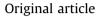
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Synthesis of 2-aziridinyl phosphonates by modified Gabriel-Cromwell reaction and their antibacterial activities

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ABSTRACT

A set of new aziridinyl phosphonates (4a-g) were synthesized by using the Gabriel–Cromwell reaction and its modified version developed in this study and their structures confirmed by HRMS, IR, and NMR spectra. All the compounds were screened for their antibacterial activity. They all showed comparable moderate to good growth inhibitory activity in reference to ampicillin and streptomycin.

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1. Introduction

Aziridines are important small molecules that are found in the structure of biologically active natural compounds and have many applications in organic chemistry [1]. They are used as precursors to amino acids, azomethine ylides, and amino alcohols. In addition they are used as monomers for polymerization [2], chiral auxiliaries [3], and chiral ligands [4]. Aziridinyl phosphonates are the precursors of α -amino phosphonates that find applications such as enzyme inhibitors [5], as haptens for catalytic antibodies [6], as antibacterial agents [7], and herbicides [8]. Although many studies have been carried out for the synthesis of aziridine-2-carboxylate esters, there are only limited number of studies reported for the synthesis of aziridinyl phosphonates [9]. One of the methods reported by Kim et al. involves the nitrene addition to cinnamoyl phosphonate derivatives which produced N-tosyl-2-aziridinyl phosphonates in 81–89% yield [10]. Another synthesis reported by Stevens et al. gives rather specific N-vinylaziridinyl phosphonate derivatives in 29–57% yield [11]. Davis et al. reported the synthesis of chiral aziridinyl phosphonates in 76% yield by reacting chiral sulfinimines with chloromethyl phosphonates [12]. Loreto et al. also studied synthesis of aziridinyl phosphonates by (ethoxycarbonyl)nitrene addition to α,β -unsaturated phosphonates which yielded aziridines in 14-45% [13]. In the study of Lesniak et al. 1,3dipolar cycloaddition of diazophosphonate with aryl imines produced aziridinyl phosphonates in 67-78% yield after a period of 15 days [14]. Very recently, Hodgson et al. reported aziridinyl phosphonate synthesis by lithiation-induced phosphonyl migration from nitrogen to carbon in terminal aziridines [15]. Due to the limited number of studies and also the limitations in the application of some of the methods such as low yields, long reaction times, or specific aziridine synthesis there is a need for the development of more general methods for the synthesis of aziridinyl phosphonates. Our group has been involved in the synthesis of new aziridine derivatives to use them as chiral ligands for metal catalyzed enantioselective synthesis of organic compounds [16]. As an extention of previous studies, here in we report the synthesis of new aziridinyl phosphonates by modified Gabriel-Cromwell reaction and also their antibacterial activities.

2. Results and discussion

2.1. Chemistry

One of the commonly used methods for the aziridine synthesis is the Gabriel-Cromwell reaction Scheme 1. A nice asymmetric application of this method was reported by Garner et al. [17]. Berlin et al. applied this method for the synthesis of diethyl 2-aziridinyl phosphonate ($\mathbf{4}, \mathbf{R} = \mathbf{H}$) which was obtained in 62% yield by heating dibromo compound 2 with ammonia in a sealed tube Scheme 1 [18]. This was the only aziridine reported by this group using the

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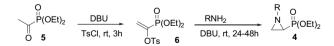
$$\overset{O}{\underset{P(OEt)_2}{\longrightarrow}} \overset{Br}{\underset{P(OEt)_2}{\longrightarrow}} \overset{Br}{\underset{P(OEt)_2}{\longrightarrow}} \overset{O}{\underset{P(OEt)_2}{\xrightarrow}} \overset{RNH_2}{\underset{(HBr)}{\xrightarrow}} \overset{O}{\underset{P(OEt)_2}{\longrightarrow}} \overset{RNH_2}{\underset{P(OEt)_2}{\longrightarrow}} \overset{R}{\underset{P(OEt)_2}{\longrightarrow}} \overset{R}{\underset{P(OEt)_2}{\longrightarrow}}$$

Scheme 1. Berlin's synthesis of aziridinyl phosphonate by Gabriel-Cromwell reaction.

Gabriel–Cromwell reaction. The steps of this reaction involves addition of Br₂ to vinyl group to form compound **2**. Then HBr elimination by triethylamine or the amine used for aziridination gives α -bromo phosphonate **3**. Michael addition of another equivalent of amine to **3** then S_N2 displacement of bromide leads to the aziridine **4**.

