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Research Article

Synthesis of ³H- and ²H₄-labelled versions of the hypoxiaactivated pre-prodrug 2-((2-bromoethyl)-2,4-dinitro-6-(((2-(phosphonooxy)ethyl)amino)carbonyl)anilino)ethyl methanesulfonate (PR-104)

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Abstract: 3 H- and 2 H₄-versions of the hypoxia-activated pre-prodrug PR-104 [2-[(2-bromoethyl)-2,4-dinitro-6-[[[2-(phosphonooxy)ethyl]amino]carbonyl]anilino]ethyl methanesulfonate], labelled in the ethylcarboxamide side chain, have been prepared, respectively, by $[^{3}$ H]NaBH₄ reduction of a precursor late stage aldehyde, and by late stage incorporation of deuterium with 2-amino[1,1,2,2- 2 H₄]ethanol. Copyright © 2007 John Wiley & Sons, Ltd.

Keywords: dinitrobenzamide mustard; hypoxia-activated prodrug; PR-104; tritium; deuterium

Introduction

The asymmetric mustard dinitrobenzamide phosphate PR-104 (1)^{1,2} is a soluble pre-prodrug designed to be converted rapidly into the prodrug 2 by serum phosphatases (Scheme 1). This alcohol then diffuses efficiently to, and is activated selectively in, hypoxic regions of solid tumors³ to reactive reduced species (e.g. 3 and 4) that are cytotoxic DNA cross-linking agents.⁴ PR-104 shows excellent *in vivo* activity against the subfraction of hypoxic cells in human tumor xenografts,^{1,2} and has recently begun Phase I clinical trials.

As part of the preclinical development of 1, a tritium-labelled version was required for drug disposition and metabolic studies. As previous work⁴ had shown that oxidative loss of one or both arms of the mustard was a minor pathway in the metabolism of compounds related to 1, it was decided to place the label in the carboxamide side chain. This had the additional benefit of being able to start with prodrug 2 as the late stage

intermediate. A stably labelled version of **1** was also required, to use as an internal standard for the mass spectrometric analysis of preclinical and clinical plasma samples. The presence of natural isotopes of the bromine in **1** necessitated a standard that differed by at least three mass units, and a synthesis that exploited the late stage introduction of commercially available tetra-deutero 2-aminoethanol was devised.

Discussion

The synthesis of tritium-labelled 1 (1-T) is shown in Scheme 2. A number of oxidizing agents were evaluated for the conversion of alcohol 2 to aldehyde 5, the best being Dess-Martin periodinane and the eventually employed pyridinium chlorochromate (PCC). Reduction of 5, used in excess) with [3H]NaBH₄ (60 Ci/mmol, diluted two-fold with unlabelled NaBH₄) in ethanol/ THF at 20°C for 20 min gave the labelled alcohol 6 in good yield, with a measured specific activity of 6.3 Ci/ mmol. Compound 6 (diluted 10-fold with unlabelled 2) was phosphorylated as previously reported for 2, with di-tert-butyl diisopropylphosphoramidite/1H-tetrazole followed by oxidation with 70% hydrogen peroxide, to provide the phosphate ester 7. This was cleaved with trifluoroacetic acid to give phosphate diacid 1-T in 65% overall yield from 6, with a measured specific activity of 0.77 Ci/mmol.

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$$O_2N$$
 NO_2
 H
 $NHOH$
 NH_2
 H
 NH_2
 NH_2
 H
 NH_2
 H
 NH_2
 NH_2
 H
 NH_2
 NH_2
 H
 NH_2
 N

(i) Serum phosphatases; (ii) cellular reductases under hypoxia.

Scheme 1 Mechanism of activation of PR-104.

$$O_2N$$
 O_2N
 O_2N

Reagents: (i)PCC; (ii) [T]NaBH₄/EtOH; (iii) i-Pr₂NP(OtBu)₂/1*H*-tetrazole, then H_2O_2 ; (iv) TFA/CH₂Cl₂

Scheme 2 Synthesis of 3H PR-104.

