

Synthesis and antitumour activity of trimethylsilylpropyl substituted benzimidazoles

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Abstract – The quaternisation of *N*-substituted benzimidazoles by heating with various alkyl, allyl, propargyl and benzyl chlorides and bromides leads to the formation of benzimidazolium salts. The interaction of *N*-monosubstituted benzimidazoles with various salts (CuCl₂, ZnCl₂, CoCl₂, PdCl₂ and AgNO₃) yielded stable solid complexes. Potential cytotoxic activity of synthesised benzimidazolium salts and benzimidazole metal complexes was tested in vitro on four monolayer tumour cell lines: MG-22A (mouse hepatoma), HT-1080 (human fibrosarcoma), B16 (mouse melanoma), Neuro 2A (mouse neuroblastoma) and normal mouse fibroblast cells. A preliminary analysis of the structure–activity relationship for the benzimidazole derivatives clearly indicates that the character of substituents in the benzimidazole ring has strong influence on the cytotoxic activity. The insertion of the silicon atom into the *N*-alkyl chain increases the cytotoxic activity of benzimidazolium salts significantly, which show a very significant potency in vitro against all studied tumour cell lines, being particularly active in experiments with B16 (mouse melanoma). TD₅₀ for the most active compounds are in the range 0.001–0.008 µg ml⁻¹. Cytotoxicity of benzimidazole metal complexes (L₂MX₂) strongly depends on the metal nature. 1-(3-Trimethylsilylpropyl)benzimidazole in dose 1 mg kg⁻¹ inhibits carcinoma S-180 tumour growth by 62% (on ICR mice). © 2001 Éditions scientifiques et médicales Elsevier SAS

benzimidazoles / benzimidazolium salts / benzimidazole complexes / cytotoxic activity / antitumour activity

1. Introduction

The benzimidazole ring is an important pharmacophore in drug discovery [1]. Copper(II) and silver(I) complexes of 2-pyridyl-1H-benzimidazoles possess considerable activity against *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Shigella flexneri* and *Candida albicans* [2]. Some *N*-(nitrophenylsulphonyl)benzimidazoles show good antiviral properties against RNA (Coxsackie B5 and Mumps) viruses [3]. A series of 1,2,4-trisubstituted benzimidazoles was evaluated in vitro as potent and selective neuropeptide Y Y1 receptor antagonists. Within this series the activity was enhanced for compounds with two appropriately positioned aminoalkyl groups (e.g. 4-piperidinoalkyl

functionalities) at either N(1) or N(1) and C(4) positions of aromatic heterocycle [4–6]. *N*-Substituted benzimidazol-2-ones have low nanomolar affinity for the human neuropeptide Y Y5 receptor and behave as functional antagonists of the Y5 receptor [7]. To discover new immunosuppressive compounds, a variety of 5-alkoxy substituted 2-(4-pyridylmethylsulfinyl)benzimidazoles were synthesised and evaluated for their ability to inhibit protein tyrosine phosphatase activities. Enzymatic analysis with several phosphatases reveals that 5-isopropoxy-2-[(4-methylthiopyridyl-2)methylsulfinyl]benzimidazole shows high specific inhibition against CD45 [8]. 1-Methyl-2-phenylethynyl-3-(3-phenylpropyn-2-yl)-benzimidazolium tetrafluoroborate and trifluoromethanesulfonate salts cause extensive cleavage of super-coiled DNA when incubated under mild conditions. Frank single-strand DNA cleavage is observed at concentrations as low as 1 µM [9]. Cytotoxicity of

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2-alkyl, 2-aryl, and 2-piperazinyl benzimidazole-4,7-dione derivatives were tested against three cancer cell lines (mouse lymphocytic leukaemia P388, and human gastric carcinoma SNU-1 and SNU-16). These compounds showed potent cytotoxicity against all three cell lines tested and especially SNU-16 was sensitive to them. 2-Aryl and 2-piperazinyl benzimidazole-4,7-dione derivatives were more potent than mitomycin C against P388 and SNU-16 [10]. Significant cytotoxic activity against several human tumour cell lines was displayed by 5,6-dichloro-2-(tetrahydropyran-2-yl)-benzimidazole [11], 6-aziridinylbenzimidazoles [12], pyrido[1,2-*a*]benzimidazoles [13], 3-substituted pyrrolo[1,2-*a*]benzimidazoles [14] and 1-(2,6-difluorophenyl)-1H,3H-thiazolo[3,4-*a*]benzimidazole [15].

Taking into account extensive DNA-cleaving ability of benzimidazolium tetrafluoroborate and trifluoromethanesulfonate salts [9], it may be concluded that they have the potential to function as effective antitumour agents. These results prompted us to synthesise new silyl substituted benzimidazolium salts to investigate their *in vitro* cytotoxicity, *in vivo* antitumour activity and the influence on cell morphology.

2. Chemistry

The general synthetic route chosen for preparation of substituted benzimidazolium salts included two stages. Among the possible methods for synthesis of *N*-alkylsubstituted imidazoles phase-transfer catalysed (PTC) *N*-alkylation is one of the simplest and most convenient routes [16]. The alkylation of benzimidazole by 1-trimethylsilylprop-3-yl iodide, allyl bromide and heptyl bromide in a two-phase ben-

zene–potassium hydroxide system in the presence of 18-crown-6 used as phase-transfer catalyst (the molar ratio of benzimidazole:alkylating agent:18-crown-6 is 1:1:0.03) affords the corresponding *N*-alkyl substituted benzimidazoles **1–3** in high yields. The following quaternisation of compounds **1** and **2** by heating with various alkyl, allyl, propargyl and benzyl chlorides and bromides leads to the formation of benzimidazolium salts **4–25** (figure 1). In the case of quaternisation by various (chloromethyl)silanes (e.g. trimethyl(chloromethyl)silane, methyldi(2-furyl)-(chloromethyl)silane, methyldi(2-thienyl)(chloromethyl)silane), the reaction with *N*-trimethylsilylpropylbenzimidazole (**1**) proceeds with desilylation of trimethylsilyl group in the alkylating agent and formation of *N*-trimethylsilylpropyl-*N'*-methylbenzimidazolyl chloride (**4**) in good yield. The ¹H-NMR spectroscopic data obtained for compound **4** are similar to those of corresponding iodide **5** prepared by heating of benzimidazole **1** with methyl iodide.

