

Preliminary communication

Structural variations of piritrexim, a lipophilic inhibitor of human dihydrofolate reductase: synthesis, antitumor activity and molecular modeling investigations

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Abstract

Piritrexim (PTX) (**1**), a lipophilic inhibitor of the human dihydrofolate reductase, has been evaluated as an anticancer agent. The synthesis of four structural variations (**2–5**) of PTX is reported. The PTX analogues **2–5** were obtained by reaction of suitable C₃-building blocks with pyrimidine-2,4,6-triamine (**14**) or with cyanacetamide (**7**) and guanidine (**10**). The evaluation of **2–4** for antitumor activity against a panel of 60 human cancer cell lines showed inhibitory effects on the growth of the cell lines. These data are supported by molecular modeling and docking studies, which show that compounds **2–4** share the same binding mode within the DHFR active site. Moreover, the estimated ligand binding energies are in good agreement with the experimental activity data.

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1. Introduction

Piritrexim (PTX) (**1**), first synthesized by Grivski et al. [1], is a lipophilic inhibitor of human dihydrofolate reductase with a pyrido[2,3-*d*]pyrimidine-2,4-diamine-structure. In contrast to methotrexate, PTX needs no active transport mechanism to reach into the cancer cell [2]. Piritrexim belongs to the lipophilic dihydrofolate reductase inhibitors of the second generation with lower affinity to histamin-*N*-methyl transferase resulting in less toxic side effects. [1,3,4]. In the NCI Cancer Test, PTX showed a significant activity against Walker 256 carcinoma, P 388 leukaemia, sarcoma 180 and the Ehrlich ascites carcinoma. In phase II studies activity against melanoma, lung cancer, colon cancer, sarcoma and head and neck cancer were reported [2].

One goal in our studies is the synthesis of new 5-deaza- and 8-deazapteridines as folate antagonists. Such a facile

synthesis of alkyl and aryl substituted pyrido[2,3-*d*]pyrimidine-2,4-diamines has been reported [5]. In previous work, Iso-PTX with a methyl group in position 7 instead of position 5 and an alternative synthesis of PTX was described [6,7].

It is well known that the pyrimidine-2,4-diamine structure is essential for inhibition of the dihydrofolate reductase by PTX and that increase of the chain to the aromatic ring in position 5 reduces activity [8–10]. In order to prepare new PTX-analogues, derivatives with a missing methyl group in position 5, an additional methyl group in position 7, and with an elongated sidechain in position 6 were prepared. Also the 5,6,7,8-tetrahydro derivative of Nor-PTX (**3**) was synthesized as a new structure modification (Fig. 1).

2. Chemistry

In the course of our syntheses of new folate antagonists with a pyrido[2,3-*d*]pyrimidine structure, different C₃-building blocks, e.g., β-dicarbonyl derivatives or α,β-unsaturated aldehydes, were cyclocondensated with different

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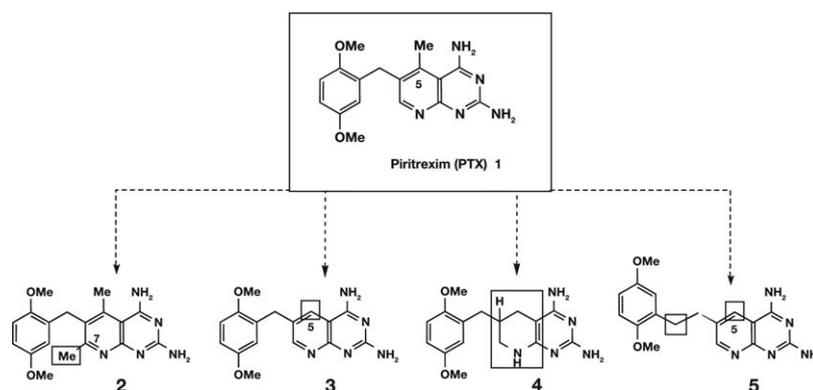


Fig. 1. Structural variation of PTX 1.

1,3-bisnucleophiles, e.g., cyanacetamide (**7**) or pyrimidine-2,4,6-triamine (**14**).

For the preparation of PTX analogue **2**, we used the same route by refluxing 1,3-diketone **6** [11] with cyanacetamide (**7**) in ethanol in the presence of piperidine [12,13]. The resulting pyridone **8** was treated with dimethylformamide and thionylchloride to yield the 2-chloronicotinonitrile **9** [1] which was reacted with guanidine (**10**) to yield 7-methyl-PTX (**2**) [14] (Fig. 2).

Our synthesis of Nor-PTX (**3**) [15,16] started from the known malonate **11** [17], which was hydrolyzed to give the acid **12** [18]. **12** was then treated with dimethylformamide and phosphoroylchloride under mild conditions and further with *N,N*-dimethylammoniumperchlorate to yield the trimethinium perchlorate **13**, representing a reactive derivative of malondialdehyde [19]. **13** cyclocondensated in good yields with pyrimidine-2,4,6-triamine (**14**) giving Nor-PTX (**3**). Recently an alternative preparation of Nor-PTX (**3**) has been published by condensation **14** and bromomalondialdehyde, followed by protection of amino groups and Pd-mediated coupling with 2,5-dimethoxybenzyl zinc chloride and final deprotection of the pivaloylated aminogroups [16] (Fig. 3).

An alternative C₃-building block for the synthesis of C-5 unsubstituted PTX derivatives **3** and **4** is the acrylaldehyde derivative **18**. The synthesis of **18** started from 3-(2,5-dimethoxyphenyl)propanol (**15**) [20] with a Swern-Oxidation with oxalylchloride and dimethylsulfoxide at -60 °C [21,22], followed by treatment with *N,N*-dimethyl-

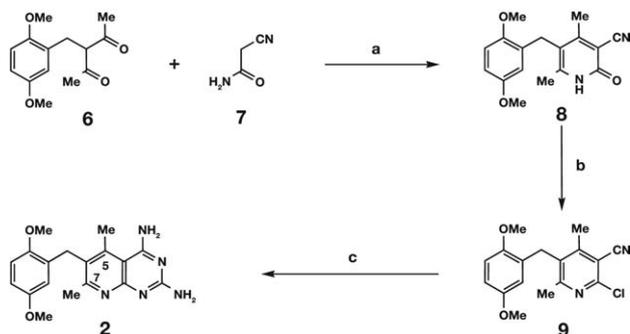


Fig. 2. Synthesis of 7-methyl-PTX reagents: (a) EtOH, piperidine, reflux; (b) SOCl₂, DMF, CH₂Cl₂, 80 °C; (c) pyridine, guanidine (10) reflux.

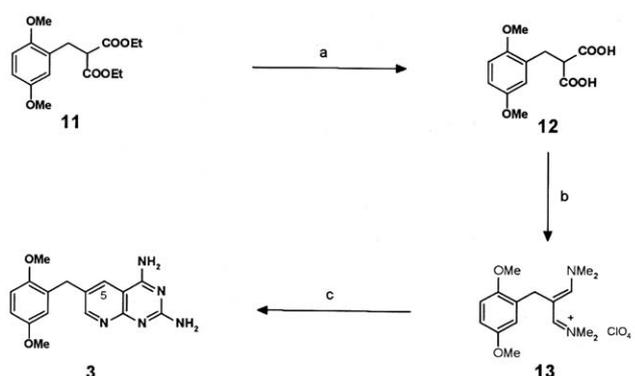


Fig. 3. Synthesis of Nor-PTX. Reagents: (a) NaOH 15% in H₂O:EtOH(1:1), 80 °C, HCl_{conc}; (b) dimethyl formamide, POCl₃, RT, Me₂NH₂ClO₄; (c) *n*-BuOH pyrimidine-2,4,6-triamine (**14**), reflux.

methylenimmoniumchloride (“Böhme-Salz”) (**16**) giving the Mannich base-hydrochloride **17** [23]. This Mannich base-HCl was unstable and eliminated dimethylamine and HCl giving the acrylaldehyde derivative **18** during purification. Reaction of **18** with pyrimidine-2,4,6-triamine (**14**) in a mixture of ethanol/acetic acid (1:1) at reflux yielded the dihydro compound **19**. Following oxidation with air in acetic acid under reflux gave Nor-PTX (**3**). Reduction of **19** in trifluoroacetic acid with hydrogen and Pd/C gave rise to 5,6,7,8-tetrahydro derivative **4** (Fig. 4).