For analysis and quantitation of the preclinical and clinical plasma samples from treatment with **1** by mass spectrometry, an internal standard, differing by at least three mass units, was required. The commercial availability of tetra-deuterated 2-aminoethanol allowed the opportunity of incorporating four D atoms in the carboxamide side chain to give **1-D4**. Rather than repeat the standard synthesis, which involved introduction of the side chain early in the synthesis, an alternative route was devised where the side chain was incorporated as late as possible (after elaboration of the mustard group) (Scheme 3).

In this procedure, the known⁵ *tert*-butyl ester **8** was reacted with diethanolamine to give the diol **9**, which was converted to the dimesylate **10** with mesyl chloride in pyridine. This was treated with lithium bromide in DMF, and the resulting bromomesylate **11** was separated from the corresponding dibromo compound and

unreacted 10 by column chromatography. The ester 11 was then cleaved with trifluoroacetic acid to give the key acid 12 in 33% overall yield from 8.

Activation of acid 12 with a number of coupling reagents (DECP, CDI, EDCI) proved unsuccessful. However, conversion of 12 to the corresponding acid chloride (oxalyl chloride, catalytic DMF) followed by reaction with 2-amino[1,1,2,2-2H₄]ethanol in the presence of triethylamine gave the labelled alcohol 13 in good yield. HPLC analysis showed this contained 4.4% of the corresponding chloromesylate, generated by chloride exchange of the bromide. The alcohol 13 was phosphorylated with di-tert-butyl diisopropylphosphoramidite/1H-tetrazole viously described for 2, and the resulting phosphate ester 14 was cleaved with trifluoroacetic acid to give phosphate diacid 1-D4, in 16% overall yield from 8. Deuterium incorporation (98%) was confirmed by high resolution FAB mass spectrometry.

Reagents: (i) HN(CH₂)CH₂OH)₂; (ii) MsCl; (iii) LiBr; (iv) TFA; (v) (COCl)₂, then H₂N(CD₂)₂OH; (vi) i-Pr₂NP(OtBu)₂/1H-tetrazole, then H₂O₂; (vii) TFA/CH₂Cl₂

Scheme 3 Synthesis of 2H4 PR-104.

Experimental section

Analyses were performed by the Microchemical Laboratory, University of Otago, Dunedin, NZ. Melting points were determined using an Electrothermal Model 9200 digital melting point apparatus, and are as read. NMR spectra were measured on a Bruker DRX-400 spectrometer, and referenced to Me₄Si. Mass spectra were recorded on a Varian VG-70SE spectrometer. HPLC was carried out using a Bondclone 10 C18 reverse-phase silica gel column, with a Phillips PU4100M gradient elution pump and a Phillips PU 4120 diode array detector, and eluting with the appropriate ratios of 95% MeCN/5% water (solvent A) and 0.45 M ammonium formate buffer (solvent B; pH 3.45). The specific activities of 6 and 1-T were determined by quantitation of ³H-labelled samples by HPLC, and subsequent off-line liquid scintillation counting (Packard 1500 Tri-Carb) of the isolated HPLC peaks.

2-((2-Bromoethyl)-2,4-dinitro-6-(((2-oxoethyl)amino)carbonyl)anilino)ethyl methanesulfonate (5)

A solution of 2-[(2-bromoethyl)-2-[[(2-hydroxyethyl)amino|carbonyl|-4,6-dinitroanilino|ethyl methanesulfonate¹ (2) (600 mg, 1.20 mmol) in dry CH₂Cl₂ (125 ml) was stirred with 3A powdered molecular sieves (1.5g) for 5 min at room temperature, then treated with powdered pyridinium chlorochromate 3.60 mmol). The mixture was stirred at room temperature for a further 15 min, then treated with water (50 ml) and filtered. The organic phase was washed with water (50 ml), dried and concentrated under reduced pressure. The residue was chromatographed on silica gel, eluting with EtOAc/petroleum ether (4:1). The middle fractions were combined and rechromatographed, eluting with EtOAc/petroleum ether (6:1) to give 5 (413 mg, 69%) as a yellow gum which was used directly: ${}^{1}H$ NMR [(CD₃)₂SO] δ 9.61 (s, 1H), 9.15 (t, $J = 5.4 \,\mathrm{Hz}$, 1H), 8.77 (d, $J = 2.8 \,\mathrm{Hz}$, 1H), 8.42 (d, $J = 2.8 \,\mathrm{Hz}$, 1H), 4.28 (t, $J = 5.4 \,\mathrm{Hz}$, 2H), 4.20 (t, $J = 5.5 \,\text{Hz}$, 2H), 3.62–3.54 (m, 2H), 3.54–3.43 (m, 4H), 3.12 (s, 3H).

