TWO ISOFLAVONES FROM JUNIPERUS MACROPODA

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Abstract—Two new isoflavones, junipegenin-B and -C, have been isolated from ethanolic extracts of *Juniperus* macropoda and their structures assigned. One of them, junipegenin-C, has been synthesized from iridin.

INTRODUCTION

In a recent communcation [1], we reported the presence of a new isoflavone (which we now designate as junipegenin-A) from the ethanolic extract of the leaves of *Juniperus macropoda* Boiss. (Cupressaceae). In the present paper we report the isolation and characterization of two further isoflavones, junipegenin-B and -C, from the same extract.

RESULTS AND DISCUSSION

The isolation of junipegenin-B and -C, having very close R_f on TLC, materialized by repeated column chromatography on Si gel of the mother liquor left after the crystallization of irigenin from irigenin-rich fractions.

Junipegenin-B, mp 188–189° is assigned the structure 5,7-dihydroxy-6-methoxy-3-(3,4-dimethoxy)-phenyl-4H-1-benzopyran-4-one. The above structure is assigned on the basis of the following spectral and chemical studies. It analysed for $C_{18}H_{16}O_7$ (M⁺ 344). The ¹H NMR spectrum (60 MHz, CDCl₃) established its nature as an isoflavone. A sharp singlet at δ 7.86 is assigned to C-2 proton. The C-8 proton is located at δ 6.50. In the diacetate this signal shifts to δ 7.20. Weakly coupled singlets at δ 7.10 and 6.97 integrating for one and two protons are identified for the 2' and 5', 6' protons, respectively.

The UV spectrum indicates the presence of 5,7dihydroxy grouping by the application of shift reagents. The structure is further confirmed by the oxidative degradation of junipegenin-B diacetate by K MnO_4 in acetone. One of the products of oxidation has been identified as veratric acid. Thus junipegenin-B is 5,7dihydroxy-6,3',4'-trimethoxy isoflavone (1).



Junipegenin-C, needles from ethyl acetate, mp $233-235^{\circ}$, analysed for $C_{10}H_{18}O_8$ (M⁺ 374). The ¹H NMR (CDCl₃ + DMSO-d₆) displays a singlet at δ 8.03 integrating for a single proton and is assigned to the C-2 proton. A singlet at δ 6.46 integrating for one proton is assigned to C-8. A singlet at δ 6.80 which integrates for two protons is assigned to the 2', 6' protons in the ring B. The C-8 proton shifts to δ 7.20 in the ¹H NMR spectrum of junipegenin-C diacetate. The presence of a 5,7-dihydroxy system is evident from the UV spectra recorded with various diagonistic shift reagents. On the basis of spectral data, junipegenin-C is 5,7-dihydroxy-6,3',4',5'-tetramethoxy isoflavone (2).

This structure is confirmed by its partial synthesis from iridin (3). 3 on methylation with CH_2N_2 , followed by acid hydrolysis resulted in the formation of 2. Its identity was confirmed by co-TLC, mmp, ¹H NMR and super-imposable IR with junipegenin-C.

EXPERIMENTAL

All mps are uncorr. The irigenin-rich fractions from column chromatography of defatted alcoholic extract were pooled together. The mother liquor (10g) left after the repeated crystallization of irigenin displayed two spots of almost identical R_f on the chromatoplates (CHCl₃-EtOAc, 3:1). Separation of these compounds was achieved by repeated column chromatography over Si gel using CHCl₃-MeOH in different proportions.

Junipegenin-B (1). Crystallizes from MeOH as golden yellow plates (80 mg), mp 188–189°, analysed for $C_{18}H_{16}O_7$. Found: C, 62.83; H, 4.65. Requires: C, 62.79; H, 4.65 %. UV (MeOH) max nm: 268, 340 sh; + NaOMe 275, 345; + dry AlCl₃ 275, 315; + AlCl₃/HCl 275, 375; + NaOAc 281, 350. IR (KBr) cm⁻¹: 3370 (-OH), 1650 (C=O), 1575, 1522, 1450, 1365, 1150, 1145, 1065, 1015, 997, 865, 825, 805, 665. ¹H NMR (60 MHz, CDCl₃): δ 3.96 (6H, s, 3'4'-OMe), 4.06 (3H, s, 6-OMe), 6.50 (1H, s, C-8), 6.97 (2H, s, 5', 6'-H), 7.10 (1H, s, 2'-H), 7.86 (1H, s, 2-H), 13.0 (2H, br s, phenolic H) which disappears on D₂O exchange. MS M⁺ m/e (rel. int.): 344 (100), 329 (51.0), 326 (42.95), 315 (8.05), 301 (61.07), 172 (16.51), 162 (3.8), 157 (4.02).

