# Synthesis and Antibacterial Activity of a New Series of Tetracyclic Pyridone Carboxylic Acids. $2^{1a}$

Yoshikazu Jinbo,\* Masahiro Taguchi, Yoshimasa Inoue, Hirosato Kondo, Tomohiro Miyasaka, Hideki Tsujishita, Fumio Sakamoto, and Goro Tsukamoto

New Drug Research Laboratories, Kanebo, Ltd., 5-90, Tomobuchi-cho 1-Chome Miyakojima-ku, Osaka 534, Japan

Received April 26, 1993®

A novel tetracyclic pyridone carboxylic acid with a thiazolidine ring, 1,2-dihydro-9,1-(epoxymethano)-7-fluoro-8-(4-methyl-1-piperazinyl)-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylic acid (4a), and variants with a nitrogen atom (4b) or carbonyl group (4c) in the place of the 10-position oxygen atom of 4a were prepared and tested for antibacterial activity and inhibitory activity on DNA gyrase from Escherichia coli KL-16. The in vitro antibacterial potency with regard to the 10-position atom was found to be of the following order;  $O > NCH_3 = C = O$ . The  $IC_{50}$  values for DNA gyrase inhibition activity for the 4a, 4b, and 4c compounds were 0.33, 0.53, and 0.67 g/mL, respectively. The activity of 4a, in which the C-3 methyl group and C-5 of ofloxacin (2a) were connected with a sulfur atom to restrict the conformation of 2a, was more potent than that of 2a against both Gram-positive and -negative bacteria, except for Pseudomonas aeruginosa. Compared to the tetracyclic pyridone carboxylic acid 1a, which has a flat thiazole ring, compound 4a showed comparable or slightly more potent activity against both Gram-positive and -negative bacteria, except for P. aeruginosa.

Since the development of norfloxacin<sup>2</sup> as a useful therapeutic agent, a large number of analogues in which a fluorine atom is introduced at the C-6 position of the quinolone ring system have been synthesized. We recently reported the synthesis and antibacterial activity of a series of tetracyclic pyridone carboxylic acids with a flat thiazole ring (1a-d).<sup>1,3</sup> The antibacterial activity of 1b was comparable to or more potent than that of 1a. The structure—activity relationship of these tetracyclic pyridone carboxylic acids (1a-d) is different from that of the tricyclic ofloxacin analogues (2a-d),<sup>4-6</sup> and the dicyclic ciprofloxacin analogues (3a,b).<sup>7</sup> In compounds 2a,b and 3a,b, the introduction of a nitrogen atom into the C-8 position of the quinolone rings resulted in an overall reduction in antibacterial activity.

In the course of our study of tetracyclic quinolone antibacterials, we focused on new tetracyclic pyridone carboxylic acids (4a-c) that have a thiazolidine ring that is not flat (Chart I).

Kondo et al.<sup>8</sup> and Segawa et al.<sup>9a</sup> have reported that tricyclic pyridone carboxylic acids (5a,b) with a non-flat thiazolidine ring exhibited potent antibacterial activity.

Ohta et al.  $^{10}$  reported that of loxacin (2a) had two stable conformations without significant energy difference. One of the stable conformers is that where the C-3 methyl moiety is perpendicular to the plane of the quinolone ring (dihedral angle  $5-4-3-11=80^{\circ}$ ) and the other is that where it is rather inclined to the plane of the ring (dihedral angle  $5-4-3-11=32^{\circ}$ ). They speculated that the latter was the active conformer, contributing to the antibacterial activity. Compound 4a is a conformationally restricted analogue of 2a, and its dihedral angle (3a-1a-1-2) would be close to that (5-4-3-11) of the active conformer of 2a. This compound 4a could therefore be expected to have potent antibacterial activity.

Here, we describe the synthesis, antibacterial activity, and inhibition of DNA gyrase supercoiling activity of a new series of tetracyclic pyridone carboxylic acids, 4a-c.

#### Chemistry

The route of synthesis of the target compound 4a is summarized in Scheme I. The starting material a 6 was reacted with 2,3-dibromopropanol in the presence of K<sub>2</sub>-CO<sub>3</sub> in DMF to give 7. Treatment of 7 with K<sub>2</sub>CO<sub>3</sub> in DMF gave the mixture of 8 and its rearranged isomer 9, the product ratio being 1:9, on the basis of the <sup>1</sup>H NMR spectrum. These compounds could not be separated, and their structures were deduced from the structures of the final products 4a and 10. The construction of 9 is considered to have arisen as follows: the hydroxyl group of 7 attacked at the C-3a position to produce thiolate 11, and this was followed by cyclization to afford the tetracyclic compound 9 (Scheme II). Attempts to convert 7 to 8 selectively, under various kinds of basic conditions (NaH/ DMF, LDA/THF, potassium tert-butoxide/DMF, and Mg, CCl<sub>4</sub>/DMF), were unsuccessful. The mixture of 8 and 9 was reacted with triacetoxyborane in acetic anhydride to isolate 13 and to afford a crude mixture containing 12. Compound 13 was reacted with 1-methylpiperazine and was then hydrolyzed under acidic conditions to give 10. Similar treatment of the crude mixture containing 12 gave 4a (contaminated with a small amount of 10). The structures of 4a and 10 were determined on the basis of the <sup>13</sup>C NMR spectra (Table I). The signals attributed to C-9 of 4a and 10 in  $D_2O$  were observed at 140.1 and 122.0 ppm (or 127.2 ppm), respectively. The higher shift of the signal of the former compared with that of the latter definitely supports the suggested structures of 4a and 10.

