Tetrahedron 67 (2011) 3724-3732

Contents lists available at ScienceDirect

Tetrahedron



journal homepage: www.elsevier.com/locate/tet

Axially chiral phosphine—oxazoline ligands in silver(I)-catalyzed asymmetric Mannich reaction of *N*-Boc aldimines with trimethylsiloxyfuran

Qian-Yi Zhao^a, Min Shi^{a,b,*}

^a Key Laboratory for Advanced Materials and Institute of Fine Chemicals, School of Chemistry and Molecular Engineering, East China University of Science and Technology, 130 Mei Long Road, Shanghai 200237, China

^b State Key Laboratory of Organometallic Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 354 Fenglin Road, Shanghai 200032, China

ARTICLE INFO

Article history: Received 28 January 2011 Received in revised form 14 March 2011 Accepted 17 March 2011 Available online 24 March 2011

Keywords: N-Boc aldimines Asymmetric Mannich reaction Axially chiral phosphine—oxazoline ligands Silver acetate Trimethylsiloxyfuran

1. Introduction

Catalytic asymmetric vinylogous Mannich (AVM)-type reaction of trimethylsiloxyfuran with aldimines has been proved to be a very powerful synthetic protocol to prepare chiral γ -butenolide derivatives bearing an amino functional group in recent years.¹ Many successful examples have been reported thus far. For example, Martin and Lopez reported the first example of catalytic asymmetric addition of trialkylsilyloxyfurans to aldimines in 1999, affording the corresponding adducts in moderate ee value along with high yield.^{1a} A few years late, Hoveyda and Snapper have developed a elegant asymmetric catalytic system with silver(I)-based catalyst using a 2-methyloxyphenyl group as aldimine substituent, leading to the corresponding product of AVM (asymmetric vinylogous Mannich) reaction of trimethylsiloxyfuran with aldimine in excellent diastereo- and enantioselectivity.^{1b,c} Later on, a new highly diastereo- and enantioselective catalytic asymmetric vinylogous Mannich-type reaction system applicable to a wide range of aldimines, which do not possess a 2-methyloxyphenyl group, with siloxyfuran using an axially chiral phosphine—oxazoline ligand [(R,S)-P-Oxa-Ph]/AgOAc/CF₃CH₂OH combination has been developed by our group.^{1e} During our ongoing investigation on this interesting asymmetric catalytic system, we attempted to explore the AVM reaction of trimethylsiloxyfuran with N-Boc (^tBuOC(O))-protected aldimine,

ABSTRACT

Axially chiral phosphine–oxazoline ligand **L6** was found to be a fairly effective chiral ligand in silver(I)catalyzed asymmetric Mannich reaction of *N*-Boc aldimines with trimethylsiloxyfuran to give the corresponding adducts in up to 97% yield, 7:1 dr and 86% ee (major diastereoisomer).

© 2011 Elsevier Ltd. All rights reserved.

which has not yet been developed. This is because asymmetric addition of nucleophiles to *N*-Boc-protected imines has provided a straightforward synthetic approach to chiral amines since its precursor (BocNH₂) as well as the *N*-Boc protected imine are easily available and the *N*-protecting group (*N*-Boc) can be readily cleaved under several convenient acidic conditions (HCl or TFA).² In this paper, we wish to report that axially chiral phosphine—oxazoline ligand **L6** derived from (*R*)-binol is a fairly effective catalyst in the silver(I)catalyzed asymmetric Mannich reaction of *N*-Boc aldimines with trimethylsiloxyfuran under mild conditions, providing the corresponding chiral *N*-Boc-protected γ -butenolides in moderate to good enantiomeric excesses and good to high yields as well as good diastereoselectivities.

2. Results and discussion

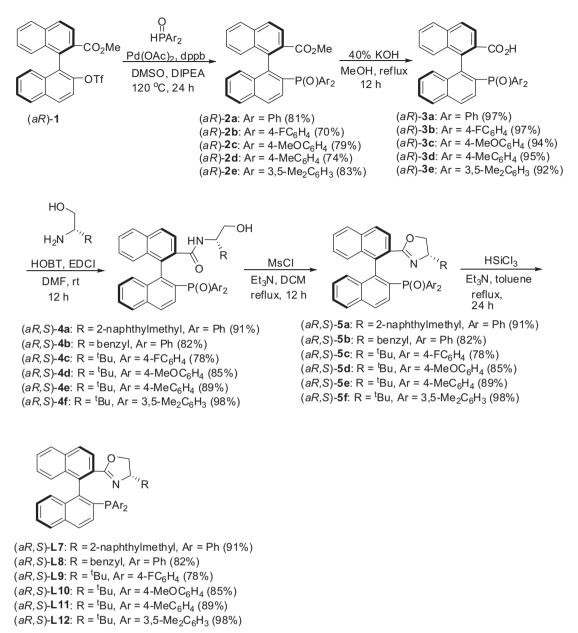
Axially chiral phosphine–oxazoline ligands **L1–L6** are known compounds and were synthesized according to the previous literature.^{1e,3} The new chiral phosphine–oxazoline ligands **L7–L12** were synthesized from (**a**R)-**1**, which is derived from (R)-binol⁴ and their synthetic route has been shown in Scheme 1. The spectroscopic data of these new chiral ligands are indicated in the experimental section.

With these above chiral ligands in hand, we set out to utilize the Lewis acid AgOAc (10 mol %)⁵ combined with chiral phosphine–oxazoline ligands **L1–L12** (10 mol %) (Fig. 1) as the catalysts



^{*} Corresponding author. Fax: +86 21 64166128; e-mail address: mshi@mail. sioc.ac.cn (M. Shi).

^{0040-4020/\$ –} see front matter @ 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2011.03.046



Scheme 1. Reaction procedure for the preparation of ligands L7-L12.

and the AVM reaction of readily available *N*-Boc aldimine **1a** (0.15 mmol) with siloxyfuran **2** (0.27 mmol) as a model reaction in dichloromethane (CH₂Cl₂) containing 0.27 mmol (1.8 equiv) of CH₃CH₂OH^{1d,e,5a} to develop the optimal reaction conditions and the results of these experiments are summarized in Table 1. It was

found that (aR,S)-*P*-Oxa-^tBu (**L6**) is the best chiral ligand in this reaction to afford the corresponding product **3a** in 97% yield and 86% ee for the major diastereoisomer (dr=6:1) in CH₂Cl₂ at -78 °C for 24 h (Table 1, entry 6). Other chiral ligands **L1**–**L5** and **L7–L12** are not as effective as **L6** in this reaction under identical conditions

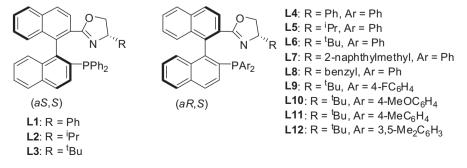
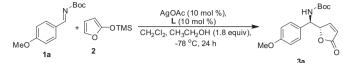


Fig. 1. Axially chiral phosphine-oxazoline ligands L1-L12.

3726

Table 1

Optimization of chiral ligands for the AVM reaction of N-Boc aldimine $\mathbf{1a}$ and siloxyfuran $\mathbf{2}$



Entry ^a	Ligand	Yield [%] ^b	anti:syn ^c	anti, ee [%] ^d
		3a	3a	3a
1	L1	90	>20:1	47
2	L2	79	7:1	53
3	L3	82	5:1	52
4	L4	98	>20:1	46
5	L5	99	>20:1	51
6	L6	97	6:1	86
7	L7	89	9:1	61
8	L8	94	9:1	68
9	L9	85	5:1	65
10	L10	88	6:1	80
11	L11	90	7:1	79
12	L12	92	4:1	79

^a Reaction conditions: **1a** (0.15 mmol), **2** (0.27 mmol), CH₃CH₂OH (0.27 mmol), AgOAc (10 mol %), Iigand (10 mol %), CH₂Cl₂ (1.0 mL), and the reaction was carried out at -78 °C for 24 h.

^b Yields of the purified *syn* and *anti* product.

^c Determined by ¹H NMR spectroscopic data of the crude product.

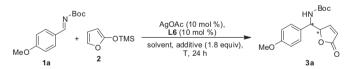
^d Determined by chiral HPLC analysis.

(Table 1, entries 1–5 and 7–12) although excellent yields and drs were achieved when **L1**, **L4**, and **L5** were used as the chiral ligands in this reaction (Table 1, entries 1, 4, and 5).

The effects of temperature, solvents, and additives were then investigated using the best chiral (aR,S)-P-Oxa- t Bu ligand **L6** in the model reaction of **1a** with **2**, the results are outlined in Table 2 combined with the result obtained in DCM for comparison (Table 2, entry 5). It can be seen from Table 2, increasing the reaction temperature from -78 °C to room temperature (20 °C) caused the decrease in enantioselectivity (from 86% to 61%) although excellent

Table 2

Optimization of the temperature, solvent, and additive for the AVM reaction of N-Boc aldimine **1a** and siloxyfuran **2**



Entry ^a	T [°C]	Solvent	Additive	Yield [%] ^b	anti:syn ^c	anti, ee [%] ^d
				3a	3a	3a
1	rt	DCM	CH ₃ CH ₂ OH	99	20:1	61
2	0	DCM	CH ₃ CH ₂ OH	67	>20:1	67
3	-20	DCM	CH ₃ CH ₂ OH	63	5:1	73
4	-40	DCM	CH ₃ CH ₂ OH	90	8:1	64
5	-78	DCM	CH ₃ CH ₂ OH	97	6:1	86
6	-78	THF	CH ₃ CH ₂ OH	85	19:1	77
7	-78	Toluene	CH ₃ CH ₂ OH	60	>20:1	74
8	-20	CH ₂ ClCH ₂ Cl	CH ₃ CH ₂ OH	83	>20:1	78
9	-20	CHCl ₂ CHC ₂	CH ₃ CH ₂ OH	77	11:1	57
10	-78	DCM	CF ₃ CH ₂ OH	80	6:1	52
11	-78	DCM	BnOH	63	3:1	65
12	-78	DCM	i-PrOH	82	5:1	85
13	-78	DCM	^t BuOH	88	11:1	74

^a Reaction conditions: **1a** (0.15 mmol), **2** (0.27 mmol), CH₃CH₂OH (0.27 mmol), AgOAc (10 mol %), ligand (10 mol %), solvent (1.0 mL), and the reaction was carried out at rt \sim -78 °C for 24 h.