However, the vinyl phosphonate **1** is difficult to synthesize [19] and highly expensive. As an alternative to this method we planned to start with acetyl phosphonate which can be synthesized easily using the Michaelis–Arbusow reaction [20]. After the synthesis of acetyl phosphonate **5**, the next step was the conversion of this compound to tosylate **6** Scheme 2. Since it was necessary to have a good leaving group at α -position of the phosphonate. This conversion was made possible by treatment of acetyl phosphonate with DBU and tosyl chloride.

In order to synthesize compound 6 in high yield, it was necessary to optimize the reaction conditions. For this reason, different solvents (THF, DCM, CH₃CN, MeOH, and CHCl₃) were tried. Among the solvents, CHCl₃ and DCM gave similar results, and tosylate **6** was obtained in about 50% yield. Product formation was not observed in methanol and the highest yield (87%) was obtained with acetonitrile. In the case of THF, yield was 3-5% lower compared to acetonitrile. We have tried two bases DBU and Et₃N but almost no product formation took place in Et₃N. Therefore, DBU was chosen as the base. After determining solvent and base, different reaction times were tried. Increasing the reaction time from 3 h to 24 h didn't change the yield considerably. Change of reaction concentration had a significant effect on the yield. The highest yield was obtained when the concentration was about 0.2 M. Reactions carried out at 0.5 M and 1.0 M gave the product in low yield. We have also tried to synthesize mesvlate instead of tosvlate but the vield was low. After determining the optimum conditions for the synthesis of tosylate **6**, the aziridine forming step was carried out Scheme 2. Again, after doing some optimization studies by using different solvents (THF, DCM, CHCl₃, and CH₃CN), different amine concentrations and reaction times, it was found that the highest yield could be obtained by stirring tosylate **6** with amine (1.4 equiv) and DBU (1.0 equiv) at rt for 24 h in CH₃CN. After optimizing the reaction conditions, different amines were used to test the applicability of this method. The same aziridines were also synthesized by the Gabriel–Cromwell reaction (Method B, Scheme 1) in order to compare the efficiency of two methods. The results of both methods are summarized in Table 1. The absolute stereochemistry at the aziridine chirality center of 4e, 4e', 4f, and 4f' were not determined.



Scheme 2. Synthesis of aziridinyl phosphonate from acetyl phosphonate.

2.2. Antibacterial studies

The newly synthesized aziridinyl phosphonates 4a-g were screened for their antibacterial activity against *Bacillus subtilis* (Gram-positive), *Escherichia coli* DH5 α (Gram-negative), and three fresh water isolates, namely Fs48 (*Gordonia* spp) (Gram-positive to variable), Fs30 (*Brevundimonas* spp) (Gram-negative), and Fs24

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| Synthesis of 2-aziridinyl phosphonates. |
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| Entry | Amine | Product | Yield (%) Method A ^a | |
|-------|---|--|------------------------------------|----|
| 1 | NH ₂ | Ph N 4a P(OEt) ₂ | 88 (86) | 85 |
| 2 | (CH ₃) ₂ CHNH ₂ | 4b P(OEt) ₂ | 88 (62) | 79 |
| 3 | NH ₂ | 4c ∧ P(OEt) ₂ | 89 (76) | 96 |
| 4 | NH ₂ | $ \begin{array}{c} $ | 50 (92) | 73 |
| 5 | HO H NH ₂ | HO N 4e, 4e' - P(OEt) ₂ | 65 (78) | 91 |
| 6 | Ph Me | $\begin{array}{c} H \\ Ph \\ \hline \\ 0 \\ \mathbf{4f}, \mathbf{f}' \\ \end{array} \begin{array}{c} H \\ N \\ P(OEt)_2 \end{array}$ | 76 (82) | 71 |
| 7 | Ph N P(OEt) ₂ | H N H 4gP_OEt OEt | 85 | |

^a Method A: Tosylate **6**, amine (1.4 equiv), and DBU (1 equiv) were stirred at rt for 24 h. In parenthesis, values obtained using acetonitrile as the solvent after 48 h stirring at rt.

^b Method B: Dibromide **2** and amine (3 equiv) were refluxed in acetonitrile for 3 h.

