Cu-Catalyzed Pyridine Synthesis via Oxidative Annulation of Cyclic Ketones with Propargylamine

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ABSTRACT: A Cu-catalyzed, easily scalable one-pot synthesis of fused pyridines by the reaction of cyclic ketones with propargylamine is described. The protocol was optimized based on the results of more than 30 experiments. The highest product yields were achieved in *i*-PrOH as a solvent in the presence of 5.0 mol % CuCl₂ in air. In contrast to the well-known Au-catalyzed protocol, our procedure is "laboratory friendly", cost-effective, and suitable for preparing dozens of grams of fused pyridine-based building blocks and does not require a high-pressure autoclave technique. Decreasing the catalyst amount in the reaction to 1.25 mol % CuCl₂ provided a yield comparable to that achieved with 5 mol % catalyst, though a longer reaction time was required. A plausible reaction mechanism was proposed. The scope and limitation of the reaction were studied using 24 different cyclic ketones as starting materials. The fused pyridine yield



decreased among cyclic ketones in the following order: six-membered \gg eight-membered > five-membered ~ seven-membered. The elaborated reaction conditions demonstrated tolerance to a number of protective functional groups in ketone such as ester, *tert*-butoxycarbonyl (Boc)-protected amine, and acetal moieties.

INTRODUCTION

Modern drug design requires screening compounds (SCs) based on sophisticated structural motifs,¹ in particular, aromatic heterocycles fused with saturated cyclic hydrocarbons or nonaromatic heterocycles,² possessing desired physicochemical properties.^{3,4} Nowadays, the development of organic synthesis is significantly determined by drug discovery,⁴ but often the preparation stage remains a rate-limiting factor for projects in medicinal chemistry.⁵ Therefore, studies of new catalytic reactions for simple, cost-effective, and easily scalable preparation of drug candidates, as well as corresponding building blocks (BBs), are strongly required. Continuing our ongoing monitoring of the commercially available space of SCs and BBs, as well as to identify existing "white spots" in "chemical space",^{6,7} we turned our attention to the development of methods for large-scale synthesis of pyridines annulated to (functionalized) saturated cycles (PASCs) because many representative examples of this group of structures are currently proceeding as active compounds in different phases of clinical trials (Figure 1).⁸⁻¹⁴

Most of the existing methods proposed for the synthesis of pyridine derivatives and suitable for preparing PASCs are retrosynthetically based on the cyclization of cycloalkanones 16 or their derivatives (parent pyridines 9 fused with cyclopentane, cyclohexane, and cycloheptane are provided in Figure 2 as representative examples). In particular, the application of the

intermolecular inverse electron demand Diels–Alder (IEDDA) reaction of 1,2,3-triazine 14 with the corresponding enamines 13 (method A)¹⁵ is one of the most promising synthetic approaches at present. Unfortunately, the poor availability of heterocycles 14 prevents multigram syntheses. Thermal oxygeninduced cyclization of oxime *O*-allyl ethers 15 (method B)¹⁶ requires harsh reaction conditions (sealed glass tube, heating at 190 °C). Besides, yields of the target compounds using such a method are usually low. That is why a scale-up possibility has not been investigated for the reaction. Application of [4 + 2]-cyclocondensation of the corresponding cyclic ketones with (*Z*)-3-aminoacrolein (17) (method C)¹⁷ is restricted by the limited availability of 17 and again troublesome scale-up of the process.

Several transition metal-mediated catalytic procedures have been elaborated for preparing PASCs from the corresponding ketones or alcohols as starting materials. Propargylamine **18** (method D),¹⁸ 1,3-diaminopropane **19** (method E),¹⁹ and 3aminopropan-1-ol **21** (method F)^{20–22} have been used as

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Figure 1. Representative examples of biologically active PASCs.



Figure 2. Known approaches (A–F) for the synthesis of pyridines fused with saturated cyclic fragments.

cyclization partners (Figure 2). However, in most cases, these methods were not suitable for the cost-effective multigram synthesis of annulated pyridines. The high cost of gold catalysts, the necessity to perform the reaction in an autoclave (method D)¹⁸ or using commercially unavailable and expensive Ru and Ir catalysts (methods E and F), $^{20-22}$ along with undetermined tolerance to commonly used protected functional groups, and, finally, the poor or unknown applicability to scaling-up are the reasons for continuing efforts in looking for more efficient reaction procedures.²³ Herein, we turned our attention to the Au-catalyzed method F. In our hands, this method could be reproduced on a 10 g scale of starting ketones with moderate preparative yields of PASCs. However, such a procedure lacked wide implementation. Despite our significant experience with the autoclave technique,²⁴ even in the presence of hazardous gases²⁵ (the autoclave was filled with oxygen for the pyridine synthesis by method D), the method could not be easily scaled up for preparing multigram quantities of the target compounds. The availability and cost efficiency of Au catalysts also left much to be desired. Thus, this study aimed to develop a method for multigram (20+ g) synthesis of PASCs from propargylamine and cyclic ketones based on noble metal-free catalysts, as well as to reveal the influence of various factors on the reaction course.

As a result, the mild "laboratory friendly" conditions for the multigram synthesis of PASCs were found. The method has prospects for further semi-industrial applications.

RESULTS AND DISCUSSION

First, we tried to avoid using the expensive NaAuCl₄ salt as the catalyst. In a seminal Abbiati paper,¹⁸ Cu salts have been tested compared to the Au-based catalysts, but the former catalysts have demonstrated significantly lower efficiency. Also, during our project, Meng and co-workers published a report, where copper(II) 1,3,5-benzenetricarboxylate (HKUST-1) is used as the reaction catalyst but with the low scaling of the reaction (only up to ca. 500 mg of a ketone).²⁶ This report is consistent with our preliminary results on the catalytic synthesis of pyridines starting from propargylamine in the presence of HKUST-1.²⁷ Therefore, the study was focused on using copper(II) salts as catalysts for the synthesis of PASCs. As a model reaction, the synthesis of ethyl 5,6,7,8-tetrahydroquinoline-6-carboxylate was chosen. This compound contained the quite sensitive ester group (potentially able to react with amines or to undergo hydrolysis), and ester tolerance to the reagents was shown. This process is quite typical in the series, and the product is a valuable building block for MedChem.²⁸ In the first step, the reaction was tested on a 1.5 g scale of the starting ketone with different Cu(II) salts using the autoclave technique similar to the method reported in the Abbiati paper.¹⁸ The yields in these experiments were lower than those obtained in our experiments using NaAuCl₄·2H₂O as the catalyst (Table 1, entries 1-7). The reaction time (8 h) was chosen based on unification reasons. In model experiments, the product yield did not change after 2 h of heating at 5% Cu(II) loading or after 4 h of heating at 1.25% Cu(II) loading, as was shown in separate experiments.