^b Yields of the purified *syn* and *anti* product.

^c Determined by ¹H NMR spectroscopic data of the crude product.

^d Determined by chiral HPLC analysis.

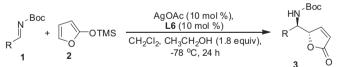
drs were observed at 0 °C and at room temperature (dr>20:1) (Table 2, entries 1–5). When solvents, such as tetrahydrofuran (THF), toluene, CH₂ClCH₂Cl, and CHCl₂CHCl₂ were used in this reaction, lower ee value (from 77% to 57%) was realized but along with good to excellent drs (from 11:1 to >20:1) (Table 2, entries 6–9). Previous studies have revealed that the addition of alcoholic additives can enhance the reactivity and enantioselectivity in this asymmetric reaction.⁶ The optimization studies on the effects of different alcoholic additives in this reaction were then performed and the results of these experiments are also summarized in Table 2 (entries 10-13). A significant decrease in enantioselectivity was observed by adding CF₃CH₂OH or BnOH (1.8 equiv) into the reaction system (Table 2, entries 10 and 11). We also examined additive effects of *i*-PrOH and ^tBuOH in this reaction under identical conditions, but no improvement was observed (Table 2, entries 12 and 13). Therefore, the best reaction conditions are those using 10 mol % of AgOAc along with 10 mol % of chiral ligand L6 as the catalyst and carrying out the reaction in CH₂Cl₂ at -78 °C for 24 h in the presence of 1.8 equiv of CH₃CH₂OH.

Having established the optimal conditions, we next examined the generality of this reaction using various N-Boc-protected imines 1 with siloxyfuran 2 and the results are summarized in Table 3. It was found that the *N*-Boc-protected imines **1b**-i bearing an aromatic ring (R group=aromatic ring) could react with 2 smoothly to give the corresponding asymmetric vinylogous Mannich-type products **3b**-i in good to high yields (79%-97%) and good diastereoselectivities (4:1–7:1 dr) as well as moderate to good enantiomeric excesses (63%–83% ee) for the major diastereoisomers whether they have electron-rich or electron-poor aromatic groups (Table 2, entries 2–9). When R is a heteroaryl group, such as 2-furyl group, similar result was obtained, affording the desired adduct 3j in 79% yield, 80% ee and 7:1 dr (Table 2, entry 10). As for alkyl N-Boc-protected imine 1k, the reactions also proceeded smoothly to give the corresponding *N*-Boc-protected γ -butenolides **3k** in 75 yield and 75% ee along with 6:1 dr value (Table 2, entry 11).

The synthetic utility of the product can be displayed by removal of the *N*-Boc-protecting group with trifluoroacetic acid (TFA) in CH_2Cl_2 at room temperature for 4 h, affording the corresponding free amine product **4** in 92% yield (Scheme 2). The absolute

Table 3

Scope and limitations of AVM reaction of N-Boc aldimines 1 and siloxyfuran 2



Entry ^a	R	Yield [%] ^b	anti:syn ^c	anti, ee [%] ^d
		3	3	3
1	<i>p</i> -МеОС ₆ Н ₄ , 1а	3a , 97 (83)	6:1	86
2	<i>m</i> -MeOC ₆ H ₄ , 1b	3b , 95 (83)	7:1	78
3	<i>p</i> -МеС ₆ Н ₄ , 1с	3c , 90 (72)	4:1	83
4	<i>m</i> -MeC ₆ H ₄ , 1d	3d, 92 (79)	6:1	71
5	3,4,5-(MeO) ₃ C ₆ H ₂ , 1e	3e, 94 (78)	5:1	76
6	Ph, 1f	3f , 82 (70)	6:1	77
7	<i>p</i> -FC ₆ H ₄ , 1g	3g , 93 (80)	6:1	63
8	<i>p</i> -BrC ₆ H ₄ 1 h	3h , 88 (75)	6:1	67
9	m-BrC ₆ H ₄ 1i	3i , 79 (69)	7:1	73
10	2-Furyl, 1j	3j , 79 (69)	7:1	80
11	Cydohexyl, 1k	3k , 88 (75)	6:1	75

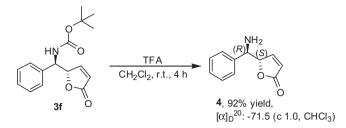
 a Reaction conditions: **1a** (0.15 mmol), **2** (0.27 mmol), CH₃CH₂OH (0.27 mmol), AgOAc (10 mol %), ligand (10 mol %), CH₂Cl₂ (1.0 mL), and the reaction was carried out at $-78\ ^\circ$ C for 24 h.

^b Yields are determined by ¹H NMR spectrum and in parentheses are isolated yields of the *anti* isomer.

^c Determined by ¹H NMR spectroscopic data of the crude product.

^d Determined by chiral HPLC analysis.

configuration was assigned to be (*R*,*S*) for the products 3a-k by comparison of the free amine product 4's ¹H NMR and ¹³C NMR spectra data as well as optical rotation with literature values.^{1b,e}



Scheme 2. Removal of the *N*-Boc group and determination of the absolute configuration of **4**.

To identify the coordination pattern between **L6** and AgOAc, we treated **L6** with AgOAc (1.0 equiv) at 25 °C in DCM- d_2 for 0.5 h, and measured their ³¹P NMR spectroscopy at 25 °C. A significant chemical shift change of their ³¹P NMR spectroscopic signals has been observed from -15.97 ppm (singlet) to 4.85 ppm (doublet, J=555 Hz) in comparison with that of **L6**, suggesting the coordination between **L6** and AgOAc (see the Supplementary data). On the basis of previous mechanistic studies by Hoveyda and our group,^{1C,e} a transition-state model can be outlined in Fig. 2. In the activated complex, to minimize the steric interactions, the substrate is bound *anti* to the bulky substituent (^fBu) on oxazoline of **L6**. The catalyst-bound imine may react with the siloxyfuran by an *endo*-type addition,⁷ and consequently, the reaction of a optically active *N*-Boc imines with siloxyfuran catalyzed by AgOAc combined with **L6** provides the (*R*, *S*) stereoisomer predominantly.

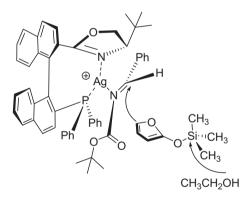


Fig. 2. Plausible transition-state model.

3. Conclusion

In summary, we have reported a silver(I)-catalyzed catalytic asymmetric Mannich reaction of *N*-Boc imines with trimethylsiloxyfuran in the presence of chiral phosphine—oxazoline ligand **L6** in CH₂Cl₂ under mild conditions, affording the corresponding adducts in good to high yields and good diastereoselectivities along with moderate to good enantioselectivities. Since the *N*-Boc group can be easily cleaved under several convenient acidic conditions (HCl or TFA),² the chiral *N*-Boc-protected γ -butenolides synthesized in this paper will be more useful in organic synthesis.

4. Experimental section

4.1. General remarks

Melting points were determined on a digital melting point apparatus and temperatures were uncorrected. Optical rotations were determined at 589 nm (sodium D line) by using a Perkin-Elmer-341 MC digital polarimeter; $[\alpha]_D$ -values are given in unit of $10 \text{ deg}^{-1} \text{ cm}^2 \text{ g}^{-11} \text{H} \text{ NMR}$ spectra were recorded on a Bruker AM-300 and AM-400 spectrometer for solution in CDCl₃ with tetramethylsilane (TMS) as an internal standard; coupling constants *J* are given in hertz.¹⁹F and ³¹P NMR spectra were recorded on a Bruker AM-300 and AM-400 spectrophotometers with complete proton decoupling spectrophotometers. Infrared spectra were recorded on a Perkin–Elmer PE-983 spectrometer with absorption in cm⁻¹. Flash column chromatography was performed using 300-400 mesh silica gel. For thin-layer chromatography (TLC), silica gel plates (Huanghai GF₂₅₄) were used. Chiral HPLC was performed on an SHIMADZU SPD-10A vp series with chiral columns (Chiralpak AD-H, OJ-H, PA-H, and IC-H columns 4.6×250 mm, (Daicel Chemical Ind., Ltd.)). Mass spectra were recorded by EI, ESI, MALDI, and HRMS was measured on an HP-5989 instrument. Organic solvents used were dried by standard methods when necessary. N-Boc aldimines 1a-k were prepared according to the previous reports.⁸ (*aR*)-2a and (*aR*)-3a are known compounds and were prepared according to literature procedures.^{1e,3}

4.2. Typical reaction procedure for the preparation of (aR)-2

To a solution of (aR)-1⁴ (115 mg, 0.25 mmol), diarylphosphine oxide (0.50 mmol), palladium diacetate (5.6 mg, 0.025 mmol), and 1,4-bis(diphenylphosphino)butane (10.7 mg, 0.025 mmol), in 1.5 mL of DMSO was added diisopropylethylamine (0.21 mL, 1.2 mmol), and the mixture was stirred at 120 °C for 24 h. After being cooled to room temperature, the reaction mixture was diluted with EtOAc, washed with H₂O, and dried over MgSO₄. Removal of the solvent followed by column chromatography on silica gel (elution with *n*-hexane/EtOAc=1/2) gave compound (*aR*)-2.