The synthesis of 4b was achieved according to the route of synthesis shown in Scheme III. The starting material  $^{1a}$  6 was reacted with N-(benzyloxycarbonyl)-N-methyl-2,3-dibromopropylamine  $^{11}$  in the presence of  $K_2CO_3$  in DMF to afford 14. Deprotection of the benzyloxycarbonyl group of 14 with HBr/HOAc gave the crude 15, followed by the cyclization of 15 without purification to give the tetracyclic compound 16. In the  $^{1}$ H NMR of 16, the doublet signal at 3.19 ppm (3 H, d, J=3 Hz), based on the coupling of N-CH<sub>3</sub> and the fluorine atom at the C-8 position, was observed, and this suggested the structure of N-CH<sub>3</sub> at

Abstract published in Advance ACS Abstracts, September 15, 1993.

$$\begin{array}{c|c} & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & \\ & & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\$$

the 10-position. On cyclization of 15, a rearranged isomer similar to 9 was not given; the reason for this was thought to be that the steric hindrance of the methylamino group prevented an attack on the C-3a position. The reaction of 16 with triacetoxyborane in acetic anhydride afforded 17. Replacement of the C-8 fluorine atom of 17 by 1-methylpiperazine, followed by hydrolysis under acidic conditions, gave 4b.

The synthesis of 4c was achieved according to the route of synthesis shown in Scheme IV. The starting material<sup>8</sup> 18 was reacted with methyl 3,4-dibromobutyrate<sup>12</sup> in the presence of  $K_2CO_3$  to give 19, followed by the reaction with triacetoxyborane in acetic anhydride to afford 20. Compound 20 was reacted with 1-methylpiperazine, and, when hydrolyzed under acidic conditions, gave 21. Finally, cyclization of 21 in polyphosphoric acid afforded 4c.

## Biological Results and Discussion

Table II shows the in vitro antibacterial activity of a series of tetracyclic pyridone carboxylic acids (4a-c), 10, 1a, 1b, and 2a (ofloxacin) against four Gram-positive bacteria (Staphylococcus aureus FDA 209P JC-1, Staphylococcus aureus IID 803, Staphylococcus epidermidis IAM 1296, and Enterococcus faecalis IID 682) and five Gram-negative bacteria (E. coli NIHJ JC-2, E. coli KC-14, Klebsiella pneumoniae B54, Pseudomonas aeruginosa IFO 3445, and Pseudomonas aeruginosa E-2). Compound 4a exhibited 2-16 times more potent antibacterial activity than 4b and 4c against both Gram-positive and -negative bacteria. The activity of 4b was comparable to that of 4c. This result clearly indicated that an oxygen atom in the 10-position of a tetracyclic pyridone carboxylic acid with a non-flat thiazolidine ring was optimal relative to a nitrogen atom or a carbonyl group. This structure-activity relationship is similar to that 4-6 noted for a series of tricyclic ofloxacin analogues and is different from that1a of tetracyclic quinolone analogues that have a flat thiazole ring (1a-c). The antibacterial activity of 4a was compared to that of the reference compounds 1a, 1b, and 2a. The activity of 4a against Gram-positive bacteria was more potent than that of 2a, comparable to that of 1a, and less potent than that of 1b. Against Gram-negative bacteria. except for P. aeruginosa, 4a was slightly more potent than 1a and 2a and comparable to 1b.

Figure 1 and Table III show the inhibition of the supercoiling activity of DNA gyrase isolated from  $E.\ coli$  KL-16, along with the in vitro activity against the same organism, of 4a-c, 10, 1a, 1b, and 2a. When the IC<sub>50</sub> values of the tetracyclic quinolone series that have a thiazoline ring (4a-c) are compared, it can be seen that replacement of the 10-position oxygen atom of 4a by a nitrogen atom or a carbonyl group somewhat reduces the inhibition of DNA gyrase. The MIC value of 4b was 16-fold greater than that of 4a, representing a much wider potency difference than the 1.6-fold differential in their IC<sub>50</sub>. This

## Scheme Ia

$$F = \begin{pmatrix} OH & CO_2Et \\ F & N \end{pmatrix} = \begin{pmatrix} OH & CO_2Et \\ OH & F \end{pmatrix} = \begin{pmatrix} CO_2Et \\ F & N \end{pmatrix} = \begin{pmatrix} CO_2Et \\ F$$

### Scheme II

$$\begin{bmatrix}
F & CO_2Et \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
CO_2Et \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
CO_2Et \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

$$\begin{bmatrix}
F & OH \\
F & OH
\end{bmatrix}$$

Table I. <sup>13</sup>C NMR Chemical Shifts (ppm) of 4a and 10 in D<sub>2</sub>O and Their Assignment

F 7 6 5a 4 CO<sub>2</sub>H  
14 13 N 8 9 N 4 HC  
15 11 2

4a 
$$\times$$
 = O,  $\times$  = S

assignment	4a	10		
1	63.8	59.8		
2	34.1	78.7		
$3a^a$	165.1	168.9		
4	103.7	93.8		
5	177.8  (d, J = 3  Hz)	180.9  (d, J = 3  Hz)		
5a	118.6  (d, J = 10  Hz)	127.2 <sup>b</sup> (br)		
6	106.2 (d, J = 25 Hz)	110.3  (d, J = 24  Hz)		
7	157.6  (d, J = 248  Hz)	159.9  (d, J = 253  Hz)		
8	133.8  (d, J = 15  Hz)	141.2 (d, J = 16 Hz)		
9	140.1  (d, J = 7  Hz)	$122.0^b  (d, J = 7  Hz)$		
9a	126.7	129.4		
11	70.2	28.6		
12ª	170.6	<b>164</b> .0		
13	56.8	56.9 (d, J = 1 Hz)		
14	49.9	49.9		
15	45.9	45.8		

a,b These assignments may be interchanged.

