

New Oxaanalogues of Spermine

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Abstract—A new isosteric charge-deficient spermine analogue, 1,12-diamino-4,9-diaza-5-oxadodecan, and *O*-(7-amino-4-azaheptyl)oxime of 3-aminopropanal, a stable analogue of the Schiff base intermediate in the enzymatic oxidation of spermine, were synthesized. The possible use of these compounds for the inhibition of spermine oxidase is discussed.

Key words: oxaspermine, polyamine analogues, polyamines, spermine oxidase

INTRODUCTION

A selective control of activity of the enzymes involved in the metabolism of polyamines is widely used at studying the role of spermine and spermidine in the processes of growth and differentiation of cells.² These investigations are of a certain scientific and applied importance due to significance and a variety of cellular functions of polyamines and their increased content in tumor cells in comparison with that on normal cells [1, 2].

The activation of the enzymes of polyamine catabolism is the most effective way of the exhaustion of the intracellular pool of spermine and spermidine [3]. Therefore, in recent years, the study of these enzymes is one of important directions in the polyamine biochemistry.

The classical pathway of spermine degradation consists in its *N*¹-acetylation catalyzed by SSAT and the subsequent oxidation of the product by PAO (Scheme 1) [4]. A selective effectively acting nonreversible inhibitor *N*¹,*N*⁴-bis(2,3-butadienyl)-1,4-diaminobutane (MDL-72.527, see the figure) is known for the well-known flavin-dependent PAO; it exhibits *K*_i 0.09 μM and τ_{1/2} 2.2 min toward to the isolated enzyme [5].

Recently, SMO has been found in animal cells; this is a new flavin-dependent enzyme that oxidizes Spm into Spd without the preliminary *N*¹-acetylation [6–11].

Therefore, the Spm degradation can proceed by two independent pathways (Scheme 1). Several SMO inhibitors are known; the best of them, *cis*-3,8,13,18,23,28,33,38,43,48-decaazapentacontene-25 (SL-11150, figure), has IC₅₀ 10⁻⁷ M [8]. A similar flavin-dependent Spm oxidase also exists in plants, whereas 1,17-bis(guanidyl)-9-azaheptadecane (guazatine, figure) inhibits the enzyme from maize with *K*_i 7.5 × 10⁻⁹ M [12]. The X-ray study of the enzyme–inhibitor complex (E–I) showed that the inhibition efficiency is due to a firm noncovalent binding of this compound in the active site of enzyme [13].

The syntheses of two new compounds, 1,12-diamino-4,9-diaza-5-oxadodecane (**I**) (Scheme 2) and *O*-(7-amino-4-azaheptyl)oxime of 3-aminopropanal (**II**) (Scheme 3), are described in this communication; we suggest the use of these compounds for the SMO inhibition and the study of the mechanism of action of this enzyme.

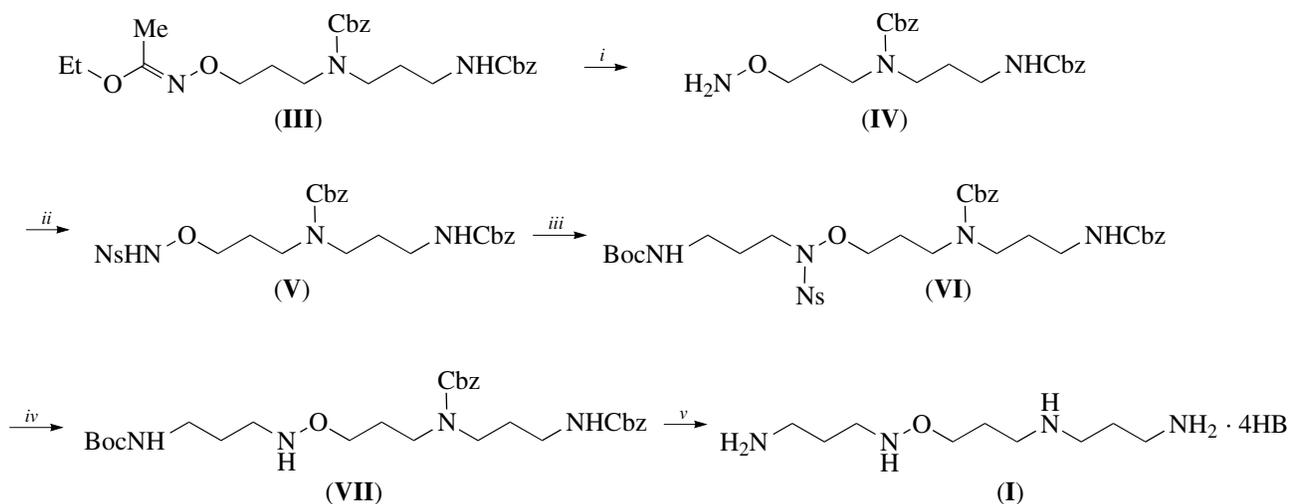
RESULTS AND DISCUSSION

The enzymatic oxidation of Spm was shown to proceed through the intermediate formation of a Schiff base, hydrolyzed to 3-aminopropanal and Spd (Scheme 4) [10].