The synthesis of PTX analogue **5** with an elongated side chain is similar to our Nor-PTX procedure. Alkylation of diethylmalonate (**21**) with 2-(2,5-dimethoxyphenyl)ethylbromide (**20**) gave the substituted malonate **22** [24], which was hydrolyzed to 2-(2,5-dimethoxyphenyl)ethylmalonic acid (**23**) [25,26]. Treatment with dimethylformamide, phosphoroylchloride and crystallisation with *N,N*-dimethyl ammonium per chlorate gave trimethinium salt **24** [27], which cyclocondensated with pyrimidine-2,4,6-triamine (**14**) to give Homo-Nor-PTX (**5**) (Fig. 5).

3. Pharmacology

The enzyme dihydrofolate reductase is responsible for the reduction of dihydrofolic acid (DHFA) to tetrahydrofolic

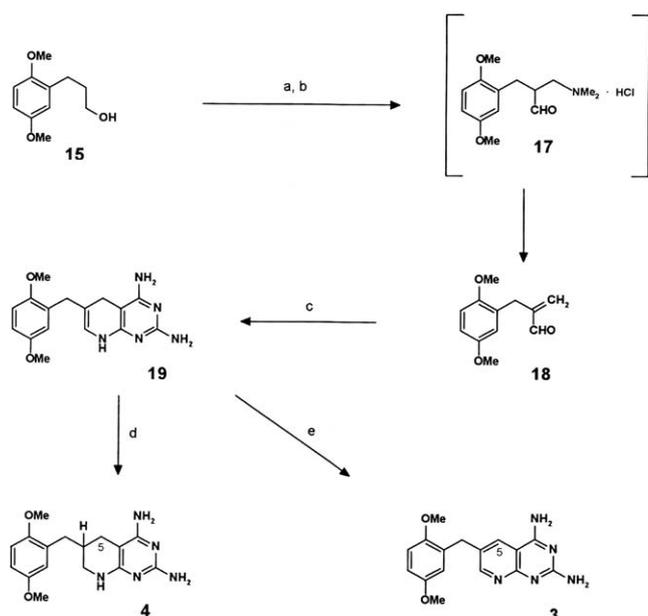


Fig. 4. Synthesis of Tetrahydro-Nor-PTX. Reagents: (a) oxalylchloride, dimethylsulfoxide, CH_2Cl_2 , -60°C ; (b) Böhme salt (16) CH_2Cl_2 , RT; (c) pyrimidine-2,4,6-triamine (14), EtOH, HOAc, reflux; (d) $\text{H}_2/\text{Pd-Ac}$ (10%), THFA, RT; (e) air, HOAc, reflux.

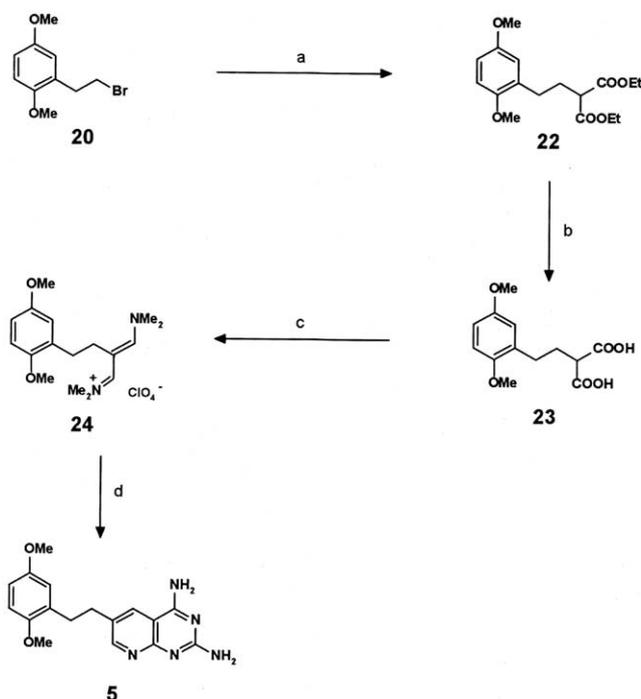


Fig. 5. Synthesis of PTX-analogue 5. Reagents: (a) diethylmalonate (21), NaH, THF; (b) 15% aqueous NaOH, 80°C ; HCl_{conc} . (c) dimethylformamide, POCl_3 , 30°C , $\text{Me}_2\text{NH}_2\text{ClO}_4$; (d) $n\text{-BuOH}$, pyrimidine-2,4,6-triamine (14), reflux.

acid (THFA), by using NADPH as cofactor. Tetrahydrofolate represents a C_1 -building block for the cellular biosynthesis of purines, pyrimidines or amino acids [8,28,29]. Especially for the synthesis of thymidilate, a permanent supply

of THFA out of DHFA is essential. Inhibition of dihydrofolate reductase results in lack of desoxythymidin-monophosphate and finally in a thymidinless cell-death [30,31].

Methotrexate, a classical folate antagonist, is well established in antitumor therapy [32]. Unfortunately the hydrophilic nature of the molecule limits transport into the cells of lung and CNS by diffusion [33]. Before methotrexate can act as an antitumor agent, an active transport mechanisms into the cell and polyglutamation by the enzyme folypolyglutamate synthase is necessary. A new class of dihydrofolate reductase inhibitors is represented by lipophilic molecules with a quinazoline or a pyrido[2,3-*d*]pyrimidine core, e.g., PTX (1). These lipophilic inhibitors do not need an active transport mechanism for diffusion into the cell and no activation as they lack a glutamate sidechain. Piritrexim represents a non-classical lipophilic inhibitor of dihydrofolate reductase of the second generation as it has no affinity to histamin-*N*-methyltransferase giving less toxic side effects [34].

The idea behind the lipophilic dihydrofolate reductase inhibitors is to overcome drug resistances. In phase II studies activity of PTX (1) against malignant melanomas, lung cancer, colon cancer, sarcoma and head and neck cancer were reported [35]. In addition, PTX (1) is active as an antipsoriasis agent and as a drug against opportunistic infections with *Pneumocystis carinii* and *Toxoplasma gondii* [33]. Secondary infections with these organisms are the main cause of death for AIDS patients.

4. Molecular modeling studies

To understand the obtained pharmacological data on a structural basis, we analysed the DHFR inhibitors MTX, PTX, and the newly synthesized and tested compounds 2–4 by molecular modeling and docking techniques. There are several crystal structures of DHFR in complex with MTX and one in complex with PTX available through the RCSB Protein Data Bank [36]. Structural comparison of the human DHFR mutant L22Y-MTX complex (Protein Data Bank entry 4DLS) and the human DHFR mutant L22F-PTX complex (Protein Data Bank entry 1DLR) by fitting the protein C_α carbon atoms revealed that both inhibitors share a common interaction pattern with the protein, despite of their very different size. The pyrido[2,3-*d*]pyrimidine moiety of PTX is able to form the same hydrogen bond binding pattern with the protein as the corresponding substructure of MTX. Moreover, the *p*-dimethoxyphenyl subunit of PTX arranges in the DHFR binding site almost parallel to the 1,4-disubstituted phenyl group of MTX.