(3H)2-((2-Bromoethyl)-2-(((2-hydroxyethyl)amino)carbonyl)-4,6-dinitroanilino)ethyl methanesulfonate (6)

A freshly prepared solution of [3H]NaBH₄ (0.84 mg, 0.02 mmol, nominally 1.2 Ci at 60 Ci/mmol) (Amersham Biosciences, UK) and unlabelled NaBH₄ (0.76 mg, 0.02 mmol) in EtOH (1.5 ml) was added dropwise to a stirred solution of 5 (99 mg, 0.20 mmol) in THF (5 ml) at 20°C over a period of 5 min. The mixture was stirred at 20°C for a further 15 min, then quenched with 0.1 N aqueous MsOH (1 ml), diluted with water (20 ml) and extracted with EtOAc (2 \times 20 ml). The combined organic layer was washed with water $(2 \times 20 \text{ ml})$, dried, and concentrated under reduced pressure. Chromatography on silica gel, eluting with EtOAc/petroleum ether (6:1), gave unreacted 5 (8 mg), then 6 (60 mg, 75%); (6.3 Ci/mmol) as a yellow gum, identical on HPLC with unlabelled 2: 1H NMR [(CD₃)₂SO] δ 8.76–8.70 (m, 2H), 8.35 (d, $J = 2.8 \,\text{Hz}$, 1H), 4.83-4.75 (m. 1H), 4.28 (t, J = 5.5 Hz, 2H), 3.61-3.52 (m, 4H), 3.51-3.42 (m, 4H), 3.39-3.31 (m, 2H), 3.13 (s, 3H).

(³H)2-((2-Bromoethyl)-2-(6-*tert*-butoxy-8,8-dimethyl-6-oxido-5,7-dioxa-2-aza-6-phosphanon-1-anoyl)-4,6-dinitroanilino)ethyl methanesulfonate (7)

A stirred solution of 6 (30 mg, 0.06 mmol; 235 GBq/ mmol [6.3 Ci/mmol] and unlabelled 2 (270 mg, 0.54 mmol) in dry DMF (1.5 ml) at 10°C was treated with 1H-tetrazole (67 mg, 0.96 mmol), followed by the dropwise addition of di-tert-butyl diisopropylphosphoramidite (0.26 ml, 0.78 mmol), 95%). The mixture was warmed to room temperature of 2h, then cooled to 0°C and treated slowly with a solution of 70% aqueous H_2O_2 (0.15 ml) in THF (0.2 ml). The mixture was stirred at 10°C for a further 30 min, then poured into cold water (25 ml) and extracted with EtOAc (2×10 ml). The combined organic layers were washed with 5% aqueous $Na_2S_2O_5$ (10 ml), water (4 × 10 ml), dried, and evaporated under reduced pressure. The residue was chromatographed on silica gel, eluting with EtOAc/ petroleum ether, and the eluant was concentrated under reduced pressure below 30°C to a small volume and diluted with hexane to precipitate 7 (303 mg, 73%) as a yellow oil: ${}^{1}\text{H NMR [(CD_{3})_{2}SO]} \delta 8.93 \text{ (t, } J = 5.6 \text{ Hz,}$ 1H), 8.75 (d, J = 2.8 Hz, 1H), 8.34 (d, J = 2.8 Hz, 1H), 4.28 (t, J = 5.4 Hz, 2H), 4.06-3.97 (m, 2H), 3.61-3.43(m, 8H), 3.13 (s, 3H), 1.43 (s, 18H).

