Synthesis of Novel 2-Alkoxy-3-amino-3-arylpropan-1-ols and 5-Alkoxy-4-aryl-1,3-oxazinanes with Antimalarial Activity

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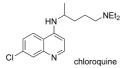
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A variety of novel *syn*-2-alkoxy-3-amino-3-arylpropan-1-ols was prepared through LiAlH₄-promoted reductive ring-opening of *cis*-3-alkoxy-4-aryl- β -lactams in Et₂O. The latter γ -aminoalcohols were easily converted into *cis*-5-alkoxy-4-aryl-1,3-oxazinanes using formaldehyde in THF. Both series of compounds were evaluated against a chloroquine sensitive strain of *Plasmodium falciparum* (D10), revealing micromolar potency for almost all representatives. Eleven compounds exhibited antimalarial activity with IC₅₀ values of $\leq 30 \ \mu$ M, and the majority of these compounds did not show cytotoxicity at the concentrations tested.

Introduction

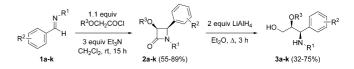
With 300–500 million clinical cases and 2 million deaths each year, malaria remains a major issue in health control, especially in developing countries.¹ Quinoline containing compounds have long been used for the treatment of malaria, and systematic modification has led to a variety of antimalarial drugs with diverse substitutions around the quinoline ring.² One of the first drugs to be prepared was the potent and inexpensive chloroquine (CQ), which is a 7-chloroquinoline with an amino substituent at position 4.³



The antimalarial activity of chloroquine appears to be linked to the haem metabolism of the causative agent, Plasmodium falciparum.⁴ However, the spread of chloroquine-resistant P. falciparum strains has dashed hopes of global malaria eradication and has complicated the clinical management of malaria in endemic areas. Consequently, many efforts are devoted to the design and synthesis of novel and structurally diverse compounds with potential antimalarial activity. Recently, 1-aminopropan-2-ols were presented as novel antimalarial agents, with activities of the best seven compounds from a library in the $1-10 \,\mu\text{M}$ range against both a chloroquine sensitive (3D7) and chloroquine resistant (FCR3) strain of the parasite.⁵ From a chemical point of view, the presented methodology was based on the ring-opening of 2-(alkoxymethyl)- and 2-(aminomethyl)epoxides by amines, resulting in the 1-aminopropan-2-ol motif due to ring-opening at the unsubstituted epoxide carbon atom.

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Scheme 1. Staudinger Synthesis of *cis*-4-Aryl- β -lactams 2 and Their Reduction toward 3-Aminopropan-1-ols 3



In the present paper, the synthesis and biological evaluation of a structurally more complex type of aminopropanols is described, i.e., new 2-alkoxy and 2-phenoxy substituted 3-amino-3-arylpropan-1-ols, starting from suitable azetidin-2-ones through reductive ring-opening. We reasoned that the combination of a γ -aminoalcohol moiety and an aryl group in these propanes might result in novel and easily accessible classes of antimalarial agents. Furthermore, cyclization of 3-amino-3-arylpropan-1-ols into 4-aryl-1,3-oxazinanes was performed in order to introduce conformational constraint into the target compounds.

Besides their biological relevance as potential antibiotics, β -lactams have acquired a prominent place in organic chemistry as synthons for further elaboration (" β -lactam synthon method").⁶ Since then, the constrained azetidin-2-one ring has been employed successfully in a large variety of different synthetic methodologies toward all kinds of nitrogen-containing target compounds.⁷ In spite of the fact that in theory a broad variety of different γ -aminoalcohols can be obtained by reductive ring-opening of β -lactams, mainly by means of complex metal hydrides, this methodology has been applied to a rather limited extend so far.⁸

Results and Discussion

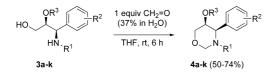
Synthesis. The synthesis of 3-alkoxy and 3-phenoxy substituted 4-aryl- β -lactams **2** was performed by means of the Staudinger reaction between the appropriate imines and ketenes. Thus, treatment of *N*-(arylmethylidene)amines **1**, obtained by condensation of different benzaldehydes in dichloromethane in the presence of MgSO₄ and Et₃N utilizing 1 equiv of the appropriate amine,⁹ with 1.1 equiv of methoxy-, phenoxy-, or benzyloxyacetyl chloride in dichloromethane in the presence of 3 equiv of triethylamine afforded the corresponding novel 4-arylazetidin-2-ones **2** in good yields at room temperature for 15 h (Scheme 1, also see Supporting Information). The relative

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stereochemistry of β -lactams **2** was assigned as *cis* based on the coupling constants between the protons at C3 and C4 in ¹H NMR (4.4–4.7 Hz, CDCl₃), in accordance with literature data.¹⁰ Subsequently, *cis*-azetidin-2-ones **2** were subjected to a reductive ring-opening by means of 2 mol equiv of lithium aluminum hydride in diethyl ether, affording the corresponding 2-alkoxy or 2-phenoxy substituted *syn*-3-amino-3-arylpropan-1-ols **3** in good yields after reflux for 3 h (Scheme 1, also see Supporting Information).