4.2.1. Compound (**aR**)-**2b**. Yield: 70%, 96 mg, white solid, mp 79–80 °C; IR (neat): ν 3056, 2949, 2884, 2835, 1718, 1589, 1497, 1277, 1225, 1190, 1158, 1113, 877, 828, 766, 662 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 3.55 (3H, s), 6.70–6.75 (2H, m), 6.86–6.91 (2H, m), 6.97 (1H, d, *J*=8.4 Hz), 7.05–7.20 (3H, m), 7.30–7.40 (3H, m), 7.45–7.51 (3H, m), 7.64 (1H, dd, *J*=8.8, 12.0 Hz), 7.72 (1H, d, *J*=8.4 Hz), 7.89 (1H, d, *J*=8.4 Hz), 7.95 (1H, d, *J*=8.8 Hz), 8.05 (1H, d, *J*=8.4 Hz); ³¹P NMR (161.94 MHz, CDCl₃, 85% H₃PO₄): δ 26.24; ¹⁹F NMR (376 MHz, CDCl₃, CFCl₃): δ –107.4 to 107.5 (1F, m), –107.8 to 107.9 (1F, m); MS (ESI) *m/z* 549.5 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₄H₂₄O₃F₂P requires (M+H⁺) 549.1426, found: 549.1439; [α]₆²⁰ +19.8 (*c* 0.44, CH₂Cl₂).

4.2.2. Compound (**aR**)-**2c**. Yield: 79%, 113 mg, white solid, mp 85–68 °C; IR (neat): ν 3057, 2976, 2885, 2836, 1719, 1595, 1501, 1277, 1244, 1177, 1115, 1023, 828, 800, 766, 695, 646 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 3.53 (3H, s), 3.71 (3H, s), 3.77 (3H, s), 6.58 (2H, dd, *J*=2.4, 8.8 Hz), 6.68 (1H, dd, *J*=2.4, 8.8 Hz), 6.97–7.02 (2H, m), 7.09–7.13 (1H, m), 7.18–7.27 (3H, m), 7.33–7.41 (3H, m), 7.49 (1H, t, *J*=8.0 Hz), 7.68–7.78 (3H, m), 7.90–7.95 (2H, m), 8.03 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 27.44; MS (MALDI) *m*/*z* 573.5 (M+H⁺, 100). HRMS (ESI) calcd for C₃₆H₃₀O₅P requires (M+H⁺) 573.1825, found: 573.1828; [*α*]_D²⁰ +15.8 (*c* 0.40, CH₂Cl₂).

4.2.3. Compound (**aR**)-**2d**. Yield: 74%, 100 mg, white solid, mp 79–80 °C; IR (neat): ν 3056, 2986, 2877, 2837, 1720, 1599, 1433, 1276, 1241, 1186, 1132, 809, 765, 747, 658, 645 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 2.20 (3H, s), 2.30 (3H, s), 3.53 (3H, s), 6.83 (2H, dd, *J*=2.4, 7.8 Hz), 6.95–7.11 (5H, m), 7.16–7.25 (3H, m), 7.34–7.40 (3H, m), 7.48 (1H, t, *J*=7.8 Hz), 7.63–7.70 (2H, m), 7.76 (1H, d, *J*=9.0 Hz), 7.89–7.94 (2H, m), 8.02 (1H, d, *J*=9.0 Hz); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 28.82; MS (ESI) *m/z* 541.6

(M+H⁺, 100). HRMS (MALDI) calcd for $C_{36}H_{30}O_3P$ requires (M+H⁺) 541.1927, found: 541.1918; $[\alpha]_D^{20}$ +6.0 (*c* 0.58, CH₂Cl₂).

4.2.4. Compound (**aR**)-**2e**. Yield: 83%, 118 mg, white solid, mp 107–110 °C; IR (CH₂Cl₂): ν 3048, 2948, 2918, 2859, 1724, 1597, 1455, 1433, 1276, 1243, 1190, 1132, 871, 852, 767, 733, 696 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 2.06 (6H, s), 2.20 (6H, s), 3.59 (3H, s), 6.70 (1H, s), 6.89 (1H, d, *J*=12 Hz), 6.95–7.02 (3H, m), 7.11–7.29 (5H, m), 7.36 (1H, td, *J*=1.2, 7.2 Hz), 7.49 (1H, t, *J*=7.2 Hz), 7.68–7.75 (3H, m), 7.90–7.95 (2H, m), 8.03 (1H, d, *J*=8.4 Hz); ³¹P NMR (161.94 MHz, CDCl₃, TMS): δ 27.08; MS (ESI) *m*/*z* 569.4 (M+H⁺, 100). HRMS (ESI) calcd for C₃₈H₃₃O₃PNa requires (M+Na⁺) 591.2060, found: 591.2071; $[\alpha]_{10}^{20}$ –8.2 (*c* 1.0, CH₂Cl₂).

4.3. Typical reaction procedure for the preparation of (aR)-3

To a solution of (*aR*)-2 (0.20 mmol) in 2.5 mL of methanol (dioxane for (*aR*)-2b) was added 500 μ L of 40% KOH solution and the mixture was refluxed for 13 h. The reaction mixture was acidified to pH=2 by addition of concd HCl at 0 °C and extracted with CH₂Cl₂ (2×20 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude product was recrystallized in CH₂Cl₂ and petroleum ether to give compound (*aR*)-3.

4.3.1. *Compound* (*aR*)-*3b*. Yield: 97%, 104 mg, white solid, mp 148–150 °C; IR (neat): ν 3057, 2989, 2884, 2829, 1588, 1497, 1386, 1226, 1159, 1134, 1013, 807, 772, 745, 663 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 6.28 (1H, d, *J*=8.4 Hz), 6.62–6.65 (2H, m), 6.80 (1H, t, *J*=7.2 Hz), 7.12–7.20 (5H, m), 7.26–7.34 (3H, m), 7.54 (1H, t, *J*=8.0 Hz), 7.68–7.77 (4H, m), 7.89–7.97 (3H, m); ³¹P NMR (161.94 MHz, CDCl₃, 85% H₃PO₄): δ 33.26 (t, *J*=1.6 Hz); ¹⁹F NMR (376 MHz, CDCl₃, CFCl₃): δ –104.9 to 105.0 (1F, m), –106.2 to 106.3 (1F, m); MS (ESI) *m*/*z* 535.5 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₃H₂₁O₃F₂PNa requires (M+Na⁺) 557.1089, found: 557.1089; [α]₆²⁰–78.7 (*c* 1.0, CH₂Cl₂).

4.3.2. *Compound* (*aR*)-*3c*. Yield: 94%, 105 mg, white solid, mp 116–117 °C; IR (neat): ν 3058, 2964, 2892, 2837, 1711, 1594, 1501, 1404, 1254, 1116, 1022, 800, 746, 650 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 3.68 (3H, s), 3.87 (3H, s), 6.18 (1H, d, *J*=8.4 Hz), 6.43 (2H, dd, *J*=2.0, 8.8 Hz), 6.68–6.72 (1H, m), 7.00–7.05 (4H, m), 7.10 (1H, d, *J*=8.4 Hz), 7.18 (1H, t, *J*=8.0 Hz), 7.24–7.28 (1H, m), 7.34 (1H, dd, *J*=8.4, 12.4 Hz), 7.51 (1H, t, *J*=8.0 Hz), 7.66–7.74 (4H, m), 7.87–7.94 (3H, m); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 34.53; MS (ESI) *m*/*z* 559.4 (M+H⁺, 100). HRMS (ESI) calcd for C₃₅H₂₇NaO₅P requires (M+Na⁺) 581.1488, found: 581.1507; [α]_D²⁰ + 86.4 (*c* 0.59, CH₂Cl₂).

4.3.3. *Compound* (*aR*)-3*d*. Yield: 95%, 100 mg, white solid, mp 245–246 °C; IR (neat): ν 3049, 2984, 2883, 2833, 1717, 1599, 1404, 1284, 1115, 1086, 1019, 826, 806, 752, 680, 649 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 2.16 (3H, s), 2.44 (3H, s), 6.18 (1H, d, *J*=8.7 Hz), 6.63–6.73 (3H, m), 6.96–7.03 (2H, m), 7.09–7.20 (2H, m), 7.24–7.37 (4H, m), 7.51 (1H, t, *J*=7.5 Hz), 7.63–7.73 (4H, m), 7.86–7.93 (3H, m); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 35.79; MS (ESI) *m/z* 527.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₅H₂₈O₃P requires (M+H⁺) 527.1771, found: 527.1766; [α]₆²⁰+119.5 (c 0.7, CH₂Cl₂).