Here we describe the substitution of oxygen atom for the methylene group of Spm in position 5, which resulted in *N,O*-dialkylhydroxylamine (**I**), in order to obtain new potential inhibitors of SMO. Since oxa-Spm (**I**) is an isostere of Spm, one can expect that this compound will be a good competitive inhibitor of SMO. Oxygen atom in the fifth position of (**I**) decreases the basicity of the adjacent NH group (*pK*_a ~ 5.5) in comparison with Spm (*pK*_a of the NH group of 7.97 [14]), and, therefore, (**I**) exists in the form of trication at the

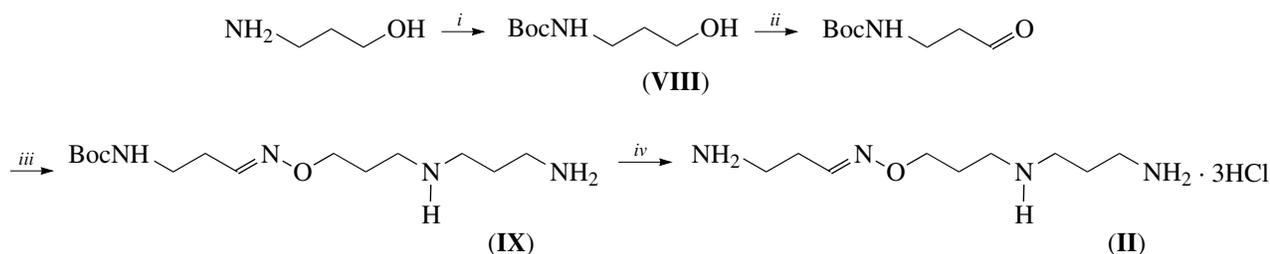
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² Abbreviations: Bpoc, 2-(4-biphenyl)prop-2-yloxy carbonyl; Mts, mesitylenesulfonyl; Ns, 2-nitrobenzenesulfonyl; PAO, polyamine oxidase (EC 1.5.3.11); Pmc, 2,2,5,7,8-pentamethyl-6-chromanesulfonyl; Spd, spermidine (1,8-diamino-4-azaoctane); Spm, spermine (1,12-diamino-4,9-diazadodecane); SMO, spermine oxidase; and SSAT, spermine/spermidine *N*¹-acetyltransferase (EC 2.3.1.57).



i, $\text{HCl}/i\text{-PrOH}/\text{H}_2\text{O}$, *ii*, $\text{Ns-Cl}/\text{CH}_2\text{Cl}_2$, *iii*, $\text{BocNH}(\text{CH}_2)_3\text{I}/\text{K}_2\text{CO}_3/\text{DMF}$, *iv*, $\text{PhSH}/\text{K}_2\text{CO}_3/\text{DMF}$, *v*, HBr/AcOH .

Scheme 2. The scheme of synthesis of 1,12-diamino-4,9-diaza-5-oxadodecane (I).



i, $\text{Boc}_2\text{O}/\text{THF}$, *ii*, $\text{CrO}_3 \cdot \text{Py} \cdot \text{HCl}/\text{Al}_2\text{O}_3/\text{CH}_2\text{Cl}_2$, *iii*, $\text{H}_2\text{NO}(\text{CH}_2)_3\text{NH}(\text{CH}_2)_3\text{NH}_2/\text{MeOH}/\text{H}_2\text{O}$, *iv*, HCl/MeOH .