In the second step, we carried out the same reaction of ketone **22c** with propargylamine **18** in the presence of Cu(II) catalysts in a round-bottom flask in air at atmospheric pressure using an efficient condenser to prevent volatilization of **18** (bp = 83 °C). To our delight, this approach gave more promising results. In the presence of 2.5 mol % CuCl₂ (Table 1, entry 12), the yield of the product was comparable to those obtained in the synthesis using

Table 1. Optimization of the Synthesis of Ethyl 5,6,7,8-Tetrahydroquinoline-6-carboxylate $23c^{a,b}$

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EtO ₂ C	\checkmark .	Conditions	EtO ₂ C
		see Table 1	
	✓ ¹ O H ₂ N 22c	18	~ N 23c
	220	10	
entry	catalyst	solve	nt $(^{\circ}C)$ $(\%)$
1^d	2.5 mol % NaAuCl ₄ ·2	H ₂ O EtOH	100 48
2 ^{<i>d</i>}	7.3 mol % HKUST-1	EtOH	100 44
3 ^d	8.7 mol % CuCl ₂ ·2H ₂	O EtOH	100 42
4 ^{<i>d</i>}	8.7 mol % CuCl ₂	EtOH	100 36
5 ^{<i>d</i>}	3.7 mol % Cu(Piv) ₂	EtOH	100 31
6 ^{<i>d</i>}	8.7 mol % Cu(NO ₃) ₂	•6H ₂ O EtOH	100 39
$7^{d,e}$	8.7 mol % CuCl ₂	EtOH	100 38
8	1.3 mol % CuCl ₂ ·2H ₂	O EtOH	79 48
9	2.5 mol % CuCl ₂ ·2H ₂	O EtOH	79 45
10	5.0 mol % CuCl ₂ ·2H ₂	O EtOH	79 50
11	1.3 mol % CuCl ₂	EtOH	79 48
12	2.5 mol % CuCl ₂	EtOH	79 41
13	5.0 mol % CuCl ₂	EtOH	79 49
14	5.0 mol % CuCl ₂ , 5.0 K ₃ [Fe(CN) ₆]	mol% EtOH	79 43
15 ^f	5.0 mol % CuCl ₂	EtOH	79 66
16	5.0 mol % Cu(OAc) ₂	EtOH	79 59
17	5.0 mol % Cu(NO ₃) ₂	•6H ₂ O EtOH	79 46
18	5.0 mol % Cu(OTf) ₂	EtOH	79 60
19	5.0 mol % HKUST-1	EtOH	79 50
20	5.0 mol % FeCl ₃	EtOH	79 41
21	2.5 mol % CuCl ₂	<i>i</i> -PrOH	83 65
22	5.0 mol % CuCl ₂	<i>i</i> -PrOH ^g	83 75
23	10.0 mol % CuCl ₂	<i>i</i> -PrOH	83 60
24 ^{<i>h</i>}	5.0 mol % CuCl ₂	<i>i</i> -PrOH	83 59
25	5.0 mol % CuCl ₂ , 5.0 K ₃ [Fe(CN) ₆]	mol % <i>i</i> -PrOH	83 62
26	5.0 mol % CuCl ₂ , 10 quinhydrone	mol % <i>i</i> -PrOH	83 57
27	5.0 mol % CuCl ₂ , 10 ascorbic acid	mol % <i>i</i> -PrOH	83 67
28	5.0 mol % Cu(NO ₃) ₂	·6H ₂ O <i>i</i> -PrOH	83 58
29	5.0 mol % CuCl	<i>i</i> -PrOH	83 61
30	5.0 mol % CuI	<i>i</i> -PrOH	83 63
31	5.0 mol % CuCl ₂	$(CF_3)_2C$	HOH 58 42
32	5.0 mol % CuCl ₂	t-BuOH	82 57
33	5.0 mol % CuCl ₂	<i>i</i> -AmOH	131 62
34	5.0 mol % CuCl ₂	dioxane	101 61

^aThe experiments for a 1.5 g scale of ketone **22c**. ^bThe reaction time was 8 h. ^cThe yield was determined according to the NMR spectra of pretreated aliquots using 1,3,5-trimethoxybenzene as a standard. ^dIn autoclave under an O₂ atmosphere. ^eKNO₃ was added as a co-oxidant. ^fPropargylamine was added in three portions with 2 h intervals. ^gThe best reaction conditions, which were chosen for subsequent research, are highlighted in bold. ^hWith barbotage of air through the reaction mixture.

autoclave techniques. As the next step of screening the reaction conditions, the Cu(II) source was optimized. In particular, it included variations of the catalyst type (CuCl₂·2H₂O, CuCl₂, Cu(OAc)₂, Cu(OTf)₂, Cu(NO₃)₂·6H₂O, HKUST-1, CuCl, CuI), catalyst loading, and solvent (EtOH, *i*-PrOH, (CF₃)₂CHOH, *t*-BuOH, (CH₃)₂CHCH₂CH₂OH, dioxane). Since the reaction scheme involved oxidation at a certain step (the plausible mechanism is presented below), the agents potentially facilitating electron transfer in a Cu^{I/II} redox couple

were also added $(K_3[Fe(CN)_6])$, quinhydrone, ascorbic acid) and respective modifications of the procedure (e.g., air barbotage, slower reagent addition, etc.) were tested. To determine the role of oxygen in the catalytic process, air bubbling through the reaction mixture was carried out. Air barbotage led to a slight decrease of product yield because it could cause the blowing off of propargylamine from the reaction vessel by air flow. More intense oxidative dimerization of propargylamine could also contribute to lower yield in this case. The main results of the optimization are summarized in Table 1. Among Cu sources, anhydrous CuCl₂ turned out to be the best choice. The optimal value of Cu(II) loading depended on the solvent in different ways (ca. 2.5 mol % in EtOH and ca. 5.0 mol % in *i*-PrOH; see Table 1, entries 11-13 and 21-23, respectively). The increase of the catalyst loading led to lower yields. This observation can be referred to as a significant contribution of side reactions in the presence of high Cu(II) salt concentration. Such side reactions can include oxidative coupling of propargylamine in the presence of $CuCl_{2}$ ²⁹ as well as self-condensation of ketones.^{30,3}

Isopropanol was found to be the optimal solvent for the reaction (Table 1, entries 12, 22, 29–32). Adding K_3 [Fe(CN)₆], quinhydrone, or ascorbic acid did not lead to rising yields. Moreover, $K_3[Fe(CN)_6]$ or ascorbic acid slightly decreased the yields, and an even more significant decrease of the yield was observed in the presence of quinhydrone (Table 1, entries 25-27). Replacement of CuCl₂ by CuCl or CuI led to similar product yields. This observation is consistent with the proposed plausible reaction mechanism, where both Cu(II) and Cu(I) played their roles in the catalytic cycle. Notably, both CuCl, in which Cu(I) can be easily oxidized by oxygen to Cu(II), and CuI, where Cu(I) is stable in air, are effective. Other deviations from the initially chosen conditions also did not make a positive effect on the reaction outcome (Table 1, entries 12, 15). Therefore, stirring of the reaction mixture under reflux in i-PrOH in the presence of 5.0 mol % CuCl₂ as a catalyst with air oxygen access proved to be the optimal protocol. Under these conditions, the model reaction of ethyl cyclohexa-4-one-1carboxylate with propargylamine yielded 75% of the desired ethyl 5,6,7,8-tetrahydroquinoline-6-carboxylate 23c (Table 1, entry 22).