finding indicates that replacement of the 10-position oxygen atom of 4a by a nitrogen atom reduced cell permeability<sup>13</sup> along with producing a decrease in inherent activity. The IC<sub>50</sub> data of 4a and 1a and 4b and 1b show that a replacement of the flat thiazole ring of tetracyclic pyridone carboxylic acid with non-flat thiazolidine ring reduces the inhibitory effect on DNA gyrase. It appears that a modification of a flat thiazole ring with a non-flat thiazolidine ring had slightly a unfavorable effect on DNA gyrase inhibitory activity. When the IC50 value of 4a was compared with that of 2a, we found that the DNA gyrase inhibitory activity of 4a was 1.6-fold more potent than that of 2a. Compound 4a exhibited more potent antibacterial activity and was a more potent inhibitor of DNA gyrase than 2a. This result might be interpreted in terms of the effects of conformational restrictions; a stable conformer of 4a fixed by connection of the 3-methyl group and the C-5 of 2a with a sulfur atom fitted the active conformer of 2a. This speculation was supported by performing conformational analysis of the model compound 22 by molecular mechanics (Chart II). The compound 22 has only one minimum-energy conformer; this has a dihedral angle (3a-1a-1-2) of 24°, and its conformer resembles the active conformer of 2a (corresponding dihedral angle (5-4-3-11) is 32°).<sup>10</sup>

Compound 10, in which the 3-sulfur atom and 10-oxygen atom of 4a were interchanged, exhibited poor antibacterial activity and inhibitory effect against DNA gyrase.

These findings suggests that a tetracyclic quinolone framework that has a non-flat thiazolidine ring, i.e., the framework of 4a, is a structural variation of the pyridone carboxylic acids that will be of value as an antibacterial agent.

#### **Experimental Section**

Melting points were determined with a Büchi capillary melting point apparatus, Model 535; all melting points are uncorrected.  $^1\mathrm{H}$  NMR spectra were recorded on a Bruker AM-300 spectrometer with TMS or 3-(trimethylsilyl)-3-propanesulfonic acid sodium salt as an internal reference in a solution of CDCl<sub>3</sub>, DMSO-d<sub>6</sub>, or D<sub>2</sub>O. IR spectra were recorded with a Hitachi IR 270-50 infrared spectrometer. Elemental analyses were performed with a Yanagimoto CHN-CORDER MT-3, and all analytical values were within  $\pm 0.4\%$  of the calculated theoretical values.

Ethyl 1,2-Dihydro-1-(hydroxymethyl)-5-oxo-7,8,9-trifluoro-5*H*-thiazolo[3,2-a]quinoline-4-carboxylate (7). A mixture of ethyl 4-hydroxy-6,7,8-trifluoro-2-mercaptoqunoline-3-carboxylate (6) (3.80 g, 12.5 mmol), 2,3-dibromopropanol (3.29 g, 15.1 mmol), and  $K_2CO_3$  (6.08 g, 44.0 mmol) in DMF (38 mL) was stirred at 50-55 °C for 4 h. The resulting precipitate was collected by filtration and washed with DMF, water, and acetone to afford 7 (2.96 g, 65%) as colorless crystals. 7 (recrystallized from DMSO): mp 266 °C dec; ¹H NMR (DMSO- $d_6$ )  $\delta$  1.27 (3 H, t, J = 7 Hz), 3.36 (1 H, d, J = 12 Hz), 3.5-3.7 (3 H, m), 4.25 (2 H, q, J = 7 Hz), 5.32 (1 H, t, J = 6 Hz), 5.61 (1 H, q, J = 6.5 Hz), 7.90 (1 H, ddd, J = 2, 8.5, 10.5 Hz). Anal. ( $C_{16}H_{12}F_3NO_4S$ ) C, H, N.

Mixture of Ethyl 1,2-Dihydro-9,1-(epoxymethano)-7,8-difluoro-5-oxo-5*H*-thiazolo[3,2-a]quinoline-4-carboxylate (8) and Ethyl 1,2-Dihydro-9,1-(epithiomethano)-7,8-difluoro-5-oxo-5*H*-oxazolo[3,2-a]quinoline-4-carboxylate (9). A mixture of 7 (5.42 g, 15.1 mmol) and  $K_2CO_3$  (10.13 g, 73.3 mmol) in DMF (50 mL) was stirred at 45–50 °C for 17 h. The resulting precipitate was collected by filtration washed with DMF, water, and acetone, and recrystallized from DMSO to afford a mixture of 8 and 9 (8:9 = 1:9, determined by the integration of the signal at  $\delta$  7.70 and 7.51) (1.16 g, 22%) as colorless crystals: mp >280 °C. Anal. ( $C_{15}H_{11}F_2NO_4S\cdot0.5H_2O$ ) C, H, N.

8:  $^{1}$ H NMR (DMSO-d<sub>6</sub>)  $\delta$  1.28 (3 H, t, J = 7 Hz), 3.27 (1 H, dd, J = 6, 11 Hz), 3.60 (1 H, dd, J = 7, 11 Hz), 4.24 (2 H, q, J = 7 Hz), 4.36 (1 H, dd, J = 9.5, 10.5 Hz), 4.8–5.0 (1 H, m), 5.02 (1 H, dd, J = 4, 10.5 Hz), 7.51 (1 H, dd, J = 7.5, 11 Hz).