To investigate the influence of PTX structure modifications on the DHFR binding behaviour, we used the molecular docking package QXP (Quick eXplore) to predict possible orientations of the various ligands within the DHFR binding site. The program allows the positioning of the fully flexible ligands within a partially flexible binding cleft by a Monte

Carlo procedure followed by energy minimization in combination with several scoring methods for binding affinity estimation [37]. To verify and check the docking procedure, we first applied the protocol to the already experimentally known MTX and PTX complexes named above. In both cases, QXP was able to predict the binding mode of the inhibitors within the DHFR active site correct (data not shown).

5. Results and discussion

Piritrexim analogues **2–4** were evaluated for antitumor activity in the NCI's in vitro disease-oriented antitumor screening against a panel of 60 human tumor cell lines derived from leukaemia, lung cancer, colon cancer, CNS cancer, melanoma, ovarian cancer, renal cancer, prostate cancer and breast cancer. The compounds were diluted to five concentrations, the highest being 10^{-4} mol/l, the others 10^{-5} , 10^{-6} , 10^{-7} and the lowest 10^{-8} mol/l. The growth inhibition power (GI_{50}) was evaluated and the results obtained are shown in Table 1.

The majority of tumors were inhibited by micromolar concentrations (10^{-6} mol/l) of the test compounds, in some cases even at concentrations of 10^{-7} or 10^{-8} mol/l. The 5,6,7,8-tetrahydro compound **4** showed the highest average activity against all the 60 cell lines with GI_{50} values between 0.024 and 89.1 $\mu\text{mol/l}$. It also was the most active compound against leukaemic cell lines with a GI_{50} concentration of 0.024 $\mu\text{mol/l}$ against leukaemia K-562, followed by a GI_{50} concentration of 0.087 $\mu\text{mol/l}$ against colon cancer HCT-116 and 0.10 $\mu\text{mol/l}$ against non-small lung cancer NHI-H460.

The GI_{50} concentrations of Nor-PTX (**3**) were between 0.135 and >100 $\mu\text{mol/l}$. It mainly had a activity against leukaemia cell lines (K-562, 0.135 $\mu\text{mol/l}$; HL-60(TB), 0.138 $\mu\text{mol/l}$; MOLT-4, 0.174 $\mu\text{mol/l}$; SR, 0.417 $\mu\text{mol/l}$; CCRF, 0.832 $\mu\text{mol/l}$). It also inhibited the growth of non-small lung cancer cells A549/ATCC ($GI_{50} = 0.295$ $\mu\text{mol/l}$) and NCI-H460 ($GI_{50} = 0.229$ $\mu\text{mol/l}$). Finally, the colon cancer cell lines HCT-116 ($GI_{50} = 0.209$ $\mu\text{mol/l}$) and SW-620 ($GI_{50} = 0.240$ $\mu\text{mol/l}$) were sensitive against compound **3**.

The 7-methyl-PTX (**2**) had GI_{50} concentrations between 0.214 and 15.448 $\mu\text{mol/l}$ in the antitumor test. **2** showed highest activity against leukaemia cell line K-562 with a GI_{50} concentration of 0.214 $\mu\text{mol/l}$ and against colon cancer HCT-116 ($GI_{50} = 0.389$ $\mu\text{mol/l}$) (Table 1).

In order to compare the binding affinity of the newly synthesized PTX analogues, we docked the compounds **1–4** into the empty binding site of the experimentally known human DHFR mutant IDLR. In these simulations, the ligand is fully flexible (free translation, rotation and rotation of functional groups about single bonds). Within the protein, the side chains Glu30 and Phe31, as well as the crystal water molecule 252, are also kept flexible to improve the interaction to the ligand. Fig. 6 shows as a representative example an

overlay of the best docking solutions with the highest predicted binding affinity for PTX and compound **2**. Both compounds share the same binding mode, which is also the same for **3** and **4** (not visualized in Fig. 6). For the estimation of the binding affinity, the non-bonded Coulomb and van der Waals interactions between protein and ligand, as well as the ligand energy relative to the estimated global minimum of the free ligand, and the binding site energy relative to the local minimum for the empty site were taken into account. All new synthesized compounds show the same binding pattern as PTX within the experimentally resolved complex IDLR. Moreover, the predicted binding affinities for the compounds **1–4** (**1**: -78.8 ; **2**: -48.3 ; **3**: -62.6 ; **4**: -60.1 ; all values in kJ mol^{-1}) are in good agreement with the experimentally determined activity data. Therefore, we are now able to rank new PTX homologues in terms of their expected affinity to the DHFR active site.

6. Conclusions

From literature and our own studies, we knew about the importance of the pyrimidine-2,4-diamine substructure for the inhibition of dihydrofolate reductase [8]. It was also reported that an increasing substituent in position 5 attenuates the potency of PTX derivatives [9]. In our studies, we focused on methyl-substituents in position 5 and 7 and hydrogenation of the pyridine ring of the molecule.

Test results showed that the activity of Iso-PTX, with a methyl group in position 7 instead of position 5 [6,7,15], is 20 times weaker than PTX (**1**). **2** with an additional methyl group in position 7 of PTX is seven times lower in activity than PTX. Nor-PTX (**3**) is five times, the tetrahydro derivative **4** four times less active than PTX in average. The shift of the methyl group from position 5–7 lowers activity and also an additional group in position 7 gives no improvement. The experimental findings are in good agreement with predicted binding affinities obtained by molecular docking studies, which allow an at least qualitative ranking of new, not yet synthesized and tested PTX homologues.

7. Experimental protocols

7.1. Chemistry

Melting points (m.p.) were determined on a BÜCHI, Typ 310, capillary apparatus and are uncorrected. Boiling points were measured on a Kugelrohr distillation apparatus from BÜCHI (Typ GRK 50). IR spectra were determined with a PERKIN–ELMER Typ 1740 infrared Fourier-transformation spectrophotometer in KBr or CHCl_3 . ^1H - and ^{13}C -NMR spectra were recorded in $\text{DMSO}-d_6$, if no other solvent is named (tetramethylsilane as internal standard), at 250.13 MHz (60.6 MHz) on a Bruker AC 250 or at 360.13 MHz (90.6 MHz) on a BRUKER AM 360. Mass

Table 1
Inhibition of in vitro tumor cell growth by PTX (1), methotrexate and PTX-derivatives 2–4