N-Monosubstituted benzimidazoles **1** and **3** were used as ligands for synthesis of complexes **26–34** by reaction with salts CuCl₂, ZnCl₂, CoCl₂, PdCl₂ and AgNO₃ (molar ratio of benzimidazole **1** or **3** with salt 2:1) in ethanol. All obtained complexes L₂MX_{*n*} are stable solids, poorly soluble in common organic solvents.

3. Results and discussion

3.1. *In vitro* cytotoxic activity

Potential cytotoxic activity of synthesised benzimidazolium salts **6–25** and benzimidazole complexes **26–34** was tested *in vitro* on four monolayer tumour cell lines: MG-22A (mouse hepatoma), HT-1080 (human fibrosarcoma), B16 (mouse melanoma), Neuro 2A (mouse neuroblastoma) and normal mouse fibroblast cells. The experimental evaluations of cytotoxicity are presented in tables I and II. A preliminary analysis of the structure–activity relationship for the cytotoxic action clearly indicates the strong influence of the substituents in the benzimidazole heterocycle on toxic effects *in vitro*. Initial *N*-trimethylsilylpropylbenzimidazole (**1**) exhibits high cytotoxic activity (TD₅₀ = 0.55–2.13 μg ml⁻¹) against all cell lines studied. The following quaternisation of this compound by various alkyl, allyl, propargyl and benzyl halides as a rule increases cytotoxic effect, which is especially expressed in the case of mouse

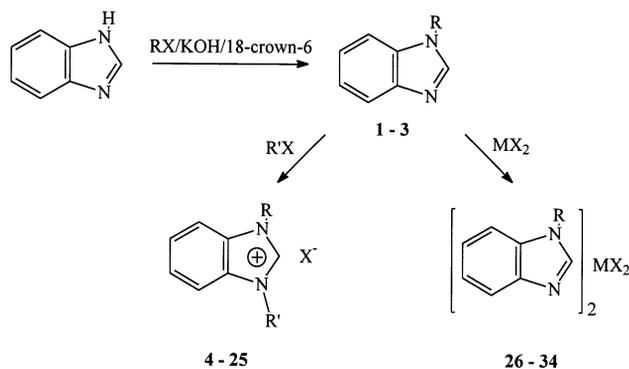


Fig. 1. Synthesis of benzimidazolium salts **4–25** and benzimidazole metal complexes **26–34**.

Table I. Cytotoxic activity of benzimidazolyl salts (TD₅₀, µg ml⁻¹).

N°	R	R'	X	HT - 1080		MG - 22A		B16		Neuro 2A		3T3
				TD ₅₀ ^a	NO, % CV ^b	TD ₅₀ ^a						
1	2	3	4	5	6	7	8	9	10	11	12	
1	Me ₃ Si(CH ₂) ₃			1.2	-	2.13	-	0.6	12	0.55	12	>1
6	Me ₃ Si(CH ₂) ₃	C ₂ H ₅	Br	0.2	250	0.45	150	0.001	40	0.5	13	0.5
7	Me ₃ Si(CH ₂) ₃	C ₉ H ₁₉	Cl	0.07	133	0.03	150	0.008	44	0.6	200	0.25
8	Me ₃ Si(CH ₂) ₃	Me ₃ Si(CH ₂) ₃	Cl	0.25	150	0.5	250	0.001	50	0.55	23	0.35
9	Me ₃ Si(CH ₂) ₃	(Me ₃ SiO) ₃ Si(CH ₂) ₃	Cl	0.2	200	0.65	250	0.008	50	0.6	21	0.55
10	Me ₃ Si(CH ₂) ₃		Cl	0.55	54	0.75	21	0.25	75	3	300	0.2
11	Me ₃ Si(CH ₂) ₃	CH ₂ =CHCH ₂	Br	0.35	150	0.35	200	0.002	30	0.8	8	0.5
12	Me ₃ Si(CH ₂) ₃	HC≡CCH ₂	Br	0.4	150	0.35	75	0.02	3	2	6	0.6
13	Me ₃ Si(CH ₂) ₃	Me ₃ SiC≡CCH ₂	Br	2.3	100	0.9	100	0.25	200	4.5	43	
14	Me ₃ Si(CH ₂) ₃	CH ₂ Ph	Br	1.4	200	1	25	0.005	29	0.85	9	0.8
15	Me ₃ Si(CH ₂) ₃		Br	0.15	200	0.35	200	0.003	40	0.85	11	0.6
16	Me ₃ Si(CH ₂) ₃		Br	0.45	100	0.3	180	0.14	150	0.85	150	
17	CH ₂ =CHCH ₂	CH ₃	I	98	6	*	3	*	5	62	31	-
18	CH ₂ =CHCH ₂	C ₂ H ₅	Br	66	19	87	11	*	20	-	-	-
19	CH ₂ =CHCH ₂	C ₄ H ₉	Cl	38	250	61	19	42	32	52	21	-
20	CH ₂ =CHCH ₂	C ₉ H ₁₉	Cl	0.65	125	0.4	175	0.4	300	0.75	250	4.7
21	CH ₂ =CHCH ₂	CH ₂ =CHCH ₂	Br	*	9	84	24	*	12	3.7	47	-
22	CH ₂ =CHCH ₂	HC≡CCH ₂	Br	43	167	100	14	*	8	4	130	-
23	CH ₂ =CHCH ₂	Me ₃ SiC≡CCH ₂	Br	14	300	82	12	70	14	-	-	-
24	CH ₂ =CHCH ₂	CH ₂ Ph	Br	21	300	45	21	29	200	-	-	-
25	CH ₂ =CHCH ₂		Br	4.2	200	32	83	5.9	300	1.8	550	>80

^a Concentration (µg/mL) providing 50% cell killing effect [(CV+MTT)/2]^b NO Concentration (%) (CV: coloration)

* No cytotoxic effect was detected.