Acetylation (Ac₂O–C₅H₅N) gave the diacetate, colourless needles from MeOH, mp 162–163°, analysed for C₂₂H₂₀O₉. ¹H NMR (60 MHz, CDCl₃): δ 2.40 and 2.46 (2 × 3H, 2 × Ar. OCOMe), 3.90 (3H, s, 6-OMe), 3.96 (6H, s, 3',4'-OMe), 6.97 (2H, br s, 5',6'-H), 7.06 (1H, s, 2'-H), 7.20 (1H, s, 8-H), 7.83 (1H, s, 2-H). Methylation of 1 with CH₂N₂ furnished the 7-methyl ether, colourless needles from MeOH, mp 178-179. Analysed for C₁₉H₁₈O₇. ¹H NMR (CDCl₃): δ 3.96 (12H, s, 4 × Ar-OMe). 6.46 (1H, s, 8-H), 6.96 (2H, br. s, 5',6'-H), 7.13 (1H, s, 2'-H), 7.86 (1H, s, 2-H). Methylation of 1 with DMS-K₃CO₃- Me₂CO gave the 5,7-dimethyl ether, colourless plates from MeOH, mp 185-186°. Analysed for C₂₀H₂₀O₇. ¹H NMR (CDCl₃): δ 3.90 (9H, s, 3 × Ar-OMe), 3.93 (6H, s, 2 × Ar-OMe), 6.66 (1H, s, 8-H), 6.96 (2H, s, 5',6'-H), 7.16 (1H, s, 2'-H), 7.80 (1H, s, 2-H).

Oxidative degradation of junipegenin-B diacetate. The diacetate (30 mg) was subjected to oxidative degradation by KMnO₄ in Me₂CO. Among other products, a compound crystallized from H₂O (7.0 mg), mp 180–181, analysed for $C_0H_{10}O_4$, and was identified as veratric acid by co-TLC, mmp and superimposable IR.

Junipegenin-C. Crystallized from EtOAc as fine needles, mp 233–235°, analysed for $C_{19}H_{18}O_8$. Found: C, 61.08; H, 4.82. Requires: C, 60.96; H, 4.81 %. UV (MeOH) max nm: 267,327 sh; + NaOMe 273, 334; + NaOAc 271, 330; + AlCl₃ 272, 316; + AlCl₃/HCl 272, 318. IR (KBr) cm⁻¹: 3450, 1670, 1625, 1590, 1460, 1260, 1225, 1190, 1019, 805, 720. ¹H NMR (60 MHz, CDCl₃ + DMSO-d₆): δ 3.96 (12H, m, 4 × Ar⁻ OMe), 6.53 (1H, s. 8-H), 6.80 (2H, s, 2',6'-H), 8.03 (1H, s, 2-H), 12.80 (1H, s, phenolic H) disappears on D₂O exchange. MS M⁺ m/e (rel. int.): 374 (100), 359 (54,47), 356 (15,12), 341 (18,37), 187 (5,69), 1,78 (4,06), 177 (6,50).

Acetylation (Ac₂O-C₅H₅N) furnished the diacetate, needles from MeOH, mp 161–162°, analysed for $C_{23}H_{22}O_{10}$. ¹H NMR (CCl₄): δ 2.46 and 2.53 (2 × 3H, 2 × 2 × Ar-OCOMe), 3.93 (12H, *m*, 4 × Ar–OMe), 6.63 (2H, s. 216'-H), 7.10 (1H, s, 8-H) and 7.80 (1H, s, 2-H).

Partial synthesis of junipegenin-C from iridin. Iridin (100 mg) was dissolved in a minimum quantity of MeOH and a solution of CH_2N_2 in dry ether was slowly added to it. The reaction mixture was kept at 0° for 2 hr and after processing yielded a product which was hydrolysed with 7°, methanolic HCl. The product of hydrolysis crystallized from EtOAc as fine needles, mp 231-232°. Co-TLC, mmp and superimposable IR showed the semisynthetic product to be identical with junipegenin-C in all respects.

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REFERENCE

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