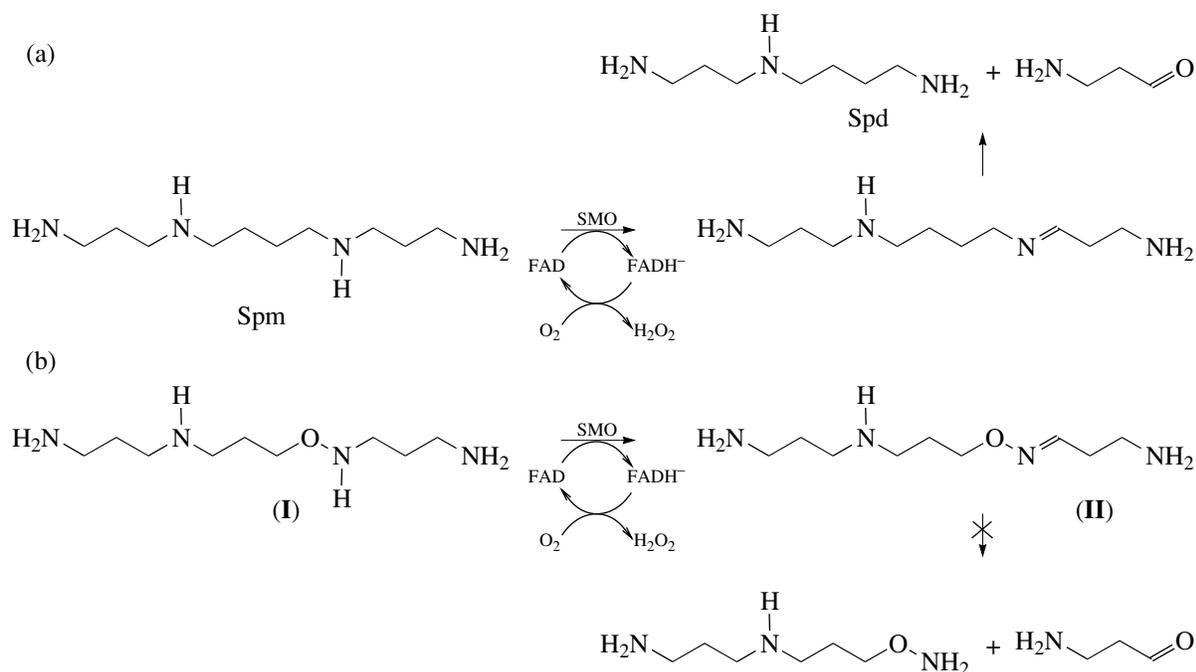
Scheme 3. The scheme of synthesis of *O*-(7-amino-4-azaheptyl)oxime of 3-aminopropanal (II).

with 3-*N*-Boc-aminopropyl iodide was chosen for the obtaining of 1,12-diamino-4,9-diaza-5-oxadodecane (I) (Scheme 2).

The *gem*-bis-Mts-derivative of *O*-substituted hydroxylamine is known to be formed under the standard conditions of the synthesis of arenesulfonylamides with the use of triethylamine as a base and only at an excess mesitylenesulfonyl chloride (Mst-Cl) [18]. The interaction of 2-nitrobenzenesulfonyl chloride (Ns-Cl) with hydroxylamine (IV) proceeds ambiguously even at the equimolar ratio $\text{Ns-Cl} : \text{RONH}_2 : \text{Et}_3\text{N}$. However, the reaction with 2.5–3.0-fold excess of *O*-substituted hydroxylamine in the absence of other bases leads to mono-NS-derivative (V) in yield of approximately 70%. The further conversions consisted in the alkyla-

tion of (V) with 3-*N*-Boc-aminopropyl iodide, selective denosylation of (VI) with thiophenol (like that described for amines [19, 20], and the removal of Cbz groups by HBr/AcOH , which resulted in the target tetrahydrobromide of 1,12-diamino-4,9-diaza-5-oxadodecane (I).

The synthesis of oxime (II) was achieved starting from 3-*N*-Boc-aminopropanol (VIII) (Scheme 3), which was oxidized at the first stage by pyridinium chlorochromate on neutral alumina to the corresponding aldehyde (Scheme 3). 3-(*N*-*tert*-Butyloxycarbonyl)aminopropanal was not isolated in pure state, but was directly subjected to the reaction with 1-amino-4-aza-7-aminooxyheptane. The resulting oxime (IX) was isolated by column chromatography on silica gel as a mixture of *syn*- and *anti*-isomers at the ratio of 40 : 60



Scheme 4. (a) Oxidative cleavage of spermine in animals [10], and (b) the presumed enzymatic oxidation of (I) to oxime (II).

(^1H NMR data). The subsequent removal of Boc-protective group by the treatment with HCl/MeOH at 0°C and recrystallization led to the desired (II) trihydrochloride; in this case, the ratio of *syn*- and *anti*-isomers remained almost unchanged (35 : 65).

Note that the reduction of (II) with NaCNBH_3 should lead to oxa-Spm (I), because the reduction of oximes is the general method of synthesis of *O,N*-dialkylhydroxylamine (see above).