- Not tested.

melanoma B16. In the series of trimethylsilylpropylbenzimidazolium salts **6–19** the TD₅₀ values change from 0.25 µg ml⁻¹ [R' = (C₄H₃S)₂MeSi(CH₂)₃ (**9**) and Me₃SiC≡CCH₂ (**13**)] to 0.001 µg ml⁻¹ [R' = Et (**6**) and Me₃Si(CH₂)₃ (**8**)] on B16 cell line. Methoxybenzyl

derivative **15** exhibits the highest cytotoxicity on B16 among benzylbenzimidazolium salts (**14–16**). For compounds with unsaturated substituent R' (salts **11–13**) the activity increases in the order: Me₃SiC≡CCH₂ < HC≡CCH₂ < CH₂=CHCH₂ for all cell lines. The

Table II. Cytotoxic activity of benzimidazole metal complexes **26–34** (TD₅₀, µg ml⁻¹).

No.	R	MX ₂	Colour	HT-1080		MG-22A		B16		Neuro 2A		3T3
				TD ₅₀ ^a	NO, %CV ^b	TD ₅₀ ^a						
26	Me ₃ Si(CH ₂) ₃	CuCl ₂	yellow	5	400	4.4	450	1.2	200	2	250	3.6
27	Me ₃ Si(CH ₂) ₃	ZnCl ₂	white	0.6	250	0.75	250	0.4	300	4.2	300	4.1
28	Me ₃ Si(CH ₂) ₃	CoCl ₂	blue	0.5	450	0.75	450	0.5	200	4.9	200	1.2
29	Me ₃ Si(CH ₂) ₃	PdCl ₂	grey	2	500	0.8	500	0.45	200	3.9	200	4.1
30	Me ₃ Si(CH ₂) ₃	AgNO ₃	white	2.9	250	3.6	250	1.5	350	5.4	250	2.2
31	C ₇ H ₁₅	CuCl ₂	yellow	1.5	450	2.5	300	1.8	333	0.25	1000	–
32	C ₇ H ₁₅	ZnCl ₂	white	35	300	21	300	–	–	–	–	–
33	C ₇ H ₁₅	CoCl ₂	blue	23	221	15	300	–	–	–	–	–
34	C ₇ H ₁₅	PdCl ₂	grey	52	400	33	158	–	–	–	–	–

^a Concentration (µg ml⁻¹) providing 50% cell killing effect [(CV+MTT)/2].

^b NO concentration (%) (CV: coloration).

–, Not tested.

influence of quaternisation is less effective on mouse hepatoma MG-22A (TD₅₀ = 1–0.03 µg ml⁻¹) and human fibrosarcoma HT-1080 (TD₅₀ = 0.55–0.07 µg ml⁻¹, with the exception of derivative **13**) being almost negligible on Neuro 2A cell line (TD₅₀ = 4.5–0.5 µg ml⁻¹). *N*-Allylbenzimidazole **2** derivatives bearing methyl (**17**), ethyl (**18**), butyl (**19**), allyl (**21**), propargyl (**22**), trimethylsilylpropargyl (**23**), and benzyl (**24**) substituent show a slight cytotoxic effect (*table I*). The substitution of a short alkyl chain by a nonyl group (salt **20**) leads to a considerable increase of activity (0.4 µg ml⁻¹ on mouse hepatoma MG-22A and mouse melanoma B16 cell lines). Comparison of the tumour growth inhibition for trimethylsilylpropyl **6–16** and allyl **17–25** benzimidazolium salts confirms a significant influence of the long lipophilic trimethylsilylpropyl group or alkyl chain on cytotoxic action.

Inspection of the results presented in *table II* for di(*N*-trimethylsilylpropylbenzimidazole) metal complexes **27–30** and its silicon free analogues **32–34** shows that the insertion of the silicon atom into the *N*-alkyl chain defines high level of cytotoxic activity. The only exception is di(*N*-trimethylsilylpropylbenzimidazole) copper dichloride **26** which is a weaker tumour growth inhibitor than the corresponding alkyl analogue **31**.

It has been found that the metal nature in the L₂MX_n complexes determine their in vitro activity. The inhibition ability of trimethylsilylpropyl substituted complexes **26–30** decreases in the following order: Zn > Co > Pd > Ag > Cu.

Under our experimental conditions several benzimidazolium salts (**1**, **11–13**) exhibit remarkable NO-inhibitor properties, however, other active *N*-trimethyl-

silylpropylbenzimidazole derivatives are medium NO-inducers. In general, benzimidazolium metal complexes **26–34** are more potent NO-inducers than the benzimidazolium salts **1–25**, di(*N*-heptylbenzimidazole) copper dichloride **31** being the most active (1000% in the mouse neuroblastoma Neuro 2A).