(*Kocuria* spp) (Gram-positive) by performing disc diffusion assays [21]. The Fs48, 30, and 24 were isolated from fresh water fish surface mucus and identified at genus level with 16s ribosomal DNA sequencing. Ampicillin and Streptomycin were used as the reference drugs. The results were recorded for each tested compound as the average diameter of inhibition zone of bacterial growth around the disks in mm (Table 2).

Results of antibacterial screening studies revealed that all the aziridinyl phosphonates showed moderate to good activity as compared to reference antibiotics. As can be seen from Table 2 aziridinyl phosphonate **4b** having benzyl group on the nitrogen showed better activity than reference antibiotics against *B. subtilis*. Aziridine **4e'** showed similar activity as amphicillin but better activity than Streptomycin against the same bacteria. In the case of *E. coli*, again, **4e'** showed the highest activity. Compounds **4b**, **4c**, **4d**, **4e'**, and **4f** showed similar activity against Fs48, **4c** showed the highest activity against this bacteria. For Fs30, all the compounds showed better activity than ampicillin and **4e** showed the highest inhibitory activity against this bacteria among the series of aziridinyl phosphonates. The compound **4e** showed the highest activity against Fs24 and the others **4a**, **4b**, **4c**, **4e'**, and **4f** showed moderate inhibitory activity against the same bacteria.

| Table 2 |
|--|
| Antibacterial activities of aziridinyl phosphonates. |

| Compounds | Inhibition zone diameters (mm) | | | | | |
|--------------|--------------------------------|------------------|------------------------------------|------------------------|------------------|--|
| | Bacillus subtilis | Escherichia coli | Fs48 Gordonia spp | Fs30 Brevundimonas spp | Fs24 Kocuria spp | |
| 4a | 10 ± 0 | 8 ± 0 | 13 ± 0 | 13.66 ± 1.15 | 18 ± 1.73 | |
| 4b | 19.33 ± 1.15 | 9 ± 0 | 15.66 ± 3.78 | 16 ± 3.60 | 19 ± 2.64 | |
| 4c | 9.66 ± 0.57 | 10 ± 0 | $\textbf{23.33} \pm \textbf{2.88}$ | 14.33 ± 0.57 | 18.33 ± 2.88 | |
| 4d | 10.33 ± 0.57 | 13.33 ± 0.88 | 14.66 ± 0.57 | 15 ± 0 | 14.66 ± 0.57 | |
| 4e | 11 ± 2 | 10.66 ± 1.15 | 11 ± 1.41 | 20 ± 0 | 25 ± 0 | |
| 4e' | 17 ± 0 | 20 ± 0 | 15.33 ± 4.61 | 13 ± 3.46 | 18.66 ± 1.51 | |
| 4f | 10 ± 0 | 13 ± 1 | 13.33 ± 1.15 | 13.66 ± 1.15 | 12 ± 2 | |
| 4f' | 11.66 ± 1.15 | 12 ± 0 | 16.33 ± 1.51 | 13.33 ± 0.57 | 19 ± 1 | |
| 4g | 11.33 ± 2.30 | 11.33 ± 2.30 | 11.66 ± 2.88 | 12.33 ± 2.51 | 12.33 ± 2.51 | |
| Ampicillin | 17.33 ± 2.3 | 16.67 ± 1.52 | ND | 10.67 ± 0.57 | 47.67 ± 0.57 | |
| Streptomycin | 14 ± 0.0 | 19.67 ± 0.57 | 29 ± 1.73 | 29.33 ± 0.57 | 26 ± 6.9 | |

It is important to note that diastereomeric aziridines **4e** and **4e'** behaved differently. While the first one showed highest activity (highest of all) against Fs24 *Kocuria spp*, second one showed the highest activity against *E. coli*. Also the diasteromers **4f** and **4f'** showed parallel activities against the first four bacteria in Table 2 but different activities against Fs24 *Kocuria spp*. Aziridine **4g** having no substituent on the nitrogen showed a low but very similar activity against all the bacteria. From these results it can be concluded that the substituent on the nitrogen as well as the stereochemistry of aziridine ring affects the antibacterial activity of these compounds.

