohol (0.204 mmol, 1.1 equiv) and FeCl₃·6H₂O/C (50 mol%) at ambient temperature. After the reaction was completed (monitored by TLC), the reaction mixture was filtered and the catalyst was washed with dichloromethane. After evaporation of the solvent under vacuum, the crude products were purified by silica gel column chromatography (petroleum ether/EtOAc = 10/1).

Spectral Data for New Compounds

Trifluoroethyl 4,6-di-O-acetyl-2,3-dideoxy-D-erythro-hex-2-enopyranoside (12b)

Viscous oil; $\alpha/\beta = 7:1$; ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.06–2.09 (m, 6 H), 3.95–4.09 (m, 3 H), 4.18–4.20 (m, 1H), 5.09 (s, 1 H), 5.31 (dd, 1 H, $J = 1.4, 9.8$ Hz), 5.82–5.94 (m, 2 H). ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 20.60, 20.82, 62.51, 64.78, 64.85, 65.06, 67.39, 94.47, 126.19, 130.34, 170.15, 170.63. MS (ESI): m/z calculated for [M+Na]⁺ C₁₂H₁₅F₃O₆Na 335.07, found 335.17. ESI-HRMS: Calcd for C₁₂H₁₅F₃O₆Na (M+Na⁺) 335.0713, found 335.0731.

2,4,6-tri-O-acetyl mequinol galactosyl-4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (15b)

Viscous oil; $[\alpha]_D^{20} = +108$ (c 0.23, CH₂Cl₂); α only; ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.05–2.18 (m, 15H), 3.78 (s, 3H), 3.92–3.95 (m, 2H), 4.10–4.21 (m, 3 H), 4.24–4.28 (m, 2H), 4.83–4.85 (d, 1H), 5.24 (s, 1H), 5.32–5.39 (m, 1H), 5.44–5.45 (d, 1 H, $J = 2.9$ Hz), 5.63–5.66 (m, 1H), 5.85–5.87 (m, 1H), 6.81–6.83

(dd, $J = 2.2, 6.9$ Hz), 6.94–6.96 (dd, $J = 2.2, 6.9$ Hz, 2H). ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 14.09, 20.65, 20.78, 20.80, 20.92, 22.67, 29.34, 29.68, 31.91, 55.65, 61.55, 62.51, 64.96, 65.30, 67.29, 69.67, 70.81, 73.30, 91.05, 101.09, 114.55, 118.58, 127.04, 129.34, 151.11, 155.71, 169.16, 170.14, 170.37, 170.65. MS (ESI): m/z calculated for [M+Na]⁺ C₂₉H₃₆O₁₅Na 647.20, found 647.33. ESI-HRMS: Calcd for C₂₉H₃₆O₁₅Na (M+Na⁺) 647.1946, found 647.1990.

Trichloroethyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-derythro-hex-2-enopyranoside
(28b)

Yellow oil; $[\alpha]_D^{20} = -130$ (c 0.16, CH₂Cl₂); α only; ¹H NMR (500 MHz, CDCl₃): δ (ppm) 6.20 (1H, dd, $J = 10.0, 5.5$ Hz), 6.12 (1H, dd, $J = 10.0, 3.0$ Hz), 5.30 (1H, d, $J = 2.5$ Hz), 5.05 (1H, dd, $J = 5.0, 2.0$ Hz), 4.45–4.42 (1H, m), 4.33–4.31 (1H, m), 4.23–4.17 (3H, m), 2.07 (3H, s), 2.06 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 170.5, 170.2, 129.2, 126.2, 96.4, 94.1, 79.3, 67.5, 62.6, 62.5, 20.7 (2C). MS (ESI): m/z calculated for [M+Na]⁺ C₁₂H₁₅Cl₃O₆Na 382.98, found 383.08. ESI-HRMS: Calcd for C₁₂H₁₅Cl₃O₆Na (M+Na⁺) 382.9826, found 382.9858.

Trifluoroethyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-derythro-hex-2-enopyranoside
(29b)

Viscous liquid; $[\alpha]_D^{20} = -104$ (c 0.14, CH₂Cl₂); α only; ¹H NMR (500 MHz, CDCl₃) δ (ppm): 6.21 (1H, dd, $J = 10.0, 5.5$ Hz), 6.07 (1H, dd, $J = 10.0, 3.0$ Hz), 5.16 (1H, d, $J = 2.5$ Hz), 5.05 (1H, dd, $J = 5.5, 2.0$ Hz), 4.34–4.33 (1H, m), 4.28–4.21 (2H, m), 4.20–3.96 (2H, m), 2.09 (3H, s), 2.08 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 170.5, 170.2, 129.1, 126.2, 94.0, 67.4, 64.6, 64.4, 62.6, 62.4, 20.7, 20.6. MS (ESI): m/z calculated for [M+Na]⁺ C₁₂H₁₅F₃O₆Na 335.07, found 335.17. ESI-HRMS: Calcd for C₁₂H₁₅F₃O₆Na (M+Na⁺) 335.0713, found 335.0727.