4.3.4. Compound (**a***R*)-**3e**. Yield: 92%, 102 mg, white solid, mp 325–328 °C; IR (CH₂Cl₂): ν 3054, 2918, 2859, 2461, 1923, 1718, 1599, 1502, 1466, 1402, 1272, 1232, 1157, 1134, 1104, 878, 852, 766, 738, 693, 644 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 2.00 (6H, s), 2.36 (6H, s), 6.24 (1H, d, *J*=8.0 Hz), 6.62 (1H, s), 6.70 (2H, d, *J*=12.8 Hz), 6.77 (1H, t, *J*=8.0 Hz), 7.10–7.17 (2H, m), 7.24–7.30 (3H, m), 7.34–7.42 (3H, m), 7.53 (1H, t, *J*=8.0 Hz), 7.65 (1H, d, *J*=8.0 Hz), 7.72 (1H, d, *J*=8.0 Hz), 7.88–7.95 (3H, m); ³¹P NMR (161.94 MHz, CDCl₃, TMS): δ 34.02; MS (ESI) *m*/*z* 555.4 (M+H⁺, 100). HRMS (ESI) calcd

for C₃₇H₃₁O₃PNa requires (M+Na⁺) 577.1903, found: 577.1911; $[\alpha]_D^{20}$ +127.4 (*c* 1.0, CH₂Cl₂).

4.4. Typical reaction procedure for the preparation of (*aR*)-4

A mixture of compound (aR)-3 (1.0 mmol), chiral aminoalcohol (2.0 mmol), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (576 mg, 3.0 mmol), and 1-hydroxy-benzotriazole (542 mg, 4.00 mmol) in *N*,*N*-dimethylformamide (20 mL) was stirred at room temperature overnight. Then, the solvent was removed from the flask under reduced pressure. The resulting residue was diluted with CH₂Cl₂, washed with saturated NaCl solution twice, and extracted by CH₂Cl₂. The organic layer was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was chromatographed on silica gel (elution with petroleum ether/EtOAc=1:1) to give compound (*aR*,*S*)-4.

4.4.1. *Compound* (*aRS*)-*4a*. Yield: 91%, 620 mg, white solid, mp 229–230 °C; IR (neat): ν 3367, 3296, 3057, 3022, 2946, 2877, 2825, 1651, 1537, 1435, 1297, 1147, 1112, 872, 822, 750, 721, 687, 639 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 1.86 (1H, dd, *J*=5.1, 13.5 Hz), 2.13 (1H, dd, *J*=10.2, 13.5 Hz), 3.32–3.40 (1H, m), 3.70–3.74 (1H, m), 4.00–4.02 (1H, m), 4.24–4.29 (1H, m), 6.66–6.73 (3H, m), 6.89–6.96 (2H, m), 7.06–7.13 (4H, m), 7.19–7.28 (2H, m), 7.34–7.38 (3H, m), 7.45 (1H, t, *J*=7.2 Hz), 7.52–7.64 (7H, m), 7.70–7.86 (4H, m), 7.91–7.98 (3H, m), 8.97 (1H, d, *J*=7.5 Hz); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 31.41; MS (ESI) *m/z* 682.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₆H₃₇NO₃P requires (M+H⁺) 682.2506, found: 682.2510; $[\alpha]_D^{20}$ –28.3 (*c* 1.1, CH₂Cl₂).

4.4.2. *Compound* (*aR,S*)-4*b*. Yield: 82%, 517 mg, white solid, mp 262–263 °C; IR (neat): ν 3406, 3263, 3059, 2995, 2879, 2825, 1649, 1543, 1435, 1295, 1147, 1090, 943, 875, 832, 816, 753, 707, 641 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 1.68 (1H, dd, *J*=5.1, 13.5 Hz), 1.94 (1H, dd, *J*=10.2, 13.5 Hz), 3.27–3.36 (1H, m), 3.68–3.73 (1H, m), 3.83–3.91 (1H, m), 4.22–4.26 (1H, m), 6.67–6.75 (3H, m), 6.89–6.97 (4H, m), 7.07–7.13 (6H, m), 7.20–7.33 (2H, m), 7.52–7.65 (6H, m), 7.79 (2H, dd, *J*=8.4, 30.0 Hz), 7.90–8.02 (4H, m), 8.85 (1H, d, *J*=7.8 Hz); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 31.49; MS (ESI) *m/z* 632.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₂H₃₅NO₃P requires (M+H⁺) 632.2349, found: 632.2342; $[\alpha]_D^{20}$ –2.6.3 (*c* 0.74, CH₂Cl₂).

4.4.3. *Compound* (*aR*)-*4c*. Yield: 78%, 494 mg, white solid, mp 225–226 °C; IR (neat): *v* 3431, 3224, 3065, 2965, 2880, 2836, 1647, 1591, 1495, 1397, 1227, 1184, 1093, 1058, 847, 825, 680 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.24 (9H, s), 3.26–3.34 (1H, m), 3.69–3.82 (2H, m), 4.25 (1H, t, *J*=6.0 Hz), 6.36–6.43 (2H, m), 6.54 (1H, d, *J*=9.0 Hz), 6.97–7.11 (3H, m), 7.19–7.25 (3H, m), 7.30–7.43 (3H, m), 7.49–7.62 (3H, m), 7.77 (1H, d, *J*=9.0 Hz), 7.83–8.02 (4H, m), 8.12 (1H, d, *J*=8.4 Hz); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 28.66 (t, *J*=1.6 Hz); ¹⁹F NMR (282 MHz, CDCl₃, CFCl₃): δ –106.3 to 106.5 (1F, m), –108.0 to 108.1 (1F, m); MS (ESI) *m/z* 634.7 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₉H₃₄NO₃F₂PNa requires (M+Na⁺) 656.2137, found: 656.2126; [α]_D²⁰ –34.0 (*c* 0.29, CH₂Cl₂).

4.4.4. Compound (**a***R*)-**4d**. Yield: 85%, 558 mg, white solid, mp 191–192 °C; IR (neat): ν 3429, 3302, 3241, 3068, 2958, 2900, 2836, 1642, 1594, 1522, 1499, 1249, 1169, 1113, 1022, 799, 765, 667, 643 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 0.25 (9H, s), 3.44 (1H, dd, *J*=6.4, 11.6 Hz), 3.64 (3H, s), 3.73–3.81 (2H, m), 3.85 (3H, s), 6.22 (2H, dd, *J*=2.4, 8.8 Hz), 6.48 (1H, d, *J*=8.4 Hz), 6.90–7.03 (5H, m), 7.20–7.31 (3H, m), 7.52–7.59 (3H, m), 7.74–7.81 (3H, m), 7.90–7.96 (3H, m), 8.09 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 29.25; MS (ESI) *m/z* 658.6 (M+H⁺, 100). HRMS (ESI) calcd

for C₄₁H₄₁NO₅P requires (M+H⁺) 658.2717, found: 658.2714; $[\alpha]_D^{20}$ –43.7 (*c* 0.39, CH₂Cl₂).

4.4.5. Compound (**aR**)-**4e**. Yield: 89%, 557 mg, white solid, mp 117–118 °C; IR (neat): ν 3425, 3305, 3239, 3055, 2973, 2950, 2877, 2830, 1645, 1550, 1502, 1308, 1165, 1113, 1093, 810, 748, 661, 646 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 0.24 (9H, s), 2.07 (3H, s), 2.41 (3H, s), 3.44 (1H, dd, *J*=6.0, 11.2 Hz), 3.74–3.82 (2H, m), 4.41 (1H, s), 6.48 (3H, d, *J*=8.4 Hz), 6.88–6.98 (3H, m), 7.19–7.33 (5H, m), 7.50–7.60 (3H, m), 7.72–7.80 (3H, m), 7.90–7.96 (3H, m), 8.08 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 31.01; MS (ESI) *m/z* 626.7 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₁H₄₁NO₃P requires (M+H⁺) 626.2819, found: 626.2818; [α]_D²⁰ –41.7 (*c* 0.52, CH₂Cl₂).

4.4.6. *Compound* (*aR*)-*4f*. Yield: 98%, 640 mg, white solid, mp 275–278 °C; IR (CH₂Cl₂): ν 3434, 3055, 2957, 2866, 1649, 1551, 1503, 1272, 1175, 1164, 1152, 1162, 876, 852, 718, 748, 696 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.25 (9H, s), 1.93 (6H, s), 2.36 (6H, s), 3.40–3.48 (1H, m), 3.73–3.81 (2H, m), 4.33 (1H, t, *J*=7.2 Hz), 6.45 (1H, s), 6.50 (1H, d, *J*=8.1 Hz), 6.65 (1H, d, *J*=12.6 Hz), 6.97 (1H, t, *J*=6.9 Hz), 7.16–7.31 (4H, m), 7.50–7.66 (5H, m), 7.75 (1H, d, *J*=8.7 Hz), 7.91–8.00 (3H, m), 8.10 (1H, d, *J*=8.7 Hz); ³¹P NMR (121.45 MHz, CDCl₃, TMS): δ 24.77; MS (ESI) *m/z* 654.4 (M+H⁺, 100). HRMS (ESI) calcd for C₄₃H₄₅NO₃P requires (M+H⁺) 654.3132, found: 654.3145; [α]²⁰_{*f*}–30.1 (*c* 1.0, CH₂Cl₂).

4.5. Typical reaction procedure for the preparation of (aR)-5

To a mixture of compound (aR)-4 (1.0 mmol), 4-dimethylaminopyridine (1.22 mg, 0.01 mmol), and triethylamine (1.8 mL, 2.0 mmol) in dichloromethane (15 mL) was added methanesulfonyl chloride (0.4 mL, 2.0 mmol) at 0 °C, and the solution was stirred for 0.5 h at this temperature. Then another portion of triethylamine (1.4 mL, 9.0 mmol) was added to the solution, and it was refluxed until the initially formed mesylation product disappeared (checked by TLC plates). On cooling to room temperature, the reaction mixture was diluted with CH₂Cl₂, then washed with saturated NaHCO₃ solution and extracted by CH₂Cl₂. The organic layer was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was chromatographed on silica gel (elution with EtOAc) to give compound (aR.S)-5.