EXPERIMENTAL

The reagents Cbz-Cl , Boc_2O , PhSH , absolute EtOH , and 1-amino-3-bromopropane were from Fluka; 2-nitrobenzenesulfonyl chloride and 3-aminopropanol, from Aldrich. 7-Aminoxy-4-aza-1-aminoheptane and 7-(1'-ethoxyethylidene)aminoxy-4-(*N*-benzyloxycarbonyl)aza-1-(*N*-benzyloxycarbonyl)aminoheptane were synthesized as described in [21], and pyridinium chlorochromate adsorbed on Al_2O_3 was synthesized as described in [22].

TLC was carried out on precoated Kieselgel 60 F_{254} plates (Merck) in the following systems: (A) 95 : 5 chloroform–methanol, (B) chloroform, (C) 4 : 2 : 1 : 2 *n*-butanol–acetic acid–pyridine–water, (D) 9 : 1 chloroform–methanol, and (E) 8 : 2 dioxane–25% ammonia. Substances were detected on the chromatograms by their absorption of UV light and by the color reaction with ninhydrin; the aminoxy compounds, in the form of fluorescent oximes of pyridoxal-5'-phosphate. Col-

umn chromatography was carried out on Kieselgel (40–63 μm , Merck), elution systems being specified in text.

NMR spectra were registered on a Bruker Avance 500 DRX instrument (Germany) with the working frequency of 500.1 MHz for ^1H and 125.8 MHz for ^{13}C nuclei. Tetramethylsilane was used as an internal standard in CDCl_3 and CD_3OD solutions and sodium salt of 3-trimethylsilyl-1-propanesulfonic acid in D_2O . Chemical shifts are given in ppm, and spin–spin coupling constants in Hz.

7-Aminoxy-4-(*N*-benzyloxycarbonyl)aza-1-(*N*-benzyloxycarbonyl)aminoheptane (IV). Concentrate HCl (1.5 ml) was added to a solution of (III) (4.7 g, 9.73 mmol) in isopropanol (10 ml), the solution was kept for 10 min at room temperature and evaporated in a vacuum to dryness. The residue was dissolved in water (5 ml), 10 M NaOH (1.5 ml) was added, and extracted with chloroform (3×5 ml). The extract was washed with a saturated solution of NaCl (2×2 ml), dried with MgSO_4 , and evaporated in a vacuum to dryness. The residue was dissolved in chloroform (6 ml) and purified by column chromatography on silica gel (60 g), successively eluting with chloroform and 97 : 3 chloroform–methanol. The target (IV) was dried in a vacuum over P_2O_5 ; viscous oil; yield 2.71 g (73%); R_f 0.37 (A); ^1H NMR (CDCl_3): 7.4–7.2 (10 H, m, Ph); 5.12 (2 H, s, CH_2Ph), 5.08 (2 H, s, CH_2Ph), 3.77 (2 H, m, H_2NOCH_2), 3.46 (2 H, m, $\text{H}_2\text{NOCH}_2\text{CH}_2\text{CH}_2\text{N}$), 3.35–3.08 [4 H, m, $\text{NCH}_2(\text{CH}_2)_2\text{NHCbz}$ +

$N(\text{CH}_2)_2\text{CH}_2\text{NHCbz}$], and 1.65–1.38 (4 H, m, $\text{H}_2\text{NOCH}_2\text{CH}_2\text{CH}_2\text{N} + \text{NCH}_2\text{CH}_2\text{CH}_2\text{NHCbz}$).

7-(*N*-2-Nitrobenzenesulfonyl)aminoxy-4-(*N*-benzyloxycarbonyl)aza-1-(*N*-benzyloxycarbonyl)aminoheptane (V). A solution of Ns-Cl (0.48 g, 2.2 mmol) in dichloromethane (6.5 ml) was added to a solution of (IV) (2.71 g, 6.5 mmol) in dichloromethane (8 ml). The reaction mixture was kept for 24 h at room temperature and evaporated in a vacuum to dryness. The residue was dissolved in 99 : 1 chloroform–methanol mixture (6 ml) and purified on a silica gel column (60 g) eluted with 99 : 1 chloroform–methanol. The target (V) was dried in a vacuum over P_2O_5 to give a viscous oil; yield 1.3 g (65% from Ns-Cl); R_f 0.57 (A); $^1\text{H NMR}$ (CDCl_3): 8.18 (1 H, m, Ns), 7.99 (1 H, m, Ns), 7.85 (1 H, m, Ns), 7.74 (1 H, m, Ns), 7.36–7.26 (10 H, m, Ph), 5.60 (1 H, s, HNO), 5.12 (2 H, s, CH_2Ph), 5.08 (2 H, s, CH_2Ph), 4.04 (2 H, m, NsHNOCH_2), 3.30–3.28 (4 H, m, $\text{H}_2\text{NOCH}_2\text{CH}_2\text{CH}_2\text{N} + \text{NCH}_2(\text{CH}_2)_2\text{NHCbz}$), 3.13 (2 H, m, $\text{N}(\text{CH}_2)_2\text{CH}_2\text{NHCbz}$), and 1.88–1.67 (4 H, m, $\text{NsHNOCH}_2\text{CH}_2\text{CH}_2\text{N} + \text{NCH}_2\text{CH}_2\text{CH}_2\text{NHCbz}$).