Panel/cell line	Cytotoxicity GI (50) in $\mu\text{mol/l}$	(^a) (^b) (^c) GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$
	1	2	3	4	Methotrexate
<i>Leukaemia</i>					
CCRF-CEM	0.040	1.175	0.832	0.537	0.025
HL-60(TB)	N.D.	0.708	0.138	0.195	0.026
K-562	0.010	0.214	0.135	0.024	0.025
MOLT-4	0.030	0.776	0.174	0.692	0.029
RPMI-8226	0.468	6.166	3.467	9.550	0.151
SR	0.030	0.537	0.417	0.776	0.025
<i>Non-small cell lung cancer</i>					
A549/ATCC	0.010	0.646	0.295	0.219	0.028
EKVX	21.878	13.183	2.188	14.454	7.586
HOP-18	N.D.	N.D.	N.D.	N.D.	7.079
HOP-62	0.219	6.166	1.950	2.512	0.034
HOP-92	16.982	15.488	1.738	N.D.	12.882
NCI-H226	4.898	13.183	13.804	89.125	58.884
NCI-H23	0.224	0.759	0.525	0.245	0.035
NCI-H322M	0.355	8.710	9.333	1.585	0.501
NCI-H460	N.D.	0.741	0.229	0.100	0.025
NCI-H522	N.D.	4.898	N.D.	N.D.	0.151
LXPL 529	N.D.	N.D.	N.D.	N.D.	0.048
<i>Small cell lung cancer</i>					
DMS 114	N.D.	N.D.	N.D.	N.D.	0.026
DMS 273	N.D.	N.D.	N.D.	N.D.	0.039
<i>Colon cancer</i>					
COLO 205	14.125	10.715	7.943	7.079	1.905
DLD-1	N.D.	N.D.	N.D.	N.D.	0.052
HCC-2998	0.063	12.882	N.D.	4.571	0.240
HCT-116	0.013	0.389	0.209	0.087	0.025
HCT-15	0.015	1.622	1.413	N.D.	0.054
HT29	0.029	0.891	0.417	0.603	0.046
KM12	0.427	2.239	2.399	0.550	0.036
KM20L2	N.D.	N.D.	N.D.	N.D.	0.089
SW-620	0.055	1.622	0.240	0.479	0.036
<i>CNS cancer</i>					
SF-268	0.145	6.547	0.933	0.178	0.032
SF-259	0.355	0.813	0.603	0.479	0.042
SF-539	1.660	N.D.	1.122	1.122	0.062
SNB-19	N.D.	2.399	14.791	1.445	0.257
SNB-75	6.761	3.311	27.542	13.183	25.704
SNB-78	N.D.	N.D.	N.D.	N.D.	251.189
U251	5.129	1.413	2.591	0.776	0.141
XF498	N.D.	N.D.	N.D.	N.D.	158.489
<i>Melanoma</i>					
LOX IMVI	0.021	0.562	0.407	0.457	0.025
MALME-3M	N.D.	5.888	12.303	13.804	5.012
M14	0.043	0.955	0.759	0.468	0.028
M19-MEL	N.D.	N.D.	N.D.	N.D.	0.708
SK-MEL-2	0.072	15.488	1.318	40.738	239.883
SK-MEL-28	1.122	10.965	14.454	11.482	4.074
SK-MEL-2	0.098	0.759	0.676	0.741	0.058
UACC257	15.488	12.023	1.995	3.890	0.871
UACC-62	0.072	0.550	0.589	0.661	0.027
<i>Ovarian cancer</i>					
IGROV1	0.013	0.977	0.631	0.851	0.071
OVCAR-3	0.050	10.471	10.000	15.136	0.363
OVCAR-4	16.982	10.471	13.804	16.218	251.189

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Table 1
(continued)

Panel/cell line	Cytotoxicity GI (50) in $\mu\text{mol/l}$	^(a) ^(b) ^(c) GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$	GI (50) in $\mu\text{mol/l}$
OVCAR-5	19.055	7.762	12.882	8.318	2.884
OVCAR-8	0.055	0.661	0.525	0.871	0.030
SK-OV-3	12.303	12.023	>100	30.903	128.825
<i>Renal cancer</i>					
786-0	0.263	0.646	0.589	0.427	0.027
A498	N.D.	10.715	2.399	18.197	2.399
ACHN	N.D.	2.455	1.072	1.202	0.046
CAKI-1	0.063	11.482	2.512	N.D.	0.030
RXF-393	2.239	15.136	1.585	13.183	32.359
RXF-631	N.D.	N.D.	N.D.	N.D.	0.059
SN12C	0.513	10.000	0.891	0.776	0.028
TK-10	12.023	11.220	11.220	5.754	93.325
UO-31	0.035	4.266	7.413	11.482	0.091
<i>Prostata cancer</i>					
PC-3	0.052	0.724	3.162	0.288	N.D.
DU-145	0.933	N.D.	2.344	3.162	N.D.
<i>Breast cancer</i>					
MCF7	12.023	3.981	1.820	2.344	N.D.
MCF/ADR-RES	4.467	9.333	63.093	9.120	N.D.
MDA-MB-231/ATCC	17.378	13.490	13.183	0.263	N.D.
HS578T	N.D.	12.023	19.055	8.318	N.D.
MDA-MB-435	0.204	0.794	0.617	0.398	N.D.
MDA-N	0.304	0.759	0.372	0.617	N.D.
BT-549	13.490	1.905	N.D.	10.715	N.D.
T-47D	15.136	N.D.	16.218	20.893	N.D.

^a Data obtained from NCI's in vitro disease-oriented tumor cells screen.

^b GI₅₀ is the molar concentration causing 50% growth-inhibition of tumor cells. Compounds with GI₅₀ > 100 $\mu\text{mol/l}$ are considered inactive.

^c N.D., not determined.

spectra were determined on a Finnigan TSQ 70 (electron impact, ionization energy 70 eV). Elemental analyses were measured on a Heraeus Typ CHN-Rapid. MPLC was performed with a Büchi pump Type B-688, Büchi glass columns and Macherey–Nagel silica gel 60, 0.04–0.063 mm, 230–400 mesh.

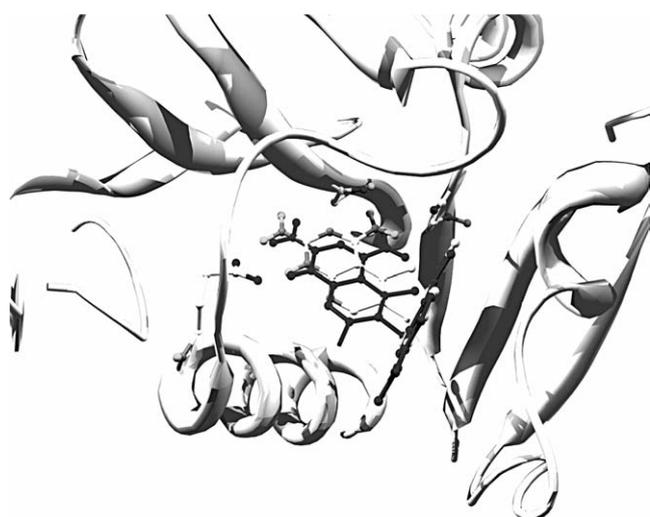


Fig. 6. Overlay of the best docking solutions of PTX (light grey) and the 7-methyl homologue **2** (dark grey) within the 1DLR-binding pocket. For clarity, the protein is visualized as a ribbon indicating the secondary structure elements without explicit sidechains.

7.1.1. 6-(2,5-Dimethoxybenzyl)-5,7-dimethylpyrido[2,3-d]-pyrimidine-2,4-diamine (**2**)

To a cold solution of sodium (506 mg, 22 mmol) in methanol (50 ml), guanidine hydrochloride (**10-HCl**) (1.84 g, 22 mmol) was added. The residue (NaCl) was separated and washed with methanol. The unified methanol phases were evaporated under reduced pressure. The resulting oil was diluted with pyridine (5 ml) and 2-chloro-5-(2,5-dimethoxybenzyl)-4,6-dimethylnicotinonitrile (**9**) (1040 mg, 3.3 mmol) was added. The mixture was refluxed for 5 h. Then water (25 ml) was added and the residue was isolated. Crystallisation from H₂O/dimethylformamide (20:1) provided 57% (630 mg) of a colourless powder; m.p. 310 °C (H₂O/DMF); IR: cm^{-1} 3339, 3167 (NH₂), 2999 (Ar-H), 1646 (C=N), 1575, 1545, 1498 (C=C); ¹H-NMR: δ 2.32 (3H, s, CH₃), 2.48 (3H, s, CH₃), 3.55 (3H, s, OCH₃), 3.82 (3H, s, OCH₃), 3.90 (2H, s, CH₂), 5.99 (1H, d, $J = 3$ Hz, 6'-H), 6.06 (2H, br, NH₂, D₂O exchangeable), 6.74 (1H, dd, $J_1 = 9$ Hz, $J_2 = 3$ Hz, 4'-H), 6.84 (2H, br, NH₂, D₂O exchangeable), 6.94 (1H, d, $J = 9$ Hz, 3'-H); ¹³C-NMR: δ 17.37 (5-CH₃), 23.80 (7-CH₃), 27.96 (CH₂), 55.06 (OCH₃), 55.74 (OCH₃), 104.02 (C-4a), 110.19, 111.13, 114.28, 128.37 (C-1', C-3', C-4', C-6'), 124.37 (C-6), 144.24 (C-5), 151.10 153.11 (C-2', C-5'), 160.54, 161.65, 162.52, 163.74 (C-2, C-4, C-7, C-8a); MS: 339 (100) [M⁺], 324 (22) [M⁺ -CH₃], 309 (58) [M⁺ -CH₃, -CH₃], 308 (40) [M⁺ -OCH₃], 293 (19) [M⁺ -CH₃,

-OCH₃]. Anal. calculated for C₁₈H₂₁N₅O₂ (339.40): C 63.7; H 6.24; N 20.6 (found: C 62.7; H 6.24; N 20.6).