4.5.1. Compound (**a***R*,**S**)-**5a**. Yield: 91%, 603 mg, white solid, mp 106–107 °C; IR (neat): ν 3050, 2986, 2888, 2836, 1631, 1504, 1435, 1363, 1308, 1189, 1113, 1057, 971, 815, 747, 697 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 2.45 (1H, dd, *J*=8.4, 13.8 Hz), 3.02 (1H, dd, *J*=5.4, 13.8 Hz), 3.58 (1H, t, *J*=7.8 Hz), 3.91 (1H, t, *J*=8.4 Hz), 4.25–4.36 (1H, m), 6.99–7.22 (10H, m), 7.27–7.52 (10H, m), 7.60–7.77 (6H, m), 7.84–7.89 (2H, m), 7.99 (1H, d, *J*=8.4 Hz); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 28.41; MS (ESI) *m*/*z* 664.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₆H₃₅NO₂P requires (M+H⁺) 664.2400, found: 664.2399; $[\alpha]_D^{20}$ +8.0 (*c* 0.30, CH₂Cl₂).

4.5.2. Compound (**a***R*,**S**)-**5***b*. Yield: 82%, 503 mg, white solid, mp 94–95 °C; IR (neat): ν 3053, 2987, 2886, 2828, 1634, 1496, 1436, 1310, 1190, 1113, 1056, 973, 818, 748, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 2.23 (1H, dd, *J*=8.8, 14.0 Hz), 2.87 (1H, dd, *J*=5.6, 14.0 Hz), 3.51 (1H, t, *J*=8.0 Hz), 3.92 (1H, t, *J*=8.8 Hz), 4.16–4.22 (1H, m), 6.94 (2H, d, *J*=6.4 Hz), 7.02–7.07 (3H, m), 7.10–7.21 (8H, m), 7.23–7.30 (2H, m), 7.34–7.39 (3H, m), 7.45 (2H, dd, *J*=8.0, 11.6 Hz), 7.53 (1H, t, *J*=7.2 Hz), 7.65–7.72 (3H, m), 7.91–7.95 (3H, m); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 27.53; MS (ESI) *m*/*z* 614.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₂H₃₃NO₂P requires (M+H⁺) 614.2243, found: 614.2235; [α]_D²⁰ +13.4 (*c* 0.57, CH₂Cl₂).

4.5.3. Compound (**aR**,**S**)-**5c**. Yield: 78%, 480 mg, white solid, mp 80–81 °C; IR (neat): *ν* 3059, 2978, 2953, 2838, 1640, 1589, 1498, 1355,

1236, 1158, 1099, 1058, 1112, 817, 741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.57 (9H, s), 3.55 (1H, t, *J*=8.4 Hz), 3.70 (1H, t, *J*=9.3 Hz), 3.93 (1H, t, *J*=9.3 Hz), 6.72–6.83 (4H, m), 7.05–7.27 (4H, m), 7.31–7.43 (5H, m), 7.50 (1H, t, *J*=7.5 Hz), 7.69–7.77 (3H, m), 7.88–7.97 (3H, m); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 27.52 (t, *J*=1.8 Hz); ¹⁹F NMR (282 MHz, CDCl₃, CFCl₃): δ –108.4 to 108.5 (2F, m); MS (ESI) *m/z* 616.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₉H₃₃NO₂F₂P requires (M+H⁺) 616.2212, found: 616.2211; [α]^D_D⁰ +72.6 (*c* 0.42, CH₂Cl₂).

4.5.4. *Compound* (*aR,S*)-*5d.* Yield: 85%, 543 mg, white solid, mp 96–97 °C; IR (neat): ν 3058, 2950, 2895, 2836, 1637, 1595, 1501, 1292, 1251, 1117, 1114, 1024, 823, 750, 663 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.56 (9H, s), 3.51 (1H, t, *J*=8.4 Hz), 3.69 (1H, t, *J*=8.4 Hz), 3.75 (3H, s), 3.76 (3H, s), 3.91 (1H, dd, *J*=8.4, 9.9 Hz), 6.56–6.65 (4H, m), 7.03–7.22 (4H, m), 7.26–7.39 (5H, m), 7.47 (1H, t, *J*=7.8 Hz), 7.68–7.96 (6H, m); ³¹P NMR (121.45 MHz, CDCl₃, 85% H₃PO₄): δ 29.04; MS (ESI) *m/z* 640.7 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₁H₃₉NO₄P requires (M+H⁺) 640.2611, found: 640.2624; [α]⁶⁰ +42.0 (*c* 0.28, CH₂Cl₂).

4.5.5. *Compound* (*aR*,*S*)-*5e.* Yield: 89%, 540 mg, white solid, mp 83–84 $^{\circ}$ C; IR (neat): ν 3050, 2973, 2898, 2868, 2836, 1635, 1600, 1359, 1186, 1112, 1055, 973, 808, 749, 658 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 0.56 (9H, s), 2.25 (3H, s), 2.26 (3H, s), 3.51 (1H, t, *J*=8.4 Hz), 3.67 (1H, t, *J*=9.6 Hz), 3.90 (1H, dd, *J*=8.4, 9.6 Hz), 6.88–6.91 (4H, m), 7.03–7.11 (3H, m), 7.17–7.37 (6H, m), 7.47 (1H, t, *J*=8.0 Hz), 7.66–7.77 (3H, m), 7.86–7.93 (3H, m); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ 28.46; MS (ESI) *m/z* 608.7 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₁H₃₉NO₂P requires (M+H⁺) 608.2713, found: 608.2714; [α]_D²⁰ +41.9 (*c* 0.45, CH₂Cl₂).

4.5.6. *Compound* (*aR*,*S*)-*5f*. Yield: 98%, 622 mg, white solid, mp 121–126 °C; IR (CH₂Cl₂): *v* 3050, 2954, 2865, 1640, 1600, 1477, 1360, 1271, 1191, 1124, 978, 870, 853, 824, 749, 669, 642 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.55 (9H, s), 2.13 (6H, s), 2.14 (6H, s), 3.54 (1H, t, *J*=9.3 Hz), 3.72 (1H, t, *J*=9.3 Hz), 3.98 (1H, t, *J*=8.7 Hz), 6.85 (2H, d, *J*=13.2 Hz), 6.96–7.22 (8H, m), 7.36 (1H, t, *J*=7.5 Hz), 7.47 (1H, t, *J*=7.5 Hz), 7.66–7.78 (3H, m), 7.86–7.93 (3H, m); ³¹P NMR (121.45 MHz, CDCl₃, TMS): δ 23.80; MS (EI) *m/z* (%): 635 [M⁺] (7.6), 578 (22.7), 378 (79.0), 365 (45.6), 257 (91.5), 210 (100.0), 209 (31.5), 183 (76.3), 91 (42.1), 57 (25.7), 43 (18.2); HRMS (EI) calcd for C₄₃H₄₂NO₂P requires (M⁺) 635.2953, found: 635.2956; [α]²⁰ +41.7 (*c* 1.0, CH₂Cl₂).

4.6. Typical reaction procedure for the preparation of L7–L12

To a mixture of compound (*aR,S*)-5 (1.0 mmol) and triethylamine (4.2 mL, 30 mmol) in toluene (20 mL) was added trichlorosilane (1.0 mL, 10.0 mmol) at 0 °C, and the solution was stirred for 0.5 h at this temperature. It was then refluxed for 24 h under nitrogen atmosphere. After cooling to room temperature, the mixture was diluted with EtOAc and quenched with small amount of saturated NaHCO₃. The resulting suspension was filtered through Celite and the filter cake was washed with EtOAc. The combined organic layer was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude product was purified by silica gel column chromatography (elution petroleum ether/EtOAc=4:1) to give chiral ligands **L7–L12**.

L7. 589 mg, Yield: 91%, white solid, mp 67–68 °C; IR (neat): ν 3049, 2973, 2889, 2837, 1632, 1504, 1477, 1432, 1308, 1261, 1091, 1055, 1024, 815, 741, 694 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 2.26 (1H, dd, *J*=8.8, 13.6 Hz), 2.91 (1H, dd, *J*=5.2, 13.6 Hz), 3.36 (1H, t, *J*=8.0 Hz), 3.60 (1H, t, *J*=8.8 Hz), 4.05–4.14 (1H, m), 6.96–7.29 (15H, m), 7.34 (1H, s), 7.40–7.49 (5H, m), 7.65–7.69 (2H, m), 7.75 (1H, d, *J*=8.0 Hz), 7.83–7.91 (3H, m), 8.02 (1H, d, *J*=8.4 Hz), 8.16 (1H, d, *J*=8.4 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ –15.38; MS (MALDI) *m/z* 648.6 (M+H⁺, 100). HRMS (ESI) calcd for

C₄₆H₃₅NOP requires (M+H⁺) 648.2451, found: 648.2457; $[\alpha]_D^{20}$ –29.9 (*c* 0.54, CH₂Cl₂).

L8. 490 mg, Yield: 82%, white solid, mp 65–66 °C; IR (neat): ν 3050, 2975, 2885, 2831, 1633, 1495, 1478, 1432, 1262, 1093, 1054, 1025, 971, 816, 741, 694 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 2.08 (1H, dd, *J*=8.8, 13.6 Hz), 2.75 (1H, dd, *J*=4.2, 13.6 Hz), 3.30 (1H, t, *J*=8.0 Hz), 3.62 (1H, t, *J*=8.8 Hz), 3.95–4.03 (1H, m), 6.89 (2H, d, *J*=6.8 Hz), 6.95–7.05 (4H, m), 7.10–7.27 (13H, m), 7.39–7.49 (3H, m), 7.84–7.91 (3H, m), 8.01 (1H, d, *J*=8.8 Hz), 8.13 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ –15.32; MS (ESI) *m/z* 598.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₂H₃₃NOP requires (M+H⁺) 598.2294, found: 598.2274; $[\alpha]_D^{20}$ –24.1 (*c* 0.61, CH₂Cl₂).