1-(*N*-*tert*-Butyloxycarbonyl)amino-4-aza-5-oxa-9-(*N*-benzyloxycarbonyl)aza-12-(*N*-benzyloxycarbonyl)aminododecane (VII). A cooled to +4°C solution of 3-bromo-1-aminopropane hydrobromide (5.5 g, 25 mmol) in water (7 ml) was mixed with 10 M NaOH (2.5 ml), dioxane (20 ml) and a solution of Boc_2O (6 g, 28 mmol) in dioxane (10 ml). The reaction mixture was stirred for 16 h at room temperature, evaporated to dryness in a vacuum, and the residue was dissolved in chloroform. The solution was successively washed with water (2 × 10 ml), 10% citric acid (2 × 10 ml), and water (3 × 10 ml), dried with MgSO_4 , and evaporated to dryness in a vacuum. The residue was dissolved in acetone (20 ml), NaI (4.1 g, 27.2 mmol) was added, and the mixture was stirred for 18 h at room temperature. The precipitate was separated, the filtrate was evaporated to dryness in a vacuum, and the residue was dissolved in chloroform. The solution was washed with water (2 × 2 ml), dried with MgSO_4 , and evaporated, and the residue was dried in a vacuum at +4°C over P_2O_5 to give 1-(*N*-*tert*-butyloxycarbonyl)amino-3-iodopropane in the form of a viscous slowly solidifying yellowish oil; yield 5.35 g (83%). A part of the semicrystalline product was triturated with a small quantity of hexane and kept overnight at +4°C; the solid was separated, washed with cold hexane, dried in a vacuum over paraffin and P_2O_5 , and obtained colorless crystals of $\text{BocNH}(\text{CH}_2)_3\text{I}$; R_f 0.36 (B); mp 41–42°C; $^1\text{H NMR}$ (CD_3OD): 4.58 (1 H, br. s, NH), 3.26 (2 H, m, NHCH_2), 3.16 (2 H, t, J 6.5, CH_2I), 2.06–1.98 (2 H, m, $\text{CH}_2\text{CH}_2\text{CH}_2$), and 1.48 [9 H, s, $\text{C}(\text{CH}_3)_3$].

A mixture of (V) (1.3 g, 2.2 mmol), K_2CO_3 (1.19 g, 8.64 mmol), and $\text{BocNH}(\text{CH}_2)_3\text{I}$ (0.8 g, 2.8 mmol) was stirred in DMF (10 ml) for 24 h at room temperature; the residue was separated; and the filtrate was evaporated in a vacuum to dryness. The residue was dissolved

in chloroform, successively washed with water (2 × 5 ml) and a saturated solution of NaCl (2 × 5 ml), and dried with MgSO_4 . The solution was evaporated in a vacuum to dryness, and the raw 1-(*N*-*tert*-butyloxycarbonyl)amino-4-(*N*-2-nitrobenzenesulfonyl)aza-5-oxa-9-(*N*-benzyloxycarbonyl)aza-12-(*N*-benzyloxycarbonyl)aminododecane (VI) was dissolved in DMF (30 ml). The solution was mixed with K_2CO_3 (1.2 g, 8.72 mmol) and PhSH (0.27 ml, 2.6 mmol) and stirred for 4 h at room temperature. The reaction mixture was evaporated to dryness in a vacuum, the residue was suspended in chloroform, and the precipitate was separated by centrifugation and washed with a small quantity of chloroform. The combined organic solutions were successively washed with water (5 ml), 10% citric acid (2 × 10 ml), 1 M NaHCO_3 (2 × 10 ml), and water (10 ml) and evaporated in a vacuum. The residue was dissolved in 100 : 1 chloroform–methanol mixture (6 ml) and purified by a chromatography on a silica gel column (60 g), eluted with 100 : 1 chloroform–methanol mixture to get (VII) as a viscous oil dried in a vacuum over P_2O_5 ; yield 0.58 g (46%); R_f 0.37 (A); $^1\text{H NMR}$ (CDCl_3): 7.36–7.26 (10 H, m, Ph), 5.12 (2 H, s, CH_2Ph), 5.09 (2 H, s, CH_2Ph), 3.62 (2 H, m, $\text{HNOCH}_2(\text{CH}_2)_2\text{NCbz}$), 3.34–3.28 ($\text{CH}_2\text{N}(\text{Cbz})\text{CH}_2$); 3.23–3.11 (4 H, m, $\text{BocNHCH}_2 + \text{CH}_2\text{NHCbz}$), 2.86 $\text{BocNH}(\text{CH}_2)_2\text{CH}_2\text{NHO}$; 1.78–1.60 (6 H, m, $\text{BocNHCH}_2\text{CH}_2\text{CH}_2\text{NHO} + \text{HNOCH}_2\text{CH}_2\text{CH}_2\text{N} + \text{NCH}_2\text{CH}_2\text{CH}_2\text{NHCbz}$), and 1.41 [9 H, s, $\text{C}(\text{CH}_3)_3$].