3. Conclusion

A new method which can be considered as a modified Gabriel—Cromwell reaction was developed for the synthesis of 2-aziridinyl phosphonates. Using this method eight new aziridines including stereoisomers were synthesized in 70–92% yield. The same aziridines were also synthesized by using classical Gabriel—Cromwell reaction in 71–96% yield to compare the efficiency of two methods. Basically both methods form the aziridinyl phosphonates in similar yields. The main advantages of the method developed in this study are the ease of availability of the starting material **6** and the fact that the aziridination reaction proceeds at rt. The only disadvantage is the longer reaction times (24-48 h).

The antibacterial activities of synthesized aziridinyl phosphonates were tested for the first time in this study by performing disc diffusion assays. These studies showed that aziridine derivatives have different activities against different bacteria. The substituent on the nitrogen has a significant effect on the activity. The stereochemistry is also important, diasteromers **4e**, **4e'** and **4f**, **4f'** varied in their activity against different bacteria. Especially for the diastereomers **4e** and **4e'**, there is a significant difference in terms of antibacterial activity.

4. Experimental

4.1. Chemistry

IR spectra were recorded on FT-IR Shimadzu 8300 spectrophotometer. ¹H, ¹³C, and ³¹P NMR spectra were recorded on a Bruker 400 MHz spectrometer in CDCl₃–CCl₄ using tetramethylsilane as internal standard and chemical shifts are reported in δ units. HRMS were recorded on a Waters Micromass Q-TOF instrument in ES⁺ mode. Thin layer chromatography was performed on pre-coated silica plates (Merck Kiesegel 60 F254) and column chromatography using silica gel (mesh 230–400). Phosphomolybdic acid in ethanol and ninhydrin was used as visualizing agents.

4.1.1. Synthesis of 1-(diethoxyphosphoryl) vinyl 4methylbenzenesulfonate (**6**)

To a dry two necked round bottom flask with a magnetic stir bar under N₂ atmosphere, diethyl acetyl phosphonate (100 µL, 0.62 mmol) and 4-methylbenzene-1-sulfonyl chloride (176 mg, 0.923 mmol) in CH₃CN (2.7 mL) was added. The reaction flask was cooled to 0 °C in an ice-water bath. Then, DBU (141 µL, 0.92 mmol) was slowly added. The resulting mixture was stirred at rt for 3 h. At the end of this time, water (10 mL) and CH₂Cl₂ (10 mL) was added to the reaction flsk. Two layers were separated and aqueous layer was extracted with CH_2Cl_2 (2 \times 10 mL). The combined organic layers were dried over MgSO₄ and concentrated. The crude product was purified by flash column chromatography on silica gel (EtOAc, $R_f = 0.69$) to give **6** in 87% yield (179 mg, 0.536 mmol) as a light yellow solid. ¹H NMR δ 7.77 (d, J = 8.3 Hz, 2H), 7.28 (d, *J* = 8.1 Hz, 2H), 5.89 (ddd, *J* = 12.8, 11.8, 2.6 Hz, 2H), 4.10–3.84 (m, 4H), 2.41 (s, 3H), 1.24 (t, J = 7.1 Hz, 6H). ¹³C NMR δ 147.21 (Ar), 144.95 (Ar), 133.43 (CH₂CP), 129.64 (Ar), 128.44 (Ar), 118.98 (d, $J_{P-C} = 23.16$ Hz), 62.86 (CH₂CH₃), 62.81 (CH₂CH₃), 21.64 (CH₃Ar), 16.18 (CH₃CH₂), 16.12 (CH₃CH₂). ³¹P NMR δ 5.24.

4.2. General procedure for the synthesis of aziridinyl phosphonates **4** (*Method A*)

To a stirred mixture of 1-(diethoxyphosphoryl)vinyl 4-methylbenzenesulfonate (**6**, 0.83 mmol) and DBU (0.83 mmol, 1.0 equiv) was added amine (1.16 mmol, 2 equiv) at rt ad stirred for 24 h. When the reactions were carried out in CH₃CN, the concentration was adjusted to ~ 2 M with respect to amine. The resulting mixture was stirred at this temperature for 48 h. The crude product was purified by column chromatography using silica gel and hexane:ethyl acetat (1:1 mixture) as the eluent.