7.1.2. 6-(2,5-Dimethoxybenzyl)pyrido[2,3-d]pyrimidine-2,4-diamine (**3**) (Nor-PTX)

Method 1: *N*-[(2*E*)-2-((2,5-dimethoxybenzyl)-3-dimethylamino)prop-2-en-1-ylidene]-*N*-methyl-methanaminium-perchlorate (**13**) (1.0 g, 2.7 mmol) was heated to reflux with pyrimidine-2,4,6-triamine (**14**) (0.33 g, 2.6 mmol) in *n*-butanol (10 ml) for 10 h. The precipitate was filtered off and crystallized from H₂O/dimethylformamide (20:1) to yield 75% (610 mg) **3** as a colourless powder.

Method 2: 6-(2,5-Dimethoxybenzyl)-5,8-dihydropyrido[2,3-*d*]pyrimidine-2,4-diamine (**19**) (500 mg, 1.6 mmol) was solved in acetic acid (30 ml) and heated to reflux for 10 h while bubbling air through the solution. The solution was diluted with water (10 ml) and neutralized with 5N aqueous sodium hydroxide. The precipitate was filtered off and crystallized from H₂O/dimethylformamide (20:1) to yield 60% (300 mg) **3** as a colourless powder; m.p. 310 °C (H₂O/MF); IR: cm⁻¹ 3429, 3354, 3157 (NH₂), 2996, 2937 (C-H), 2834 (OCH₃), 1675, 1651, (C=N), 1601, 1580, 1504 (C=C); ¹H-NMR: δ 3.69 (3H, s, OCH₃), 3.72 (3H, s, OCH₃), 3.91 (2H, s, CH₂), 6.75–6.92 (3H, m, 3'-H, 4'-H, 6'-H), 7.19 (2H, s, 4-NH₂, D₂O exchangeable), 8.28 (2H, s, 2-NH₂, D₂O exchangeable), 8.39 (1H, d, *J* = 2.5 Hz, 5-H), 8.59 (1H, d, *J* = 2.5 Hz, 7-H); ¹³C-NMR: δ 33.20 (CH₂), 55.90 (OCH₃), 56.37 (OCH₃), 105.02 (C-4a), 112.41, 112.54, 117.05, 129.86 (C-1', C-3', C-4', C-6'), 131.82, 133.75 (C-5, C-6), 151.61, 153.64 (C-2', C-5'), 154.26 (C-4), 156.50 (C-7), 159.62 (C-8a), 163.70 (C-2); MS: 311 (100) [M⁺], 296 (15) [M⁺ -CH₃], 280 (29) [M⁺ -(CH₃O)]. Anal. C₁₆H₁₇N₅O₂ (C,H,N).

7.1.3. 6-(2,5-Dimethoxybenzyl)-5,6,7,8-tetrahydropyrido[2,3-*d*]pyrimidin-2,4-diamine (**4**)

A mixture of 6-(2,5-dimethoxybenzyl)-5,8-dihydropyrido[2,3-*d*]pyrimidine-2,4-diamine (**19**) (0.1 g, 0.032 mol) trifluoroacetic acid (5 ml) and palladium/carbon (10%) (25 mg) was hydrogenated at 400 kPa. The catalyst was separated over celtite and the filtrate concentrated under reduced pressure. The resulting oil was diluted with water (2 ml) and neutralized with concentrated aqueous sodium hydroxide. The precipitate was filtered off and crystallized from H₂O/dimethylformamide (20:1) to yield 60% **4** (60 mg) of a colourless powder; m.p. 268–269 °C (H₂O/DMF); IR: cm⁻¹ 3447, 3341, 3181 (NH₂, NH), 2990, 2927 (C-H), 2834 (OCH₃), 1581 (C=N), 1500 (C=C); ¹H-NMR: δ 1.93–2.07 (2H, m, CH₂), 2.29–2.68 (3H, m, 5-H, 6-H), 2.72–3.05 (2H, m, 7-H), 3.69 (3H, s, OCH₃), 3.72 (3H, s, OCH₃), 5.10 (2H, s, NH₂, D₂O exchangeable), 5.38 (2H, s, NH₂, D₂O exchangeable), 5.91 (1H, s, NH, D₂O exchangeable), 6.70 (1H, d, *J* = 3 Hz, 6'-H), 6.74 (1H, dd, *J*₁ = 8.5 Hz, *J*₂ = 3 Hz, 4'-H), 6.88 (1H, d, *J* = 8.5 Hz, 3'-H); MS: 315 (28) [M⁺], 284 (100) [M⁺ -OCH₃]. Anal. C₁₆H₂₁N₅O₂ (C,H,N).

7.1.4. 6-[2-(2,5-Dimethoxyphenyl)ethyl]pyrido[2,3-*d*]pyrimidine-2,4-diamine (**5**)

N-[(2*E*)-2-[2-(2,5-Dimethoxyphenyl)ethyl]-3-(dimethylamino)prop-2-en-1-ylidene]-*N*-methylmethanaminium-perchlorate (**24**) (0.8 g, 2.1 mmol) and pyrimidine-2,4,6-triamine (**14**) (0.24 g, 1.9 mmol) in *n*-butanol (10 ml) were heated to reflux for 10 h. The precipitate was filtered off and crystallized from H₂O/dimethylformamide (20:1) to yield 80% **5** (0.5 g) of a colourless powder; m.p. 303–305 °C (H₂O/DMF); IR: cm⁻¹ 3460, 3344, 3352 (NH₂), 2992, 2937 (C-H), 2834 (OCH₃), 1695, 1654, 1619 (C=N), 1598, 1580, 1553, 1500 (C=C); ¹H-NMR: δ 2.89 (4H, s, 1'-H, 2'-H), 3.66 (3H, s, OCH₃), 3.71 (3H, s, OCH₃), 6.70–6.90 (3H, m, 3''-H, 4''-H, 6''-H), 7.11 (2H, br, 4-NH₂, D₂O exchangeable), 8.24 (2H, br, 2-NH₂, D₂O exchangeable), 8.40 (1H, d, *J* = 2.5 Hz, 5-H), 8.53 (1H, d, *J* = 2.5 Hz, 7-H); ¹³C-NMR: δ 31.57 (CH₂CH₂), 32.52 (CH₂CH₂), 55.93 (OCH₃), 56.42 (OCH₃), 104.96 (C-4a), 112.18, 112.43, 116.82 (C-3', C-4', C-6'), 130.34 (C-1'), 132.73, 133.40 (C-5, C-6), 151.90, 153.52 (C-2', C-5'), 154.24, 156.50, 159.63, 163.81 (C-2, C-4, C-7, C-8a); MS: 325 (6) [M⁺], 310 (1) [M⁺ -CH₃], 174 (100) [M⁺ -(CH₃O)₂ C₆H₂CH₂]. Anal. C₁₇H₁₉N₅O₂ (C, H, N).