3.2. Cell morphology

The influence of complexes **26–28** on tumour cell morphology was examined. *Figure 2* presents the morphologic structure of human fibrosarcoma HT-1080 cells at 30 °C (control). The copper complex of *N*-trimethylsilylpropylbenzimidazole **26** induces the full necrosis of tumour cells (*figure 3*). In the case of zinc dichloride complex **27**, the cell size does not change after

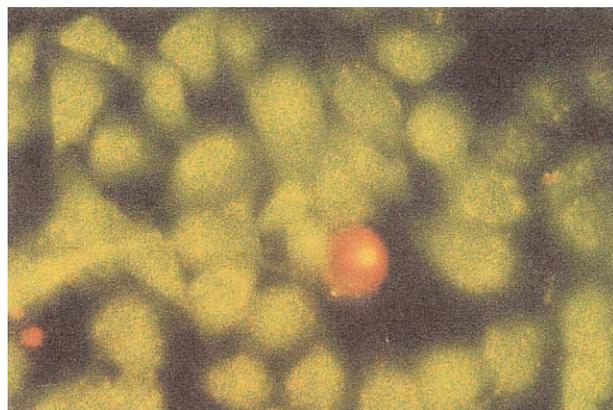


Fig. 2. View of human fibrosarcoma HT-1080 cell fenotype (30 °C, 24h).

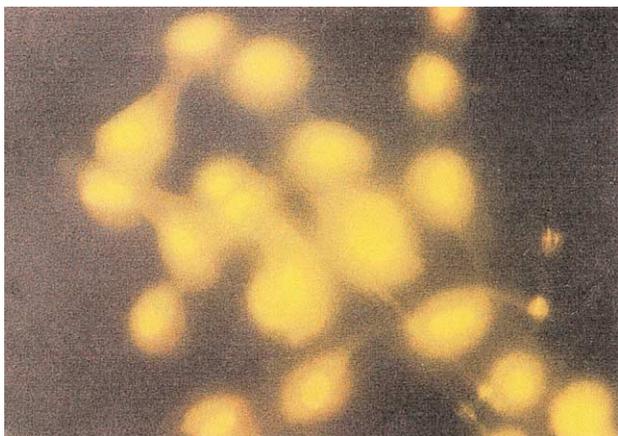


Fig. 3. View of HT-1080 cell phenotype with CU complex **26** (30 °C, 24 h).



Fig. 4. View of HT-1080 cell phenotype with Zn complex **27** (30 °C, 24 h).

apoptosis (figure 4). Action of complexes **26** and **28** results in HT-1080 cells culture necrosis with cell nucleus fragmentation (figure 5).

3.3. *In vivo* antitumour activity

The antitumour activity of *N*-trimethylsilylpropylbenzimidazole (**1**) was tested against sarcoma S-180 in male ICR mice (18–20 g). In dose 1 mg kg⁻¹ benzimidazole (**1**) inhibits the tumour growth by 62%. The higher concentration (3 mg kg⁻¹) decreases the tumour growth inhibition to 22% (figure 6).

4. Conclusions

N-Trimethylsilylpropylbenzimidazole (**1**), and its benzimidazolium salts and complexes show a very significant potency *in vitro* against different tumour cell lines, being particularly active in experiments with B16 (mouse melanoma) cell line (TD₅₀ for the most active compounds are in the range 0.001–0.008 μg ml⁻¹). *In vivo* evaluation of benzimidazole (**1**) also confirms high antitumour activity against sarcoma S-180.

5. Experimental

¹H-NMR spectra were recorded on a Varian Mercury 200 spectrometer at 200.06 MHz at 303 K. The chemical shifts are given relative to TMS from solvent (CDCl₃) signal (δ_H = 7.25). The melting points

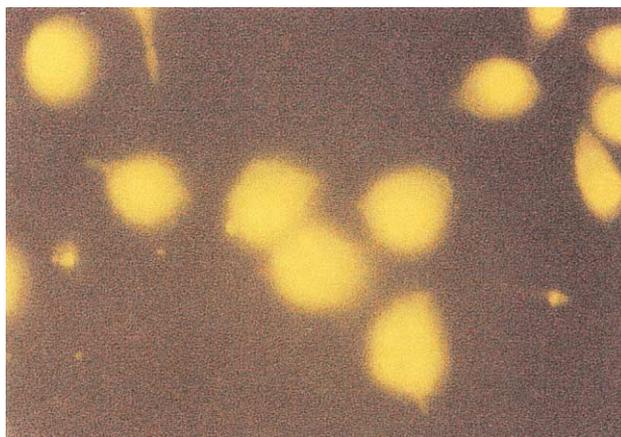


Fig. 5. View of HT-1080 cell phenotype with Co complex **28** (30 °C, 24 h).

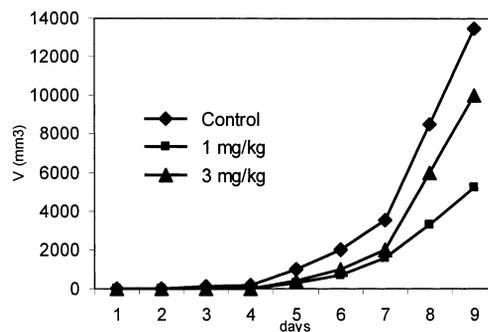


Fig. 6. Antitumour activity of 1-(3-trimethylsilylpropyl)-benzimidazole (**1**).

were determined on a 'Digital melting point analyser' (Fisher), and the results are given without correction.

5.1. Chemistry

5.1.1. General method for synthesis of *N*-substituted benzimidazoles **1–3**

The mixture of benzimidazole (0.02 mol), 1-trimethylsilylprop-3-yl iodide (allyl bromide or heptyl bromide) (0.02 mol), 18-crown-6 (0.6 mmol) and KOH (0.06 mol) in benzene (200 ml) was stirred for 6 h at reflux temperature (GLC control). After reaction completion the mixture was filtered. Benzene was evaporated and residue was dried in vacuum to give products as solids or oils.