L9. 467 mg, Yield: 78%, white solid, mp 59–60 °C; IR (neat): ν 3058, 2958, 2899, 2867, 1639, 1586, 1493, 1359, 1224, 1158, 1097, 1054, 1014, 811, 747 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 0.52 (9H, s), 3.45 (1H, t, *J*=8.0 Hz), 3.61 (1H, t, *J*=8.0 Hz), 3.71 (1H, t, *J*=8.8 Hz), 6.78–6.83 (3H, m), 6.93–7.00 (5H, m), 7.21–7.24 (4H, m), 7.35–7.45 (3H, m), 7.84–7.88 (3H, m), 7.99 (1H, d, *J*=8.8 Hz), 8.10 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ –17.54 (t, *J*=4.5 Hz); ¹⁹F NMR (282 MHz, CDCl₃, CFCl₃): δ –112.9 to 113.0 (1F, m), –113.4 to 113.5 (1F, m); MS (ESI) *m*/*z* 600.6 (M+H⁺, 100). HRMS (MALDI) calcd for C₃₉H₃₃NOF₂P requires (M+H⁺) 600.2262, found: 600.2268; [α]_D²⁰ +11.9 (*c* 0.31, CH₂Cl₂).

L10. 530 mg, Yield: 85%, white solid, mp 68–69 °C; IR (neat): ν 3059, 2960, 2899, 2834, 1635, 1592, 1496, 1282, 1244, 1176, 1093, 1028, 821, 796, 649 cm⁻¹; ¹H NMR (400 MHz, CDCl3, TMS): δ 0.52 (9H, s), 3.44 (1H, t, *J*=8.0 Hz), 3.57 (1H, dd, *J*=8.0, 10.0 Hz), 3.70 (1H, dd, *J*=8.0, 10.0 Hz), 3.71 (3H, s), 3.77 (3H, s), 6.64 (2H, d, *J*=8.0 Hz), 6.81–6.86 (3H, m), 6.90–6.97 (3H, m), 7.16–7.24 (4H, m), 7.33–7.46 (3H, m), 7.81–7.86 (3H, m), 7.97 (1H, d, *J*=8.8 Hz), 8.11 (1H, d, *J*=8.4 Hz); ³¹P NMR (161.94 MHz, CDCl₃, 85% H₃PO₄): δ –18.36; MS (ESI) *m*/*z* 624.7 (M+H⁺, 100). HRMS (MALDI) calcd for C₄₁H₃₉NO₃P requires (M+H⁺) 624.2662, found: 624.2661; [α]_D²⁰ –18.1 (*c* 0.32, CH₂Cl₂).

L11. 526 mg, Yield: 89%, white solid, mp 71–72 °C; IR (neat): ν 3047, 2971, 2896, 2836, 1634, 1496, 1359, 1262, 1099, 1054, 972, 804, 746, 693 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 0.52 (9H, s), 2.25 (3H, s), 2.32 (3H, s), 3.42 (1H, t, *J*=8.4 Hz), 3.54 (1H, t, *J*=9.6 Hz), 3.67 (1H, dd, *J*=8.4, 9.6 Hz), 6.90–6.91 (5H, m), 6.95–7.00 (1H, m), 7.08 (2H, d, *J*=7.6 Hz), 7.14–7.20 (4H, m), 7.35–7.43 (2H, m), 7.48 (1H, dd, *J*=2.8, 8.4 Hz), 7.82 (2H, d, *J*=8.4 Hz), 7.87 (1H, d, *J*=8.0 Hz), 7.98 (1H, d, *J*=8.8 Hz), 8.11 (1H, d, *J*=8.8 Hz); ³¹P NMR (161.93 MHz, CDCl₃, 85% H₃PO₄): δ –17.06; MS (MALDI) *m*/*z* 592.7 (M+H⁺, 100). HRMS (ESI) calcd for C₄₁H₃₉NOP requires (M+H⁺) 592.2764, found: 592.2762; [α]_D²⁰ –11.2 (*c* 0.5, CH₂Cl₂).

L12. 607 mg, Yield 98%, white solid, mp 86–91 °C; IR (CH₂Cl₂): ν 3050, 2953, 2866, 1640, 1598, 1582, 1477, 1359, 1264, 1124, 1101, 1055, 974, 846, 818, 746, 693, 524; ¹H NMR (300 MHz, CDCl₃, TMS): δ 0.51 (9H, s), 2.10 (6H, s), 2.23 (6H, s), 3.48 (1H, t, *J*=7.8 Hz), 3.63 (1H, t, *J*=9.6 Hz), 3.77 (1H, dd, *J*=7.8, 9.6 Hz), 6.60 (2H, d, *J*=7.8 Hz), 6.76 (1H, s), 6.83–6.98 (5H, m), 7.19–7.21 (2H, m), 7.35–7.45 (2H, m), 7.54 (1H, dd, *J*=5.7, 8.4 Hz), 7.85 (3H, t, *J*=8.4 Hz), 7.98 (1H, d, *J*=8.4 Hz), 8.10 (1H, d, *J*=8.4 Hz); ³¹P NMR (121.45 MHz, CDCl₃, TMS): δ –13.69; MS (EI) *m/z* (%): 619 [M⁺] (27.9), 535 (100.0), 493 (49.4), 430 (50.4), 408 (84.7), 378 (22.2), 362 (27.7), 281 (20.4), 241 (29.3), 57 (45.2), 41 (23.3); HRMS (EI) calcd for C₄₃H₄₂NOP (M⁺): 619.3004, found: 619.3010; [α]_D²⁰ –8.0 (*c* 1.0, CHCl₃).

4.7. General procedure for the reaction of *N*-Boc aldimines 1 with siloxyfuran 2 in the presence of AgOAc and chiral phosphine–oxazoline ligand L6

AgOAc (2.50 mg, 0.015 mmol) and chiral phosphine—oxazoline ligand **L6** (8.4 mg, 0.015 mmol) were added into a Schlenk tube and then DCM (0.5 mL) was added into the reaction vessel. The resulting solution was stirred for 0.5 h at room temperature. *N*-Boc aldimine

1 (0.15 mmol), CH₃CH₂OH (16 μ L, 0.27 mmol) was added followed by another 0.5 mL of DCM. The mixture was cooled to -78 °C, and stirred for 0.5 h, then 2-trimethylsiloxyfuran (**2**) (46 μ L, 0.27 mmol) was added. The mixture was allowed to stir at -78 °C for 24 h, and then it was allowed to warm to room temperature (22 °C). A saturated aqueous solution of NaHCO₃ was added and the aqueous layer was washed with EtOAc (3×5 mL), dried over MgSO₄, and the volatiles were removed under reduced pressure and the residue was purified by flash column chromatography on silica gel (elution with petroleum ether/EtOAc=4:1) to give **3**.

4.7.1. *Compound anti-3a*. Yield: 83%, 40 mg, white solid, mp 138–139 °C; IR (neat): ν 3364, 2990, 2938, 2841, 1767, 1693, 1506, 1282, 1242, 1152, 1091, 1030, 906, 826, 787, 634 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.43 (9H, s), 3.77 (3H, s), 5.01–5.02 (1H, m), 5.39–5.41 (2H, m), 5.97 (1H, s), 6.84 (2H, d, *J*=8.4 Hz), 7.15 (2H, d, *J*=8.4 Hz), 7.34 (1H, d, *J*=5.6 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 28.2, 55.1, 55.8, 80.2, 84.6, 114.0 (2C), 122.8, 127.5, 128.2 (2C), 153.2, 154.9, 159.4, 172.4; MS (ESI) *m/z* 337.2 (M+NH₄₊, 100). HRMS (ESI) calcd for C₁₇H₂₁NO₅Na requires (M+Na⁺) 342.1312, found: 342.1318; [α]_D²⁰ –100.7 (*c* 1.0, CH₂Cl₂, 86% ee). Enantiomeric excess was determined by HPLC with a Chiralcel PA-H column (*n*-hexane/*i*-PrOH=50/50, 0.7 mL/min, 214 nm, *t*_{minor}=21.02 min, *t*_{major}=16.99 min).

4.7.2. *Compound anti-3b.* Yield: 83%, 40 mg, white solid, mp 128–129 °C; IR (neat): *v* 3389, 2977, 2934, 2836, 1748, 1687, 1599, 1516, 1494, 1249, 1161, 1101, 1039, 884, 830, 750, 663 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.39 (9H, s), 3.81 (3H, s), 5.06–5.13 (2H, m), 5.33 (1H, s), 6.15 (1H, dd, *J*=2.0, 6.0 Hz), 6.86 (1H, dd, *J*=2.0, 8.0 Hz), 6.94–6.99 (2H, m), 7.27–7-31 (1H, m), 7.53 (1H, s); ¹³C NMR (CDCl₃, 100 MHz): δ 28.1, 55.1, 55.2, 80.2, 85.1, 113.0, 113.5, 119.3, 122.2, 129.9, 139.4, 154.4, 155.2, 159.9, 172.9; MS (ESI) *m/z* 342.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₇H₂₁NO₅Na requires (M+Na⁺) 342.1312, found: 342.1320; [α]_D²⁰ –83.6 (*c* 1.0, CH₂Cl₂, 78% ee). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column (*n*-hexane/*i*-PrOH=70/30, 0.6 mL/min, 214 nm, *t_{minor}=12.77 min*, *t_{maior}=10.15 min*).

