

Notes

Synthesis of Steroidal Nitrosoureas with Antitumor Activity

Hing-Yat P. Lam,* Asher Begleiter, Gerald J. Goldenberg,

Department of Medicine, University of Manitoba and the Manitoba Institute of Cell Biology, Winnipeg, Manitoba, R3E 0V9, Canada

and Chiu-Ming Wong

Department of Chemistry, University of Manitoba, Winnipeg, Manitoba R3T 2N2, Canada. Received August 1, 1978

Four steroidal nitrosoureas with structures which may permit specific binding to estrogen receptor were synthesized. Inhibitory activity was observed against the growth of the DMBA-induced transplantable rat mammary tumor 13762.

Among the cancer chemotherapeutic agents, two of the best known groups are the steroid hormones and the alkylating agents. In the last two decades several reports have appeared using estrogenic steroid hormones as carriers of alkylating agents in an attempt to produce antitumor agents with greater specificity of action.¹⁻⁵ However, results from this approach have been disappointing. Only two agents, namely, estramustine phosphate and estradiol mustard, have been used in clinical trials.⁶⁻⁸

The discovery of estrogen receptor (ER) with specific binding affinity in 50-60% of human breast cancer⁹ provided a new approach in this field. More specific antitumor activity may be achieved by taking advantage of the presence of ER which may selectively accumulate certain cytotoxic agents with binding affinity for ER.

It has been established that in order to obtain the highest binding affinity for ER the estrogenic steroid hormone with an *estra-1,3,5(10)*-triene skeleton must have the 3- and 17 β -hydroxyl groups available, presumably to form hydrogen bonds with ER protein at the binding site.^{10,11} In estramustine phosphate and estradiol mustard, these hydroxyl groups are used to link the steroid to the alkylating moiety. Binding sites of these two agents suggested that they have very little, if any, binding affinity for ER. It has been suggested that the relatively weak antitumor activity of these agents against breast cancer is due partly to their inability to bind to ER and thereby to accumulate in ER-positive tumor cells.^{12,13}

We have been interested in the synthesis of steroidal nitrosoureas with moieties which may permit binding to ER and may demonstrate antitumor activity. This paper reports the synthesis of two groups of steroidal nitrosoureas which are analogues of 3,17 β -estradiol. In one group (5a,b) the 3- and 17 β -hydroxyl functions were preserved and in the other group (6a,b) the 17 β -hydroxyl function was replaced by a ureidal-NH function so that the hydrogen-bond-forming ability at this position might be retained.

Chemistry. Synthesis of steroidal nitrosoureas was carried out according to Hardegger's one-step approach for the synthesis of streptozotocin and its analogues.¹⁴ Reaction of an amine with an *N*-alkyl-*N*-nitrosocarbonyl azide gave directly a nitrosourea having the nitroso group in the desired position. The initial step in the synthesis of 5a and 5b was conversion of the 3-hydroxyl group of estrone to the corresponding benzyl ether. Reaction of 3-(benzyloxy)estra-1,3,5(10)-trien-17-one (1) with dimethylsulfonium methylide yielded the 17 β -spirooxirane 2 which was converted into the hydroxy azide derivative

Table I. Antitumor Activity of Nitrosourea Derivatives from Estrogenic Steroid Hormone against Rat DMBA-Induced, Transplantable Mammary Tumor 13762

