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Solid-Phase Synthesis of Muramyl Dipeptide (MDP) Derivatives Using a Multipin Method

Gang Liu,* Shuo-De Zhang, Shu-Quan Xia and Zhen-Kai Ding

Institute of Pharmacology and Toxicology, Academy of Military Medical Science, Beijing 100850, China

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Abstract—Solid-phase synthetic method of muramyl dipeptide derivatives is reported. A diverse library of muramyl dipeptides could be potentially synthesized by acylation, reductive alkylation, sulfonamide formation, urea formation, *N*-alkylation, amine addition, or component Ugi reactions based on this method for drug screening. © 2000 Elsevier Science Ltd. All rights reserved.

Muramyl dipeptide (MDP) is the minimum structure of the cell wall of Gram-positive bacteria that can be recognized by human immunological response.¹ Since it can stimulate non-specifically human macrophage to become active against virus, bacteria, and tumors, it represents a potential therapeutic agent. Drawbacks to its clinical use, however, include poor penetration of macrophages,² pyrogenicity,³ nonspecific induction of autoimmune responses and inflammatory reaction.⁴ Many derivatives of MDP have been synthesized by solution-phase methods and these compounds have been further studied by in vitro and in vivo assays.⁵ A few of them, including GMDP,⁶ threonyl-MDP,^{7,8} MTP-PE,^{9,10} Murabutide,¹¹ Romurtide,¹² and B30-MDP¹³ are now in clinical trials. In this paper, we are the first to report a solid-phase Multipin parallel synthesis¹⁴ of MDP derivatives method that can potentially be used to make a diverse MDP derivative library with potential application for lead drug screening.

The macro crowns with a loading capacity of $5-8 \mu mol/$ pin from Chiron Mimotopes were used for the synthesis of MDP derivatives. In order to explore the solid-phase synthesis on pins, two critical building blocks were synthesized (protected muramic acid¹⁵ and Fmoc-D-isoGln 7 as Scheme 1).

Benzyl ester protected D-glutamic acid **2** was made from D-glutamic acid **1** and benzyl-alcohol in the presence of H_2SO_4 (58.9%). The crude Boc-N₃ that was synthesized from NaNO₂, Boc-N₂H₃ in HOAc and H₂O was used to protect amino group of **2** to yield Boc-D-Glu(OBzl)-OH

3. Compound **3** was further activated by DCC and HOSu to form Boc-D-Glu(OBzl)-OSu **4** active ester in anhydrous THF. Compound **4** was reacted by 25% NH₄.OH to give Boc-D-isoGln(Bzl) **5**. The benzyl group was removed by a 5% Pd/C in HOAc for 3 days to yield Boc-D-isoGln-OH **6**. After removal of Boc by 50% TFA in DCM, Fmoc-OSu was reacted with **6** for an additional 3 days at rt in the presence of 20% NaHCO₃ to yield Fmoc-D-isoGln **7** (76.7% totally, mp. 204-205 °C).

The peptide assembly was performed by a standard Fmoc peptide synthetic strategy ¹⁶ as shown in Scheme 2. Fmoc protected crowns were treated by 20% piperidine in DMF twice, each for 5 min and 15 min respectively. After thoroughly washing by DMF and methanol separately, the crowns were suspended in a 3fold-time excess of Fmoc-Lys(Dde)-OH, BOP, HOBt and NMM stock solution for 4 h. These peptide coupling steps were repeated until the protected muramic acid was coupled onto the peptide. The Dde protected group of lysine was then removed by a treatment of 2% NH₂NH₂ in DMF twice, each for 3 min. After repeated washing steps, the exposed free amino group was further acylated with various organic acid building blocks using a BOP as coupling reagent, and HOBt plus NMM as additives at rt overnight. The final products were cleaved off from solid support by a reaction of crowns with 95% TFA/H₂O for 2 h at rt. After removal of TFA by lyophilization, all products were analyzed by HPLC at 214 nm wavelength with a gradient: B from 0-100% over 20 min. A: 0.1% TFA in water, B: 70% ACN in 0.1% TFA water. Sixty compounds were synthesized, all of them have a purity over 75% and correct molecular weight by electrospray ionization (ESI) mass spectrometry without further purification. Table 1 shows

^{*}Corresponding author at current address UC Davis Cancer Center, 4645 2nd Ave., Suite 1300, Sacramento, CA 95817, USA. Tel.: +1-916-734-6414; fax: +1-916-734-6415; e-mail: gangliu27@yahoo.com

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9a-n

Scheme 2. Synthetic muramyl dipeptide derivatives on the crown.