To reveal the rate of product accumulation in the reaction mixture and to get insight into the reaction mechanism (for example, the formation of any intermediate, which slowly transforms into the target compound), we studied the dependence of the product yield on the reaction time in two systems: i-PrOH-1.25 mol % CuCl₂ and i-PrOH-5.0 mol % CuCl₂. In both cases, the reaction proceeded relatively fast and only compound 23c was detected by gas chromatography-mass spectrometry (GCMS) (this analysis does not exclude the possible formation of some minor impurities, for example, condensation products). Several conclusions can be made from the analysis of the curves. All reactions leading to 23c demonstrated comparable rates. The yield of 23c did not decrease after its formation, proving its stability. Thus, the lower yield upon addition of 10 mol % CuCl₂ (entry 23) can be explained by quick competitive reactions rather than degradation of the product 23c. There was no oxygen deficit under the reaction conditions, as the yield of 23c quickly reached the highest value in the case of 5 mol % CuCl₂. Finally, in conditions of 2-fold propargylamine excess, its loss due to volatilization did not make a significant influence on the product yield, because the yield of 23c in the experiment with 1.25 mol % CuCl₂ was

comparable to that in the reaction with 5 mol % catalyst, though a longer reaction time was necessary (Figure 3).



Figure 3. Dependence of the yield of **23c** on time for different loadings of CuCl₂. Solid lines are to guide the eye.

The product concentration vs time dependencies for 23c formation could not be fitted by the first- or the second-order kinetic equations. The values of turnover frequency (average TOF, the ratio of the quantity of 23c formed to CuCl₂ content) calculated for the initial parts of the yield vs time curves (0–25 min range) were equal to 0.38 min⁻¹ for 1.25 mol % CuCl₂ loading and 1.0 min⁻¹ for 5 mol % CuCl₂ loading. Taking into account failure to fit the kinetic curve by simple equations along with the fact that TOF was not proportional to the catalyst loading (4 times increase of the catalyst loading resulted in 2.6 times increase of the TOF value), we conclude that the reaction mechanism is complex and there is no single rate-limiting stage.

To evaluate the scope and limitations of the developed protocol, a set of ketones were tested in the reaction under the above-mentioned optimized conditions (Scheme 1). Among 24 substrates, the desired products were not detected in the reaction mixture only for 7 ketones: ethyl 2-oxocyclohexanecarboxylate **22f**,³² dihydrothiophen-3(2*H*)-one **22j**, 2-fluorocyclohexanone 22t, monoprotected 1,3-cyclohexanedione 22s, furan-2,4(3H,5H)-dione **22u**, and α,β -unsaturated cyclic ketones 22v and 22w.^{33,34} In other cases, the desired fused pyridines formed in 5-65% yields (see Table 2). Interestingly, among all ketones, the best results were obtained for the sixmembered species: the yields were ca. 60% except for sulfurcontaining ketones 22n and 22o. Generally, the yields as a function of the ketone ring size decreased in the following order: six-membered $(22a) \gg$ eight-membered (22e) > fivemembered $(22b) \sim$ seven-membered ketones (22d). This order is in agreement with the yields of enamines reported for the reaction of the corresponding ketones with dialkylamines.^{35,36} In the case of possible formation of regioisomers, the reaction proceeded at the more CH-acidic center (products 23k, 23p, and 23u). The CH-acidity dominated over steric hindrance from two possible reactive centers in ketone 22p, defining the annulation regioselectivity. In the case of ketone 22g, the steric hindrance became more important and the product of annulation at the less sterically hindered center was isolated as the major one. Also, the reaction conditions appeared to be tolerant to acid-sensitive protecting groups, such as a tertbutoxycarbonyl (Boc) substituent at nitrogen (ketones 22m, 22q, and 22p) or acetal protection of the second keto-function (ketone 22r).

Scheme 1. Set of Substrates for Cu-Catalyzed Pyridine Annulation Used in This Study



The most important achievement of this study is the possibility of easy scale-up for the $CuCl_2$ -catalyzed reaction to ca. 0.15–0.4 mol of the starting ketones (Table 2, entries 1, 11, 15, and 16; see the Experimental Section). This feature was also illustrated by the multigram synthesis of a few building blocks formed by deprotection of compounds **23** (Scheme 2). It makes the reaction very promising for the design and synthesis of MedChem-relevant BBs.

Based on our observations and the current state of the art evaluated by analyzing the literature data, we can propose an alternative mechanism of the reaction, which differs from that originally proposed by Abbiati and other authors.^{18,40} As suggested in the seminal Abbiati paper, the formation of the pyridine ring proceeded through the sequential amination of the carbonyl compounds, followed by regioselective 6-endo-dig cyclization of the N-propargylenamine intermediate with further aromatization. Both steps, amination and 6-endo-dig cyclization, were catalyzed by Au(III) salts, but a more detailed description of the catalyst role was not proposed. In the recent paper of Yamauchi³⁹ dealing with transition metal (TM)-free thermal reactions in nitrobenzene at 190 °C, it has been postulated that after the enamine formation the reaction proceeds by a more complicated mechanism. It includes an aza-Claisen rearrangement, followed by tautomerization of N-propargylenamine providing the allenic enamine. After the 1,5-sigmatropic hydrogen shift, 6π -electron cyclization to the dihydropyridine takes place, followed by oxidation of the latter to the corresponding pyridine. The propargyl aza-Claisen rearrangement is a logical step of the proposed mechanism. The thermal propargyl Claisen rearrangements need harsh conditions due to the disfavored geometry of the starting molecule bearing an alkyne fragment. The best catalysts for such types of rearrangements are Au(I) compounds,^{41,42} but Cu(I) derivatives also