9:  $^{1}$ H NMR (DMSO- $^{2}$ d<sub>6</sub>)  $\delta$  1.24 (3 H, t, J = 7 Hz), 3.37 (1 H, dd, J = 10.5, 12.5 Hz), 3.73 (1 H, dd, J = 2.5, 12.5 Hz), 4.19 (2 H, q, J = 7 Hz), 4.55 (1 H, dd, J = 8.5, 11 Hz), 4.8–5.0 (1 H, m), 5.16 (1 H, t, J = 8.5 Hz), 7.70 (1 H, dd, J = 8.5, 10.5 Hz).

1,2-Dihydro-9,1-(epoxymethano)-7-fluoro-8-(4-methyl-1piperazinyl)-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylic Acid Hydrochloride (4a) and Diacetoxy [[[1,2-dihydro-9,1-(epithiomethano)-7-fluoro-5-oxo-5H-oxazolo[3,2a]quinolin-4-yl]carbonyl]oxy]borane (13). The mixture described above (8 and 9) (500 mg, 1.47 mmol) was added to a suspension of triacetoxyborane (420 mg, 2.23 mmol) in acetic anhydride (6 mL), and the mixture was stirred at room temperature for 68 h. After addition of isopropyl ether (6 mL) to the mixture, the resulting precipitate was collected by filtration and washed with isopropyl ether to give the crude crystals of 13. The filtrate was concentrated in vacuo, and the residue was washed with isopropyl ether to give the precipitate (189 mg). The mixture of this precipitate and 1-methylpiperazine (0.15) mL, 1.35 mmol) in DMSO (1.9 mL) was stirred at room temperature for 67 h, and then the solvent was evaporated in vacuo. HCl (2 N, 2 mL) was added to the residue, and the mixture was stirred at room temperature for 1 h. The solvent was

#### Scheme IIIa

F CO<sub>2</sub>Et a F CO<sub>2</sub>Et c F CO<sub>2</sub>R<sup>2</sup> a F CO<sub>2</sub>R<sup>2</sup> CO<sub>2</sub>R<sup>2</sup> CO<sub>2</sub>R<sup>2</sup> Ab 
$$CH_3$$
  $CH_3$   $CH_3$ 

a (a) N-(Benzyloxycarbonyl)-N-methyl-2,3-dibromopropylamine, K<sub>2</sub>CO<sub>3</sub> in DMSO; (b) HBr/AcOH; (c) Et<sub>3</sub>N; (d) B(OAc)<sub>3</sub> in Ac<sub>2</sub>O; (e) (i) 1-methylpiperazine, (ii) HCl.

#### Scheme IVa

FOR 
$$CO_2Et$$
 $A$ 
 $CO_2CH_3$ 
 $CO_2CH_3$ 

<sup>a</sup> (a) Methyl 3,4-dibromobutylate, K<sub>2</sub>CO<sub>3</sub> in DMF; (b) B(OAc)<sub>3</sub> in Ac<sub>2</sub>O; (c) (i) 1-methylpiperazine, (ii) HCl; (d) (i) PPA, (ii) HCl.

Table II. In Vitro Antibacterial Activitya (Minimum Inhibitory Concentration, µg/mL) of Tetracyclic Pyridone Carboxylic Acids 4a-c, 10, 1a, 1b, and 2a

4a-c

		microorganism <sup>b</sup>								
		Gram-positive			Gram-negative					
compd	X	Sa(F)	Sa(I)	Se	Ef	Ec(N)	Ec(K)	Кр	Pa(I)	Pa(E)
4a <sup>c</sup>	0	0.10	0.10	0.20	0.78	0.05	0.025	0.05	1.56	1.56
4b	$NCH_3$	0.78	0.39	0.78	1.56	0.78	0.78	0.39	6.25	6.25
4c	C=0	0.78	0.78	3.13	3.13	0.78	0.39	0.78	6.25	3.13
10	S(3-O)	1.56	3.13	6.25	6.25	1.56	1.56	1.56	50	60
1a		0.10	0.10	0.20	0.39	0.10	0.10	0.10	0.78	0.78
1 <b>b</b>		0.05	0.05	0.05	0.10	0.05	0.05	0.05	0.39	0.78
2a (OFL)		0.39	0.39	0.78	1.56	0.10	0.10	0.10	1.56	1.56

<sup>a</sup> All values were obtained from duplicate or triplicate experiments. <sup>b</sup> Microorganism: Sa(F), Staphylococcus aureus FDA 209P JC-1; Sa(I), Staphylococcus aureus IID 803; Se, Staphylococcus epidermidis IAM 1296; Ef, E. faecalis IID 682; Ec(N), E. coli NIHJ JC-2; Ec(K), E. coli KC-14; Kp, Klebsiella pneumoniae B54; Pa(I), Pseudomonas aeruginosa IFO 3445; Pa(E), Pseudomonas aeruginosa E-2. Contaminated with a small amount (8% from NMR) of 10.

evaporated in vacuo, and the residue was recrystallized from methanol to afford 4a (7 mg, 11%) as colorless crystals. The above crude crystals of 13 were recrystallized twice from acetonitrile to give 13 (209 mg, 35%) as colorless crystals.

4a: mp >280 °C; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.99 (3 H, s), 3.2-3.3 (3 H, m), 3.4-3.8 (7 H, m), 4.15 (1 H, t, J = 11 Hz), 4.6-4.8 (1 H, m), 4.98 (1 H, dd, J = 3.5, 11 Hz), 7.01 (1 H, d, J = 12 Hz); IR (KBr):1698, 1482 cm<sup>-1</sup>. Anal. (C<sub>18</sub>H<sub>19</sub>ClFN<sub>3</sub>O<sub>4</sub>S·1.5H<sub>2</sub>O) C, H, N.