4.7.3. *Compound anti-***3***c*. Yield: 72%, 33 mg, white solid, mp 153–154 °C; IR (neat): *v* 3377, 2977, 2925, 2854, 1770, 1696, 1511, 1361, 1277, 1248, 1160, 1089, 1049, 909, 815, 743 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.43 (9H, s), 2.31 (3H, s), 5.03 (1H, s), 5.31 (1H, d, *J*=8.4 Hz), 5.41 (1H, s), 5.96 (1H, s), 7.09–7.13 (4H, m), 7.32 (1H, d, *J*=6.4 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 21.0, 28.2, 56.0, 80.3, 84.6, 122.9, 126.9 (2C), 129.3 (2C), 132.4, 138.1, 153.1, 154.9, 172.4; MS (ESI) *m/z* 326.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₇H₂₁NO₄Na requires (M+Na⁺) 326.1363, found: 326.1362; [α]_D²⁰ –101.6 (*c* 1.0, CH₂Cl₂, 83% ee). Enantiomeric excess was determined by HPLC with a Chiralcel OJ-H column (*n*-hexane/*i*-PrOH=80/20, 0.7 mL/min, 230 nm, *t*_{minor}=14.18 min, *t*_{major}=11.63 min).

4.7.4. *Compound syn*-**3c**. Yield: 18%, 9 mg, white solid, mp 144–145 °C; IR (neat): ν 3384, 2981, 2923, 2851, 1748, 1687, 1509, 1367, 1246, 1161, 1099, 1040, 888, 815, 750, 651 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.39 (9H, s), 2.35 (3H, s), 4.97 (1H, d, *J*=9.2 Hz), 5.03–5.05 (1H, m), 5.32 (1H, s), 6.15 (1H, dd, *J*=2.0, 5.6 Hz), 7.18 (2H, d, *J*=8.0 Hz), 7.29 (2H, d, *J*=8.0 Hz), 7.53 (1H, d, *J*=4.8 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 21.1, 28.2, 54.9, 80.2, 85.3, 122.2, 127.0 (2C), 129.5 (2C), 134.9, 138.2, 154.5, 155.2, 172.9; MS (ESI) *m/z* 326.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₇H₂₁NO₄Na requires (M+Na⁺) 326.1363, found: 326.1367; [α]_D²⁰ –1.7 (*c* 0.50, CH₂Cl₂, 7% ee). Enantiomeric excess was determined by HPLC with a Chiralcel IC-H column (*n*-hexane/*i*-PrOH=80/20, 1.0 mL/min, 214 nm, *t*_{minor}=22.73 min, *t*_{major}=10.58 min).

4.7.5. *Compound anti-3d*. Yield: 79%, 36 mg, white solid, mp 146–147 °C; IR (neat): *v* 3374, 2984, 2922, 2851, 1780, 1755, 1690, 1514, 1362, 1246, 1151, 1091, 885, 832, 703, 641 cm⁻¹; ¹H NMR (300 MHz,

CDCl₃, TMS): δ 1.43 (9H, s), 2.32 (3H, s), 5.02 (1H, s), 5.36–5.39 (2H, m), 5.97 (1H, d, *J*=2.1 Hz), 7.01–7.03 (2H, m), 7.09 (1H, d, *J*=7.5 Hz), 7.18–7-23 (1H, m), 7.34 (1H, d, *J*=5.4 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 21.3, 28.2, 56.3, 80.3, 84.5, 122.8, 124.0, 127.7, 128.5, 129.1, 135.5, 138.4, 153.0, 154.9, 172.3; MS (ESI) *m/z* 326.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₇H₂₁NO₄Na requires (M+Na⁺) 326.1363, found: 326.1360; [α]_D²⁰ –79.5 (*c* 1.0, CH₂Cl₂, 71% ee). Enantiomeric excess was determined by HPLC with a Chiralcel IC-H column (*n*-hexane/*i*-PrOH=70/30, 0.7 mL/min, 214 nm, *t*_{minor}=29.48 min, *t*_{major}=12.13 min).

4.7.6. *Compound anti-***3***e*. Yield: 78%, 44 mg, white solid, mp 68–69 °C; IR (neat): *v* 3366, 2972, 2930, 2851, 1751, 1697, 1595, 1509, 1461, 1286, 1237, 1160, 1124, 1094, 1054, 1109, 828, 683 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.45 (9H, s), 3.81 (3H, s), 3.84 (6H, s), 4.98 (1H, s), 5.33 (1H, d, *J*=7.2 Hz), 5.45 (1H, s), 6.00 (1H, s), 6.45 (2H, s), 7.32 (1H, d, *J*=5.6 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 28.2, 56.1 (2C), 56.5, 60.7, 80.5, 84.4, 104.1 (2C), 122.9, 131.3, 137.9, 153.0, 153.4 (2C), 154.9, 172.4; MS (ESI) *m/z* 402.4 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₉H₂₅NO₇Na requires (M+Na⁺) 402.1523, found: 402.1527; [α]_D²⁰ –30.1 (*c* 1.0, CH₂Cl₂, 76% ee). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column (*n*-hexane/*i*-PrOH=90/10, 0.5 mL/min, 214 nm, *t*_{minor}=13.89 min, *t*_{maior}=11.64 min).

4.7.7. *Compound anti*-**3f**. Yield: 70%, 30 mg, white solid, mp 159–160 °C; IR (neat): ν 3385, 2984, 2924, 2852, 1751, 1686, 1513, 1498, 1355, 1246, 1165, 1102, 889, 825, 804, 702, 653, 680 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 1.39 (9H, s), 5.02–5.12 (2H, m), 5.36 (1H, s), 6.17 (1H, dd, *J*=1.8, 5.7 Hz), 7.34–7.41 (5H, m), 7.56 (1H, d, *J*=5.1 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 28.1, 55.1, 80.3, 85.2, 122.2, 127.1 (2C), 128.3, 128.9 (2C), 137.8, 154.5, 155.2, 172.9; MS (ESI) *m/z* 312.2 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₆H₁₉NO₄Na requires (M+Na⁺) 312.1206, found: 312.1212; $[\alpha]_D^{20}$ –71.3 (*c* 1.0, CH₂Cl₂, 77% ee). Enantiomeric excess was determined by HPLC with a Chiralcel OD-H column (*n*-hexane/*i*-PrOH=80/20, 0.6 mL/min, 214 nm, *t*_{minor}=13.53 min, *t*_{major}=10.93 min).

4.7.8. Compound anti-**3g**. Yield: 80%, 37 mg, white solid, mp 151–152 °C; IR (neat): ν 3387, 2983, 2920, 2851, 1754, 1682, 1504, 1349, 1277, 1249, 1226, 1158, 1102, 888, 822, 805, 680 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.39 (9H, s), 5.03–5.08 (2H, m), 5.32 (1H, s), 6.17 (1H, dd, *J*=1.2, 6.0 Hz), 7.07 (2H, t, *J*=8.4 Hz), 7.38–7.42 (2H, m), 7.54 (1H, d, *J*=2.4 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 28.1, 54.4, 80.4, 85.0, 115.7 (2C, d, *J*=20.9 Hz), 122.3, 128.9 (2C, d, *J*=8.2 Hz), 133.8 (d, *J*=3.1 Hz), 154.3, 155.1, 162.4 (d, *J*=246.2 Hz), 172.8; ¹⁹F NMR (376 MHz, CDCl₃, CFCl₃): δ – 113.9 to 114.0 (1F, m); MS (ESI) *m*/*z* 330.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₆H₁₈FNO₄Na requires (M+Na⁺) 330.1112, found: 330.1113; $[\alpha]_D^{20}$ –65.7 (*c* 1.0, CH₂Cl₂, 63% ee). Enantiomeric excess was determined by HPLC with a Chiralcel OJ-H column (*n*-hexane/*i*-PrOH=90/10, 0.7 mL/min, 230 nm, *t*_{minor}=24.37 min, *t*_{major}=28.97 min).

4.7.9. *Compound anti*-**3h**. Yield: 75%, 41 mg, white solid, mp 165–166 °C; IR (neat): ν 3347, 2978, 2924, 2852, 1752, 1683, 1517, 1494, 1367, 1276, 1167, 1063, 1018, 922, 886, 833, 797, 655 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 1.39 (9H, s), 5.04–5.07 (2H, m), 5.31 (1H, s), 6.18 (1H, dd, *J*=2.1, 5.4 Hz), 7.29 (2H, d, *J*=8.4 Hz), 7.49–7.52 (3H, m); ¹³C NMR (CDCl₃, 75 MHz): δ 28.1, 54.5, 80.5, 84.8, 122.3, 122.5, 128.9 (2C), 132.0 (2C), 137.0, 154.1, 155.1, 172.7; MS (ESI) *m/z* 366.2 (M–H⁺, 100). HRMS (ESI) calcd for C₁₆H₁₇NO₄Br requires (M–H⁺) 366.0346, found: 366.0359; $[\alpha]_{D}^{20}$ –40.5 (*c* 0.5, CH₂Cl₂, 67% ee). Enantiomeric excess was determined by HPLC with a Chiralcel IC-H column (*n*-hexane/*i*-PrOH=70/30, 0.6 mL/min, 230 nm, *t*_{minor}=36.59 min, *t*_{major}=32.47 min).