Cholesteryl 4,6-di-O-acetyl-2,3-dideoxy- α -D-threo-hex-enopyranoside (30b)

White solid; $[\alpha]_D^{20} = -45$ (c 0.08, CH₂Cl₂); α only; ¹H NMR (500 MHz, CDCl₃) δ (ppm): 6.09 (1H, dd, $J = 10.0, 5.4$ Hz), 6.01 (1H, dd, $J = 10.0, 3.0$ Hz), 5.34 (1H, d, $J = 4.9$ Hz), 5.20 (1H, d, $J = 2.6$ Hz), 5.00 (1H, dd, $J = 5.2, 2.5$ Hz), 4.40–4.39 (1H, m), 4.22–4.20 (2H, m), 3.54–3.59 (1H, m), 2.43–2.39 (1H, m), 2.34–2.29 (1H, m), 2.06 (3H, s), 2.05 (3H, s), 2.01–0.66 (41H, m); ¹³C NMR (125 MHz, CDCl₃) δ (ppm): 170.6, 170.3, 130.7, 124.7, 95.6, 80.7, 66.6, 63.2, 63.0, 48.9, 43.2, 34.3, 31.7, 25.6, 23.1, 22.4, 21.1, 20.8 (2C), 16.1. MS (ESI): m/z calculated for [M+Na]⁺ C₃₇H₅₈O₆Na 621.43, found 621.50. ESI-HRMS: Calcd for C₃₇H₅₈O₆Na (M+Na⁺) 621.4126, found 621.4141.

Methyl 4,6-di-O-acetyl-2,3-dideoxy- α -D-threo-hex-2-enopyranoside (31b)

White solid; $[\alpha]_{\text{D}}^{20} = -159$ (c 0.16, CH_2Cl_2); α only; ^1H NMR (500 MHz, CDCl_3) δ (ppm): 6.09 (1H, dd, $J = 10.1, 5.2$ Hz), 6.04 (1H, dd, $J = 10.1, 2.9$ Hz), 5.13 (1H, d, $J = 2.6$ Hz), 5.00 (1H, dd, $J = 5.2, 2.5$ Hz), 4.41–4.38 (1H, m), 4.22–4.19 (2H, m), 3.45–3.40 (1H, m), 2.19 (1H, m), 2.07 (3H, s), 2.06 (3H, s), 1.65–0.65 (17H, m); ^{13}C NMR (125 MHz, CDCl_3) δ (ppm): 170.6, 170.3, 130.7, 124.7, 95.6, 80.7, 66.6, 63.2, 63.0, 48.9, 43.2, 34.3, 31.7, 25.6, 23.1, 22.4, 21.1, 20.8(2C), 16.1. MS (ESI): m/z calculated for $[\text{M}+\text{Na}]^+ \text{C}_{20}\text{H}_{32}\text{O}_6\text{Na}$ 391.21, found 391.33. ESI-HRMS: Calcd for $\text{C}_{20}\text{H}_{32}\text{O}_6\text{Na}$ ($\text{M}+\text{Na}^+$) 391.2091, found 391.2100.

2,4,6-tri-O-acetyl mequinol galactosyl-4,6-di-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (33b)

Viscous liquid; $[\alpha]_{\text{D}}^{20} = -47$ (c 0.33, CH_2Cl_2); α only; ^1H NMR (500 MHz, CDCl_3) δ (ppm): 6.95–6.94 (2H, d, $J = 9.0$ Hz), 6.95–6.94 (2H, d, $J = 9.1$ Hz), 6.10–6.07 (1H, m), 5.86–5.83 (1H, dd, $J = 10.1, 3.0$ Hz), 5.47–5.46 (1H, d, $J = 3.1$ Hz), 5.38–5.34 (1H, m), 5.03–5.01 (1H, dd, $J = 5.7, 1.9$ Hz), 4.85–4.83 (1H, d), 4.28–4.22 (2H, m), 4.20–4.17 (2H, m), 4.14–4.11 (2H, m), 3.97–3.96 (1H, m), 3.99–3.97 (1H, m), 3.78 (3H, s), 2.18–2.05 (15H, m). ^{13}C NMR (125 MHz, CDCl_3) δ (ppm): 170.65, 170.38, 170.30, 170.26, 169.26, 155.68, 151.10, 129.92, 125.28, 118.55, 114.53, 101.09, 90.44, 72.78, 70.82, 69.60, 67.16, 65.21, 62.26, 62.23, 61.61, 60.36, 55.62, 29.66, 20.75, 20.73, 20.63, 14.16. MS (ESI): m/z calculated for $[\text{M}+\text{Na}]^+ \text{C}_{29}\text{H}_{36}\text{O}_{15}\text{Na}$ 647.20, found 647.33. ESI-HRMS: Calcd for $\text{C}_{29}\text{H}_{36}\text{O}_{15}\text{Na}$ ($\text{M}+\text{Na}^+$) 647.1946, found 649.1974.

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46. XRD experiments were attempted to characterize our catalyst but failed with the low dosage of Ferric ion in the catalyst. The baking operation was used to remove the ethanol residue in the catalyst; otherwise, it might affect the reaction as a glycosyl acceptor.