	R	MWt (Calcd/found)		R	MWt (Calcd/found)
9a		1049.7/1049.6	9h		854.1/854.2
9b	CH30	922.2/922.4	9i		865.1/865.4
9c		854.1/8554.4	9j		917.9/918.3
9d	HOHO	862.1/862.2	9k	N OCH ₂ C ₆ H ₅	1063.9/1064.1
9e	CH ₃ (CH ₂) ₁₄ -	948.4/948.5	91	Murml acyl	1075.4/1075.3
9f	NHCOCH ₂ CH ₂ -	943.3/943.6	9m	HO- NHCOCH ₂ CH ₂ -	901.1/901.4
9g	CH ₃ CONH- NHCOCH ₂ CH ₂ -	942.2/942.2	9n	NHCOCH ₂ CH ₂ -	899.2/899.3

Table 1. The synthetic muramyl dipeptide derivatices

examples of 14 typical compounds with correct molecular weight.

In conclusion, we have developed a solid-phase synthetic method of muramyl dipeptide derivatives using the Multipin approach. Further more, a diversity library synthesis is under way by acylation, reductive alkylation, sulfonamide formation, urea formation, *N*-alkylation, amine addition, or component Ugi reactions based on this solid-phase synthetic method.

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References

- 1. Pabst, M. J.; Beranova-Giorgianni, S.; Krueger, J. M. Neuro. Immuno. Modulation **1999**, *6*, 261.
- 2. Merhi, G.; Coleman, A. W.; Devissaguet, J. P.; Barratt, G. M. J. Med. Chem. 1996, 39, 4483.
- 3. Riveau, G.; Masek, K.; Parant, M.; Chedid, L. J. Exp. Med. 1980, 152, 869.
- 4. Koga, T.; Kakimoto, K.; Kotani, S.; Sumiyoshi, A.; Saisho, K. *Microbiol. Immunol.* **1986**, *30*, 717.

- 5. Baschang, G. Tetrahedron 1989, 45, 6331.
- 6. Palache, A. M.; Beyer, W. E.; Hendriksen, E.; Gerez, L.; Aston, R.; Ledger, P. W.; de Regt, V.; Kerstens, R.; Rothbarth, P. H.; Osterhaus, A. D. *Vaccine* **1996**, *14*, 1327.
- 7. Hart, M. K.; Palker, T. J.; Matthews, T. J.; Langlois, A. J.; Lerche, N. W.; Martin, M. E.; Scearce, R. M.; McDanal, C.; Bolognesi, D. P.; Haynes, B. F. J. Immunol. **1990**, 145, 2677.
- 8. Ivins, B. E.; Welkos, S. L.; Little, S. F.; Crumrine, M. H.; Nelson, G. O. Infect. Immun. 1992, 60, 662.
- 9. Kahn, J. O.; Sinangi, F.; Baenziger, J.; Murcar, N.; Wynne, D.; Coleman, R. L.; Steimer, K. S.; Dekker, C. L.; Chernoff, D. J. Infec. Dis. **1994**, *170*, 1288.
- 10. Keefer, M. C.; Graham, B. S.; McElrath, M. J.; Matthews, T. J.; Stablein, D. M.; Corey, L.; Wright, P. F.; Lawrence, D.;
- Fast, P. E.; Weinhold, K.; Hsieh, R. H.; Chernoff, D.; Dekker,
- C.; Dolin, R. AIDS Res. Hum. Retroviruses 1996, 12, 683.
- 11. Bahr, G. M.; Darcissac, E.; Bevec, D.; Dukor, P.; Chedid, L. Int. J. Immunopharmacol. **1995**, *17*, 117.
- 12. Namba, K.; Nakajima, R.; Otani, T.; Azuma, I. Vaccine **1996**, 14, 1149.
- 13. Kaji, M.; Kaji, Y.; Kaji, M.; Ohkuma, K.; Honda, T.; Oka, T.; Sakoh, M.; Nakamura, S.; Kurachi, K.; Sentoku, M. *Vaccine* **1992**, *10*, 663.
- 14. Geysen, M. H.; Meloen, R. H.; Barteling, S. J. Proc. Natl. Acad. Sci. USA 1984, 81, 3998.
- 15. Gross, P. H.; Rimpler, M. Liebigs Ann. Chem. 1986, 37.
- 16. Liu, G.; Mu, S. F.; Yun, L. H.; Ding, Z. K.; Sun, M. J. J. Pept. Res. 1999, 54, 480.