13: mp 253 °C dec; <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.90 (3 H, s), 1.91 (3 H, s), 3.59 (1 H, dd, J = 10.5, 13 Hz), 3.88 (1 H, dd, J = 2.5,13 Hz), 4.94 (1 H, dd, J = 9, 11.5 Hz), 5.32 (1 H, m), 5.52 (1 H, t, J = 9 Hz), 8.04 (1 H, dd, J = 8, 10 Hz). Anal. ( $C_{17}H_{12}BF_{2}$ -NO<sub>8</sub>S-0.5H<sub>2</sub>O) C, H, N.

1,2-Dihydro-9,1-(epithiomethano)-7-fluoro-5-oxo-5H-oxazolo[3,2-a]quinoline-4-carboxylic Acid Hydrochloride (10). A mixture of 13 (300 mg, 0.68 mmol) and 1-methylpiperazine (0.25 mL, 2.25 mmol) in DMSO (3 mL) was stirred at room temperature for 2 h, and then the solvent was evaporated in vacuo. HCl 2 N, 4 mL) was added to the residue, and the mixture was stirred at room temperature for 1 h. The resulting precipitate was collected by filtration and recrystallized from 1 N HCl to afford 10 (72 mg, 24%) as colorless crystals: mp >280 °C; ¹H NMR (D<sub>2</sub>O)  $\delta$  3.00 (3 H, s), 3.2–3.7 (7 H, m), 4.66 (1 H, dd, J =8.5, 11.5 Hz), 4.8-5.0 (1 H, m), 5.34 (1H, t, J = 8.5 Hz), 7.26 (1 H, d, J = 11.5 Hz); IR (KBr) 1714, 1511 cm<sup>-1</sup>. Anal. (C<sub>18</sub>H<sub>19</sub>-CIFN<sub>3</sub>O<sub>4</sub>S-1.5H<sub>2</sub>O) C, H, N.

Ethyl 1-[[N-(Benzyloxycarbonyl)-N-methylamino]methyl]-1,2-dihydro-5-oxo-7,8,9-trifluoro-5H-thiazolo[3,2-a]quinoline-4-carboxylate (14). A mixture of ethyl 4-hydroxy-6,7,8-trifluoro-2-mercaptoquinoline-3-carboxylate (6) (1.20 g, 3.96 mmol), N-(benzyloxycarbonyl)-N-methyl-2,3-dibromopropylamine (1.80 g, 4.93 mmol), and K<sub>2</sub>CO<sub>3</sub> (2.70 g, 19.5 mmol) in DMF (15 mL) was stirred at 50 °C for 3 h, and then the solvent was evaporated in vacuo. The residue was added to water and extracted with CHCl<sub>3</sub>. The CHCl<sub>3</sub> extract was washed with water and dried over Na<sub>2</sub>SO<sub>4</sub>. Evaporation of solvents in vacuo, purification of the residue by silica gel column chromatography (silica gel 60, 230–400 mesh, Merck; CHCl<sub>3</sub> as an eluent), and recrystallization from EtOAc/cyclohexane gave 14 (1.10 g, 55%) as colorless crystals: mp 152.5-153.0 °C; ¹H NMR (CDCl<sub>3</sub>) δ 1.43

Figure 1. Inhibitory patterns of 4a-c, 10, 1a, 1b, and 2a against the supercoiling activity of  $E.\ coli\ KL-16\ DNA$  gyrase: differences between 4a and other compounds. \* p<0.05, \*\* P<0.01 vs 4a.

**Table III.** Inhibitory Effects  $(IC_{50})^a$  of 4a-c, 10, 1a, 1b, and 2a on Supercoiling Activity of DNA Gyrase from  $E.\ coli$  KL-16 and in Vitro Antibacterial Activity  $(MIC)^b$  against the Same Strain

compd	IC <sub>50</sub> , μg/mL	MIC, μg/mL	compd	IC <sub>50</sub> , μg/mL	MIC, μg/mL
4a	0.33	0.05	1a	0.14	0.05
4b	0.53	0.78	1b	0.16	0.05
4c	0.67	0.39	2a	0.53	0.10
10	2.61	1.56			

 $^a$  Calculated by the quantitative measurement of the supercoiled DNA peak in an agarose gel by densitometric assay.  $^b$  See footnote in Table II.

(3 H, t, J = 7 Hz), 3.7-3.9 (1 H, m), 4.43 (2 H, q, J = 7 Hz), 4.9-5.3 (2 H, m), 5.7-5.9 (1 H, m), 7.3-7.4 (5 H, m), 8.08 (1 H, ddd, J = 2, 9, 10 Hz). Anal.  $(C_{24}H_{21}F_3N_2O_5S)$  C, H, N.

Ethyl 7,8-Difluoro-1,2-dihydro-9,1-[(N-methylimino)methano]-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylate (16). A mixture of 14 (1.00 g, 1.98 mmol) and 30% HBr/AcOH (10 mL) was stirred at room temperature for 2 h. The solvent was evaporated in vacuo, and the residue was washed with methanol to give the crystals (450 mg). Triethylamine (0.55 mL, 3.95 mmol) was added to a solution of these crystals in CH<sub>3</sub>CN (20 mL), and the mixture was stirred at 70 °C for 18 h. The mixture was diluted with CHCl<sub>3</sub>, washed with dilute HCl and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated in vacuo, and the residue was recrystallized from CH<sub>3</sub>CN/CH<sub>3</sub>OH to afford 16 (200 mg, 28%) as colorless crystals: mp >280 °C; ¹H NMR (CDCl<sub>3</sub>)  $\delta$  1.43 (3 H, t, J = 7 Hz), 3.15 (1 H, dd, J = 11, 12.5 Hz), 3.19 (3 H, d, J = 3 Hz), 3.36 (1 H, dd, J = 7, 11 Hz), 3.44 (1 H, dd, J= 10, 13 Hz), 3.58 (1 H, dd, J = 3.5, 13 Hz), 4.40 (2 H, q, J = 7 Hz), 4.68 (1 H, m), 7.58 (1 H, dd, J = 8, 10.5 Hz). Anal.  $(C_{16}H_{14}F_2N_2O_3S)$  C, H, N.