(³H)2-((2-Bromoethyl)-2,4-dinitro-6-(((2-(phosphonooxy)ethyl)-amino)carbonyl)anilino)ethyl methanesufonate (1-T)

A solution of **7** (299 mg, 0.43 mmol) in CH₂Cl₂ (2 ml) was treated with TFA (2 ml) at room temperature for 1 h, then concentrated to near dryness under reduced pressure below 30°C. The residual gum was dissolved in dry MeCN (0.2 ml) and the solution was diluted with dry CH₂Cl₂ (10 ml), filtered, seeded, and refrigerated at 5°C for 16 h. The separated solid was collected, washed with CH₂Cl₂ and dried under high vacuum to give **1-T** (223 mg, 89%; 98.5% pure by HPLC) (0.77 Ci/mmol) as yellow crystals. ¹H NMR [(CD₃)₂SO] δ 11.0 (br, 2H), 8.97–8.90 (m, 1H), 8.74 (d, J=2.8 Hz, 1H), 8.37 (d, J=2.8 Hz, 1H), 4.29 (t, J=5.4 Hz, 2H), 3.98 (q, J=6.2 Hz, 2H), 3.62–3.43 (m, 8H), 3.13 (s, 3H).

tert-Butyl 2-(bis(2-hydroxyethyl)amino)-3,5-dinitrobenzoate (9)

A solution of *tert*-butyl 2-chloro-3,5-dinitrobenzoate⁵ (8) (3.27 g, 10.8 mmol) in dioxane (10 ml) at 10 °C was

treated with diethanolamine (2.84 g, 27 mmol) and vigorously stirred at room temperature for 3 h. Dilution with water provided a solid which was purified by column chromatography on silica gel, eluting with EtOAc/petroleum ether (4:1), followed by recrystallization from EtOAc/iPr₂O to give *tert*-butyl 2-[bis(2-hydroxyethyl)amino]-3,5-dinitrobenzoate (**9**) (3.78 g, 94%) as a yellow solid: m.p. 117–118°C. ¹H NMR [(CD₃)₂SO] δ 8.69 (d, J = 2.8 Hz, 1H), 8.40 (d, J = 2.8 Hz, 1H), 4.53 (t, J = 5.6 Hz, 2H), 3.57 (q, J = 5.9 Hz, 4H), 3.19 (t, J = 6.1 Hz, 4H), 1.60 (s, 9 H). Analytically required for C₁₅H₂₁N₃O₈: C, 48.52; H, 5.7; N, 11.32. Found: C, 48.54; H, 5.8; N, 11.31.

tert-Butyl 2-(bis(2-((methylsulfonyl)oxy)ethyl)amino)-3,5-dinitrobenzoate (10)

A stirred solution of **9** (3.76 g, 10.1 mmol) in pyridine (5 ml) at 0°C was treated dropwise with MsCl (1.90 ml, 24.5 mmol), and then allowed to come to room temperature for 1 h. The mixture was diluted with water (50 ml) and the resulting solid was purified by column chromatography on silica gel, eluting with EtOAc, followed by recrystallization from EtOAc/iPr₂O to give **10** (4.79 g, 90%) as a yellow solid: m.p. $105.5-106^{\circ}$ C. ¹H NMR [(CD₃)₂SO] δ 8.81 (d, J=2.8 Hz, 1H), 8.53 (d, J=2.8 Hz, 1H), 4.30 (t, J=5.5 Hz, 4H), 3.49 (t, J=5.5 Hz, 4H), 3.13 (s, 6H), 1.61 (s, 9 H). Analytically required for C₁₇H₂₅N₃O₁₂S₂: C, 38.71; H, 4.78; N, 7.97; S, 12.16. Found: C, 38.78; H, 4.82; N, 7.96; S, 12.30.

tert-Butyl 2-((2-bromoethyl)-(2-((methylsulfonyl)oxy) ethyl)amino)-3,5-dinitrobenzoate (11)

A mixture of **10** (5.00 g, 9.48 mmol) and LiBr (0.99 g, 11.4 mmol) in DMF (10 ml) was stirred at 50 °C for 1.5 h, then diluted with water and refrigerated. The precipitated semi-solid was dried and chromatographed on silica gel, eluting with EtOAc/petroleum ether (9:11) to give firstly *tert*-butyl 2-[bis(2-bromoethyl)amino]-3, 5-dinitrobenzoate (1.60 g) as a yellow solid: 1 H NMR [(CD₃)₂SO] δ 8.80 (d, J = 2.8 Hz, 1H), 8.50 (d, J = 2.8 Hz, 1H), 3.61–3.54 (m, 4H), 1.61 (s, 9 H).