To obtain molecular diversity within γ -aminoalcohols **3**, a variety of different representatives of this class of compounds was prepared by altering the amino group (R¹ = *i*Pr, *n*Pr, *i*Bu, *t*Bu, *c*Hex, Ph, Bn), the aryl moiety (R² = H, 4-Me, 3-OMe, 4-OMe, 4-Cl, 2-Br, 2-F) and the alkoxy or phenoxy substituent (R³ = Me, Ph, Bn). This small library of novel γ -aminoalcohols **3** was then screened for potential antimalarial activity (see section Biological Evaluation).

With the intention to introduce conformational constraint into the target compounds, 3-aminopropan-1-ols **3** were subsequently converted into new *cis*-5-alkoxy-4-aryl-1,3-oxazinanes **4** by treatment with 1 equiv of formaldehyde (37% in water) in THF at room temperature for 6 h (Scheme 2, also see Supporting Information). There are only a few reports in the literature on the synthesis of oxazinanes,¹¹ either as biologically relevant targets¹² or as synthons for further elaboration.^{13,14} Oxazinanes **4** appear to be highly stable, as these compounds can be stored (neat) for prolonged times without any loss of purity. Furthermore, oxazinanes **4** were shown to be stable under hydrolytic conditions, as these compounds were recovered completely and without loss of purity after stirring for 15 h at room temperature in water or in a water/DMSO (1/1) system.

In accordance with literature data, the methylene group between oxygen and nitrogen in oxazinanes **4** appeared as two doublets with characteristic chemical shifts between 4 and 5 ppm (with a $\Delta \delta > 0.5$ ppm) and a coupling constant of 9 Hz (¹H NMR, CDCl₃).¹⁵ Also, in ¹³C NMR, this methylene carbon resonated at 79–84 ppm (CDCl₃), in accordance with the literature.¹⁵ The *cis*-relationship of the alkoxy group with respect to the aryl moiety is a direct consequence of the stereodefined Staudinger formation of *cis*- β -lactams **2**, followed by transfer of the stereochemical information through the following reaction steps toward 2-alkoxy or 2-phenoxy substituted *syn*-3-amino-3-arylpropan-1-ols **3** and *cis*-1,3-oxazinanes **4**.

This small library of novel oxazinanes **4**, in which different substitution patterns have been incorporated, was then screened for potential antimalarial activity (see section Biological Evaluation).

Further evidence for the presence of a methylene bridge in 1,3-oxazinanes **4** was provided by reductive ring-opening of 5-benzyloxy-3-isobutyl-4-(4-methylphenyl)-1,3-oxazinane **4b** as a selected example by means of 2 equiv of sodium borohydride in methanol under reflux for 3 h, resulting in the corresponding 3-(*N*-methylamino)propan-1-ol **5** in 83% yield (Scheme 3).

Biological Evaluation. Subsequently, 3-aminopropan-1-ols **3a–k**, **5**, and 1,3-oxazinanes **4a–k** were screened for in vitro antiplasmodial activity. These samples were tested in duplicate

Scheme 3. Reductive Ring-Opening of 5-Benzyloxy-3isobutyl-4-(4-methylphenyl)-1,3-oxazinane **4b** to 3-(*N*-Methylamino)propan-1-ol **5** by Means of NaBH₄

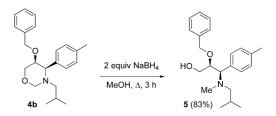


 Table 1. In Vitro Antiplasmodial Activity against P. falciparum (CQS)

 D10 Strain

compd	IC50 (µM)	IC ₉₀ (µM)	compd	IC50 (µM)	IC ₉₀ (µM)
3a	45.89	164.73	4b	30.34	ND^{a}
3b	15.3	70.76	4c	42.57	ND
3c	175.1	ND	4d	19.35	137.52
3d	6.36	12.48	4e	121.46	ND
3e	ND	ND	4f	14.08	38.55
3f	10.46	43.09	4g	16.81	118.46
3g	25.13	223.44	4h	84.87	ND
3h	85.22	ND	4i	129.02	ND
3i	13.73	251.63	4j	87.08	ND
3j	16.42	47.78	4k	ND	ND
3k	25.24	101.9	5	81.97	ND
4a	66.31	ND	CQ $(n=8)^b$	0.017	0.067
		1			

^{*a*} ND = not determined. ^{*b*} n = number of data sets averaged.

on one occasion against a chloroquine sensitive (CQS) strain of *P. falciparum* (D10). Continuous in vitro cultures of asexual erythrocyte stages of *P. falciparum* were maintained using a modified method of Trager and Jensen.¹⁶

Quantitative assessment of antiplasmodial activity in vitro was determined via the parasite lactate dehydrogenase assay using a modified method described by Makler.¹⁷