1,12-Diamino-4,9-diaza-5-oxadodecane tetrabromobromide (I). A 32.3% solution of HBr in AcOH (3 ml) was added to a solution of (VII) (0.58 g, 1.01 mmol) in AcOH (5 ml). The mixture was kept at room temperature until the evolution of CO_2 ceased. The precipitated solid was separated, washed with diethyl ether, and dried in a vacuum over $\text{P}_2\text{O}_5/\text{KOH}$ to give (I); yield 0.36 g (69%); R_f 0.27 (B); mp 169–171°C (decomp.); $^1\text{H NMR}$ (D_2O): 4.11 (2 H, t, J 6.0, HNOCH_2), 3.31 (2 H, t, J 7.6, $\text{OHNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 3.17 J 7.9, $\text{HNO}(\text{CH}_2)_2\text{CH}_2\text{NH}_2$; 3.14 J 8.0, $\text{CH}_2\text{HN}(\text{CH}_2)_2\text{CH}_2\text{NH}_2$; 3.08 (2 H, t, J 7.4, $\text{HNOCH}_2\text{CH}_2\text{CH}_2$), 3.07 [2 H, t, J 7.4, $\text{CH}_2\text{HNCH}_2(\text{CH}_2)_2\text{NH}_2$]; and 2.10–2.01 (6 H, m, $\text{OHNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2 + \text{HNOCH}_2\text{CH}_2 + \text{CH}_2\text{HNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$); $^{13}\text{C NMR}$ (D_2O): 73.83 (t, HNOCH_2), 49.84 (t, $\text{OHNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 47.79 (t, $\text{HNOCH}_2\text{CH}_2\text{CH}_2$), 47.62 [t, $\text{CH}_2\text{HNCH}_2(\text{CH}_2)_2\text{NH}_2$], 40.09 [t, $\text{HNO}(\text{CH}_2)_2\text{CH}_2\text{NH}_2$], 39.52 [t, $\text{CH}_2\text{HN}(\text{CH}_2)_2\text{CH}_2\text{NH}_2$], 27.48 (t, $\text{HNOCH}_2\text{CH}_2$), 26.73 (t, $\text{CH}_2\text{HNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), and 25.56 (t, $\text{OHNCH}_2\text{CH}_2\text{CH}_2\text{NH}_2$).

Found, %: C 20.51, H 5.28, N 10.50. $\text{C}_9\text{H}_{28}\text{Br}_4\text{N}_4\text{O}$. Calc., %: C 20.47, H 5.35, N 10.61.

3-(*N*-*tert*-Butyloxycarbonyl)aminopropanol (VIII). A solution of Boc_2O (1.5 g, 7 mmol) in THF (7 ml) was added to a solution of 3-aminopropanol (0.5 g, 7 mmol)

in THF (7 ml). The mixture was kept for 12 h at room temperature and evaporated in a vacuum to dryness. The residue was dissolved in chloroform; the solution was successively washed with 10% citric acid (2 × 5 ml), 1 M NaHCO₃ (2 × 5 ml), and water (5 ml); and was evaporated in a vacuum to dryness. The residue was dried in a vacuum over P₂O₅, to get (**VIII**) as a viscous oil; yield 1 g (82%); *R_f* 0.5 (D); ¹H NMR (CDCl₃): 4.82 (1 H, s, NH), 3.65 (2 H, m, CH₂OH), 3.27 (2 H, m, NHCH₂), 3.08 (1 H, s, OH), 1.65 (2 H, m, CH₂CH₂OH), and 1.43 [9 H, s, C(CH₃)₃].