Diacetoxy[[[7,8-difluoro-1,2-dihydro-9,1-[(N-methylimino)methano]-5-oxo-5H-thiazolo[3,2-a]quinolin-4-yl]carbonyl]oxy]borane (17). A mixture of 16 (1.00 g, 2.84 mmol) and triacetoxyborane (0.80 g, 4.26 mmol) in acetic anhydride (20 mL) was stirred at 70 °C for 1 h. The solvent was evaporated in vacuo, and the residue was washed with isopropyl ether (50 mL) to give 17 (1.10 g, 85%) as colorless crystals: mp 269 °C dec; ¹H NMR (DMSO- $d_6$ )  $\delta$  1.90 (6 H, s), 3.21 (3 H, d, J = 4 Hz), 3.54 (1 H, dd, J = 11.5, 13.5 Hz), 3.67 (1 H, dd, J = 10.5, 13 Hz), 3.8–3.9 (2 H, m), 5.2–5.4 (1 H, m), 7.59 (1 H, dd, J = 7.5, 10 Hz). Anal. (C<sub>18</sub>H<sub>15</sub>BF<sub>2</sub>N<sub>2</sub>O<sub>7</sub>S·1.5H<sub>2</sub>O) C, H, N.

1,2-Dihydro-7-fluoro-9,1-[(N-methylimino)methano]-8-(4-methyl-1-piperazinyl)-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylic Acid Hydrochloride (4b). A mixture of 17 (230 mg, 0.509 mmol) and 1-methylpiperazine (0.23 mL, 2.07 mmol) in DMSO (2 mL) was stirred at 60 °C for 15 h. Ethanol (6 mL) was to the mixture, and the mixture was filtered. The filtrate was concentrated in vacuo, and 2 N HCl (1 mL) was added to the residue. The mixture was stirred at room temperature for 15 min. After addition of acetone (20 mL) to the mixture, the resulting precipitate was collected by filtration and recrystallized from methanol to afford 4b (58 mg, 25%) as colorless crystals:



 $^{o}$  The dihedral angles 1 (3a-1a-1-2) and 2 (11-10-9-9a) of the stable conformation were 24° and 21°, respectively.

mp >280 °C dec; ¹H NMR ( $D_2O$ )  $\delta$  2.89 (3 H, s), 3.02 (3 H, s), 2.9–3.1 (2 H, m), 3.2–3.4 (3 H, m), 3.4–3.8 (7 H, m), 4.4–4.6 (1 H, m), 6.74 (1 H, d, J = 12 Hz); IR (KBr) 1689, 1473 cm<sup>-1</sup>. Anal. ( $C_{19}H_{22}CIFN_4O_3S\cdot1.25H_2O$ ) C, H, N.

Ethyl 7,8-Difluoro-1,2-dihydro-1-[(methoxycarbonyl)methyl]-5-oxo-5*H*-thiazolo[3,2-a]quinoline-4-carboxylate (19). A mixture of ethyl 6,7-difluoro-4-hydroxy-2-mercaptoquinoline-3-carboxylate (18) (3.00 g, 10.5 mmol), methyl 3,4-dibromobutylate (3.00 g, 11.5 mmol), and  $K_2CO_3$  (4.38 g, 31.7 mmol) in DMF (30 mL) was stirred at 50-55 °C for 3 h. The solvent was evaporated in vacuo, and the residue was washed with  $H_2O$  and acctone to give 19 (1.91 g, 47%) as colorless crystals. 19 (recrystallized from methanol): mp 183.0-184.4 °C; ¹H NMR (DMSO- $d_6$ )  $\delta$  1.28 (3 H, t, J = 7 Hz), 2.69 (1 H, dd, J = 2.5, 16.5 Hz), 2.88 (1 H, dd, J = 10, 16.5 Hz), 3.42 (1 H, d, J = 12 Hz), 3.62 (3 H, s), 3.71 (1 H, dd, J = 7.5, 12 Hz), 4.24 (2 H, q, J = 7 Hz), 5.6-5.7 (1 H, m), 7.94 (1 H, dd, J = 6.5, 12 Hz), 7.99 (1 H, dd, J = 9, 10.5 Hz). Anal. ( $C_{17}H_{15}F_2NO_5S$ ) C, H, N.

Diacetoxy [[(7,8-difluoro-1,2-dihydro-1-[(methoxycarbonyl)methyl]-5-oxo-5H-thiazolo[3,2-a]quinolin-4-yl)carbonyl]oxy]borane (20). A mixture of 19 (1.81 g, 4.72 mmol) and triacetoxyborane (1.34 g, 7.12 mmol) in acetic anhydride (9 mL) was stirred at room temperature for 25 min and then at 45–50 °C for 135 min. After addition of ether (100 mL) to the mixture, the resulting precipitate was collected by filtration, washed with ether, and recrystallized from CHCl<sub>3</sub>/ether to give 20 (1.55 g, 67%) as colorless crystals: mp 215 °C dec; 'H NMR (DMSO- $d_6$ )  $\delta$  1.90 (3 H, s), 1.91 (3 H, s), 2.82 (1 H, dd, J = 2.5, 16.5 Hz), 3.07 (1 H, dd, J = 9.5, 16.5 Hz), 3.62 (3 H, s), 3.72 (1 H, d, J = 12 Hz), 4.02 (1 H, dd, J = 8.5, 12 Hz), 6.05 (1 H, m), 8.33 (1 H, dd, J = 8.5, 10 Hz), 8.52 (1 H, dd, J = 6.5, 12 Hz). Anal. (C<sub>19</sub>H<sub>16</sub>BF<sub>2</sub>-NO<sub>9</sub>S-2H<sub>2</sub>O) C, H, N.