Later fractions gave **11** (2.11 g, 43%) as a yellow solid: m.p. (EtOAc/iPr₂O) 97–98°C. ¹H NMR [(CD₃)₂SO] δ 8.81 (d, J = 2.8 Hz, 1H), 8.52 (t, J = 2.8 Hz, 1H), 4.29 (t, J = 5.5 Hz, 2H), 3.62–3.55 (m, 2H), 3.54–3.46 (m, 4H), 3.12 (s, 3H), 1.60 (s, 9 H). Analytically required for C₁₆H₂₂BrN₃O₉S: C, 37.51; H, 4.33; N, 8.2; Br, 15.60. Found: C, 37.65; H, 4.51; N, 8.12; Br, 15.69.

Further elution with EtOAc gave unreacted 10 (0.92 g).

2-((2-Bromoethyl)-(2-((methylsulfonyl)oxy)ethyl) amino)-3.5-dinitrobenzoic acid (12)

A solution of 11 (2.70 g, 5.27 mmol) in TFA (12 ml) was stirred at room temperature for 3h, then diluted with water (35 ml) to complete precipitation of the product. Recrystallization from EtOAc/iPr₂O gave **12** (2.19 g, 91%) as a yellow solid: m.p. 137-138°C. ¹H NMR [(CD₃)₂SO] δ 14.18 (br, 1H), 8.82 (d, J = 2.8 Hz, 1H), 8.60 (d, $J = 2.8 \,\text{Hz}$, 1H), 4.27 (t, $J = 5.4 \,\text{Hz}$, 2H), 3.59-3.46 (m, 6H), 3.12 (s, 3H). Analytically required for C₁₂H₁₄BrN₃O₉S.0.5H₂O: C, 30.98; H, 3.25; N, 9.03. Found: C, 32.12;32.17; H, 3.10;3.07; N, 9.18;9.23.

2-((2-Bromoethyl)-2-(((2-hydroxy(1,1,2,2-2H₄)ethyl)amino)carbonyl)-4.6-dinitroanilino)ethyl methanesulfonate (13)

A suspension of powdered 12 (3.12g, 6.84 mmol) in CH_2Cl_2 (40 ml) was treated with oxalvl chloride (0.88 ml, 10.3 mmol) and DMF (1 drop) and stirred at room temperature for 1 h. Evaporation of the volatiles under reduced pressure below 25°C, followed by azeotroping with benzene gave the crude acid chloride. This was dissolved in THF (15 ml) and added dropwise to a stirred solution of 2-amino[1,1,2,2-2-H₄lethanol (0.58 g, 8.91 mmol) (Cambridge Isotope Laboratories, Andover, MA 01810, USA) and Et₃N (1.43 ml, 10.3 mmol) in dioxane/THF (1:1) (25 ml) at 0°C. The mixture was stirred at 0°C for a further 10 min, then poured into ice-cold 0.1 N aqueous MsOH (120 ml) and extracted with EtOAc (2×80 ml). The combined organic phase was washed with water (2 × 40 ml), dried, and concentrated under reduced pressure. Chromatography on silica gel, eluting with EtOAc, followed by precipitation of the product from a CH₂Cl₂ solution with hexane, gave 13 (2.96 g, 86%) as a yellow gum; ¹H NMR [(CD₃)₂SO] δ 8.74 (d, $J = 2.8 \,\mathrm{Hz}$, 1H), 8.71 (s, 1H), 8.35 (d, J =2.8 Hz, 1H), 4.76 (s, 1H), 4.28 (t, J = 5.5 Hz, 2H), 3.60-3.54 (m, 2H), 3.51-3.43 (m, 4H), 3.13 (s, 3H). ¹³C NMR [(CD₃)₂SO] δ 165.3, 145.8, 145.3, 140.9, 136.2, 127.5, 122.1, 67.5, 58.4 (br, CD₂), 5.43, 51.0, 41.3 (br CD₂), 36.5, 29.7. HRMS(FAB) calculated, for $C_{14}H_{16}D_4^{79}BrN_4O_9S$ (MH⁺) m/z 503.0385; found, 503.0375.