5.1.1.1. *N*-(3-Trimethylsilylpropyl)benzimidazole (**1**)

Yield 83%, m.p. 63–64 °C. Impurities: <1% (HPLC on Symmetry C₁₈₃ 3.9×150 mm. Mobile phase: 80% acetonitrile+20% H₂O. Detector UV 254 nm). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 9H, Me₃Si), 0.4–0.64 (m, 2H, CH₂Si), 1.65–2.04 (m, 2H, CH₂), 4.15 (t, 2H, *J* = 7.6 Hz, CH₂N), 7.22–7.38 (m, 3H, arom), 7.33–7.89 (m, 2H, arom). Anal (C₁₃H₂₀N₂Si) C, H, N.

5.1.1.2. *N*-Allylbenzimidazole (**2**)

Yield 80%. Impurities: <1% (HPLC on Symmetry C₁₈, 3.9×150 mm. Mobile phase: 80% acetonitrile+20% H₂O. Detector UV 254 nm). ¹H-NMR (CDCl₃, TMS) δ (ppm): 4.73 (dt, 2H, *J* = 5.4 Hz, *J* = 1.4 Hz, CH₂N), 5.09–5.26 (m, 2H, CH₂), 5.88–6.04 (m, 1H, CH), 7.24–7.36 (m, 4H, arom), 7.87 (s, 1H, CH). Anal (C₁₀H₁₀N₂) C, H, N.

5.1.1.3. *N*-Heptylbenzimidazole (**3**)

Yield 80%. Impurities: <1% (HPLC on Symmetry C₁₈, 3.9×150 mm. Mobile phase: 80% acetonitrile+20% H₂O. Detector UV 254 nm). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.85 (t, 3H, *J* = 6.8 Hz Me), 1.27 (m, 8H, (CH₂)₄), 1.85 (m, 2H, CH₂), 4.12 (t, 2H, *J* = 7.0 Hz, CH₂N), 7.24–7.29 (m, 3H, arom), 7.78–7.82 (m, 1H, arom), 7.88 (s, 1H, CH). Anal (C₁₄H₂₀N₂) C, H, N.

5.1.2. Preparation of benzimidazolium salts

The mixture of *N*-substituted benzimidazole (0.02 mol) in toluene and alkylhalogenide (0.02 mol) was refluxed with stirring for about 4 h. Then it was allowed to stand at room temperature overnight to give a solid product. The precipitate was filtered, washed with toluene and recrystallised from ethanol.

5.1.2.1. *N*-Methyl-*N'*-(3-trimethylsilylpropyl)-benzimidazolyl chloride (**4**)

Yield 67%, m.p. 221 °C, TLC *R*_f = 0.36 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 18H, Me₃Si), 0.45–0.65 (m, 2H, CH₂Si), 1.85–2.10 (m, 2H, CH₂), 4.44 (s, 3H, CH₃N), 4.67 (t, 6H, *J* = 7.8 Hz, CH₂N), 7.6–7.71 (4H, m, arom), 11.67 (s, 1H, CH). Anal. (C₂₂H₃₉ClN₂Si) C, H, N.

5.1.2.2. *N*-Methyl-*N'*-(3-trimethylsilylpropyl)-benzimidazolyl iodide (**5**)

Yield 87%, m.p. 98–99 °C, TLC *R*_f = 0.38 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.07 (s, 18H, Me₃Si), 0.60–0.69 (m, 2H, CH₂Si), 2.07–2.12 (m, 2H, CH₂), 4.39 (s, 3H, CH₃N), 4.71 (t, 6H, *J* = 7.8 Hz, CH₂N), 7.77–7.81 (m, 4H, arom), 11.20 (s, 1H, CH). Anal (C₁₄H₂₃IN₂Si) C, H, N.

5.1.2.3. *N*-(3-Trimethylsilylpropyl)-*N'*-(ethyl)-benzimidazolyl bromide (**6**)

Yield 85%, m.p. 207–208 °C, TLC *R*_f = 0.70 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.03 (s, 9H, Me₃Si), 0.48–0.7 (m, 2H, CH₂Si), 1.86–2.28 (m, 4H, CH₃+CH₂), 4.67 (t, 4H, *J* = 7.6 Hz, CH₂N+CH₂N), 7.67–7.76 (m, 4H, arom), 11.51 (s, 1H, CH). Anal (C₁₅H₂₅BrN₂Si) C, H, N.

5.1.2.4. *N*-Nonyl-*N'*-(3-trimethylsilylpropyl)-benzimidazolyl chloride (**7**)

Yield 80%, m.p. 164–165 °C, TLC *R*_f = 0.43 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 18H, Me₃Si); 0.42–0.66 (m, 2H, CH₂Si); 0.90 (t, 3H, *J* = 6.3 Hz, CH₃); 1.20 (m, 14H, (CH₂)₇); 1.90–2.10 (m, 2H, CH₂); 4.67 (t, 6H, *J* = 7.8 Hz, CH₂N); 7.40 (m, 4H, arom); 11.68 (s, 1H, CH). Anal (C₂₂H₃₉ClN₂Si) C, H, N.

5.1.2.5. *N,N'*-Bis(3-trimethylsilylpropyl)benzimidazolyl chloride (**8**)

Yield 76%, m.p. >250 °C; TLC *R*_f = 0.53 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 18H, Me₃Si), 0.4–0.64 (m, 4H, CH₂Si), 1.84–2.04 (m, 4H, CH₂), 4.62 (t, 4H, *J* = 7.8 Hz, CH₂N), 7.40 (m, 4H, arom), 11.62 (s, 1H, CH). Anal (C₁₉H₃₅ClN₂Si₂) C, H, N.

5.1.2.6. *N*-(3-Trimethylsilylpropyl)-*N'*-[3-tris(trimethylsilyloxy)silylpropyl]benzimidazolyl chloride (**9**)

Yield 96%, m.p. 94–95 °C (from benzene). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 9H, Me₃Si), 0.03 (s, 27H, Me₃SiO), 0.51–0.61 (m, 4H, CH₂Si), 1.93–2.01 (m,

4H, CH₂), 4.10 (m, 4H, CH₂N), 7.54–7.70 (m, 4H, arom), 11.61 (s, 1H, CH). Anal (C₂₅H₅₃ClN₂O₃Si₅) C, H, N.