4.2.1. Diethyl 1-benzylaziridin-2-ylphosphonate (4a)

Colorless oil, (EtOAc, $R_f = 0.1$), yield 88% (197 mg, 0.732 mmol); ¹H NMR δ 7.40–7.07 (m, 5H), 4.14–3.71 (m, 4H), 3.55 (d, J = 13.0 Hz, 1H), 3.24 (d, J = 13.0 Hz, 1H), 2.12 (dd, J = 9.2, 3.5 Hz, 1H), 1.77–1.39 (m, 2H), 1.20 (t, J = 7.0 Hz, 3H), 1.13 (t, J = 7.1 Hz, 3H). ¹³C NMR δ 137.78 (CH), 128.38 (CH), 128.28 (CH), 127.34 (CH), 65.27 (d, J = 7.3 Hz, CH₂Ph), 62.29 (d, J = 6.2 Hz, CH₂CH₃), 61.96 (d, J = 6.1 Hz, CH₂CH₃), 31.92 (d, J = 5.3 Hz, CH₂N), 31.71 (d, $J_{P-C} = 216.9$ Hz, PCH), 16.39 (CH₃), 16.33 (CH₃). ³¹P NMR δ 22.48. IR (neat, cm⁻¹) 3062, 2981, 2930, 2906, 1454, 1019, 766. HRMS-EI (m/z): calcd for C₁₃H₂₁NO₃P (M + H⁺): 270.1259; found: 270.1254.

4.2.2. Diethyl 1-isopropylaziridin-2-ylphosphonate (4b)

Colorless oil, (EtOAc, $R_f = 0.14$), yield 88.4% (165 mg, 0.746 mmol); ¹H NMR δ 4.17–3.94 (m, 4H), 2.00 (ddd, J = 8.9, 3.5, 1.0 Hz, 1H), 1.45 (td, J = 7.1, 1.1 Hz, 1H), 1.41–1.32 (m, 2H), 1.27 (t,

J = 7.1 Hz, 6H), 1.10 (d, *J* = 8.9 Hz, 3H), 1.08 (d, *J* = 8.9 Hz, 3H). ¹³C NMR δ 62.36 (d, *J* = 7.1 Hz, CH(CH₃)₂), 62.18 (d, *J* = 6.4 Hz, CH₂CH₃), 61.67 (d, *J* = 6.3 Hz, CH₂CH₃), 31.36 (d, *J*_{P-C} = 219.3 Hz, PCH), 31.07 (d, *J* = 5.2 Hz, CH₂N), 21.87 (C), 16.36 (CH₃), 16.30 (CH₃), 16.23 (CH₃). ³¹P NMR δ 23.11. IR(neat, cm⁻¹) 3345, 2970, 2931, 2874, 1370, 1234, 1024, 970. HRMS-EI (*m*/*z*): calcd for C₉H₂₁NO₃P (M + H⁺): 222.1259; found: 222.1251.

4.2.3. Diethyl 1-cyclohexylaziridin-2-ylphosphonate (4c)

Colorless oil, (EtOAc, $R_f = 0.29$), yield 89% (210 mg, 0.804 mmol); ¹H NMR δ 4.15–3.99 (m, 4H), 2.04–1.95 (m, 1H), 1.73 (m, 4H), 1.58–1.31 (m, 5H), 1.27 (t, J = 7.0 Hz, 3H), 1.26 (t, J = 7.0 Hz, 3H), 1.21–0.97 (m, 4H). ¹³C NMR δ 70.08 (d, J = 6.7 Hz, CHNCHP), 62.29 (d, J = 6.3 Hz, CH₂CH₃), 61.82 (d, J = 6.3 Hz, CH₂CH₃), 32.41 (CH₂CHCH₂), 30.78 (d, $J_{P-C} = 219.2$ Hz, PCH), 30.59 (d, J = 5.2 Hz, CH₂NCH), 25.93 (CH₂CH₂CH), 24.46 (CH₂CH₂CH₂CH), 16.45 (d, J = 6.1 Hz, CH₃CH₂), 16.36 (d, J = 6.2 Hz, CH₃CH₂). ³¹P NMR δ 23.36. IR(neat, cm⁻¹) 2980, 2927, 2854, 1449, 1245, 1022, 961. HRMS-EI (m/z): calcd for C₁₂H₂₅NO₃P (M + H⁺): 262.1572; found: 262.1569.