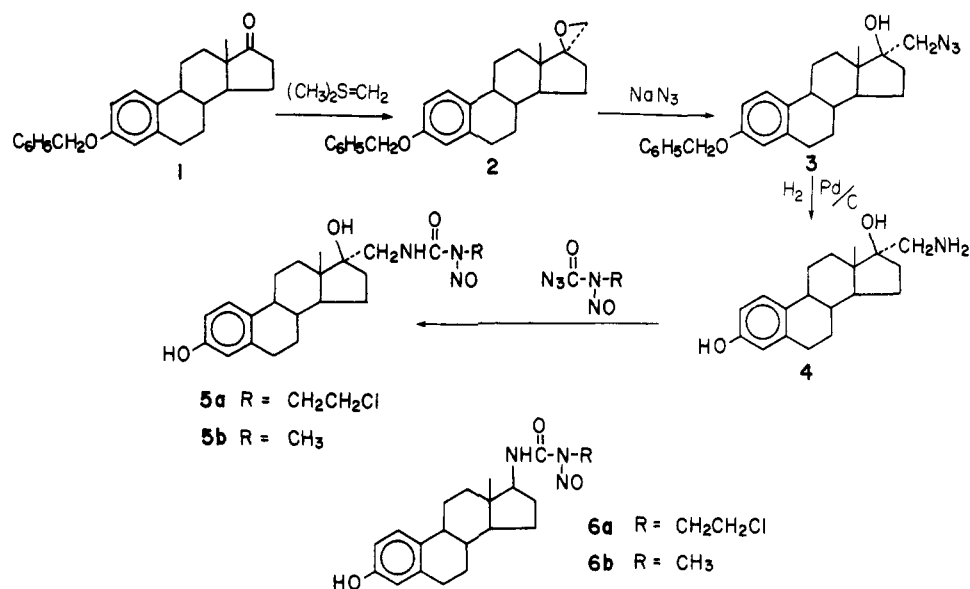
compd	dose, (mg/kg)/day	% inhibition ^a
5a	2	0
	10	35
	40	85
5b	2	0
	10	10
	40	23
6a	2	0
	10	50
	40	100
6b	2	4
	10	15
	40	15
CCNU	10	80

^a Tumor growth in groups of six rats each, treated with the dose of compound indicated, was compared to that in 12 control rats treated with sesame oil, and the result is expressed as mean percent inhibition (% inhibition = $C - T/C \times 100$, where T is tumor size in treated rats and C that in control rats).

3 by treatment with NaN₃ and H₃BO₃. Hydrogenation reduced the azide group of 3 to amine and at the same time cleaved the benzylic ether to form the amino alcohol 4, which reacted with the alkyl nitrosocarbonyl azide to yield the corresponding nitrosoureas 5a and 5b. Similarly, reaction of the 17 β -amino-3-hydroxyestra-1,3,5(10)-triene, obtained from reduction of estrone oxime, with the *N*-alkyl-*N*-nitrosocarbonyl azide furnished 6a and 6b (see Scheme I).

Biological Data. Antitumor activity of compounds 5a,b and 6a,b was evaluated against the 13762 DMBA induced and transplantable rat mammary adenocarcinoma in Fischer/344 rats. The rats were implanted with 2- to 3-mm³ fragments of tumor. Treatment was initiated 1 day after transplantation and continued for 20 days. Compounds were administered subcutaneously as suspensions in sesame oil. During the first 10 days, groups of six rats each were treated daily with the dose indicated and then with double the dose every 2 days from day 11 through day 19. On day 21, tumor size was measured with a caliper and expressed as the arithmetic mean of two perpendicular diameters. The antitumor activity of the four compounds is presented in Table I. Compounds 5a, 6a (at doses of 10 and 40 (mg/kg)/day), and 5b (at 40 (mg/kg)/day)

Scheme 1



showed definite inhibition of tumor growth; results were statistically significant, as evaluated by the Student's *t* test. Since *N*-(2-chloroethyl)-*N'*-cyclohexyl-*N*-nitrosourea (CCNU), a nonsteroidal nitrosourea, also showed inhibition of tumor growth, a more detailed pharmacological evaluation will be required to determine whether these steroidal nitrosoureas will demonstrate a higher therapeutic index. Binding studies of these compounds to tumor ER will be undertaken.

Experimental Section

All melting points are uncorrected. IR spectra were recorded on a Perkin-Elmer 701A spectrometer; NMR spectra were obtained with a Varian A-56/60A spectrometer using CDCl₃ as solvent unless specified otherwise and tetramethylsilane as internal standard; mass spectra were obtained on a Finnigan 1015, RF-quadrupole mass spectrometer. Purity of oily products was established by demonstrating a single spot on TLC. Microanalysis was performed by Dr. C. Daesale, Organic Microanalysis, Montreal.