The samples were prepared to a 2 mg/mL stock solution in 10% DMSO and sonicated to enhance solubility. Samples were tested as a suspension if not completely dissolved. Stock solutions were stored at -20 °C. Further dilutions were prepared on the day of the experiment. Chloroquine (CQ) was used as the reference drug in all experiments. A full dose-response was performed for all compounds to determine the concentration inhibiting 50% of parasite growth (IC₅₀ value). Test samples were tested at a starting concentration of 100 μ g/mL, which was then serially diluted 2-fold in complete medium to give 10 concentrations, with the lowest concentration being $0.2 \,\mu \text{g/mL}$. The same dilution technique was used for all samples. CQ was tested at a starting concentration of 100 ng/mL. The highest concentration of solvent to which the parasites were exposed had no measurable effect on the parasite viability (data not shown). The IC₅₀ values were obtained using a nonlinear dose-response curve fitting analysis via Graph Pad Prism v.4.0 software (see Supporting Information). The results of this biological study are presented in Table 1.

Interestingly, these results show micromolar potency for almost all representatives, pointing to the significant biological potential of γ -aminoalcohols **3** and 1,3-oxazinanes **4**. Moreover, 11 compounds exhibited antimalarial activity with IC₅₀ values of \leq 30 μ M against *P. falciparum* (CQS) D10 strain, and compounds **3b**, **3d**, **3f**, **3i** and **4f** showed promising biological activity with IC₅₀ values \leq 15 μ M. 2-Alkoxy-3-amino-3-arylpropan-1-ols **3** were generally more active than the corresponding oxazinanes **4**, with the most active compound being **3d**. It is noteworthy that these compounds were synthesized in racemic form, and it is conceivable that enantiomerically pure variants could deliver superior activities. Nevertheless, 2-alkoxy-

Table 2. IC_{50} Values of Compounds Tested in Vitro for Antiplasmodial Activity and Cytotoxicity

compd	D10: IC ₅₀ (µM)	CHO: IC ₅₀ (µM)	SI^a
3b	15.3	143.5	9.4
3d	6.36	NT^{c}	ND^d
3f	10.46	NT	ND
3i	13.73	326.38	23.8
3ј	16.42	144.58	8.8
4f	14.08	NT	ND
4g	16.81	NT	ND
emetine $(n = 2)^b$		0.108	ND

^{*a*} Selectivity index (SI) = IC_{50} CHO/IC₅₀ D10. ^{*b*} *n* = number of data sets averaged. ^{*c*} NT = not toxic at 100 µg/mL. ^{*d*} ND = not determined.

3-amino-3-arylpropan-1-ols **3** can be considered as a potential new class of antimalarial agents.

In addition to the above-described antimalarial screenings, cytotoxicity studies were performed on the samples that showed an antimalarial activity of less than 17 μ M. Thus, γ -aminoalcohols 3b, 3d, 3f, 3i, and 3j and 1,3-oxazinanes 4f and 4g were subjected to further testing using the MTT-assay [3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide-assay]. The MTT-assay is used as a colorimetric assay for cellular growth and survival and compares well with other available assays.18 The tetrazolium salt MTT was used to measure all growth and chemosensitivity. The test samples were tested in triplicate on one occasion. The test samples were prepared to a 2 mg/mL stock solution in 10% methanol or 10% DMSO and were tested as a suspension if not properly dissolved. Test compounds were stored at -20 °C until use. Emetine was used as the reference drug in all experiments. The initial concentration of emetine was 100 μ g/mL, which was serially diluted in complete medium with 10-fold dilutions to give six concentrations, the lowest being 0.001 μ g/mL. The same dilution technique was applied to all the test samples. The highest concentration of solvent to which the cells were exposed had no measurable effect on the cell viability (data not shown). The 50% inhibitory concentration (IC₅₀) values were obtained from full dose-response curves, using a nonlinear dose-response curve fitting analysis via Graph Pad Prism v.4.0 software (see Supporting Information).

The results of this biological study are presented in Table 2. Interestingly, the in vitro cytotoxicity results showed that only compounds **3b** and **3j** have SI's of 9 (Table 2), whereas all other compounds did not show cytotoxicity at the concentrations tested.

In summary, the relevance of 3-alkoxyazetidin-2-ones as synthetic precursors for the biologically important class of 2-alkoxy-3-aminopropan-1-ols has been demonstrated by the preparation of a variety of novel *syn*-2-alkoxy-3-amino-3-arylpropan-1-ols. Cyclization of the latter γ -aminoalcohols by means of formaldehyde afforded a convenient entry into the corresponding new *cis*-5-alkoxy-4-aryl-1,3-oxazinanes. The biological importance of these classes of compounds was demonstrated by evaluation of their in vitro antiplasmodial activity and cytotoxicity, pointing to the promising potential of *syn*-2-alkoxy-3-amino-3-arylpropan-1-ols **3** as a novel type of antimalarial agents.