4.2.4. Diethyl 1-(furan-2-ylmethyl)aziridin-2-ylphosphonate (4d)

Colorless oil, (EtOAc, $R_f = 0.24$), yield 50% (108 mg, 0.415 mmol); ¹H NMR δ 7.29 (dd, J = 1.8, 0.8 Hz, 1H), 6.25 (dd, J = 3.2, 1.8 Hz, 1H), 6.21 (d, J = 3.2 Hz, 1H), 4.07–3.92 (m, 4H), 3.62 (d, J = 13.9 Hz, 1H), 3.29 (d, J = 13.9 Hz, 1H), 2.09 (ddd, J = 9.2, 3.2, 1.2 Hz, 1H), 1.70–1.56 (m, 2H), 1.24 (t, J = 7.1 Hz, 3H), 1.19 (t, J = 7.1 Hz, 3H). ¹³C NMR δ 151.22 (OCCH), 142.09 (OCHCH), 110.25 (CHCO), 108.35 (CHCHO), 62.33 (d, J = 6.2 Hz, CH₂CH₃), 62.01 (d, J = 6.1 Hz, CH₂CH₃), 56.27 (d, J = 7.5 Hz, CH₂CCH), 31.23 (d, J = 5.4 Hz, CH₂CHP), 31.03 (d, $J_{P-C} = 216.7$ Hz), 16.43 (CH₃), 16.37 (CH₃). ³¹P NMR δ 22.40. IR(neat, cm⁻¹) 3113, 2983, 2931, 2908, 1505, 1243, 1017, 796. HRMS-EI (m/z): calcd for C₁₁H₁₉NO₄P (M + H⁺): 260.1052; found: 260.1058.

4.2.5. Diethyl (1-(1-hydroxybutan-2-yl)aziridin-2-yl)phosphonate (**4e** and **4e**')

4e: Colorless oil, $[\alpha]_D^{21} = +38$ (c, 0.1, CH₂Cl₂), (EtOAc/ MeOH = 10:1, R_f = 0.32), yield 33% (69 mg, 0.274 mmol); ¹H NMR δ 4.20-4.01 (m, 4H), 3.68-3.53 (m, 2H), 3.33 (s, 1H), 2.01 (dd, J = 8.9, 3.6 Hz, 1H), 1.72 (ddd, J = 20.5, 6.6, 3.6 Hz, 1H), 1.57 (t, J = 7.1 Hz, 1H), 1.50–1.42 (m, 2H), 1.37–1.32 (m, 1H), 1.30 (t, J = 7.1 Hz, 3H), 1.26 (t, J = 7.1 Hz, 3H), 0.88 (t, J = 7.5 Hz, 3H).¹³C NMR δ 72.52 (d, J = 6.8 Hz, CHCH₂OH), 65.52 (CH₂OH), 63.01 $(d, J = 6.0 Hz, CH_2CH_3), 62.08 (d, J = 6.6 Hz, CH_2CH_3), 31.11$ (d, $J_{P-C} = 218.3 \text{ Hz}$), 29.53 (d, J = 5.9 Hz, CH₂N), 24.65 (CH₂CHN), 16.35 (d, J = 2.8 Hz, CH₃CH₂O), 16.30 (d, J = 2.2 Hz, CH₃CH₂O), 10.47 (CH₃CH₂CH). ³¹P NMR δ 23.84. IR(neat, cm⁻¹) 3403, 2977, 2933, 2877, 1233, 1019, 965, 795. HRMS-EI (m/z): calcd for C₁₀H₂₃NO₄P (M + H⁺): 252.1364; found: 252.1359. 4e': Colorless oil, $[\alpha]_D^{21} = +44$ (*c*, 0.1, CH₂Cl₂), (EtOAc/MeOH = 10:1 $R_f = 0.25$), yield 45% (94 mg, 0.374 mmol); ¹H NMR δ 4.15–4.00 (m, 4H), 3.68–3.55 (m, 2H), 2.37 (s, 1H), 2.09 (dd, *J* = 9.1, 3.6 Hz, 1H), 1.67 (t, *J* = 7.1 Hz, 1H), 1.64–1.57 (m, 1H), 1.55–1.43 (m, 2H), 1.37–1.31 (m, 1H), 1.31–1.28 (m, 3H), 1.28–1.24 (m, 3H), 0.89 (t, J = 7.5 Hz, 3H). ¹³C NMR δ 73.08 (d, J = 6.7 Hz, CHCH₂OH), 63.85 (CH₂OH), 62.52 (d, J = 6.4 Hz, CH₂O), 62.00 (d, J = 6.3 Hz, CH₂O), 31.34 (CH₂N), 29.73 (d, $J_{P-C} = 219.5 \text{ Hz}$, 23.93 (CH₂CHN), 16.45 (d, $J_{P-C} = 5.9 \text{ Hz}$, CH3CH2), 16.39 (d, $J_{P-C} = 6.1$ Hz, CH₃CH₂), 10.26 (CH₃CH₂CH). ³¹P NMR δ 22.86. IR(neat, cm⁻¹) 3394, 2979, 2931, 2878, 1233, 1019, 964, 795. HRMS-EI (m/z): calcd for C₁₀H₂₃NO₄P (M + H⁺): 252.1364; found: 252.1354.