3-(Benzyloxy)estra-1,3,5(10)-trien-17-one (1). Estrone (12 g, 44 mmol) was refluxed for 44 h with benzyl chloride (120 mL, 1.0 mol) in acetone (600 mL) containing a suspension of pulverized K₂CO₃ (60 g). The mixture was cooled to room temperature, and 200 mL of CHCl₃ was added. The precipitated K₂CO₃ was removed by filtration. The solvent and excess benzyl chloride were removed by evaporation in vacuo. Crystallization from CHCl₃-MeOH provided the ketone **1** (15 g, 94% yield) as a colorless crystal: mp 127–128 °C, lit.¹⁵ mp 130–131 °C; IR (CH₂Cl₂) 1735 cm⁻¹ (C=O); NMR δ 0.92 (s, 3 H, C₁₈-H), 5.08 (s, 2 H, benzylic H), 6.75–7.60 (m, unresolved, 8 H, Ar H).

(17*S*)-Spiro[3-(benzyloxy)estra-1,3,5(10)-trien-17,2'-oxirane] (2). A solution of ketone **1** (15 g, 42 mmol) in THF (100 mL) was added to a solution of freshly prepared dimethylsulfonium methylide (125 mmol) in Me₂SO (100 mL) and THF (100 mL) at 0 °C.^{16,17} The mixture was stirred at 0 °C for 10 min and then at room temperature for 1 h. It was then poured into ice water and extracted with ether. The extract was washed (water and brine), dried (MgSO₄), and evaporated to dryness. Crystallization from CHCl₃-EtOAc furnished **2** as a colorless crystalline product in three crops (15 g, 95% yield): mp 155–157 °C; IR (CH₂Cl₂) 1030 cm⁻¹ (oxirane); NMR δ 0.93 (s, 3 H, C₁₈-H), 2.88 (AB quartet, *J* = 5 Hz, 2 H, oxirane CH₂), 5.08 (s, 2 H, benzylic H), 6.75–7.60 (m, unresolved, 8 H, Ar H); MS *m/e* 374 (M⁺ - CH₂O), 91 (C₆H₅CH₂⁺).

17α-(Azidomethyl)-3-(benzyloxy)-17β-hydroxyestra-1,3,5(10)-triene (3). A mixture of the oxirane **2** (8.8 g, 23 mmol), activated NaN₃ (15 g, 0.23 mol), and pulverized H₃BO₃ (6 g, 97 mmol) in DMF (100 mL) was refluxed under N₂ for 4 h.¹⁸ The

mixture was poured into H₂O and extracted with ether. The extract was washed (water and brine), dried (MgSO₄), and evaporated to give the azide **3** as a pale-yellow oil (9.2 g, 94% yield), which showed a single spot on TLC (silica; benzene-EtOAc, 9:1; *R_f* 0.49) and was adequate for further transformation. Crystallization from CHCl₃-hexane gave pale-yellow crystals: mp 96–98 °C; IR (CH₂Cl₂) 3600 (OH), 2120 cm⁻¹ (N₃); NMR δ 0.93 (s, 3 H, C₁₈-H), 3.44 (AB quartet, *J* = 12 Hz, 2 H, 17α-CH₂-), 5.02 (s, 2 H, benzylic H), 6.75–7.60 (m, unresolved, 8 H, Ar H).

17α-(Aminomethyl)-3,17β-dihydroxyestra-1,3,5(10)-triene (4). A solution of the azide **3** (5.2 g, 13 mmol) in absolute ethanol (250 mL) was hydrogenated over 600 mg of Pd/C at atmospheric pressure and at room temperature for 39 h. The catalyst was removed by filtration, and the filtrate was evaporated to yield a colorless solid. Crystallization from MeOH-CHCl₃ provided a colorless crystalline product (2.6 g, 70%): mp 242–243 °C; IR (Nujol) 3280, 3170 cm⁻¹ (NH₂); MS *m/e* 301 (M⁺). Anal. (C₁₉H₂₇N₂O₂) C, H, N.