possess catalytic activity in this process.^{43,44} This fact can explain the advantages of Au(I)-based catalysts in some cases (Table 2, entries 2, 5). To the best of our knowledge, the formation of π type complexes for Cu(II) species with alkynes required for the catalytic activity in the propargyl Claisen rearrangement is still unknown.^{45,46} However, the generation of Cu(I) compounds from Cu(II) ones has been well documented for the copper(II) acetate-accelerated azide-alkyne cycloaddition. It proceeds via Cu(II) reduction and oxidation of alcohols, used as solvents, or as a result of a Glaser reaction of terminal alkynes employed as substrates.47 Therefore, it would be logical to propose that Cu(II) compounds play an important role in the catalytic cycle, as they are known catalysts for imine formation,⁴⁸ as well as for dihydropyridine oxidative aromatization.^{49,50} The modified plausible mechanism proposed by us is represented in Scheme 3. In the first step, imine 29 is formed from the corresponding ketone 22 and propargylamine 18. This reaction is obviously catalyzed by Cu(II). Imine 29 exists in equilibrium with the corresponding enamine 30. The next step of transformation is the Cu(I)-catalyzed propargyl aza-Claisen rearrangement, leading to allene 33. The following steps are similar to the TM-free reaction and include tautomerization, 1,5-sigmatropic hydrogen shift, and 6π -electron cyclization to dihydropyridine. The latter is oxidized to a pyridine derivative directly by O_2 or through a Cu(II)-catalyzed process. Such a sequence of steps explains the advantage of Au-based catalysts over other ones because in this case the transformations of 30 into 33 are very efficient. In addition, the imine-enamine tautomerism can determine the regioselectivity of the reactions with the substituents stabilizing the corresponding enamine form (see ketones 22k, 22p, and 22u). In the reaction course, Cu(I) could be oxidized to Cu(II) by air, but this step was not critical. The proposed mechanism implies that the reaction requires some

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Table 2. Cu-Catalyzed Pyridine Annulation to Cyclic Ketones

Entr y	Substrate	Product	Scale, mol ^a	Yield, % ^b	Previously reported results
1	22a	23a	0.031	65	-
2	22b	23b	0.036	22	77%, NaAuCl ₄ ·2H ₂ O ¹⁸ ; 89%, HetPh ₂ PAuCl ^{c 37}
3	22c	EtO ₂ C	0.15	63	Yield n.d., NaAuCl ₄ ·2H ₂ O 28
4	22d	23d	0.027	20	-
5	22e	23e	0.024	29	67%, NaAuCl ₄ ·2H ₂ O ^{18;} 78%, HetPh ₂ PAuCl ^{c 34}
6	22g	EtO ₂ C 23g 2:1 23g'	0.017	58 ^d	-
7	22h	0 23h	0.006	5 ^e	-
8	22i	Boc N 23i	0.006	6 ^e	-
9	22k	000 S 23k	0.022	44	-
10	221	0 231	0.030	34	-
11	22m	Boc N N 23m	0.38	64	74%, NaAuCl ₄ ·2H ₂ O ³⁸
12	22n	S 23n	0.006	25 ^e	-
13	220		0.006	5 ^e	-
14	22p	BOC N 23p	0.015	48	-
15	22q	BocHN 23q	0.23	61	-
16	22r	23r	0.30	59	-
17	22u		0.028	52	32%, TM free, 190 °C, nitrobenzene, air ³⁹

^{*a*}Amount of the starting ketone 22. ^{*b*}Isolated yields. ^{*c*}HetPh₂P = 1-(2-(diphenylphosphino)thiophen-3-yl)-1H-imidazole; EtOH, 120 °C. ^{*d*}Preparative yield for the mixture of regioisomers; separation failed. ^{*e*}Yield according to GCMS of the reaction mixture; isolation failed.

Scheme 2. Synthesis of MedChem-Relevant Building Blocks as a Follow-Up of Cu-Catalyzed Formation of Pyridines



Scheme 3. Plausible Mechanism of Cu-Catalyzed Pyridine Annulation



quantity of Cu(II) and Cu(I) to be simultaneously present in the reaction mixture. However, the proposed above-mentioned sequence of steps is still declarative, and more thorough mechanistic studies are needed, which are beyond the scope of the current paper.

CONCLUSIONS

As a result of this study, we demonstrated that the reaction of cyclic ketones with propargylamine catalyzed by $CuCl_2$ in air led to the formation of fused pyridines with high yields. The approach was efficiently used for the synthesis of fused cycloalkenopyridines on a multigram scale. The optimal conditions for pyridine annulation included refluxing the starting materials in *i*-PrOH in the presence of 5.0 mol % $CuCl_2$ with free access to air oxygen. For the tested ketones, the elaborated reaction conditions were suitable for preparing a wide range of fused pyridines but did not lead to the formation of pyridines starting from 2-oxocycloalkane carboxylates, dihydrothiophen-3(2H)-one, 2-fluorocyclohexanone, monoprotected 1,3-cyclohexandione, furan-2,4(3H,5H)-dione, and α,β -unsaturated cyclic ketones. The product yields in the case of the

cycloalkanones with ring size from 5 to 8, as well as their oxa-, aza-, and thia-analogues, were significantly different. The best results were found in the case of six-membered ketones, which afforded the desired fused pyridines in ca. 60% yield with an exception of thiopyranone and its oxidized derivative. The dependence of product yields on ketone ring size is in the following order: six-membered \gg eight-membered > fivemembered \sim seven-membered. In the case of unsymmetrical ketones, two factors can control regiochemistry: "push-pull" stabilization of the intermediate enamine and steric hindrance, with the major role of the former. The reaction conditions were tolerant to several protecting functional groups in ketones, such as ester, Boc-protected amine, and acetal. These groups were not affected under the used reaction conditions, but afterward, the products could be easily deprotected producing functionalized fused cycloalkyl pyridines.

The elaborated catalytic procedure avoids harsh conditions, problems with availability of starting materials, using autoclave techniques, and expensive catalysts based on noble metals. In addition, the method includes only two operationally simple steps and relies on readily available and nontoxic starting

materials. It provides 40-60% overall yields of the target products, has been efficiently used on an up to several dozen gram scale, and, based on our experience, has the potential for further scale-up. This makes functionalized fused cycloalkyl pyridines readily available to the chemical community, most importantly for their use as building blocks in medicinal chemistry programs.