Experimental Section

¹H NMR spectra were recorded at 300 MHz (JEOL ECLIPSE+) with CDCl₃ as solvent and tetramethylsilane as internal standard. ¹³C NMR spectra were recorded at 75 MHz (JEOL ECLIPSE+) with CDCl₃ as solvent. Mass spectra were obtained with a mass spectrometer Agilent 1100, 70 eV. IR spectra were measured with a Spectrum One FT-IR spectrophotometer. Elemental analyses were performed with a PerkinElmer series II CHNS/O analyzer 2400. Dichloromethane was distilled over calcium hydride, while diethyl ether was dried over sodium benzophenone ketyl. Other solvents were used as received from the supplier. The purity of all new compounds has been assessed by means of gas chromatographic analysis (purity >95%).

Synthesis of *cis*-4-Aryl- β -lactams (2). General procedure: To an ice-cooled solution of *N*-(arylmethylidene)amine 1 (10 mmol) and triethylamine (30 mmol) in dichloromethane (25 mL) was added dropwise a solution of alkoxyacetyl chloride (11 mmol) in CH₂Cl₂ (10 mL). After stirring for 15 h at room temperature, the reaction mixture was poured into water (30 mL) and extracted with CH₂Cl₂ (2 × 25 mL). Drying (MgSO₄), filtration of the drying agent, and removal of the solvent afforded *cis*-3-alkoxy-4-aryl- β -lactam 2, which was purified by recrystallization from ethanol or column chromatography on silica gel.

cis-3-Benzyloxy-1-isopropyl-4-phenylazetidin-2-one (2a). Recrystallization from absolute EtOH; mp = 101.3 °C. ¹H NMR (300 MHz, CDCl₃): δ 1.04 and 1.25 (6H, 2 × d, *J* = 6.6 Hz), 3.83 (1H, septet, *J* = 6.6 Hz), 4.10 and 4.25 (2H, 2 × d, *J* = 11.3 Hz), 4.74 (1H, d, *J* = 4.4 Hz), 4.78 (1H, d, *J* = 4.4 Hz), 6.91–6.95, 7.18–7.21 and 7.36–7.44 (10H, 3 × m). ¹³C NMR (75 MHz, CDCl₃): δ 20.2, 21.4, 44.8, 60.9, 72.2, 82.7, 127.8, 128.2, 128.3, 128.5, 128.7, 135.4, 136.5, 166.6. IR (NaCl, cm⁻¹): $\nu_{C=0} = 1739$. MS (70 eV): *m/z* (%) 296 (M⁺+1, 100). Anal. calcd for C₁₉H₂₁NO₂: C 77.26, H 7.17, N 4.74; found: C 77.40, H 7.42, N 4.61.

Synthesis of *syn*-2-Alkoxy-3-amino-3-arylpropan-1-ols (3). General procedure: To an ice-cooled solution of *cis*-3-alkoxy-4-aryl- β -lactam 2 (10 mmol) in dry diethyl ether (50 mL) was added lithium aluminum hydride (20 mmol) in small portions. After reflux for 3 h, the reaction mixture was cooled to 0 °C and water (10 mL) was added in order to quench the excess of LiAlH₄. The resulting suspension was filtered over celite and washed with diethyl ether (40 mL), and the filtrate was poured into water (50 mL) and extracted with Et₂O (3 × 30 mL). Drying (MgSO₄), filtration of the drying agent, and removal of the solvent in vacuo afforded *syn*-2-alkoxy-3-amino-3-arylpropan-1-ol **3**, which was purified by means of column chromatography on silica gel or recrystallization from absolute EtOH.

syn-2-Benzyloxy-3-isopropylamino-3-phenylpropan-1-ol (3a). $R_f = 0.14$ (hexane/EtOAc 4/1). ¹H NMR (300 MHz, CDCl₃): δ 1.01 (6H, d, J = 6.2 Hz), 2.58 (1H, septet, J = 6.2 Hz), 3.46–3.49 (1H, m), 3.79 and 3.99 (2H, 2 × (d × d), J = 12.2, 3.3, 2.2 Hz), 4.05 (1H, d, J = 3.4 Hz), 4.35 and 4.60 (2H, 2 × d, J = 11.8 Hz), 7.15–7.20 and 7.23–7.38 (10H, 2 × m). ¹³C NMR (75 MHz, CDCl₃): δ 21.4, 24.3, 45.0, 63.3, 63.7, 71.6, 81.0, 127.4, 127.6, 127.9, 128.3, 128.4, 138.1, 140.6. IR (NaCl, cm⁻¹): $\nu_{NH,OH} = 3329$. MS (70 eV): m/z (%) 300 (M⁺ + 1, 100). Anal. calcd for C₁₉H₂₅NO₂: C 76.22, H 8.42, N 4.68; found: C 76.05, H 8.57, N 4.52.