(2-Chloroethyl)nitrosocarbamoyl Azide. A solution of 2-chloroethyl isocyanate (20 g, 0.19 mol) in dry CCl₄ (200 mL) at 0 °C was bubbled with dry HCl for 3 h. The solvent was evaporated in vacuo to yield (2-chloroethyl)carbonyl chloride as an oily product (27 g, 99% yield): IR (CH₂Cl₂) 1700 (C=O), 3430 cm⁻¹ (NH).

Activated NaN₃ (8.0 g, 0.12 mol) was added to a solution of (2-chloroethyl)carbonyl chloride (13 g, 89 mmol) in acetone (100 mL) at 0 °C. The mixture was stirred at 0 °C for 3 h and then filtered. The precipitate was washed exhaustively with ether. The filtrate was concentrated in vacuo to a small volume and then combined with the washings. The combined organic solution was washed (ice-cold water and brine), dried (MgSO₄), and evaporated to dryness. Crystallization from benzene-hexane gave (2-chloroethyl)carbonyl azide as a colorless crystalline product (15 g, 82% yield): mp 48–49 °C, lit.¹⁹ mp 49.6–50.2 °C; IR 1710 (C=O), 2150, 2170 (N₃), 3420 cm⁻¹ (NH); NMR δ 3.62 (m, unresolved, 4 H, ClCH₂CH₂N-), 5.93 (br s, 1 H, NH).

To a solution of (2-chloroethyl)carbonyl azide (1.2 g, 8.1 mmol) in pyridine, 5 M NOCl (3 mL) in glacial acetic acid was added dropwise over a period of 5 min while the temperature was kept at 0 °C. The mixture was stirred at 0 °C for a further 5 min and then poured into ice water and extracted with ether. The ether layer was washed (ice-cold 2 N HCl two times, 10% NaHCO₃, brine successively) and dried (MgSO₄). Evaporation of the solvent gave a yellow oil which was applied to a column containing 50 g of silica gel. Elution of the column with ether-hexane (1:3) yielded 0.70 g of (2-chloroethyl)nitrosocarbonyl azide (49% yield) as a yellow oil, which showed a single spot on TLC (silica; hexane-ether, 6:1; *R_f* 0.32): IR (CH₂Cl₂) 2180 (N₃), 1720 (C=O), 1530 cm⁻¹ (NO); NMR δ 3.52 and 4.17 (A₂B₂ system, 2 t, *J* = 7 Hz, 4 H, -N(NO)CH₂CH₂Cl), lit.¹⁹ δ 3.50 and 4.15.

N-(2-Chloroethyl)-N'-[(3,17 β -dihydroxyestra-1,3,5(10)-trien-17 α -yl)methyl]-N-nitrosoarea (5a). To a solution of the amine **4** (0.70 g, 2.3 mmol) in pyridine (15 mL) at 0 °C, (2-chloroethyl)nitrosocarbonyl azide (0.42 g, 2.3 mmol) in ether (1.3 mL) was added dropwise with stirring. The mixture was stirred at 0 °C for 3 h and then ice water was added, followed by extraction with ether. The ether extract was washed (ice-cold 2 N HCl, 10% NaHCO₃, and brine successively), dried (MgSO₄), and evaporated to dryness. Crystallization from ether yielded the nitrosoarea **5a** as a pale-yellow crystalline product (0.70 g, 70% yield): mp 116–119 °C dec; IR (CH₂Cl₂) 3560 (OH), 3410 (NH), 1720 (C=O), 1525 (CNH), 1490 cm⁻¹ (NO); NMR (acetone-d₆) δ 1.00 (s, 3 H, C₁₈-H), 3.66, 4.28 (A₂B₂ system, 2 t, J = 7 Hz, 4 H, -N(NO)CH₂CH₂Cl), 3.70 (m, 2 H, 17 α -CH₂N-), 6.60–7.20 (m, 3 H, Ar H); MS m/e 327 (M⁺ - HON=NCH₂CH₂Cl). Anal. (C₂₉H₃₀ClN₃O₄) C, H, Cl, N.