EXPERIMENTAL SECTION

General Information. The solvents were purified according to the standard procedures. Absolute ethanol and isopropanol were used. All starting materials were obtained from Enamine Ltd. Melting points were measured on an automated melting point system. ¹H and ¹³C NMR spectra were measured on a Bruker Avance 500 spectrometer (at 500 MHz for ¹H and 126 MHz for ¹³C nuclei) and a Varian Unity Plus 400 spectrometer (at 400 MHz for ¹H and 101 MHz for ¹³C nuclei). Tetramethylsilane (¹H, ¹³C) was used as an internal standard. Elemental analyses were performed at the Laboratory of Organic Analysis, Institute of Organic Chemistry, National Academy of Sciences of Ukraine. Their results were found to be in good agreement $(\pm 0.4\%)$ with the calculated values. Preparative high-performance liquid chromatography (HPLC) analyses were done on an Agilent 1200. Mass spectra were measured on an Agilent 1100 LCMSD SL instrument (atmospheric pressure chemical ionization (APCI)) and an Agilent 5890 series II 5972 GCMS instrument (electron impact ionization (EI)). Kinetic experiments were carried out using reaction mixtures containing 1 M 22c and 2 M propargylamine and 1.25 or 5 mol % CuCl₂ in isopropanol. The total volume of the reaction mixtures at the beginning of the experiment was 10 mL. Aliquots (0.2 mL) were taken after certain time periods, and these aliquots were quickly cooled to room temperature, purified from CuCl₂ using flash chromatography on silica gel (the column was washed out with 10 mL of chloroform, followed by 25 mL of chloroform and then 25 mL of ethyl acetate, to elute the organic compounds). The liquids were concentrated under reduced pressure and analyzed by GCMS. The GC signal intensities were calibrated using solutions with known concentrations of 22c and 23c. n-Dodecane was added as a standard to five aliquots. However, it was found that the 22c/23c ratio was sufficiently accurate in determining the product yield. This procedure does not ensure the preservation of all propargylamine in the aliquot. Error bars were evaluated by analysis of 10 specially prepared model solutions with specified quantities of 22c, 23c, and CuCl₂ (5 mol %), which were treated as described above for the aliquots; the error bar equals the highest deviations of 23c concentration from the specified values, observed in these experiments. The same error bars were used for all measurements.

General Procedure for the Synthesis of Pyridines 23. $CuCl_2$ · 2H₂O (0.262 g, 1.52 mmol) was placed in a 25 mL beaker on a preheated 150 °C plate and heated for 2–4 min to brown color. Propargylamine (3.36 g, 61.0 mmol, 3.90 mL) and isopropanol (30 mL) were placed in a 100 mL round-bottom flask. The flask was placed in an oil bath heated to 60 °C, and anhydrous $CuCl_2$ and a solution of ketone 22 (30.6 mmol) in isopropanol (5.0 mL) were added under vigorous stirring. A water-cooled condenser was installed. The mixture was boiled under vigorous stirring overnight (12 h) at 84 °C. The reaction mixture was evaporated under reduced pressure, and the residue was dissolved in CHCl₃ (100 mL). The crude product was prepurified by flash chromatography (eluent CHCl₃, then EtOAc). The resulting solution was evaporated, and the crude product was purified by column chromatography (eluent hexane/EtOAc (0–50% EtOAc)) to obtain desired product 23.

5,6,7,8-Tetrahydroquinoline (*23a*). The compound was obtained starting from cyclohexanone (3.0 g, 30.6 mmol), prop-2-yn-1-amine (3.36 g, 61.0 mmol), and CuCl₂·2H₂O (0.262 g, 1.52 mmol) in 35 mL of *i*-PrOH. Yield = 2.63 g (65%). Purified by column chromatography (hexane/EtOAc 9:1). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, *J* = 4.7 Hz, 1H), 7.30 (d, *J* = 7.7 Hz, 1H), 6.97 (dd, *J* = 7.7, 4.6 Hz, 1H), 2.88 (t, *J* = 6.4 Hz, 2H), 2.72 (t, *J* = 6.2 Hz, 2H), 1.86 (p, *J* = 6.3 Hz, 2H), 1.77 (p, *J* = 6.1 Hz, 2H). ¹³C{¹H} NMR (101 MHz,

CDCl₃) δ 157.3, 146.7, 136.7, 132.2, 120.8, 32.5, 28.7, 23.0, 22.7. Liquid chromatography-mass spectrometry (LCMS), positive mode, m/z: 134 [M + H]⁺. Anal. calcd for C₉H₁₁N: C, 81.16; H, 8.32; N, 10.52. Found: C, 81.22; H, 8.48; N, 10.20.

6,7-Dihydro-5H-cyclopenta[b]pyridine (**23b**). The compound was obtained starting from cyclopentanone (3.0 g, 35.7 mmol), propargylamine (3.98 g, 71.3 mmol), and CuCl₂·2H₂O (0.304 g, 1.78 mmol) in 35 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 9:1). Yield = 0.92 g (22%). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.29 (d, *J* = 5.1 Hz, 1H), 7.45 (d, *J* = 7.0 Hz, 1H), 6.97 (dd, *J* = 7.5, 5.1 Hz, 1H), 2.98 (t, *J* = 7.6 Hz, 2H), 2.90 (t, *J* = 7.4 Hz, 2H), 2.09 (p, *J* = 7.6, 7.2 Hz, 2H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 165.5, 147.3, 136.9, 132.0, 120.9, 34.2, 30.7, 23.0. LCMS, positive mode, *m*/*z*: 120 [M + H]⁺. Anal. calcd for C₈H₉N: C, 80.63; H, 7.61; N, 11.75. Found: C, 80.70; H, 7.71; N, 11.59.

Ethyl 5,6,7,8-Tetrahydroquinoline-6-carboxylate (23c). The compound was obtained starting from ethyl 4-oxocyclohexane-1-carboxylate (25.0 g, 0.147 mol), propargylamine (16.2 g, 0.294 mol), and CuCl₂·2H₂O (1.25 g, 7.35 mmol) in 150 mL of *i*-PrOH. Yield = 19.1 g (63%). Purified by column chromatography (hexane/EtOAc 7:3). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.33 (d, *J* = 4.7 Hz, 1H), 7.36 (d, *J* = 7.7 Hz, 1H), 7.01 (dd, *J* = 7.7, 4.8 Hz, 1H), 4.15 (q, *J* = 7.1 Hz, 2H), 3.07–2.85 (m, 4H), 2.79–2.66 (m, 1H), 2.33–2.20 (m, 1H), 2.02–1.87 (m, 1H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 174.8, 156.0, 147.2, 136.8, 130.2, 121.2, 60.7, 39.4, 31.4, 30.9, 25.8, 14.2. LCMS, positive mode, *m/z*: 206 [M + H]⁺. Anal. calcd for C₁₂H₁₅NO₂: C, 70.22; H, 7.37; N, 6.82. Found: C, 70.19; H, 7.24; N, 6.67.