Synthesis of *cis*-5-Alkoxy-4-aryl-1,3-oxazinanes (4). General procedure: To a solution of *syn*-2-alkoxy-3-amino-3-arylpropan-1-ol **3** (5 mmol) in THF (20 mL) was added formaldehyde (5 mmol, 37% solution in H_2O) at room temperature. The resulting mixture was stirred for 6 h at room temperature, after which the solvent was removed in vacuo. The crude *cis*-5-alkoxy-4-aryl-1,3-oxazinane **4** was purified by column chromatography on silica gel.

cis-5-Benzyloxy-3-isopropyl-4-phenyl-1,3-oxazinane (4a). $R_{\rm f}$ = 0.19 (hexane/EtOAc 6/1). ¹H NMR (300 MHz, CDCl₃): δ 0.84 and 1.15 (6H, 2 × d, J = 6.6 Hz), 3.05 (1H, septet, J = 6.6 Hz), 3.36–3.39 (1H, m), 3.62 (1H, d × d, J = 12.0, 2.0 Hz), 3.88 (1H, d, J = 3.3 Hz), 4.10 (1H, d × d, J = 12.0, 2.7 Hz), 4.12 (1H, d, J = 8.8 Hz), 4.22 (2H, s), 4.74 (1H, d, J = 8.8 Hz), 6.89–6.93, 7.12–7.17, 7.24–7.37, and 7.50–7.54 (10H, 4 × m). ¹³C NMR (75 MHz, CDCl₃): δ 14.7, 21.3, 47.6, 65.5, 69.1, 71.9, 74.4, 79.1, 127.2, 127.4, 127.9, 127.95, 128.04, 129.3, 137.9, 139.3. IR (NaCl, cm⁻¹): $\nu_{\rm max}$ = 2965, 2868, 1182, 1076, 1067, 1052, 1028, 738, 697. MS (70 eV): m/z (%) 312 (M⁺ + 1, 100). Anal. calcd for C₂₀H₂₅NO₂: C 77.14, H 8.09, N 4.50; found: C 76.96, H 8.32, N 4.58.

Synthesis of syn-2-Benzyloxy-3-(*N*-isobutyl-*N*-methylamino)-3-(4-methylphenyl)propan-1-ol (5). To an ice-cooled solution of 5-benzyloxy-3-isobutyl-4-(4-methylphenyl)-1,3-oxazinane **4b** (5 mmol) in methanol (20 mL) was added sodium borohydride (10 mmol), after which the resulting suspension was heated under reflux for 3 h. Afterward, the reaction mixture was poured into water (25 mL), extracted with dichloromethane (3 \times 20 mL), and dried (MgSO₄). Filtration of the drying agent and removal of the solvent yielded the crude *syn*-2-benzyloxy-3-(*N*-isobutyl-*N*-methylamino)-3-(4-methylphenyl)propan-1-ol **5**, which was purified by means of column chromatography on silica gel (hexane/EtOAc 6/1).

$$\begin{split} R_{\rm f} &= 0.07 \; (\text{hexane/EtOAc } 6/1). \ ^{\rm H} \text{NMR} \; (300 \; \text{MHz}, \text{CDCl}_3): \; \delta \\ 0.88 \; \text{and} \; 0.90 \; (6\text{H}, 2 \times \text{d}, J = 6.4 \; \text{Hz}), \; 1.70-1.94 \; (1\text{H}, \text{m}), \; 2.07 \\ \text{and} \; 2.17 \; (2\text{H}, 2 \times (\text{d} \times \text{d}), J = 12.0, \; 7.2, \; 6.9 \; \text{Hz}), \; 2.27 \; (3\text{H}, \text{s}); \\ 2.33 \; (3\text{H}, \text{s}), \; 3.59 \; (1\text{H}, \text{d} \times \text{d}, J = 11.8, \; 4.7 \; \text{Hz}), \; 3.74 \; (1\text{H}, \text{d}, J = \\ 4.9 \; \text{Hz}), \; 3.83 \; (1\text{H}, \text{d} \times \text{d}, J = 11.8, \; 4.1 \; \text{Hz}), \; 3.89-3.93 \; (1\text{H}, \text{m}), \\ 4.70 \; (2\text{H}, \text{s}), \; 7.11-7.41 \; (9\text{H}, \text{m}). \; ^{13}\text{C} \; \text{NMR} \; (75 \; \text{MHz}, \; \text{CDCl}_3): \; \delta \\ 20.76, \; 20.81, \; 21.1, \; 25.9, \; 38.9, \; 63.5, \; 64.3, \; 71.3, \; 72.4, \; 79.0, \; 127.6, \\ 127.8, \; 128.4, \; 128.6, \; 129.9, \; 132.4, \; 137.0, \; 138.6. \; \text{IR} \; (\text{NaCl}, \, \text{cm}^{-1}): \\ \nu_{\text{OH}} = 3403. \; \text{MS} \; (70 \; \text{eV}): \; m/z \; (\%) \; 342 \; (\text{M}^+ + 1, 100). \; \text{Anal. calcd} \\ \text{for } \text{C}_{22}\text{H}_{31}\text{NO}_2: \; \text{C} \; 77.38, \; \text{H} \; 9.15, \; \text{N} \; 4.10; \; \text{found: C} \; 77.59, \; \text{H} \; 9.33, \\ \text{N} \; 3.96. \end{split}$$