N-Methyl-N'-[(3,17 β -dihydroxyestra-1,3,5(10)-trien-17 α -yl)methyl]-N-nitrosoarea (5b). Following the general procedure described for the synthesis of **5a**, reaction of the amine **4** (1.4 g, 4.8 mmol) with methylnitrosocarbonyl azide¹⁴ (4.8 mmol) gave compound **5b** as a pale-yellow crystalline product after crystallization from CH₃OH-ether-hexane (1.1 g, 60% yield): mp 144–146 °C dec; NMR (acetone-d₆) δ 1.00 (s, 3 H, C₁₈-H), 3.22 (s, 3 H, -N(NO)CH₃), 3.40 (s, ~3 H, CH₃ of CH₃OH), 3.65 (m, 2 H, 17 α -CH₂N), 6.60–7.20 (m, 3 H, Ar H); MS m/e 327 (M⁺ - HON=NCH₃). Anal. (C₂₁H₂₉N₃O₄·CH₃OH) C, H, N.

N-(2-Chloroethyl)-N'-[(3-hydroxyestra-1,3,5(10)-trien-17 β -yl)-N-nitrosoarea (6a). Following the general procedure described above, reaction of 17 β -amino-3-hydroxyestra-1,3,5(10)-triene²⁰ (2.5 g, 9.2 mmol) with (2-chloroethyl)nitrosocarbonyl azide (9.2 mmol), after crystallization from CH₃OH, gave **6a** as a colorless crystal (1.6 g, 43% yield): mp 87–90 °C dec; NMR δ 0.86 (s, 3 H, C-18), 3.66, 4.26 (A₂B₂ system, 2 t, J = 7 Hz, 4 H, -N(NO)CH₂CH₂Cl), 4.00 (m, 1 H, C₁₇-H), 6.60–7.35 (m, 3 H, Ar H); MS m/e 405 (M⁺), 297 (M⁺ - HON=NCH₂CH₂Cl), 255 (M⁺ - side chain). Anal. (C₂₁H₂₈ClN₃O₃) C, H, Cl, N.

N-Methyl-N'-[(3-hydroxyestra-1,3,5(10)-trien-17 β -yl)-N-nitrosoarea (6b). Following the general procedure described above, reaction of 17 β -amino-3-hydroxyestra-1,3,5(10)-triene (2.4 g, 8.9 mmol) and methylnitrosocarbonyl azide (8.9 mmol) furnished **6b** (1.7 g, 54% yield): mp 140–142 °C dec; NMR δ 1.00 (s, 3 H, C₁₈-H), 3.24 (s, 3 H, -N(NO)CH₃), 4.20 (m, 1 H, C₁₇-H), 6.60–7.35 (m, 3 H, Ar H); MS m/e 357 (M⁺), 297 (M⁺ - HON=NCH₃), 255 (M⁺ - side chain). Anal. (C₂₀H₂₇N₃O₃) C, H, N.

Acknowledgment. We thank Dr. A. E. Bogden, Mason Research Institute, Worcester, Mass., for providing the

13762 tumor, Evelyn Froese for her excellent technical assistance, and Dorothy Faulkner and Pearl Emery for typing the manuscript. This work was supported by a grant from the National Cancer Institute of Canada.