5.1.2.7. *N*-(3-Trimethylsilylpropyl)-*N'*-[3-methyl-di-(2-thienyl)silylpropyl]benzimidazolyl chloride (**10**)

Yield 76%, m.p. 150–151 °C, TLC R_f = 0.53 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 12H, Me₃Si, SiMe), 0.4–0.64 (m, 2H, CH₂Si), 1.15–1.20 (m, 2H, CH₂Si), 1.94–2.10 (m, 4H, CH₂), 4.66 (m, 4H, CH₂N), 7.20–7.29 (m, 4H, arom), 7.47–7.67 (m, 6H, arom), 11.67 (s, 1H, CH). Anal (C₂₅H₃₅ClN₂S₂Si₂) C, H, N.

5.1.2.8. *N*-(3-Trimethylsilylpropyl)-*N'*-(allyl)-benzimidazolyl bromide (**11**)

Yield 91%, m.p. 124–125 °C, TLC R_f = 0.54 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.04 (s, 9H, Me₃Si), 0.47–0.64 (m, 2H, CH₂Si), 1.96–2.12 (m, 4H, CH₂, CH₂), 4.65 (dt, 2H, J = 4, J = 7.6 Hz, CH₂N), 5.3–5.56 (m, 2H, CH₂N), 7.64–7.75 (m, 4H, arom), 11.47 (s, 1H, CH). Anal (C₁₆H₂₅BrN₂Si) C, H, N.

5.1.2.9. *N*-(3-Trimethylsilylpropyl)-*N'*-(propargyl)-benzimidazolyl bromide (**12**)

Yield 90%, m.p. 164–168 °C; TLC R_f = 0.57 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.03 (s, 9H, Me₃Si), 0.46–0.72 (m, 2H, CH₂Si), 1.88–2.15 (m, 2H, CH₂), 2.64 (t, 2H, J = 3 Hz, CH₂), 4.61 (m, 2H, CH₂N); 5.71 (d, 1H, J = 3 Hz, CH), 7.55–7.97 (m, 4H, arom), 11.57 (s, 1H, CH). Anal (C₁₆H₂₃BrN₂Si) C, H, N.

5.1.2.10. *N*-(3-Trimethylsilylpropyl)-*N'*-(trimethylsilylpropargyl)benzimidazolyl bromide (**13**)

Yield 90%, m.p. 90–191 °C, TLC R_f = 0.34 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 9H, Me₃Si), 0.14 (s, 9H, Me₃Si), 0.46–0.62 (m, 2H, CH₂Si), 1.88–2.15 (m, 2H, CH₂), 4.56 (m, 2H, CH₂N), 5.65 (s, 2H, CH₂N), 7.67–7.72 (m, 4H, arom), 11.54 (s, 1H, CH). Anal (C₁₉H₃₁BrN₂Si₂) C, H, N.

5.1.2.11. *N*-(3-Trimethylsilylpropyl)-*N'*-benzylbenzimidazolyl bromide (**14**)

Yield 85%, m.p. 128–130 °C, TLC R_f = 0.38 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.04 (s, 9H, Me₃Si), 0.44–0.7 (m, 2H, CH₂Si), 1.90–2.06 (m, 2H, CH₂), 4.48 (t, 2H, J = 7.6 Hz, CH₂N), 5.93 (s, 2H, CH₂), 7.35–7.7 (m, 9H, arom), 11.71 (s, 1H, CH). Anal (C₂₀H₂₇BrN₂Si) C, H, N.

5.1.2.12. *N*-(3-Trimethylsilylpropyl)-*N'*-(*o*-methoxybenzyl)benzimidazolyl bromide (**15**)

Yield 85%, m.p. 175–176 °C, TLC R_f = 0.46 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.04 (s, 9H, Me₃Si), 0.46–0.75 (m, 2H, CH₂Si), 1.95–2.26 (m, 2H, CH₂), 3.94 (s, 3H, MeO), 4.73 (t, 2H, J = 7.6 Hz, CH₂N), 5.80 (s, 2H, CH₂); 6.92–7.7 (m, 9H, arom); 11.42 (s, 1H, CH). Anal (C₂₁H₂₉BrN₂O₂Si) C, H, N.

5.1.2.13. *N*-(3-Trimethylsilylpropyl)-*N'*-(*p*-bromobenzyl)benzimidazolyl bromide (**16**)

Yield 75%, m.p. 154–155 °C, TLC R_f = 0.44 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.02 (s, 9H, Me₃Si), 0.44–0.71 (m, 2H, CH₂Si), 1.90–2.28 (m, 2H, CH₂), 4.62 (q, 2H, J = 6.4 Hz, CH₂N), 5.98 (s, 2H, CH₂), 7.51–7.86 (m, 8H, arom), 11.53 (s, 1H, CH). Anal (C₁₆H₂₅Br₂N₂Si) C, H, N.

5.1.2.14. *N*-Methyl-*N'*-allylbenzimidazolyl iodide (**17**)

Yield 91%, m.p. 144–145 °C, TLC R_f = 0.19 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 4.29 (s, 3H, CH₃), 5.27–5.30 (m, 2H, CH₂), 5.46–5.60 (m, 2H, CH₂), 6.10–6.19 (m, 1H, CH), 7.63–7.83 (m, 4H, arom), 10.75 (s, 1H, CH). Anal (C₁₁H₁₃N₂I) C, H, N.