6,7,8,9-Tetrahydro-5H-cyclohepta[b]pyridine (**23d**). The compound was obtained starting from cycloheptanone (3.0 g, 26.8 mmol), propargylamine (2.98 g, 63.5 mmol), and CuCl₂·2H₂O (0.220 g, 1.30 mmol) in 25 mL of *i*-PrOH. Yield = 0.78 g (20%). Purified by column chromatography (hexane/EtOAc 19:1). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.24 (d, *J* = 5.0 Hz, 1H), 7.32 (d, *J* = 7.5 Hz, 1H), 6.96 (dd, *J* = 7.5, 4.9 Hz, 1H), 3.12–2.86 (m, 2H), 2.82–2.64 (m, 2H), 1.83 (p, *J* = 5.8 Hz, 2H), 1.73–1.56 (m, 4H). ¹³C{H} NMR (101 MHz, CDCl₃) δ 163.2, 146.1, 138.1, 136.4, 121.1, 39.4, 35.3, 32.5, 27.9, 26.4. LCMS, positive mode, *m*/*z*: 148 [M + H]⁺. Anal. calcd for C₁₀H₁₃N: C, 81.58; H, 8.90; N, 9.51. Found: C, 81.41; H, 8.76; N, 9.31.

5,6,7,8,9,10-Hexahydrocycloocta[b]pyridine (**23e**). The compound was obtained starting from cyclooctanone (3.0 g, 23.7 mmol), propargylamine (2.65 g, 48.1 mmol), and CuCl₂·2H₂O (0.202 g, 1.18 mmol) in 22 mL of *i*-PrOH. Yield = 1.12 g (29%). Purified by column chromatography (hexane). Light yellow oil. ¹H NMR (500 MHz, CDCl₃) δ 8.37 (d, *J* = 4.7 Hz, 1H), 7.36 (d, *J* = 7.5 Hz, 1H), 7.04 (dd, *J* = 7.6, 4.8 Hz, 1H), 2.96 (t, *J* = 6.2 Hz, 2H), 2.75 (t, *J* = 6.2 Hz, 2H), 1.85–1.73 (m, 2H), 1.73–1.60 (m, 2H), 1.42–1.30 (m, 4H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 161.0, 147.0, 136.5, 136.2, 121.4, 34.6, 32.1, 31.9, 30.6, 25.9, 25.9. LCMS, positive mode, *m*/*z*: 162 [M + H]⁺. Anal. calcd for C₁₁H₁₅N: C, 81.94; H, 9.38; N, 8.69. Found: C, 81.77; H, 9.24; N, 8.86.

Ethyl 5,6,7,8-Tetrahydroquinoline-7-carboxylate (23q). The compound was obtained starting from ethyl 3-oxocyclohexane-1-carboxylate (5.0 g, 0.029 mol), propargylamine (3.2 g, 0.059 mol), and CuCl₂. 2H₂O (0.25 g, 1.47 mmol) in 50 mL of *i*-PrOH as a gray oil and mixture of two regioisomers in 2:1 ratio. Total yield of the mixture = 3.5 g (58%). ¹H NMR (400 MHz, CDCl₃) δ 8.38 (dd, J = 4.7, 1.9 Hz, 1H), 8.34* (d, J = 4.8 Hz, 1H), 7.47* (d, J = 7.8 Hz, 1H), 7.34 (d, J = 7.7 Hz, 1H), 7.07-7.03 (m, 1H), 7.05-6.97* (m, 1H), 4.21-4.09* (m, 4H), 3.76* (t, J = 5.8 Hz, 1H), 3.25-3.02 (m, 2H), 3.03-2.83 (m, 2H), 2.87-2.75 (m,3H), 2.25-2.08 (m, 2H), 2.08-1.93 (m, 2H), 1.93-1.77 (m, 2H), 1.29-1.21 (m, 3H), 1.23-1.20* (m, 3H) (*-another isomer ethyl 5,6,7,8-tetrahydroquinoline-5-carboxylate). $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR (101 MHz, CDCl₃) δ 174.8, 173.9*, 157.2*, 155.3, 147.9, 147.2*, 137.3*, 136.5, 131.0, 129.1*, 121.2*, 121.0, 61.0*, 60.6, 44.4*, 39.9, 34.5, 32.2*, 27.5*, 26.1, 25.3, 20.5*, 14.2 (*-another isomer ethyl 5,6,7,8-tetrahydroquinoline-5-carboxylate). LCMS, positive mode, m/z: 206 [M + H]⁺.

2,3-Dihydrothieno[3,2-b]pyridine 1,1-Dioxide (23k). The compound was obtained from dihydrothiophen-3(2*H*)-one 1,1-dioxide (3.0 g, 22.4 mmol), propargylamine (2.46 g, 44.8 mmol), and CuCl₂· 2H₂O (0.191 g, 1.12 mmol) in 20 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 4:1). Yield = 1.66 g (44%). Light yellow solid; m.p. = 120–121 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.77 (dd, *J* = 4.9, 1.7 Hz, 1H), 8.02 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.38 (dd, *J* = 7.9, 4.8 Hz, 1H), 3.61–3.55 (m, 2H), 3.55–3.48 (m, 2H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 157.0, 154.9, 133.5, 130.2, 123.1, 50.7, 28.7. LCMS, positive mode, *m*/*z*: 170 [M + H]⁺. Anal. calcd for C₇H₇NO₂S: C, 49.69; H, 4.17; N, 8.28; S, 18.95. Found: C, 49.98; H, 4.24; N, 8.02; S, 19.10.

7,8-Dihydro-5H-pyrano[*4,3-b*]*pyridine* (*23*). The compound was obtained starting from tetrahydro-4*H*-pyran-4-one (3.0 g, 29.9 mmol), propargylamine (3.30 g, 60.0 mmol), and CuCl₂·2H₂O (0.255 g, 1.50 mmol) in 30 mL of *i*-PrOH. Yield = 1.38 g (34%). Purified by column chromatography (hexane/EtOAc 3:2). Light yellow oil. ¹H NMR (500 MHz, CDCl₃) δ 8.41 (d, *J* = 5.0 Hz, 1H), 7.28 (d, *J* = 7.9 Hz, 1H), 7.09 (dd, *J* = 7.7, 4.8 Hz, 1H), 4.75 (s, 2H), 4.06 (t, *J* = 5.8 Hz, 2H), 3.01 (t, *J* = 5.9 Hz, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 153.5, 147.8, 132.3, 130.3, 121.3, 67.1, 65.7, 31.7. LCMS, positive mode, *m*/*z*: 136 [M + H]⁺. Anal. calcd for C₈H₉NO: C, 71.09; H, 6.71; N, 10.36. Found: C, 71.39; H, 6.82; N, 10.18.

tert-Butyl 7,8-Dihydro-1,6-naphthyridine-6(5H)-carboxylate (**23m**). The compound was obtained starting from *tert*-butyl 4oxopiperidine-1-carboxylate (75.0 g, 0.376 mol), propargylamine (41.4 g, 0.752 mol), and CuCl₂·2H₂O (3.20 g, 18.8 mmol) in 350 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 7:3). Yield = 56.2 g (64%). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.41 (d, *J* = 4.8 Hz, 1H), 7.40 (d, *J* = 7.8 Hz, 1H), 7.12 (dd, *J* = 7.8, 4.8 Hz, 1H), 4.58 (s, 2H), 3.75 (t, *J* = 6.0 Hz, 2H), 3.00 (t, *J* = 6.1 Hz, 2H), 1.49 (s, 9H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 154.7, 147.6, 147.6, 134.1, 129.1, 121.4, 80.2, 32.1, 31.7, 28.4, 28.3. LCMS, positive mode, *m/z*: 235 [M + H]⁺. Anal. calcd for C₁₃H₁₈N₂O₂: C, 66.64; H, 7.74; N, 11.96. Found: C, 66.63; H, 7.88; N, 11.73.