References and Notes

- (1) L. N. Owens, M. H. Benn, and A. M. Creighton, *Cancer Res. Campaign, Annu. Rep.*, **32**, 417 (1954).
- (2) T. Nogrady, K. M. Vagi, and V. W. Adamkiewicz, *Can. J. Chem.*, **40**, 2126 (1962).
- (3) I. Niculescu-Duvaz, A. Chambanis, and E. Tannauceanu, *J. Med. Chem.*, **10**, 172 (1967).
- (4) M. E. Wall, G. S. Abernethy, Jr., F. I. Carroll, and D. J. Taylor, *J. Med. Chem.*, **12**, 810 (1969).
- (5) F. I. Carroll, A. Philip, J. T. Blackwell, D. J. Taylor, and M. E. Wall, *J. Med. Chem.*, **15**, 1158 (1972).
- (6) Groupe European du Cancer du Sera, *Eur. J. Cancer*, **5**, 1 (1969).
- (7) E. P. Vollmer, D. J. Taylor, I. J. Masnyk, D. Cooney, B. Levine, and C. Piczak, *Cancer Chemother. Rep., Part 3*, **4**, 121 (1973).
- (8) D. D. Van Hoff, M. Rozencweig, M. Slavik, and F. M. Muggia, *J. Urol.*, **117**, 464 (1977).
- (9) W. L. McGuire, P. P. Carbone, M. E. Sears, and G. C. Eachar, in "Estrogen Receptors in Human Breast Cancer", W. L. McGuire, P. P. Carbone, and E. P. Vollmer, Eds., Raven Press, New York, N.Y., 1975, p 1.
- (10) H. P. Weber and E. Galantry, *Helv. Chim. Acta*, **55**, 544 (1972).
- (11) R. Hahnel and E. Twaddle, *J. Steroid Biochem.*, **5**, 119 (1974).
- (12) G. LeClercq, M. C. Deboel, and J. C. Heuson, *Int. J. Cancer*, **18**, 750 (1976).
- (13) G. LeClercq, J. C. Heuson, and M. C. Deboel, *Eur. J. Drug Metab. Pharmacokinet.*, **1**, 77 (1976).
- (14) A. Meier, F. Stoos, D. Martin, G. Buyuk, and E. Hardegger, *Helv. Chim. Acta*, **57**, 2622 (1974).
- (15) D. Schulster, J. K. Whitehead, and A. E. Kellie, *Biochem. J.*, **93**, 512 (1964).
- (16) E. J. Corey and M. Chaykovsky, *J. Am. Chem. Soc.*, **87**, 1353 (1965).
- (17) K. Ponsold, M. Hubner, H. Kasch, and Z. Noack, *Z. Chem.*, **11**, 106 (1971).
- (18) D. N. Kirk and M. A. Wilson, *J. Chem. Soc. C*, 414 (1971).
- (19) G. Eisenbrand, H. H. Fiebig, and W. J. Zeller, *Z. Krebsforsch. Klin. Onkol.*, **86**, 279 (1976).
- (20) O. H. Wheeler and C. Reyes-Zamora, *Can. J. Chem.*, **47**, 160 (1969).

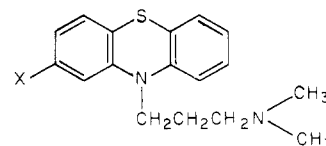
Phototoxicity of Chlorpromazine

Nigel J. Bunce,* Yogesh Kumar, and Luis Ravanal

Chemistry Department, University of Guelph, Guelph, Ontario N1G 2W1, Canada. Received May 22, 1978

The constitution of chlorpromazine has been studied in the context of its phototoxicity. Electron transfer from the side chain to the aromatic nucleus of the drug contributes to its instability to light. Even without the side chain, however, chlorphenothiazines appear to be very photolabile, so that it is unlikely that nonphototoxic analogues of chlorpromazine can be prepared merely by altering the constitution of the side chain.

Phototoxicity has been noted as a side effect in chlorpromazine therapy for a number of years,¹ but only recently has the photochemical breakdown of the drug been studied. Grant² observed that chlorpromazine (**1**) was reduced to promazine (**2**) and also underwent substitution to the hydroxy compound **3** upon illumination in aqueous solution. Formation of **3** was deduced to be a photonucleophilic process, because other nucleophiles such as alcohols and amines could undergo analogous reactions. Reduction and substitution are known photoreactions of other aryl halides.³



- 1, X = Cl
2, X = H
3, X = OH

Photodechlorination of chloro aromatic compounds has been extensively studied in other systems.³⁻⁶ Two reaction