4.2.6. Diethyl 1-(1-phenylethyl) aziridin-2-ylphosphonate (**4f** and **4f**)

4f: Colorless oil, $[\alpha]_D^{21} = +44$ (*c*, 0.1, CH₂Cl₂), (EtOAc, *R*_f = 0.38), yield 49.2% (225 mg, 0.794 mmol); ¹H NMR δ 7.34–7.08 (m, 5H),

4.21–4.01 (m, 4H), 2.35 (q, J = 6.5 Hz, 1H), 1.97 (dd, J = 8.9, 3.3 Hz, 1H), 1.55 (ddd, J = 19.3, 6.8, 3.6 Hz, 1H), 1.45 (t, J = 7.0 Hz, 1H), 1.39 (d, J = 6.5 Hz, 3H), 1.29 (t, J = 7.0 Hz, 6H). ¹³C NMR δ 143.79 (Ph), 128.28 (Ph), 127.14 (Ph), 126.58 (Ph), 70.97 (d, J = 7.1 Hz, CHPh), 62.41 (d, J = 6.5 Hz, CH₂OP), 62.19 (d, J = 6.3 Hz, CH₂OP), 32.59 (d, $I_{P-C} = 218.6$ Hz, CHP), 31.53 (d, I = 5.2 Hz, CH₂N), 23.54 (CH₃CH), 16.52 (d, $J_{P-C} = 5.7$ Hz, CH₃CH₂), 16.46 (d, $J_{P-C} = 5.8$ Hz, CH₃CH₂). ³¹P NMR δ 22.76. IR(neat, cm⁻¹) 3060, 2978, 2929, 2906, 1245, 1021, 961. HRMS-EI (m/z): calcd for C₁₄H₂₁NO₃PNa (M + Na⁺): 306.1235; found: 306.1237. **4f**': Colorless oil, $[\alpha]_D^{21} = +26$ (c, 0.1, CH₂Cl₂), (EtOAc, $R_f = 0.14$), yield 32.6% (149 mg, 0.526 mmol); ¹H NMR δ 7.40–7.10 (m, 5H), 3.96–3.54 (m, 4H), 2.32 (q, J = 6.5 Hz, 1H), 2.19 (dd, J = 9.1, 3.6 Hz, 1H), 1.60 (t, J = 7.0 Hz, 1H), 1.52–1.42 (m, 1H), 1.40 (d, J = 6.6 Hz, 3H), 1.12 (t, J = 7.1 Hz, 3H), 1.03 (t, J = 7.1 Hz, 3H).NMR δ 143.21 (Ph), 128.23 (Ph), 127.34 (Ph), 127.13 (Ph), 71.42 (d, J = 6.9 Hz, CH₃CH), 62.14 (d, J = 6.2 Hz, CH₂CH₃), 61.48 (d, J = 6.0 Hz, CH₂CH₃), 31.97 (d, J = 5.2 Hz, CH₂N), 31.28 (d, $J_{P-C} = 216.4$ Hz, CHP), 22.93 (CH₃CH), 16.30 (CH₃CH₂), 16.23 (CH₃CH₂). ³¹P NMR δ 21.49. IR (neat, cm⁻¹) 3060, 2978, 2929, 2906, 1246, 1022, 954. HRMS-EI (*m*/ *z*): calcd for C₁₄H₂₁NO₃PNa (M + Na⁺): 306.1235; found: 306.1225.