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Supporting Information Available: Spectroscopic data of compounds 2b-2k, 3b-3g, and 4b-4k, and full dose-response curves. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- Guinovart, C.; Navia, M. M.; Tanner, M.; Alonso, P. L. Malaria: Burden of disease. *Curr. Mol. Med.* 2006, 6, 137–140.
- (2) (a) Kumar, A.; Katiyar, S. B.; Agarwal, A.; Chauhan, P. M. S. Perspective in antimalarial chemotherapy. *Curr. Med. Chem.* 2003, 10, 1137–1150. (b) Bruce-Chwatt, L. J., Black, R. H., Canfield, C. J., Clyde, D. F., Peters, W., Wernsdorfer, W. H., Eds. *Chemotherapy of Malaria*, 2nd ed.; World Health Organization: Geneva, 1986.
- (3) (a) Chauhan, P. M. S.; Srivastava, S. K. Present trends and future strategy in chemotherapy of malaria. *Curr. Med. Chem.* 2001, *8*, 1535–1542. (b) Jefford, C. W. Why artemisinin and certain synthetic peroxides are potent antimalarials. Implications for the mode of action. *Curr. Med. Chem.* 2001, *8*, 1803–1826. (c) Sweeney, T. R. The present status of malaria chemotherapy—mefloquine, a novel antimalarial. *Med. Res. Rev.* 1981, *1*, 281–301.
- (4) Fitch, C. D. Ferriprotoporphyrin IX, phospholipids, and the antimalarial actions of quinoline drugs. *Life Sci.* 2004, *74*, 1957–1972.
- (5) Robin, A.; Brown, F.; Bahamontes-Rosa, N.; Wu, B.; Beitz, E.; Kun, J. F. J.; Flitsch, S. L. Microwave-assisted ring opening of epoxides: A general route to the synthesis of 1-aminopropan-2-ols with anti malaria parasite activities. J. Med. Chem. 2007, 50, 4243–4249.
- (6) Ojima, İ.; Delaloge, F. Asymmetric synthesis of building-blocks for peptides and peptidomimetics by means of the beta-lactam synthon method. *Chem. Soc. Rev.* **1997**, 377–386.
- (7) (a) Ojima, I. Recent advances in the beta-lactam synthon method. Acc. Chem. Res. 1995, 28, 383–389. (b) Fisher, J. F.; Meroueh, S. O.; Mobashery, S. Bacterial resistance to beta-lactam antibiotics: Compelling opportunism, compelling opportunity. Chem. Rev. 2005, 105, 395– 424. (c) Alcaide, B.; Almendros, P. Selective bond cleavage of the beta-lactam nucleus: Application in stereocontrolled synthesis. Synlett 2002, 381–393. (d) Singh, G. S. Recent progress in the synthesis and chemistry of azetidinones. Tetrahedron 2003, 59, 7631–7649. (e) France, S.; Weatherwax, A.; Taggi, A. E.; Lectka, T. Advances in the catalytic, asymmetric synthesis of beta-lactams. Acc. Chem. Res. 2004, 37, 592–600. (f) Alcaide, B.; Almendros, P.; Aragoncillo, C. Betalactams: Versatile building blocks for the stereoselective synthesis of non-beta-lactam products. Chem. Rev. 2007, 107, 4437–4492.
- (8) (a) Speeter, M. É.; Maroney, W. H. The action of lithium aluminum hydride on a β-lactam. J. Am. Chem. Soc. 1954, 76, 5810–5811. (b) Sammes, P. G.; Smith, S. On the synthesis of azetidines from 3-hydroxypropylamines. J. Chem. Soc., Chem. Commun. 1983, 682–684. (c) Ojima, I.; Zhao, M.; Yamato, T.; Nakahashi, K.; Yamashia, M.; Abe, R. Azetidines and bisazetidines. Their synthesis and use as the key intermediates to enantiomerically pure diamines, amino alcohols, and polyamines. J. Org. Chem. 1991, 56, 5263–5277. (d)