1,2-Dihydro-7-fluoro-1-[(methoxycarbonyl)methyl]-8-(4-methyl-1-piperazinyl)-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylic acid Hydrochloride (21). A mixture of 20 (500 mg, 1.03 mmol) and 1-methylpiperazine (0.49 mL, 4.42 mmol) in DMSO (4.4 mL) was stirred at room temperature for 75 min. The solvent was evaporated in vacuo, 2 N HCl (10 mL) was added to the residue, and the mixture was stirred at room temperature for 1 h. The resulting precipitate was collected by filtration and washed with 2 N HCl, water, and acetone to afford the hydrochloride of 21 (310 mg, 63%) as colorless crystals. 21 (recrystallized from water): mp 235 °C dec; ¹H NMR (D<sub>2</sub>O)  $\delta$  2.75 (1 H, d, J = 16 Hz), 3.01 (1 H, dd, J = 9, 16 Hz), 3.06 (3 H, s), 3.3-3.5 (4 H, m), 3.53 (1 H, d, J = 12 Hz), 3.73 (3 H, s), 3.7-4.0 (5 H, m), 5.81 (1 H, m), 6.88 (1 H, d, J = 7 Hz), 7.37 (1 H, d, J = 13 Hz). Anal. (C<sub>20</sub>H<sub>24</sub>ClFN<sub>3</sub>O<sub>5</sub>S-1H<sub>2</sub>O) C, H, N.

6-Fluoro-5-(4-methyl-1-piperazinyl)-4,8-dioxo-2,2a,3,4-tetrahydro-8*H*-benzo[*ij*]thiazolo[2,3,4-*de*]quinolizine-9-carboxylic Acid Hydrochloride (4c). A mixture of 21 (350 mg, 0.740 mmol) and polyphosphoric acid (3.5 g) was heated at 130 °C for 18 h. Water (12 mL) and 2 N HCl (4 mL) were added to the mixture, and the mixture was stirred at room temperature for 30 min. The resulting precipitate was collected by filtration, washed with 2 N HCl and acetone, and recrystallized from dilute HCl to give 4c (84 mg, 25%) as colorless crystals: mp >280 °C dec;  $^1$ H NMR (DMSO- $^1$ d<sub>6</sub>)  $^3$ 2.85 (3 H, s), 3.1-3.6 (9 H, m), 3.7-3.9 (2 H, m), 3.82 (1 H, dd,  $^1$ d = 7.5, 11.5 Hz), 5.07 (1 H, m), 8.08 (1 H, d,  $^1$ d = 13.5 Hz), 10.92 (1 H, br), 15.66 (1 H, s); IR (KBr) 1693, 1574, 1487 cm<sup>-1</sup>. Anal. ( $^1$ 1gClFN<sub>3</sub>O<sub>4</sub>S-0.5H<sub>2</sub>O) C, H, N.

In Vitro Antibacterial Activity. The MIC (minimum inhibitory concentrations) of compounds tested in this study were determined according to the standard method by a serial 2-fold agar dilution technique, using Sensitivity Disk agar (Nissui;

Tokyo, Japan). <sup>14</sup> The inoculum size was approximately 10<sup>6</sup> colony forming units/mL. The MIC of a compound was defined as the lowest concentration that prevented visible growth of bacteria after incubation at 37 °C for 18 h.

Inhibitory Effect on Supercoiling Activity of DNA Gyrase Isolated from E. coli KL-16. This assay was carried out according to a method reported previously. 15 Statistical analysis of data was performed by means of a simple linear regression analysis of the inhibitory patterns of DNA gyrase according to a logistic model, with a comparison of regression intercepts of 4a and other compounds. Planned pairwise comparisons were made by Student's t test.