4.3. Aziridin-2-ylphosphonate (4g)

Diethyl 1-benzylaziridin-2-ylphosphonate (**4a**, 50 mg, 0.19 mmol) and Pd–C (25 mg 10%) was stirred in CH₃OH (0.63 mL) under N₂ atmosphere. Then H₂ gas filled into the reaction flask having a baloon attached to it. After stirring for 30 min tlc analysis showed no starting material. The reaction mixture was filtered through celite and the solvent was removed by rotary evaporator to give pure compound **4g** (28.6 mg, 0.160 mmol) in 85% yield as a colorless oil. ¹H NMR δ 4.11–3.91 (m, 4H), 1.89 (d, J = 11.1 Hz, 1H), 1.79–1.61 (m, 2H), 1.30–1.15 (m, 6H). ¹³C NMR δ 62.03 (t, J = 5.5 Hz), 22.54 (d, J = 2.4 Hz), 21.97 (d, $J_{P-C} = 195.6$ Hz), 16.38 (CH₃), 16.32 (CH₃). ³¹P NMR δ 27.15. IR (neat, cm⁻¹) 3450, 3256, 2983, 2909, 1235, 1018, 958. HRMS-EI (*m*/*z*): calcd for C₆H₁₅NO₃P (M + H⁺): 180.0783; found: 180.0789.

4.4. Synthesis of 1,2-dibromoethylphosphonate (2)

Diethyl vinylphosphonate (**1**, 0.2 mL, 1.24 mmol) was added into a pre-dried two necked flask and was dissolved in CH₂Cl₂ (13 mL) and the reaction mixture cooled to 0 °C Br₂ (0.830 mL, 1.67 mmol, in 2.4 mL CH₂Cl₂) was added to this solution. After 30 min stirring at rt, tlc showed no starting material. The reaction mixture was applied directly to column chromatography (EtOAc) to yield **2** (382 mg, 1.18 mmol) in 95% yield as a light yellow oil. ¹H NMR δ 4.26–4.12 (m, 4H), 4.03–3.88 (m, 2H), 3.61–3.48 (m, 1H), 1.33 (t, *J* = 7.0 Hz, 6H). ¹³C NMR δ 64.01 (d, *J* = 7.0 Hz, CH₂CH₃), 63.70 (d, *J* = 6.9 Hz, CH₂CH₃), 41.88 (d, *J*_{P–C} = 150.8 Hz), 31.72 (CH₂Br), 16.37 (CH₃CH₂), 16.32 (CH₃CH₂). ³¹P NMR δ 15.27.

4.5. Synthesis of aziridinyl phosphonates **4a**–**4g** by (Method B)

Diethyl 1,2-dibromoethylphosphonate (**2**, 329 mg, 1.02 mmol) was weighed into a pre-dried two necked flask and dissolved by adding CH₃CN (1.85 mL). After adding Et₃N (0.170 mL, 1.22 mmol) at rt, white solids were observed. Then the amine (3.05 mmol) was added and the resulting mixture was refluxed at 80–85 °C for 3 h. At the end of this time, the reaction mixture was treated with 0.1 N HCl (10 mL) and CH₂Cl₂ (10 mL) was added. Two layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (2 × 10 mL). The combined organic layers were dried over Na₂SO₄, concentrated under reduced pressure, and purified by flash column chromatography on silica gel using hexane:ethyl acetat (1:1 mixture) as the eluent.

4.6. Antibacterial assays

The prepared compounds 4a-g were evaluated for their antibacterial activities against Bacillus subtilis, Escherichia coli DH5a, isolate Fs48 (Gordonia spp), Fs30 (Brevundimonas spp), Fs24 (Kocuria spp) by performing disc diffusion assays [20]. The 100 µl volumes from liquid cultures were spreaded onto nutrient agar in plates (Merck, Germany). A 100 ul volume from each test compound (we assumed that 1 μ l compound is 1 μ g) was dissolved in 400 µl 25% DMSO (DMSO final concentration was 20%). A 50 µl volumes from DMSO dissolved test compounds were incorporated in sterile disc filters (Whatman No. 1). The discs containing test compound and only 25% DMSO (control) were introduced into the middle of the bacteria inoculated agar surfaces in petri plates. The cultures were incubated 24 h at 28 °C. The experiments were performed in triplicate. Ampicillin and Streptomycin were used as the reference drugs (10 µg per disk). The results were recorded for each tested compound as the average diameter of bacterial growth inhibition zones around the disks in mm (Table 2).

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