5.1.2.15. *N*-Ethyl-*N'*-allylbenzimidazolyl bromide (**18**)

Yield 85%, m.p. 174–175 °C, TLC R_f = 0.21 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 1.74 (t, 3H, J = 7.4 Hz, CH₃), 1.69 (t, 2H, J = 7.4 Hz, CH₂), 5.34–5.37 (m, 2H, CH₂), 5.43–5.56 (m, 2H, CH₂), 6.07–6.21 (m, 1H, CH), 7.63–7.83 (m, 4H, arom), 11.19 (s, 1H, CH). Anal (C₁₂H₁₅N₂Br) C, H, N.

5.1.2.16. *N*-Butyl-*N'*-allylbenzimidazolyl chloride (**19**)

Yield 85%, m.p. 95–97 °C, TLC R_f = 0.21 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.98 (t, 3H, J = 7.2 Hz, CH₃), 1.40–1.47 (m, 2H, CH₂), 1.97–2.03 (m, 2H, CH₂), 4.59–4.68 (m, 2H, CH₂N), 5.33–5.38 (m, 2H, CH₂), 5.42–5.53 (m, 2H, CH₂), 6.04–6.13 (m, 1H, CH), 7.65–7.76 (m, 4H, arom), 11.61 (s, 1H, CH). Anal (C₁₄H₁₉N₂Cl) C, H, N.

5.1.2.17. *N*-Nonyl-*N'*-allylbenzimidazolyl chloride (**20**)

Yield 88%, m.p. 95–97 °C; TLC R_f = 0.21 (2-propanol). ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.98 (t, 3H, J = 6.4 Hz, CH₃), 1.23–1.36 (m, 10H, CH₂), 2.05 (m, 2H, CH₂), 2.59 (m, 2H, CH₂), 4.58–4.64 (m, 2H, CH₂N), 5.36–5.39 (m, 2H, CH₂); 5.42–5.51 (2H, m, CH₂); 6.10–6.13 (1H, m, CH); 7.66–7.71 (4H, m, arom); 11.57 (1H, s, CH). Anal (C₁₉H₂₉N₂Cl) C, H, N.

5.1.2.18. *N,N'*-Bis(allyl)benzimidazolyl bromide (**21**)

Yield 78%, m.p. 95–97 °C, TLC R_f = 0.21 (2-propanol). $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 5.28–5.31 (m, 4H, CH_2 , CH_2), 5.41–5.51 (m, 4H, CH_2 , CH_2), 6.00–6.19 (m, 2H, CH, CH), 7.56–7.74 (m, 4H, arom), 11.31 (s, 1H, CH). Anal ($\text{C}_{13}\text{H}_{15}\text{N}_2\text{Br}$) C, H, N.

5.1.2.19. *N*-Allyl-*N'*-(3-propargyl)benzimidazolyl bromide (**22**)

Yield 83%, m.p. 177–178 °C, TLC R_f = 0.31 (2-propanol). $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 1.94 (d, 1H, J = 1.4 Hz, CH), 5.29–5.32 (m, 2H, CH_2), 5.48–5.86 (m, 2H, CH_2), 5.66–5.68 (m, 2H, CH_2), 6.03–6.21 (m, 1H, CH), 7.64–7.97 (m, 4H, arom), 11.45 (s, 1H, CH). Anal ($\text{C}_{13}\text{H}_{13}\text{N}_2\text{Br}$) C, H, N.

5.1.2.20. *N*-Allyl-*N'*-(3-trimethylsilylpropargyl)benzimidazolyl bromide (**23**)

Yield 88%, m.p. 144–145 °C, TLC R_f = 0.26 (2-propanol). $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 0.14 (s, 9H, Me_3Si), 5.29–5.32 (m, 2H, CH_2), 5.47–5.49 (m, 2H, CH_2), 5.57–5.61 (m, 2H, CH_2), 6.04–6.21 (m, 1H, CH), 7.64–7.95 (m, 4H, arom), 11.37 (s, 1H, CH). Anal ($\text{C}_{16}\text{H}_{21}\text{N}_2\text{BrSi}$) C, H, N.

5.1.2.21. *N*-Allyl-*N'*-benzylbenzimidazolyl bromide (**24**)

Yield 82%, m.p. 200–201 °C, TLC R_f = 0.23 (2-propanol). $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 5.29–5.32 (m, 2H, CH_2), 5.45–5.49 (m, 2H, CH_2), 5.89 (s, 2H, CH_2), 6.04–6.20 (m, 1H, CH), 7.27–7.59 (m, 9H, arom), 11.66 (s, 1H, CH). Anal ($\text{C}_{17}\text{H}_{17}\text{N}_2\text{Br}$) C, H, N.

5.1.2.22. *N*-Allyl-*N'*-(*p*-bromobenzyl)benzimidazolyl bromide (**25**)

Yield 85%, m.p. 178–180 °C, TLC R_f = 0.24 (2-propanol). $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 5.26–5.29 (m, 2H, CH_2), 5.45–5.54 (m, 2H, CH_2), 5.94 (s, 2H, CH_2), 6.07–6.16 (m, 1H, CH), 7.47–7.72 (m, 8H, arom), 11.57 (s, 1H, CH). Anal ($\text{C}_{17}\text{H}_{16}\text{N}_2\text{Br}_2$) C, H, N.

5.1.3. Preparation of metal complexes in ethanol–water

A 5 ml aqueous solution of metal salts (0.02 mol) and 0.04 mol ligand in 10 ml of ethanol in a reaction tube was stirred vigorously. The mixture was refluxed by stirring for about 4 h. The mixture was then allowed to stand at room temperature overnight to give a solid product. This was then filtered, washed with water and ethanol, and dried in vacuo over anhydrous CaCl_2 .

5.1.3.1. Di[*N*-(3-trimethylsilylpropyl)benzimidazole] copper dichloride (**26**)

Yield 81%, m.p. 243–245 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): –0.03 (s, 9H, Me_3Si), 0.26–0.48 (m, 2H, CH_2Si), 1.91–2.16 (m, 2H, CH_2), 4.43 (t, 2H, J = 7.6 Hz, CH_2N), 7.22–8.38 (m, 4H, arom), 9.78 (s, 1H, arom). Anal ($\text{C}_{26}\text{H}_{40}\text{N}_4\text{Cl}_2\text{CuSi}_2$) C, H, N.