7.1.5. 5-(2,5-Dimethoxybenzyl)-4,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile (**8**)

3-(2,5-Dimethoxybenzyl)pentan-2,4-dione (**6**) (1.6 g, 6.4 mmol) was heated to reflux with cyanacetamide (**7**) (530 mg, 6.3 mmol) in ethanol (5 ml) with 10 drops of piperidine for 5 h. After cooling, the residue was filtered off and crystallized from ethanol and gave (**8**) with 58% (1.11 g) yield as colourless crystals, m.p. 151 °C (ethanol); IR: cm⁻¹ 3311, 3150 (NH), 2998, 2937 (C-H), 2842 (OCH₃), 2224 (CN), 1652 (C=N); ¹H-NMR: δ 2.19 (3H, s, CH₃), 2.20 (3H, s, CH₃), 3.62 (3H, s, OCH₃), 3.67 (2H, s, CH₂), 3.77 (3H, s, OCH₃), 6.22 (1H, d, *J* = 3 Hz, 6'-H), 6.76 (1H, dd, *J*₁ = 9 Hz, *J*₂ = 3 Hz, 4'-H), 6.93 (1H, d, *J* = 9 Hz, 3'-H), 12.33 (1H, br, N-H, D₂O exchangeable); MS: 297 (76) [M⁺], 42 (100). Anal. C₁₇H₁₈N₂O₃ (C, H, N).

7.1.6. 2-Chloro-5-(2,5-dimethoxybenzyl)-4,6-dimethylnicotinonitrile (**9**)

To a solution of *N,N*-dimethylformamide (7.3 ml, 0.095 mol) in absolute chloroform (50 ml), cooled in an ice bath, thionylchloride (11.9 g, 0.1 mol) in absolute chloroform (20 ml) was added drop wise at a maximum temperature of 5 °C. At the end of the exothermic reaction, 5-(2,5-dimethoxybenzyl)-4,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile (**8**) (3.0 g, 0.01 mol) was added within 10–15 min in small portions. The mixture was allowed to warm at room temperature and heated to reflux for 3 h. After cooling, an ethanolic solution of potassium hydroxide was added slowly, then the mixture was diluted with water and the organic layer was separated. The aqueous phase was extracted with chloroform for three times. The unified organic phases were washed three times with water (10 ml), dried over Na₂SO₄ and concentrated under reduced pressure. The

residue was purified by MPLC (cyclohexane/acetic acid (8:2)) and following crystallization from ethanol to yield 79% (2.6 g) of **9** as colourless crystals, m.p. 121–122 °C (ethanol); IR: cm^{-1} 3435 (NH), 3076, 3027 (Ar-H), 2997, 2950 (C-H), 2834 (OCH₃), 2230 (CN), 1721 (C=N), 1611, 1592, 1563, 1545, 1499 (C=C); ¹H-NMR: δ 2.40 (3H, s, CH₃), 2.42 (3H, s, CH₃), 3.60 (3H, s, OCH₃), 3.78 (3H, s, OCH₃), 3.96 (2H, s, CH₂), 6.14 (1H, d, $J = 3$ Hz, 6'-H), 6.79 (1H, dd, $J_1 = 9$ Hz, $J_2 = 3$ Hz, 4'-H), 6.95 (1H, d, $J = 9$ Hz, 3'-H); MS: 316/318 (100/33) [M⁺], 301/303 (25/8) [M⁺-CH₃], 285/287 (25/9) [M⁺-OCH₃]. Anal. C₁₇H₁₇N₂O₂Cl (C, H, N).

7.1.7. 2,5-Dimethoxybenzylmalonic acid (**12**)

Diethyl 2,5-dimethoxybenzylmalonate (**11**) (3.9 g, 0.019 mol) and 15 g NaOH in 100 ml H₂O/EtOH (1:1) were heated to 80 °C for 2 h. After cooling, water (100 ml) was added; the solution was acidified with HCl_{conc} and extracted with ether (20 ml) for three times. The organic layer was washed with water (20 ml) three times, dried with Na₂SO₄, concentrated under reduced pressure and crystallized from toluol to yield 43% (1.37 g) of colourless crystals, m.p. 155–157 °C (toluol), IR: cm^{-1} 3200–2500 (COOH), 3060 (Ar-H), 2956, 2911 (C-H), 2836 (OCH₃), 1718, 1713 (C=O), 1506 (C=C) 1293 (O-H); ¹H-NMR: δ 2.96 (2H, d, $J = 7.5$ Hz, CH₂), 3.55 (1H, t, $J = 7.5$ Hz, H-2), 3.66 (3H, s, OCH₃), 3.73 (3H, s, OCH₃), 6.69 (1H, d, $J = 3$ Hz, 6'-H), 6.75 (1H dd, $J = 9$ Hz, $J = 3$ Hz, 4'-H) 6.86 (1H, d, $J = 9$ Hz, 3'-H), 12.64 (2H, br, OH, D₂O exchangeable); ¹³C-NMR: δ 29.76 (CH₂), 55.66 (OCH₃), 56.20 (OCH₃), 111.96, 112.15, 117.08, 127.49 (C-1', C-3', C-4', C-6'), 151.77 (C-5'), 153.14 (C-2'), 170.69 (COOH); MS: 254 (34) [M⁺], 210 (100) [M⁺-CO₂]; Anal C₁₂H₁₄O₆ (C, H).

7.1.8. N-[(2E)-2-(2,5-dimethoxybenzyl)-3-dimethylamino]prop-2-en-1-ylidene]-N-methyl-methanum-perchlorate (**13**)

N,N-Dimethylformamide (2.1 g, 0.029 mol) was stirred and cooled in an ice bath. Phosphorochloride (2.09 g, 0.014 mol) was added drop wise. To this mixture, 2,5-dimethoxybenzyl-malonic acid (**12**) (2.55 g, 0.01 mol) was added in small portions and stirred for 24 h at room temperature, then 30 min under reduced pressure. After cooling in an ice bath, ethanol (4 ml) and a solution of *N,N*-dimethyl ammoniumperchlorate (1.45 g, 0.01 mol) in ethanol (4 ml) were added. The precipitate was separated and crystallized from ethanol to yield **13**, 32% (1.2 g) of colourless crystals, m.p. 154 °C (ethanol); IR: cm^{-1} 3018 (Ar-H), 2970, 2943 (C-H), 2841 (OCH₃), 1597 (C=N), 1495 (C=C); ¹H-NMR: δ 3.01 (6H, br, NCH₃), 3.23 (6H, br, NCH₃), 3.70 (3H, s, OCH₃), 3.72 (2H, s, CH₂), 3.78 (3H, s, OCH₃), 6.52 (1H, d, $J = 2.2$ Hz, 6'-H), 6.84 (1H, dd, $J_1 = 6.0$ Hz, $J_2 = 2.2$ Hz, 4'-H), 6.97 (1H, d, $J = 6.0$ Hz, 3'-H), 7.67 (2H, s, 1-H, 3-H); ¹³C-NMR: δ 24.63 (CH₂), 48.66 (CH₃), 55.24 (OCH₃), 55.74 (OCH₃), 98.36 (C-2), 110.90, 111.44, 115.98, 129.65 (C1', C-3', C-4', C-6'), 150.21, 153.40 (C-2', C-5'), 166.40 (C-1,

C-3); MS: 277 (8) [M⁺-ClO₄⁻], 232 (16) [M⁺-ClO₄⁻-(CH₃)₂NH], 188 (100). Anal. C₁₆H₂₅ClN₂O₆ (C, H, N).