tert-Butyl 3,4-Dihydro-1,5-naphthyridine-1(2H)-carboxylate (**23p**). The compound was obtained from *tert*-butyl 3-oxopiperidine-1-carboxylate (3.0 g, 15.0 mmol), propargylamine (1.65 g, 30.0 mmol), and CuCl₂·2H₂O (0.255 g, 1.50 mmol) in 20 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 4:1). Yield = 1.68 g (48%). Light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.16 (dd, *J* = 4.7, 1.5 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 1H), 7.04 (dd, *J* = 8.4, 4.7 Hz, 1H), 3.70 (t, *J* = 5.8 Hz 2H), 2.93 (t, *J* = 6.7 Hz, 2H), 1.97 (p, *J* = 6.5 Hz, 2H), 1.49 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 153.6, 149.3, 143.7, 135.2, 130.8, 120.9, 81.4, 44.5, 30.8, 28.3, 22.7. LCMS, positive mode, *m/z*: 235 [M + H]⁺. Anal. calcd for C₁₃H₁₈N₂O₂: C, 66.64; H, 7.74; N, 11.96. Found: C, 66.92; H, 8.13; N, 11.84.

tert-Butyl (*5*,*6*,*7*,*8*-*Tetrahydroquinolin-6-yl*)*carbamate* (**23***q*). The compound was obtained from *tert*-butyl (4-oxocyclohexyl)carbamate (50.0 g, 0.234 mol), propargylamine (25.8 g, 0.468 mol), and CuCl₂· 2H₂O (1.99 g, 11.7 mmol) in 240 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 7:3). Yield = 35.3 g (61%). Colorless solid; m.p. = 101–102 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.35 (d, *J* = 4.8 Hz, 1H), 7.32 (d, *J* = 7.7 Hz, 1H), 7.02 (dd, *J* = 7.7, 4.8 Hz, 1H), 4.60 (br s, 1H), 3.97 (br s, 1H), 3.10 (dd, *J* = 16.3, 4.9 Hz, 1H), 3.01 (t, *J* = 6.8 Hz, 2H), 2.63 (dd, *J* = 16.3, 8.3 Hz, 1H), 2.18–2.07 (m, 1H), 1.91–1.76 (m, 1H), 1.43 (s, 9H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 155.8, 155.3, 147.5, 137.2, 129.5, 121.2, 79.5, 45.7, 35.4, 30.3, 28.9, 28.4. LCMS, positive mode, *m*/*z*: 249 [M + H]⁺. Anal. calcd for C₁₄H₂₀N₂O₂: C, 67.72; H, 8.12; N, 11.28. Found: C, 67.36; H, 7.86; N, 11.00.

T',8'-Dihydro-5'H-spiro[[1,3]dioxolane-2,6'-quinoline] (**23r**). The compound was obtained from 1,4-dioxaspiro[4.5]decan-8-one (50.0 g, 0.30 mol), propargylamine (33.0 g, 0.60 mol), and CuCl₂·2H₂O (2.56 g, 15.0 mmol) in 280 mL of *i*-PrOH. Purified by column chromatography (hexane/EtOAc 4:1). Yield = 33.8 g (59%). Light yellow oil. ¹H NMR (500 MHz, CDCl3) δ 8.38 (d, *J* = 4.6 Hz, 1H), 7.34 (d, *J* = 7.5 Hz, 1H), 7.05 (dd, *J* = 7.7, 4.8 Hz, 1H), 4.04 (s, 4H), 3.14 (t, *J* = 6.8 Hz, 2H), 3.00 (s, 2H), 2.06 (t, *J* = 6.6 Hz, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 155.8, 147.3, 137.0, 129.4, 121.0, 107.5, 64.6,

Article

38.5, 31.5, 30.9. LCMS, positive mode, m/z: 192 [M + H]⁺. Anal. calcd for C₁₁H₁₃NO₂: C, 69.09; H, 6.85; N, 7.32. Found: C, 69.48; H, 7.16; N, 7.34.

7,8-Dihydroquinolin-5(6H)-one (23u). The compound was obtained from cyclohexane-1,3-dione (3.0 g, 26.8 mmol), propargylamine (2.95 g, 53.5 mmol), and CuCl₂·2H₂O (0.228 g, 1.34 mmol) in 23 mL of *i*-PrOH. Yield = 2.05 g (52%). Purified by column chromatography (hexane/EtOAc 4:1). Light yellow oil. ¹H NMR (500 MHz, dimethyl sulfoxide (DMSO)- d_6) δ 8.64 (dd, *J* = 4.8, 2.0 Hz, 1H), 8.23 (dd, *J* = 7.9, 1.9 Hz, 1H), 7.25 (dd, *J* = 8.0, 4.9 Hz, 1H), 3.13 (t, *J* = 6.3 Hz, 2H), 2.66 (t, *J* = 6.5 Hz, 2H), 2.17 (p, *J* = 6.5 Hz, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 197.9, 163.6, 153.4, 135.0, 128.1, 122.2, 38.5, 32.5, 21.8. LCMS, positive mode, *m*/*z*: 148 [M + H]⁺. Anal. calcd for C₉H₉NO: C, 73.45; H, 6.16; N, 9.52. Found: C, 73.46; H, 6.23; N, 9.70.

5,6,7,8-Tetrahydroquinoline-6-carboxylic Acid (24). A solution of NaOH (8.80 g, 0.22 mol) in H₂O (200 mL) was added to a stirred solution of ethyl 5,6,7,8-tetrahydroquinoline-6-carboxylate (30.0 g, 0.146 mol) in MeOH (300 mL). The reaction mixture was refluxed for 4 h in an oil bath and cooled, and then a solution of NaHSO₄ (26.4 g, 0.220 mol) in H₂O (150 mL) was added. The mixture was stirred for 30 min and evaporated. The residue was treated with boiling dry EtOH (2 × 500 mL), inorganic salts were filtered, and the organic solution was evaporated. The crude product was treated with dry EtOH (50 mL) and filtered. The product was washed with CH_3CN (2 × 30 mL) to give the desired product as a white powder. Yield = 21.2 g (82%). ¹H NMR (400 MHz, DMSO- d_6) δ 12.38 (brs, 1H), 8.29 (d, J = 4.6 Hz, 1H), 7.49 (d, J = 7.7 Hz, 1H), 7.12 (dd, I = 7.7, 4.7 Hz, 1H), 3.00–2.80 (m, 4H), 2.77– 2.64 (m, 1H), 2.24–2.04 (m,1H), 1.93–1.76 (m, 1H). ¹³C{¹H} NMR (151 MHz, DMSO-*d*₆) δ 176.3, 156.2, 147.1, 137.1, 130.7, 121.6, 38.8, 31.3, 30.7, 25.7. LCMS, positive mode, *m*/*z*: 178 [M + H]⁺. Anal. calcd for C₁₀H₁₁NO₂: C, 67.78; H, 6.26; N, 7.90. Found: C, 67.44; H, 6.29; N. 8.29.