4.7.10. Compound anti-**3i**. Yield: 69%, 38 mg, white solid, mp 161–162 °C; IR (neat): *v* 3347, 2960, 2923, 2849, 1754, 1685, 1518,

1490, 1368, 1245, 1168, 1060, 1010, 922, 888, 833, 797, 650 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, TMS): δ 1.40 (9H, s), 4.98–5.07 (2H, m), 5.33 (1H, s), 6.19 (1H, dd, *J*=2.0, 5.6 Hz), 7.24–7.28 (1H, m), 7.34–7.36 (1H, m), 7.46–7.49 (1H, m), 7.52–7.57 (2H, m); ¹³C NMR (CDCl₃, 75 MHz): δ 28.2, 54.5, 80.6, 84.8, 122.6, 125.8, 130.2, 130.5, 131.5, 133.9, 140.2, 154.1, 155.1, 172.6; MS (ESI) *m/z* 390.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₆H₁₈NO₄BrNa requires (M+Na⁺) 390.0311, found: 390.0315; [α]_D²⁰–71.7 (*c* 0.34, CH₂Cl₂, 73% ee). Enantiomeric excess was determined by HPLC with a Chiralcel IC-H column (*n*-hexane/*i*-PrOH=70/30, 0.6 mL/min, 230 nm, *t*_{minor}=14.66 min, *t*_{major}=12.26 min).

4.7.11. Compound anti-**3***j*. Yield: 69%, 29 mg, white solid, mp 136–137 °C; IR (neat): ν 3385, 2956, 2923, 2852, 1758, 1690, 1520, 1362, 1290, 1156, 1092, 1053, 1005, 894, 833, 778, 756, 690 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.45 (9H, s), 5.20–5.21 (2H, m), 5.36 (1H, s), 6.10 (1H, d, *J*=3.2 Hz), 6.27 (1H, d, *J*=2.8 Hz), 6.32 (1H, s), 7.34 (1H, s), 7.42 (1H, dd, *J*=1.2, 6.0 Hz); ¹³C NMR (CDCl₃, 75 MHz): δ 28.2, 50.5, 80.7, 83.5, 108.3, 110.5, 123.1, 142.5, 149.1, 152.6, 154.8, 172.2; MS (ESI) *m/z* 302.1 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₄H₁₇NO₅Na requires (M+Na⁺) 302.0999, found: 302.1013; [α]₂₀²⁰ –42.7 (*c* 0.5, CH₂Cl₂, 80% ee). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column (*n*-hexane/*i*-PrOH=70/30, 0.7 mL/min, 214 nm, *t*_{minor}=12.48 min, *t*_{maior}=11.02 min).

4.7.12. Compound anti-**3k**. Yield: 75%, 33 mg, white solid, mp 163–164 °C; IR (neat): ν 3366, 2981, 2920, 2852, 1740, 1694, 1518, 1366, 1285, 1163, 1094, 1039, 1021, 901, 836, 779, 703, 616 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.16–1.191 (H, m), 1.27–1.31 (1H, m), 1.45 (9H, s), 1.61–1.80 (9H, m), 3.60–3.65 (1H, m), 4.57 (1H, d, *J*=9.2 Hz), 5.00 (1H, d, *J*=7.6 Hz), 6.17 (1H, dd, *J*=1.6, 5.6 Hz), 7.49 (1H, d, *J*=5.2 Hz); ¹³C NMR (CDCl₃, 100 MHz): δ 25.8, 25.9, 26.0, 27.0, 28.2, 30.3, 38.6, 56.8, 80.0, 83.0, 122.0, 154.9, 155.5, 172.6; MS (ESI) *m/z* 318.3 (M+Na⁺, 100). HRMS (ESI) calcd for C₁₆H₂₅NO₄Na requires (M+Na⁺) 318.1676, found: 318.1681; $[\alpha]_{20}^{20}$ –40.0 (*c* 0.5, CH₂Cl₂, 75% ee). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column (*n*-hexane/*i*-PrOH=90/10, 0.6 mL/min, 214 nm, *t*_{minor}=25.72 min, *t*_{maior}=18.54 min).

4.8. General procedure for the removal of *N*-Boc group

A solution of *anti*-**3f** (233 mg, 0.81 mmol) in dichloromethane (25 mL) and TFA (5 mL) was stirred at ambient temperature for 4 h and then the solution was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (elution with petroleum ether/EtOAc=4:1) to provide the title compound **4** (141 mg, 92%).

This is a known compound.^{1b,e} 141 mg, Yield: 92%, orange oil; IR (CH₂Cl₂): ν 3381, 3318, 1754, 1600, 1494, 1453, 1331, 1161, 1089, 1029, 897, 825, 764, 702 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.64 (2H, s), 4.45 (1H, d, *J*=4.8 Hz), 5.22 (1H, ddd, *J*=1.2, 2.0, 4.8 Hz), 6.13 (1H, dd, *J*=2.0, 6.0 Hz), 7.31 (1H, dd, *J*=1.2, 6.0 Hz), 7.32–7.40 (5H, m); ¹³C NMR (100 MHz, CDCl₃, TMS): δ 56.9, 87.0, 123.2, 126.6, 128.1, 128.8, 139.3, 153.2, 172.7; $[\alpha]_D^{20}$ –71.5 (*c* 1.0, CHCl₃); MS (EI) *m/z* (100): 172 [M⁺–NH₃] (6.2), 144 (1.8), 115 (4.6), 107 (8.6), 106 (100), 104 (7.8), 79 (30.7), 77 (16.3); HRMS (EI) calcd for C₁₁H₁₁NO₂ (M⁺): 189.0790, found: 189.0792.

Acknowledgements

We thank the Shanghai Municipal Committee of Science and Technology (08dj1400100-2), National Basic Research Program of China (973)-2010CB833302, the Fundamental Research Funds for the Central Universities and the National Natural Science Foundation of China for financial support (21072206, 20872162, 20672127, 20732008, 20821002, and 20702013).

Supplementary data

Spectroscopic data and chiral HPLC traces of the compounds shown in Tables 1–3 are included in the Supplementary data. Supplementary data related to this article can be found online at doi:10.1016/j.tet.2011.03.046.

References and notes

- 1. Successful examples on the asymmetric vinylogous Mannich (AVM)-type reactions: (a) Martin, S. F.; Lopez, O. D. Tetrahedron Lett. **1999**, 40, 8949–8953; (b) Carswell, E. L.; Snapper, M. L.; Hoveyda, A. H.; Tang, J. Angew. Chem., Int. Ed. 2006, 45, 7230-7233; (c) Mandai, H.; Mandai, K.; Snapper, M. L.; Hoveyda, A. H. J. Am. Chem. Soc. 2008, 130, 17961–17969; (d) Yuan, Z. L.; Jiang, J. J.; Shi, M. Tetrahedron **2009**, 65, 6001–6007; (e) Deng, H. P.; Wei, Y.; Shi, M. Adv. Synth. Catal. **2009**, 351, 2897–2902; (f) Liu, T. Y.; Cui, H. L.; Long, J.; Li, B. J.; Wu, Y.; Ding, L. S.; Chen, Y. C. J. Am. Chem. Soc. 2007, 129, 1878-1879; (g) Sichert, M.; Schneider, C. Angew. Chem., Int. Ed. 2008, 47, 3631–3634 For a review, see: (h) Martin, S. F. Acc. Chem. Res. 2002, 35, 895-904.
- 2. (a) Greene, T. W.; Wuts, P. G. Protective Groups in Organic Synthesis; John Wiley & Sons: New York, NY, 1999; 518–525; (b) The use of α -carbamoyl sulfones for the in situ generation of N-Boc imines, see: Petrini, M. Chem. Rev. 2005, 105, 3949-3977; (c) Liu, Z.; Shi, M. Tetrahedron 2010, 66, 2619-2623.
- 3. (a) Ogasawara, M.; Yoshida, K.; Kamei, H.; Kato, K.; Uozumi, Y.; Hayashi, T. Tetrahedron: Asymmetry **1998**, 9, 1779–1787; (b) Hatano, M.; Yamanaka, M.; Mikami, K. Eur. J. Org. Chem. 2003, 2552–2555.
 Ko, D.-H.; Kim, K. H.; Ha, D.-C. Org. Lett. 2002, 4, 3759–3762.
- 5 (a) Zhao, Q.-Y.; Yuan, Z.-L.; Shi, M. Tetrahedron: Asymmetry 2010, 21, 943-951 Reviews for the silver-catalyzed asymmetric reactions, please see: (b) Naodovic, M.; Yamamoto, H. *Chem. Rev.* **2008**, *108*, 3132–3148; (c) Yanagisawa, A.; Arai, T. *Chem. Commun.* **2008**, 1165–1172; (d) Kaur, P.; Pindi, S.; Wever, W.; Rajale, T.; Li, G. Chem. Commun. 2010, 4330–4333; (e) Kaur, P.; Pindi, S.; Wever, W.; Rajale, T.; Li, G. J. Org. Chem. 2010, 75, 5144–5148; (f) Pindi, S.; Kaur, P.; Shakya, G.; Li, G. *Chem. Biol. Drug Des.* **2011**, 75, 20–25.
- (a) See Ref. 1g; For a review on the effect of additives in asymmetric catalysis, 6. please see: (b) Vogl, E. M.; Groger, H.; Shibasaki, N. Angew. Chem., Int. Ed. 1999, 38, 1570–1577; (c) Giera, D. S.; Sichert, M.; Schneider, C. Org. Lett. 2008, 10, 4259-4262.
- 7. Burr, S. K.; Martin, S. F. Org. Lett. 2000, 2, 3445-3447.
- (a) Wenzel, A. G.; Jacobsen, E. N. J. Am. Chem. Soc. 2002, 124, 12964–12965; (b) Song, J.; Wang, Y.; Deng, L. J. Am. Chem. Soc. 2006, 128, 6048–6049.