7.1.9. 2-(2,5-Dimethoxybenzyl)acrylaldehyde (**18**)

To a stirred solution of oxalylchloride (3.1 g, 0.024 mol) in 50 ml dry methylene chloride, that was cooled to -70 °C, dimethylsulfoxide (3.72 ml, 0.048 mol) in 20 ml dry methylene chloride was added drop wise. After 10 min, 3-(2,5-dimethoxy)phenylpropanol (**15**) (4.28 g, 0.022 mol) was added in small portions at -60 °C within 5 min. The solution was stirred for further 15 min. Then triethylamine (12.4 ml, 0.090 mol) was added cautiously. After the solution had warmed at room temperature *N,N*-dimethyl(methylene)-immoniumchloride ("Böhme-Salz") (**16**) (4.12 g, 0.044 mol) was added and the solution was stirred for 15 h. After the addition of methylenechloride (30 ml), the organic layer was washed three times with a saturated aqueous solutions of NaHCO₃ and NaCl, then washed with water (20 ml), dried with Na₂SO₄ and concentrated under reduced pressure giving 2.83 g of a yellow oil (**18**) in 63% yield, that was purified by vacuum distillation, b.p. 190–210 °C (0.4 kPa), IR: cm^{-1} 3017 (Ar-H), 2955, 2911 (C-H), 2836 (OCH₃), 1693 (CHO), 1503 (C=C), ¹H-NMR: (CDCl₃) δ 3.53 (2H, dd, $J_1 = 0.5$ Hz, $J_2 = 0.5$ Hz, CH₂), 3.75 (6H, s, OCH₃), 6.01 (1H, dt, $J_1 = 1$ Hz, $J_2 = 0.5$ Hz, 3-H), 6.03 (1H, dt, $J_1 = 1$ Hz, $J_2 = 0.5$ Hz, 3-H), 6.69–6.82 (3H, m, 3'-H, 4'-H, 6'-H), 9.61 (1H, s, CHO); ¹³C-NMR: δ 32.67 (CH₂), 55.22 (OCH₃), 55.68 (OCH₃), 147.93 (C-2), 135.71 (C-3), 111.67, 111.72, 116.51, 127.25 (C-1', C-3', C-4', C-6'), 151.21 (C-2'), 152.96 (C-5'), 194.68 (C-1); MS: 206 (90) [M⁺], 175 (100) [M⁺-OCH₃]. Anal. C₁₂H₁₄O₃ (C, H).

7.1.10. 5,8-Dihydro-6-(2,5-dimethoxybenzyl)pyrido[2,3-d]pyrimidine-2,4-diamine (**19**)

2-(2,5-Dimethoxybenzyl)acrylaldehyde (**18**) (911 mg, 4.4 mmol) was heated under reflux with pyrimidine-2,4,6-triamine (**14**) (428 mg, 3.4 mmol) in a mixture of ethanol (5 ml) and acetic acid (5 ml) for 5 h. After cooling, the residue was filtered off and gave **19** as a colourless powder, yield 30% (320 mg), m.p. 262–263 °C; IR: cm^{-1} 3322, 3161 (NH₂, NH), 3058 (Ar-H), 2944 (C-H), 2833 (OCH₃), 1700, 1658 (CHO), 1584, 1528, 1498 (C=C); ¹H-NMR: δ 2.90 (2H, s, CH₂), 3.09 (2H, s, 5-H), 3.68 (3H, s, OCH₃), 3.72 (3H, s, OCH₃), 5.33 (2H, s, NH₂, D₂O exchangeable), 5.61 (2H, s, NH₂, D₂O exchangeable), 5.67 (1H, d, $J = 5$ Hz, 7-H), 6.73 (1H, d, $J = 3.5$ Hz, 6'-H), 6.74 (1H, dd, $J_1 = 9$ Hz, $J_2 = 3.5$ Hz, 4'-H), 6.88 (1H, d, $J = 9$ Hz, 3'-H), 7.51 (1H, d, $J = 5$ Hz, 8-NH, D₂O exchangeable); MS: 313 (59) [M⁺], 312 (59) [M⁺-1], 298 (5) [M⁺-CH₃], 282 (76) [M⁺-OCH₃], 162 (100). Anal. C₁₆H₁₉N₅O₂ (C, H, N).

7.1.11. Diethyl [2-(2,5-dimethoxyphenyl)ethyl]malonate (**22**)

Diethylmalonate (**21**) (2.72 g, 0.017 mol) was dissolved in dry tetrahydrofuran (25 ml). To this solution, a 60% dispersion of sodium hydride in petroleum (0.72 g, 0.018 mol)

and 2-(2,5-dimethoxyphenyl)ethylbromide (**20**) (4.13 g, 0.017 mol) were added at room temperature. Then the mixture was heated to reflux for 12 h. After the addition of water (20 ml), the aqueous phase was extracted with ether (30 ml) for three times. The unified ether fractions were washed with water (20 ml) three times, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was purified by MPLC (cyclohexane/acetic acid (8:2)) to yield 3.3 g (60%) of a colourless oil (**22**), b.p. 230–250 °C (0.2–0.4 kPa); IR: cm⁻¹ 3029 (Ar–H), 2986, 2941, 2909 (C–H), 2874, 2836 (OCH₃), 1746, 1729 (COOEt), 1610, 1591, 1504, 1500 (C=C); ¹H-NMR: δ 1.18 (6H, t, *J* = 7 Hz, CH₃), 2.01 (2H, m, CH₂), 2.53 (2H, m, CH₂), 3.35 (1H, t, *J* = 7.5 Hz, CH), 3.68 (3H, s, OCH₃), 3.70 (3H, s, OCH₃), 4.11 (4H, q, *J* = 7 Hz, OCH₂), 6.69 (1H, d, *J* = 3 Hz, 6'-H), 6.74 (1H, dd, *J*₁ = 9 Hz, *J*₂ = 3 Hz, 4'-H), 6.87 (1H, d, *J* = 9 Hz, 3'-H); ¹³C-NMR: δ 13.73 (OCH₂CH₃), 27.12 (CH₂CH₂), 28.45 (CH₂CH₂), 50.60 (C-2), 50.81 (OCH₂), 55.20 (OCH₃), 55.66 (OCH₃), 111.49, 111.60, 115.92 (C-3', C-4', C-6'), 129.52 (C-1'), 151.15, 152.96 (C-2', C-5'), 168.76 (COOR); MS: 324 (4) [M⁺], 164 (100). Anal. C₁₇H₂₄O₆ (C, H).

7.1.12. [2-(2,5-Dimethoxyphenyl)ethyl]malonic acid (**23**)

Diethyl [2-(2,5-dimethoxyphenyl)ethyl]malonate (**22**) (3.24 g, 0.010 mol) was added to a solution of 50 ml 15% aqueous sodium hydroxide and heated to 80 °C for several hours. After addition of conc. hydrochloric acid to pH 4–5 and extraction with 50 ml ethylacetate for three times, the unified organic phases were dried over Na₂SO₄ and concentrated under reduced pressure. The resulting residue was crystallized from toluol yielding 85% of **23** (2.27 g) as colourless crystals, m.p. 111–112 °C (toluol); IR: cm⁻¹ 3079, 3014 (Ar–H), 2950, 2927 (C–H), 2874, 2836 (OCH₃), 1708 (COOH), 1607, 1507 (C=C); ¹H-NMR: δ 1.90–2.02 (2H, m, CH₂), 2.49–2.58 (2H, m, CH₂), 3.14 (1H, t, *J* = 7.5 Hz, CH), 3.68 (3H, s, OCH₃), 3.71 (3H, s, OCH₃), 6.68 (1H, d, *J* = 3 Hz, 6'-H), 6.74 (1H, dd, *J*₁ = 9 Hz, *J*₂ = 3 Hz, 4'-H), 6.87 (1H, d, *J* = 9 Hz, 3'-H), 10.50 (2H, br, COOH); MS: 268 (7) [M⁺], 224 (66) [M⁺ - CO₂], 152 (100). Anal. C₁₃H₁₆O₆ (C, H).