HPLC analysis of the product indicated the presence of 3.4% of the corresponding chloromesylate.

2-((2-Bromoethyl)-2-((3,3,4,4- 2 H₄)-6-*tert*-butoxy-8, 8-dimethyl-6-oxido-5.7-dioxa-2-aza-6-phosphanon-1-anoyl)-4,6-dinitroanilino)ethyl methanesulfonate

A stirred solution of 13 (1.55g, 3.08 mmol) and 1Htetrazole (345 mg, 4.93 mmol) in dry DMF (8 ml) at 10°C was treated dropwise with di-tert-butyl diisopropylphosphoramidite (1.33 ml, 4.00 mmol, 95%). The mixture was warmed at room temperature for 2 h, then cooled to 0°C and treated slowly with a solution of 70% aqueous H_2O_2 (0.8 ml) in THF (1 ml). The mixture was warmed to 10°C for 0.5 h, then poured into ice-cold water (100 ml) and extracted with EtOAc (2×40 ml). The combined organic phases were washed with 5% aqueous $Na_2S_2O_5$ (40 ml) and water (4 × 40 ml), dried, and concentrated under reduced pressure below 30°C. The residue was chromatographed on silica gel, eluting with EtOAc/petroleum ether (4:1), and the eluant was concentrated under reduced pressure below 30°C to a small volume and diluted with hexane to give 14 (1.44 g, 67%) as an unstable yellow oil. ¹H NMR [(CD₃)₂SO] δ 8.92 (s, 1H), 8.76 (d, J=2.8 Hz, 1H), 8.34 (d, J = 2.8 Hz, 1H), 4.28 (t, J = 5.4 Hz, 2H), 3.58 (t, J =7.3 Hz, 2H), 3.51-3.43 (m, 4H), 3.13 (s, 3H), 1.43 (s, 18H).

2-((2-Bromoethyl)-2,4-dinitro-6-(((2-(phosphonooxy) (1,1,2,2-2H₄)ethyl)-amino)carbonyl)anilino)ethyl methanesufonate (1-D4)

A stirred solution of **14** (730 mg, 1.05 mmol) in CH₂Cl₂ (5 ml) was treated with TFA (5 ml) for 1 h at room temperature, then concentrated to near dryness under reduced pressure below 30°C. The residual gum was dissolved in dry MeCN (0.5 ml) and the solution was diluted with dry CH₂Cl₂ (25 ml), filtered, and then refrigerated at 5°C for 30 h. The separated solid was collected, washed with CH2Cl2, and dried under high vacuum at room temperature to give 1-D4 (529 mg, 86%) as yellow crystals: m.p. 133-135°C. HPLC analysis indicated the product was 94.7% pure, with 4.7% of the corresponding chloromesylate. ¹H NMR [(CD₃)₂SO] δ 8.90 (s, 1H), 8.75 (d, J = 2.8 Hz, 1H), 8.37 (d, $J = 2.8 \,\text{Hz}$, 1H), 4.28 (t, $J = 5.4 \,\text{Hz}$, 2H), 3.58 (t, $J = 7.0 \,\text{Hz}$, 2H), 3.51–3.43 (m, 4H), 3.13 (s, 3H). P(OH)₂ signals not observed. ¹³C NMR [(CD₃OD] δ 168.4, 147.7, 147.6, 143.5, 137.8, 128.9, 123.8, 69.2, 65.0, (br, CD₂), 56.9, 53.4, 41.0 (br, CD₂), 37.3, 29.7. HRMS(FAB) calculated for C₁₄H₁₇D₄⁷⁹BrN₄O₁₂PS (MH⁺) m/z 583.0049; found, 583.0050.

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