O-(7-Amino-4-aza)heptyloxime of 3-(*N*-tert-butylloxycarbonyl)aminopropanal (IX). Pyridinium chlorochromate on neutral Al₂O₃ (7 g) was added to a solution of (**VIII**) (0.7 g, 4 mmol) in anhydrous benzene (40 ml). The mixture was stirred for 4 h at 20°C, the precipitate was separated, and the filtrate was evaporated in a vacuum to dryness. The residue was dissolved in methanol (5 ml) and treated with a neutral solution of 7-aminooxy-4-aza-1-aminoheptane trihydrochloride (0.256 g, 1 mmol) in 50% methanol (10 ml). The reaction mixture was stirred for 2 h at room temperature, evaporated to dryness in a vacuum, and the residue was dissolved in 95 : 5 dioxane–25% ammonia (6 ml). The purification on a silica gel column (60 g) successively eluted with 95 : 5 dioxane–25% ammonia and 8 : 2 dioxane–25% ammonia gave, after drying the target fraction in a vacuum over P₂O₅, 0.2 g (66% from 7-aminooxy-4-aza-1-aminoheptane trihydrochloride) of (**IX**) as a viscous oil; *R_f* 0.5 (E). ¹H NMR of isomer I (CDCl₃): 7.38 (1 H, t, *J* 5.4, NHCH₂CH₂CH=NO), 4.09 (2 H, t, *J* 6.2, NOCH₂CH₂CH₂NH), 3.32 (2 H, br. m, NHCH₂CH₂CH=NO), 2.79 (2 H, t, *J* 6.3, NHCH₂CH₂CH₂NH₂), 2.70 [4 H, m, NHCH₂(CH₂)₂NH₂ + NO(CH₂)₂CH₂NH], 2.37 (2 H, dt, *J* 5.9, NHCH₂CH₂CH=NO), 1.83 (2 H, m, NOCH₂CH₂CH₂NH), 1.66 (2 H, m, NHCH₂CH₂CH₂NH₂), and 1.44 [9 H, s, C(CH₃)₃]; isomer II: ¹H NMR (CDCl₃): 6.69 (1 H, t, *J* 5.5, NHCH₂CH₂CH=NO), 4.14 (2 H, t, *J* 6.1, NOCH₂CH₂CH₂NH), 3.28 (2 H, br. m, NHCH₂CH₂CH=NO), 2.51 (2 H, dt, *J* 5.9, NHCH₂CH₂CH=NO), and 1.86 (2 H, m, NOCH₂CH₂CH₂NH); chemical shifts of other protons coincide with those of isomer I. The isomer I/isomer II ratio was 60 : 40.

O-(7-Amino-4-azaheptyl)oxime of 3-aminopropanal trihydrochloride (II). A solution of (**IX**) (0.15 g, 0.5 mmol) in absolute methanol (5 ml) cooled to 0°C was treated with 4 M HCl/MeOH (5 ml) and kept overnight at 4°C. The reaction mixture was evaporated to dryness in a vacuum, and the residue was coevaporated with MeOH, recrystallized from MeOH–EtOH, and dried in a vacuum over P₂O₅/KOH. Crystalline (**II**) was obtained; yield 0.106 g (68%); *R_f* 0.1 (E);

mp 193–194°C (decomp.); isomer I: ¹H NMR (D₂O): 7.59 (1 H, t, *J* 5.3, CH=NO), 4.18 (2 H, t, *J* 6.0, NOCH₂), 3.26 (2 H, t, *J* 7.0, NH₂CH₂CH₂CH=NO), 3.22–3.16 [4 H, m, NHCH₂(CH₂)₂NH₂ + NO(CH₂)₂CH₂NH], 3.12 (2 H, t, *J* 5.7, NHCH₂CH₂CH₂NH₂), 2.65 (2 H, dt, *J* 7.0, NH₂CH₂CH₂CH=NO), and 2.15–2.04 (4 H, m, NHCH₂CH₂CH₂NH₂ + NOCH₂CH₂CH₂NH); ¹³C NMR (D₂O): 152.58 (d, N=CH), 73.24 (t, CH₂O), 48.10 [t, CH₂(CH₂)₂O], 47.48 [t, NH₂(CH₂)₂CH₂], 39.43 [t, NH₂CH₂(CH₂)₂], 39.10 (t, NH₂CH₂CH₂CH=N), 29.86 (t, CH₂CH=N), 28.04 (t, CH₂CH₂O), and 26.57 (t, NH₂CH₂CH₂CH₂); isomer II: ¹H NMR (D₂O): 6.96 (1 H, t, *J* 5.5, CH=NO), 4.23 (2 H, t, *J* 6.0, NOCH₂), 3.24 (2 H, t, *J* 7.3, NH₂CH₂CH₂CH=NO), and 2.77 (2 H, dt, *J* 7.3, NH₂CH₂CH₂CH=NO); ¹³C NMR (D₂O): 151.95 (d, N=CH), 73.34 (t, CH₂O), 47.96 [t, CH₂(CH₂)₂O], 47.45 [t, NH₂(CH₂)₂CH₂]; 39.14 (o, NH₂CH₂CH₂CH=N); 28.26 (t, CH₂CH₂O), 26.68 (t, CH₂CH=N), and 26.60 (t, NH₂CH₂CH₂CH₂); chemical shifts of other nuclei coincide with those of isomer I. The isomer I/isomer II ratio was 65 : 35.