7.1.13. N-[(2E)-2-[2-(2,5-Dimethoxyphenyl)ethyl]-3-(dimethylamino)prop-2-en-1-ylidene]-N-methylmethanaminium-perchlorate (**24**)

N,N-dimethylformamide (1.4 g, 19.2 mmol) was stirred and cooled in an ice bath. Phosphoroxochloride (1.39 g, 9.1 mmol) was added drop wise. This mixture was stirred for 30 min at room temperature. Then [2-(2,5-dimethoxyphenyl)ethyl]malonic acid (**23**) (1.0 g, 3.7 mmol) was added in small portions and stirred for 12 h at 30 °C; finally 30 min under reduced pressure. After cooling on ice, ethanol (8 ml) and a solution of *N,N*-dimethylammoniumperchlorate (725 mg, 5.0 mmol) in ethanol (4 ml) were added. The precipitate was separated and crystallized from ethanol to yield **24**, 69% (1.0 g) of colourless crystals, m.p. 161–162 °C (ethanol); IR: cm⁻¹ 3001 (Ar–H), 2931 (C–H), 2836 (OCH), 1588 (C=N), 1505 (C=C); ¹H-NMR: δ 2.59–2.71 (4H, m,

CH₂), 3.32 (12H, s, NCH₃), 3.69 (3H, s, OCH), 3.72 (3H, s, OCH₃), 6.74–6.93 (3H, m, 3'-H, 4'-H, 6'-H), 7.44 (2H, s, 1-H, 3-H); ¹³C-NMR: δ 33.03 (CH₂CH₂), 33.34 (CH₂CH₂), 48.66 (NCH₃), 55.29 (OCH₃), 55.51 (OCH₃), 102.97 (C-2), 111.39, 111.47, 116.34 (C-3', C-4', C-6'), 129.25 (C-1'), 151.02, 153.07 (C-2', C-5'), 165.50 (C-1, C-3); MS: 291 (4) [M⁺ - ClO₄⁻], 246 (7) [M⁺ - ClO₄⁻ - (CH₃)₂NH], 202 (100). Anal. C₁₇H₂₇ClN₂O₆ (C, H, N).

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References

- [1] E.M. Grivsky, S. Lee, C.W. Sigel, D.S. Duch, C.A. Nichol, J. Med. Chem. 23 (1980) 327–329.
- [2] E.M. Berman, L.M. Werbel, J. Med. Chem. 34 (1991) 479–485.
- [3] D.S. Duch, M.P. Edelstein, C.A. Nichol, Mol. Pharmacol. 18 (1980) 100–104.
- [4] D.S. Duch, M.P. Edelstein, S.W. Bowers, C.A. Nichol, Cancer Res. 42 (1982) 3987–3994.
- [5] R. Troschütz, T. Dennstedt, Arch. Pharm. (Weinheim) 327 (1994) 221–224.
- [6] R. Troschütz, M. Zink, T. Dennstedt, Arch. Pharm. (Weinheim) 328 (1995) 535–540.
- [7] R. Troschütz, M. Zink, R. Gnihl, J. Heterocyclic Chem. 36 (1999) 703–706.
- [8] J.M. Blaney, C. Hansch, C. Slipio, A. Vittoria, Chem. Rev. 84 (1984) 333–407.
- [9] B.S. Hurlbert, R. Ferone, T.A. Herrmann, G.H. Hitchings, M. Barnett, S.R.M. Bushby, J. Med. Chem. 11 (1968) 711–717.
- [10] T.-L. Su, Y.-K. Yang, J.-T. Huang, W.-Y. Ren, K.A. Watanabe, T.-C. Chou, J. Heterocyclic Chem. 30 (1993) 1437–1443.
- [11] C.M. Wong, D. Popien, R. Schwenk, J. TeRaa, Can. J. Chem. 49 (1971) 2712–2718.
- [12] J.B. Paine III, J. Heterocyclic Chem. 24 (1987) 351–355.
- [13] A.H. Tracy, R.C. Elderfield, J. Org. Chem. 6 (1941) 63–69.
- [14] F. Pochat, F. Lavelle, C. Fizames, A. Zerial, Eur. J. Med. Chem. 22 (1987) 135–137.
- [15] A. Gangjee, U.S. Patent (1996) 5,346,900.
- [16] A. Rosowsky, A. Chen, H. Fu, S.F. Queener, Biorg. Med. Chem. 11 (2003) 59–67.
- [17] G. Wegner, T.F. Keyes, N. Nakabayashi, H.G. Cassidy, J. Org. Chem. 34 (1969) 2822–2826.
- [18] R. Baltzly, J.S. Buck, J. Am. Chem. Soc. 62 (1940) 161–164.
- [19] J. Kučera, Z. Arnold, Collect. Czech. Chem. Commun. 32 (1967) 3792–3793.
- [20] R.A. Glennon, S.M. Liebowith, E.C. Mack, J. Med. Chem. 21 (1978) 822–825.
- [21] Houben- Weyl, in: Methoden der Organischen Chemie, J. Falbe (Hrsg.) Band E3, G. Thieme Verlag, Stuttgart, 1983, pp. 279–281.
- [22] A.J. Mancuso, S.L. Huang, D. Swern, J. Org. Chem. 43 (12) (1978) 2480–2482.
- [23] S. Takano, K. Inomata, K. Samizu, S. Tomita, M. Yanase, M. Suzuki, et al., Chem. Lett. 7 (1989) 1283–1284.
- [24] D.J. Abraham, P.E. Kennedy, A.S. Mehanna, D.C. Patwa, F.L. Williams, J. Med. Chem. 27 (1984) 967–978.
- [25] M. Conrad, Liebigs, Ann. Chem. 204 (1880) 174–176.

- [26] H. Brederick, F. Effenberger, G. Simchen, *Chem. Ber.* 96 (1963) 1350–1355.
- [27] C. Jutz, E. Schweiger, *Chem. Ber.* 107 (1974) 2383–2396.
- [28] W. Friedrich, in: *Handbuch der Vitamine*, Urban und Schwarzenberg Verlag, München, 1987, pp. 398–483.
- [29] P. Karlson, *Kurzes Lehrbuch der Biochemie für Mediziner und Naturwissenschaftler*, G. Thieme Verlag, Stuttgart, New York, 1988.
- [30] L. Stryer, *Biochemie*, Spektrum Akad, Verlag, Heidelberg, Berlin, New York, 1991 p. 640.
- [31] R.B. Silverman, *Medizinische Chemie für Organiker, Biochemiker und pharmazeutische Chemiker*, Verlag Chemie, Weinheim, 1994 p. 211.
- [32] J.R. Piper, G.S. McCaleb, J.A. Montgomery, R.L. Kisliuk, Y. Gaumont, F.M. Sirotnak, et al., *Chem.* 29 (1986) 1080–1087.
- [33] L.F. Kuyper, D.P. Baccanari, M.L. Jones, R.N. Hunter, R.L. Tansik, S.S. Joyner, et al., *J. Med. Chem.* 39 (1996) 892–903.
- [34] E.M. Berman, L.M. Werbel, *J. Med. Chem.* 34 (1991) 479–485.
- [35] E.G.E. de Vries, J.A. Gietma, P. Workman, J.E. Scott, A. Crawshaw, H.J. Dobbs, et al., *Br. J. Cancer* 68 (1993) 661.
- [36] H.M. Berman, J. Westbrook, Z. Feng, G. Gilliland, T.N. Bhat, H. Weissig, et al., *Nucl. Acids Res.* 28 (2000) 235–242. Available from <http://www.rcsb.org/pdf>.
- [37] C. McMartin, R.S. Bohacek, *J. Comp. Aided Mol. Des* 11 (1997) 333–344.