Bose, A. K.; Mathur, C.; Wagle, D. R.; Naqvi, R.; Manhas, M. S. Studies on lactams. 95. Chiral β -lactams as synthons. Stereospecific synthesis of a 6-epi-lincosamine derivative. Heterocycles 1994, 39, 491-496. (e) Watanabe, N.; Anada, M.; Hashimoto, S.-i.; Ikegami, S. Enantioselective intramolecular C-H insertion reactions of N-alkyl-N-tert-butyl-a-methoxycarbonyl-a-diazoacetamides catalyzed by dirhodium(II) carboxylates: catalytic, asymmetric construction of 2-azetidinones. Synlett 1994, 103, 1-1033. (f) Baldwin, J. E.; Adlington, R. M.; Crouch, N. P.; Mellor, L. C.; Morgan, N.; Smith, A. M.; Sutherland, J. D. Synthesis of $(2R,3S)[4-^{2}H_{3}]$ value: application to the study of the ring expansion of penicillin N by deacetoxycephalosporin C synthase from Streptomyces clavuligerus. Tetrahedron 1995, 51, 4089-4100. (g) Alcaide, B.; Aly, M. F.; Sierra, M. A. The unusual Baeyer-Villiger rearrangement of β -lactam aldehydes: totally stereoselective entry to cis-3-substituted 4-formyloxy-2-azetidinones. Tetrahedron Lett. 1995, 36, 3401-3404. (h) Lee, H. K.; Chun, J. S.; Pak, C. S. Facile transformation of 3,4-disubstituted 2-azetidinones to chiral 5,6dihydro-2-pyridones. Tetrahedron Lett. 2001, 42, 3483-3486. (i) Lee, H. K.; Chun, J. S.; Pak, C. S. Facile Transformation of 2-Azetidinones to 2-Piperidones: Application to the Synthesis of the Indolizidine Skeleton and (8S,8aS)-Perhydro-8-indolizinol. J. Org. Chem. 2003, 68, 2471-2474. (j) Lee, H. K.; Chun, J. S.; Pak, C. S. Facile conversion of 2-azetidinones to 2-piperidones: application to a formal synthesis of Prosopis and Cassia alkaloids. Tetrahedron 2003, 59, 6445-6454. (k) Birch, N. J.; Parsons, P. J.; Scopes, D. I. C. A convenient route to imino sugars and analogues of siastatin B. Synlett 2004, 277, 6-2778. (1) Kvrno, L.; Werder, M.; Hauser, H.; Carreira, E. M. Synthesis and in Vitro Evaluation of Inhibitors of Intestinal Cholesterol Absorption. J. Med. Chem. 2005, 48, 6035-6053. (m) Huguenot, F.; Brigaud, T. Convenient Asymmetric Synthesis of β -Trifluoromethyl- β -amino Acid, β -Amino Ketones, and γ -Amino Alcohols via Reformatsky and Mannich-Type Reactions from 2-Trifluoromethyl-1,3-oxazolidines. J. Org. Chem. 2006, 71, 2159-2162. (n) Del Buttero, P.; Molteni, G.; Roncoroni, M. Reductive ring opening of 2-azetidinones promoted by sodium borohydride. Tetrahedron Lett. 2006, 47, 2209-2211. (o) Mishra, R. K.; Coates, C. M.; Revell, K. D.; Turos, E. Synthesis of 2-Oxazolidinones from β -Lactams: Stereospecific Total Synthesis of (-)-Cytoxazone and All of Its Stereoisomers. Org. Lett. 2007, 9, 575-578. (p) Alcaide, B.; Almendros, P.; Cabrero, G.; Ruiz, M. P. Stereocontrolled Access to Orthogonally Protected anti, anti-4-Aminopiperidine-3,5-diols through Chemoselective Reduction of Enantiopure β -Lactam Cyanohydrins. J. Org. Chem. 2007, 72, 7980–7991. (q) Zhang, Y.-R.; He, L.; Wu, X.; Shao, P.-L.; Ye, S. Chiral N-Heterocyclic Carbene Catalyzed Staudinger Reaction of Ketenes with Imines: Highly Enantioselective Synthesis of N-Boc β -Lactams. Org. Lett. 2008, 10, 277-280. (r) D'hooghe, M.; Dekeukeleire, S.; De Kimpe, N. Reactivity of N-(ω -haloalkyl)- β -lactams with regard to lithium aluminium hydride: novel synthesis of 1-(1-aryl-3-hydroxypropyl)aziridines and 3-aryl-3-(N-propylamino)propan-1-ols. Org. Biomol. Chem. 2008, 6, 1190-1196. (s) Taubinger, A. A.; Fenske, D.; Podlech, J. Synthesis of β , β' -diamino acids from α -amino acid derived β -lactams. Synlett 2008, 539–542. (t) Sakaguchi, H.; Tokuyama, H.; Fukuyama, T. Total synthesis of (-)-kainic acid via intramolecular Michael addition: a second-generation route. Org. Lett. 2008, 10, 1711-1714