Found, %: C 34.84, H 8.11, N 17.54. C₉H₂₅Cl₃N₄O. Calc., %: C 34.68, H 8.08, N 17.98.

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REFERENCES

1. Seiler, N., *Curr. Drug Targets*, 2003, vol. 4, pp. 537–564.
2. Seiler, N., *Curr. Drug Targets*, 2003, vol. 4, pp. 565–585.
3. Casero, R.A. and Woster, P.M., *J. Med. Chem.*, 2001, vol. 44, pp. 1–26.
4. Cohen, S.S., *A Guide to the Polyamines*, New York: Oxford Univ. Press, 1998.
5. Bey, P., Bolkenius, F.N., Seiler, N., and Casara, P., *J. Med. Chem.*, 1985, vol. 28, pp. 1–2.
6. Wang, Y., Devereux, W., Woster, P.M., Murray-Stewart, T., Hacker, A., and Casero, R.A., *Cancer Res.*, 2001, vol. 61, pp. 5370–5373.
7. Murray-Stewart, T., Wang, Y., Devereux, W., and Casero, R.A., *Biochem. J.*, 2002, vol. 368, pp. 673–677.
8. Wang, Y., Murray-Stewart, T., Devereux, W., Hacker, A., Frydman, B., Woster, P.M., and Casero, R.A., *Biochem. Biophys. Res. Commun.*, 2003, vol. 304, pp. 605–611.
9. Devereux, W., Wang, Y., Murray-Stewart, T., Hacker, A., Smith, R., Frydman, B., Valasinas, A.L., Reddy, V.K., Marton, L.J., Ward, T.D., Woster, P.M., and Casero, R.A., *Cancer Chem. Pharm.*, 2003, vol. 52, pp. 383–390.
10. Vujcic, S., Diegelman, P., Bacchi, C.J., Kramer, D.L., and Porter, C.W., *Biochem. J.*, 2002, vol. 367, pp. 665–675.

11. Cervelli, M., Polticelli, F., Federico, R., and Mariotini, P., *J. Biol. Chem.*, 2003, vol. 278, pp. 5271–5276.
12. Federico, R., Leone, L., Botta, M., Binda, C., Angelini, R., Venturini, G., and Ascenzi, P., *J. Enzyme Inhib.*, 2001, vol. 16, pp. 147–155.
13. Binda, C., Angelini, R., Federico, R., Ascenzi, P., and Mattevi, A., *Biochemistry*, 2001, vol. 40, pp. 2766–2776.
14. Templeton, D.M. and Sarkar, B., *Can. J. Chem.*, 1985, vol. 63, pp. 3122–3128.
15. Wolfenden, R., *Annu. Rev. Biophys. Bioeng.*, 1976, vol. 5, pp. 271–306.
16. Lee, Y.B., Park, M.H., and Folk, J.E., *J. Med. Chem.*, 1995, vol. 38, pp. 3053–3061.
17. Lin, P.K., Maguire, N.M., and Brown, D.M., *Tetrahedron Lett.*, 1994, vol. 35, pp. 3605–3608.
18. Kuksa, V., Buchan, R., and Lin, P.K., *Synthesis*, 2000, no. 9, pp. 1189–1207.
19. Siaugue, J.-M., Segat-Dioury, F., Favre-Reguillon, A., Madic, Ch., Foos, J., and Guy, A., *Tetrahedron Lett.*, 2000, vol. 41, pp. 7443–7446.
20. Fukuyama, T., Jow, Ch.K., and Cheung, M., *Tetrahedron Lett.*, 1995, vol. 36, pp. 6373–6374.
21. Khomutov, A.R., Vepsalainen, J., Shvetsov, A.S., Hyvonen, T., Keinanen, T.A., Pustobaev, V.N., Eloranta, T.O., and Khomutov, R.M., *Tetrahedron*, 1996, vol. 52, pp. 13 751–13 766.
22. Tietze, L.F. and Eicher, T., *Reactionen und Synthesen im Organisch-chemischen Praktikum und Forschungslaboratorium*, Stuttgart: Georg Thieme, 1991. Translated under the title *Preparativnaya organicheskaya khimiya. Reaktsii i sintezy v praktikume organicheskoi khimii i nauchno-issledovatel'skoi laboratorii*, Moscow: Mir, 1999.