5,6,7,8-Tetrahydro-1,6-naphthyridine-1,6-diium Chloride (25). HCl (6 N, 50 mL) was added to a stirred solution of *tert*-butyl 7,8-dihydro-1,6-naphthyridine-6(*SH*)-carboxylate (25.0 g, 0.107 mol) in MeOH (200 mL). The reaction mixture was refluxed for 2 h in an oil bath and evaporated. The solid residue was treated with hot dry MeOH (100 mL), cooled, filtered, washed with MeOH (2 × 20 mL), and dried to give the desired product as a white powder. Yield = 20.4 g (92%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.33 (brs, 2H), 8.75 (d, *J* = 5.5 Hz, 1H), 8.33 (d, *J* = 8.0 Hz, 1H), 7.84 (dd, *J* = 8.0, 5.5 Hz, 1H), 4.43 (s, 2H), 3.49 (brs, 2H), 3.40 (t, *J* = 6.2 Hz, 2H) (Py-H⁺ was not observed due to its strong broadening at 7.0–12.0). ¹³C{¹H} NMR (151 MHz, DMSO-*d*₆) δ 148.8, 142.8, 142.4, 129.7, 124.8, 42.6, 40.9, 25.0. LCMS, positive mode, *m/z*: 135 [M – 2HCl + H]⁺. Anal. calcd for C₈H₁₂Cl₂N₂: C, 46.40; H, 5.84; N, 13.53; Cl, 34.23. Found: C, 46.59; H, 5.63; N, 13.32; Cl, 33.95.

1,2,3,4-Tetrahydro-1,5-naphthyridine Chloride (**26**). HCl (6 N, 50 mL) was added to a stirred solution of *tert*-butyl 5,6,7,8-tetrahydro-1,6-naphthyridine-6-carboxylate (25.1 g, 0.107 mol) in MeOH (200 mL). The reaction mixture was refluxed for 2 h in an oil bath and evaporated. The solid residue was treated with hot dry MeOH (100 mL), cooled, filtered, washed with MeOH (2 × 20 mL), and dried to give the desired product as a white powder. Yield = 16.4 g (90%). ¹H NMR (400 MHz, CDCl₃) δ = 7.84 (d, *J* = 4.6, 1H), 6.87 (dd, *J* = 8.0, 4.7, 1H), 6.70 (dd, *J* = 8.2, 1.6, 1H), 3.91 (s, 1H), 3.31–3.24 (m, 2H), 2.92 (t, *J* = 6.5, 2H), 2.01 (p, *J* = 6.4, 2H). ¹³C{¹H} NMR (151 MHz, CDCl₃) δ = 142.7, 140.9, 137.8, 121.9, 120.1, 77.3, 77.1, 76.9, 41.4, 30.3, 21.7. LCMS, positive mode, *m/z*: 135 [M + H]. Anal. calcd for C₈H₁₀N₂: C, 71.61; H, 7.51; N, 20.88. Found: C, 71.80; H, 7.44; N, 20.83.

5,6,7,8-Tetrahydroquinolin-6-aminium Chloride (27). HCl (6 N, 50 mL) was added to a stirred solution of *tert*-butyl *N*-(5,6,7,8-tetrahydroquinolin-6-yl)carbamate (26.6 g, 0.107 mol) in MeOH (200 mL). The reaction mixture was refluxed for 2 h in an oil bath and evaporated. The solid residue was treated with hot dry MeOH (100 mL), cooled, filtered, washed with MeOH (2 × 20 mL), and dried to give the desired product as a white powder. Yield = 18.4 g (93%). ¹H NMR (400 MHz, DMSO- d_6) δ = 8.66 (d, *J* = 5.8, 4H), 8.36 (d, *J* = 7.9, 1H), 7.80 (dd, *J* = 7.9, 5.7, 1H), 3.37–3.24 (m, 2H), 3.28–3.12 (m, 1H), 3.06 (dd, *J* = 17.1, 9.1, 1H), 2.54 (s, 0H), 2.25 (s, 1H), 2.10–1.95

(m, 1H). ¹³C{¹H} NMR (101 MHz, DMSO- d_6) δ = 151.5, 146.0, 140.2, 133.9, 124.53, 44.9, 31.4, 25.6, 24.9. LCMS, positive mode, *m*/*z*: 149 [M + H]. Anal. calcd for C₉H₁₃ClN₂: C, 58.54; H, 7.10; N, 15.17; Cl, 19.20. Found: C, 58.90; H, 6.96; N, 15.02; Cl, 19.07.

7,8-Dihydroquinolin-6(5H)-one (28). Water (400 mL), 85% phosphoric acid (200.0 g), and 7',8'-dihydro-5'H-spiro[1,3]dioxolane-2,6'-quinoline (35.0 g, 0.183 mol) were added to a 1 L flask. The reaction mixture was stirred and heated to 80 °C for 8 h in an oil bath. The reaction solution was adjusted to pH = 6 with sodium carbonate solution and then extracted with EtOAc $(3 \times 200 \text{ mL})$. The extract was dried over anhydrous sodium sulfate, and the solvent was removed under reduced pressure. The crude product was purified by gravity column chromatography (hexane/EtOAc 7:3) to give the desired product as a white powder. Yield = 22.95 g (85%). Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (d, J = 5.0, 1H), 7.38 (d, J = 7.6 Hz, 1H), 7.12 (dd, J = 7.7, 4.9 Hz, 1H), 3.56 (s, 2H), 3.23 (t, J = 6.9 Hz, 2H), 2.62 (t, J = 6.9 Hz, 2H). ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 208.6, 156.7, 147.7, 136.0, 128.3, 122.1, 43.7, 37.7, 31.1. LCMS, positive mode, m/z: 148 [M + H]⁺. Anal. calcd for C₉H₉NO: C, 73.45; H, 6.16; N, 9.52. Found: C, 73.44; H, 6.28; N, 9.35.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.0c03038.

Copies of ¹H and ¹³C NMR spectra of all new compounds (PDF)

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Notes

The authors declare no competing financial interest.

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