- (9) Dejaegher, Y.; Mangelinckx, S.; De Kimpe, N. Rearrangement of 2-Aryl-3,3-dichloroazetidines: Intermediacy of 2-Azetines. J. Org. Chem. 2002, 67, 2075–2081.
- (10) (a) Barrow, K. D.; Spotswood, T. M. Stereochemistry and PMR spectra of beta-lactams. *Tetrahedron Lett.* **1965**, *37*, 3325–3335. (b) Banik, B. K.; Barakat, K. J.; Wagle, D. R.; Manhas, M. S.; Bose, K. A. Microwave-induced organic reaction enhancement (MORE) chemistry. Part 13. Microwave-assisted rapid and simplified hydrogenation. *J. Org. Chem.* **1999**, *64*, 5746–5753. (c) Alcaide, B.; Almendros, P.; Rodríguez-Vicente, A.; Ruiz, M. P. Free radical synthesis of benzofused tricyclic beta-lactams by intramolecular cyclization of 2-azetidinone-tethered haloarenes. *Tetrahedron* **2005**, *61*, 2767–2778.
- (11) (a) Ghorai, M. K.; Das, K.; Kumar, A. Lewis acid mediated S_N2-type nucleophilic ring opening followed by [4 + 2] cycloaddition of N-tosylazetidines with aldehydes and ketones: synthesis of chiral 1,3-oxazinanes and 1,3-amino alcohols. *Tetrahedron Lett.* 2007, 48, 4373–4377. (b) Zanatta, N.; Squizani, A. M. C.; Fantinel, L.; Nachtigall, F. M.; Borchhardt, D. M.; Bonacorso, H. G.; Martins, M. A. P. Synthesis of N-substituted 6-trifluoromethyl-1,3-oxazinanes. J. Braz. Chem. Soc. 2005, 16, 1255–1261. (c) Hashmi, S. M. A.; Wazeer, M. I. M.; Hussain, M. S.; Reibenspies, J. H.; Perzanowski, H. P.; Ali, Sk. A. Conformational assignments and a nitrogen inversion process in some 3-acyloxy-1,3-oxazinanes by NMR and X-ray analysis. J. Chem. Soc., Perkin Trans. 2 1999, 87, 7–883.
- (12) (a) Cassady, J. M.; Chan, K. K.; Floss, H. G.; Leistner, E. Recent developments in the maytansinoid antitumor agents. *Chem. Pharm. Bull.* 2004, 52, 1–26. (b) Meyers, A. I.; Roland, D. M.; Comins, D. L.; Henning, R.; Fleming, M. P.; Shimizu, K. Progress toward the total

synthesis of maytansinoids—synthesis of (±)-4,5-deoxymaysine (nmethylmaysenine). J. Am. Chem. Soc. **1979**, 101, 4732–4734. (c) Gormley, G., Jr.; Chan, Y. Y.; Fried, J. Studies of the cyclic amidoacetal carbamate moiety of the maytansinoids. J. Org. Chem. **1980**, 45, 1447–1454. (d) Widdison, W. C.; Wilhelm, S. D.; Cavanagh, E. E.; Whiteman, K. R.; Leece, B. A.; Kovtun, Y.; Goldmacher, V. S.; Xie, H.; Steeves, R. M.; Lutz, R. J.; Zhao, R.; Wang, L.; Blaettler, W. A.; Chari, R. V. J. Semisynthetic maytansine analogues for the targeted treatment of cancer. J. Med. Chem. **2006**, 49, 4392–4408.

- (13) Singh, K.; Deb, P. K.; Venugopalan, P. Modified Pictet-Spengler reaction. A highly diastereoselective approach to 1,2,3-trisubstituted-1,2,3,4-tetrahydro-beta-carbolines using perhydro-1,3-heterocycles. *Tetrahedron* 2001, *57*, 7939–7949.
- (14) (a) Singh, H.; Singh, K. Carbon transfer reactions with heterocycles. V. A facile synthesis of nifedipine and analogs. *Tetrahedron* **1989**, 45, 3967–3974. (b) Alberola, A.; Alvarez, M. A.; Andres, C.; Gonzalez, A.; Pedrosa, R. Nucleophilic ring opening of 3-benzyl-1,3oxazinanes by Reformatsky reagents. A synthesis of β-amino ester derivatives. *Synthesis* **1990**, 1057–1058.
- (15) (a) Bartoli, G.; Cimarelli, C.; Palmieri, G. Convenient procedure for the reduction of beta-enamino ketones—synthesis of gamma-

amino alcohols and tetrahydro-1,3-oxazines. J. Chem. Soc., Perkin. Trans. 1 **1994**, 537–543. (b) Cimarelli, C.; Giuli, S.; Palmieri, G. Stereoselective synthesis of enantiopure gamma-aminoalcohols by reduction of chiral beta-enaminoketones. *Tetrahedron Asymm.* **2006**, *17*, 1308–1317.

- (16) Trager, W.; Jensen, J. B. Human malaria parasite in continuous culture. *Science* **1976**, *193*, 673–675.
- (17) Makler, M. T.; Ries, J. M.; Williams, J. A.; Bancroft, J. E.; Piper, R. C.; Gibbins, B. L.; Hinrichs, D. J. Parasite lactate dehydrogenase as an assay for *Plasmodium falciparum* drug sensitivity. *Am. J. Trop. Med. Hyg.* **1993**, 48, 739–741.
- (18) (a) Mosmann, T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. J. Immunol. Methods 1983, 65, 55–63. (b) Rubinstein, L. V.; Shoemaker, R. H.; Paull, K. D.; Simon, R. M.; Tosini, S.; Skehan, P.; Scudiero, D. A.; Monks, A.; Boyd, M. R. Comparison of in vitro anticancerdrug-screening data generated with a tetrazolium assay against a diverse panel of human tumor cell lines. J. Natl. Cancer Inst. 1990, 82, 1113–1118.

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