### References

- (1) (a) Jinbo, Y.; Kondo, H.; Inoue, Y.; Taguchi, M.; Tsujishita, H.; Kotera, Y.; Sakamoto, F.; Tsukamoto, G. Synthesis and Antibacterial Activity of a New Series of Tetracyclic pyridonecarboxylic Acid. J. Med. Chem., in press. (b) Kotera, Y.; Inoue, Y.; Ohashi, M.; Ito, K.; Tsukamoto, G. Antibacterial Activity of a New Tetracyclic Quinolone, No.5290, against Norfloxacin- and ciprofloxacin-Resistant Strains of Staphylococcus aureus. Chem. Pharm. Bull. 1991 39, 2644-2646.
- (2) Koga, H.; Itoh, A.; Murayama, S.; Suzue, S.; Irikura, T. Structure-Activity Relationships of Antibacterial 6,7- and 7,8-Disubstituted 1-Alkyl-1,4-dihydro-4-oxoquinoline-3- carboxylic Acids. J. Med. Chem. 1980, 23, 1358-1363.
- (a) Taguchi, M.; Kondo, H.; Inoue, Y.; Kawahata, Y.; Jinbo, Y.; Sakamoto, F.; Tsukamoto, G. Synthesis and Antibacterial Activity of New Tetracyclic Quinolone Antibacterials. J. Med. Chem. 1992, 35, 94-99. (b) Kotera, Y.; Mitsuhashi,S. In Vitro and In vivo Antibacterial Activities of KB-5246, a New Tetracyclic Quinolone. Antimicrob. Agents Chemother. 1989, 33, 1896-1900.
- (a) Hayakawa, I.; Hiramitsu, T.; Tanaka, Y. Synthesis and Antibacterial Activities of Substituted 7-Oxo-2,3-dihydro-7Hpyrido[1,2,3-de][1,4]benzoxazine-6-carboxylic Acids. Chem. Pharm. Bull. 1984, 32, 4907-4913. (b) Sato, K.; Matsuura, Y.; Inoue, M.; Une, T.; Osada, Y.; Ogawa, H.; Mitsuhashi, S. In Vitro and In Vivo Activity of DL-8280, a New Oxazine Derivative. Antimicrob. Agents Chemother. 1982, 22, 548-553. Hayakawa, I.; Hiramitsu, T.; Sasaki, T.; Ebata, T.; Imamura, M.;
- Tanaka, Y. Syntheses and Antibacterial Activities of Pyrido[1,2,3de]-1,4-benzoxazine-6-carboxylic Acids and Their Analogues. Abstracts of the 5th Symposium on Medicinal Chemistry; Kyoto, December 9-10, 1983; Abstract No. G-4.
- (6) Asahara, M.; Tsuji, A.; Goto, S.; Masuda, K.; Kiuchi, A. In Vitro and In Vivo Activities of QA-241, a New Tricyclic Quinolone Derivative. Antimicrob. Agents Chemother. 1989, 33, 1144-1152.

- (7) Sanchez, J. P.; Domagala, J. M.; Hagen, S. E.; Heifetz, C. L.; Hutt, M. P.; Nichols, J. B.; Trehan, A. K. Quinolone Antibacterial Agents. Synthesis and Structure-Activity Relationships of 8-Substitued Quinoline 3-carboxylic Acids and 1,8-Naphthyridine-3-carboxylic Acids. J. Med. Chem. 1988, 31, 983-991.
- (8) Kondo, H.; Taguchi, M.; Inoue, Y.; Sakamoto, F.; Tsukamoto, G. Synthesis and Antibacterial Activity of Thiazolo-, Oxazolo-, and Imidazolo[3,2-a][1,8]naphthyridinecarboxilic Acids. J. Med. Chem. 1990, 33, 2012-2015.
- (a) Segawa, J.; Kitano, M.; Kazuno, K.; Tsuda, M.; Shirahase, I.; Ozaki, M.; Matsuda, M.; Kise, M. Studies on Pyridonecarboxylic Acids [1]. 2. Synthesis and Antibacterial Activity of 8-Substituted-7-fluoro-5-oxo-5H-thiazolo[3,2-a]quinoline-4-carboxylic Acids. J. Heterocycl. Chem. 1992, 29, 1117-1123. (b) Segawa, J.; Kitano, M.; Kazuno, K.; Matsuoka, M.; Shirahase, I.; Ozaki, M.; Matsuda, M.; Tomii, Y.; Kise, M. Studies on Pyridonecarboxylic Acids. 1. Synthesis and Antibacterial Evaluation of 7-Substituted 6-halo-4-oxo-4H-[1,3]thiazeto-[3,2-a]quinoline-3-carboxylic Acids. J. Med. Chem. 1992, 35, 4727-4738.
- (10) Ohta, M.; Koga, H. Three-Dimensional Structure-Activity Relationships and Receptor Mapping of N1-Substituents of Quinolone Antibacterials. J. Med. Chem. 1991, 34, 131-139.
- (11) N-(Benzyloxycarbonyl)-N-methyl-2,3-dibromopropylamine was prepared by the reaction of N-methylallylamine with benzyl chloroformate followed by bromination with bromine
- (12) Corey, E. J. Polar Effects in N-Bromosuccinimide Brominations. J. Am. Chem. Soc. 1953, 75, 2251-2252.
- (a) Domagala, J. M.; Hanna, L. D.; Heifetz, C. L.; Hutt, M. P.; Mich, T. F.; Sanchez, J. P.; Solomon, M. New Structure-Activity Relationships of the Quinolone Antibacterials Using the Target Enzyme. The Development and Application of a DNA Gyrase Assay. J. Med. Chem. 1986, 29, 394-404. (b) Wentland, M. P.; Perni, R. B.; Dorff, P. H.; Rake, J. B. Synthesis and Bacterial DNA Gyrase Inhibitory Properties of a Spirocyclopropylquinolone Derivative. J. Med. Chem. 1988, 31, 1694-1697.
- (14) Goto, S.; Jo, K.; Kawakita, T.; Kosakai, N.; Mitsuhashi, S.; Nishino, T.; Ohsawa, N.; Tanami, H. Saisho-hatsuikusoshi-nodo (MIC) Sokutei-ho Sai-kaitei Ni Tsuite; About the Re-revised Method of Determination of MIC. Chemotherapy 1981, 29, 76-79.
- (a) Inoue, Y.; Sato, K.; Fujii, T.; Hirai, K.; Inoue, M.; Iyobe, S.; Mitsuhashi, S. Some Properties of Subunits of DNA Gyrase from Pseudomonas aeruginosa PAO1 and Its Nalidixic Acid-Resistant Mutant. J. Bacteriol. 1987, 169, 2322-2325. (b) Sato, K.; Inoue, Y.; Fujii, T. Aoyama, H.; Inoue, M.; Mitsuhashi, S. Purification and Properties of DNA Gyrase from a Fluoroquinolone-Resistant Strain of Escherichia coli. Antimicrob. Agents Chemother. 1986, 30, 777-780.