5.1.3.2. Di[*N*-(3-trimethylsilylpropyl)benzimidazole] zinc dichloride (**27**)

Yield 85%, m.p. 154–156 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): –0.04 (s, 9H, Me_3Si), 0.31–0.48 (m, 2H, CH_2Si), 1.80–2.06 (m, 2H, CH_2), 4.30 (t, 2H, J = 7.2 Hz, CH_2N), 7.27–8.11 (m, 4H, arom), 8.53 (s, 1H, arom). Anal ($\text{C}_{26}\text{H}_{40}\text{N}_4\text{Cl}_2\text{Si}_2\text{Zn}$) C, H, N.

5.1.3.3. Di[*N*-(3-trimethylsilylpropyl)benzimidazole] cobalt dichloride (**28**)

Yield 78%, m.p. 140–143 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): –0.05 (9H, 5, Me_3Si); 0.31–0.48 (2H, m, CH_2Si); 1.64–1.98 (2H, m, CH_2); 5.62 (2H, t, J = 7.2 Hz, CH_2N); 7.27–8.04 (4H, m, arom); 7.76 (1H, s, arom). Anal ($\text{C}_{26}\text{H}_{40}\text{N}_4\text{Cl}_2\text{CoSi}_2$) C, H, N.

5.1.3.4. Di[*N*-(3-trimethylsilylpropyl)benzimidazole] palladium dichloride (**29**)

Yield 87%, m.p. 210–212 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): –0.03 (s, 9H, Me_3Si), 0.43–0.50 (m, 2H, CH_2Si), 1.81 (m, 2H, CH_2), 4.35 (t, 2H, J = 7.2 Hz, CH_2N), 7.38–8.37 (m, 4H, arom), 8.80 (s, 1H, arom). Anal ($\text{C}_{26}\text{H}_{40}\text{N}_4\text{Cl}_2\text{Si}_2\text{Pd}$) C, H, N.

5.1.3.5. Di[*N*-(3-trimethylsilylpropyl)benzimidazole] silver nitrate (**30**)

Yield 89%, m.p. 160–162 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): –0.02 (s, 9H, Me_3Si), 0.28–0.44 (m, 2H, CH_2Si), 1.90–2.12 (m, 2H, CH_2), 4.40 (t, 2H, J = 7.6 Hz, CH_2N), 7.12–8.38 (m, 4H, arom), 9.55 (s, 1H, arom). Anal ($\text{C}_{26}\text{H}_{40}\text{N}_5\text{AgO}_3\text{Si}_2$) C, H, N.

5.1.3.6. Di[*N*-(heptyl)benzimidazole] copper dichloride (**31**)

Yield 81%, m.p. 198–200 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 0.91 (t, 3H, J = 6 Hz, CH_3), 1.23 (m, 8H, $(\text{CH}_2)_4$), 1.81 (m, 2H, CH_2), 3.34 (m, 2H, CH_2N), 7.30–7.87 (m, 4H, arom), 8.22 (s, 1H, CH). Anal ($\text{C}_{28}\text{H}_{40}\text{N}_4\text{Cl}_2\text{Cu}$) C, H, N.

5.1.3.7. Di[*N*-(heptyl)benzimidazole] zinc dichloride (**32**)

Yield 80%, m.p. 134–136 °C; $^1\text{H-NMR}$ (CDCl_3 , TMS) δ (ppm): 0.82 (t, 3H, J = 6 Hz, CH_3), 1.20 (m,

8H, (CH₂)₄), 1.79 (m, 2H, CH₂), 4.36 (t, 2H, *J* = 7.1 Hz, CH₂N), 7.30–7.87 (m, 4H, arom), 8.64 (s, 1H, CH). Anal (C₂₈H₄₀N₄Cl₂Zn) C, H, N.

5.1.3.8. Di[*N*-(heptyl)benzimidazole] cobalt dichloride (33)

Yield 90%, m.p. 120–121 °C; ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.89 (t, 3H, *J* = 6 Hz, CH₃), 1.31 (m, 8H, (CH₂)₄), 2.53 (m, 2H, CH₂), 3.57 (m, 2H, CH₂N), 7.30–7.87 (m, 4H, arom), 8.12 (s, 1H, CH). Anal (C₂₈H₄₀N₄Cl₂Co) C, H, N.

5.1.3.9. Di[*N*-(heptyl)benzimidazole] palladium dichloride (34)

Yield 92%, m.p. 178–180 °C; ¹H-NMR (CDCl₃, TMS) δ (ppm): 0.83 (t, 3H, *J* = 6.6 Hz, CH₃), 1.22 (m, 8H, (CH₂)₄), 2.52 (m, 2H, CH₂), 4.37 (t, 2H, *J* = 7.1 Hz, CH₂N), 7.38–8.10 (m, 4H, arom), 8.74 (s, 1H, CH). Anal (C₂₈H₄₀N₄Cl₂Pd) C, H, N.

5.2. *In vitro* cytotoxicity assay

Monolayer tumour cell lines: MG-22A (mouse hepatoma), HT-1080 (human fibrosarcoma), B16 (mouse melanoma), Neuro 2A (mouse neuroblastoma) and normal mouse fibroblast cells were cultivated with standard medium DMEM without an indicator ('Sigma') supplemented with 10% heat inactivated foetal bovine serum ('Sigma'). After the ampoule was defrosted the cells were used only from 1 to 4 passage. Cells in the range of 2–5 × 10⁴ (cells ml⁻¹) (depending on line nature) were placed on 96-well plates immediately after compounds were inoculated to wells. The control cells without test compounds were cultivated on separate plates. The plates were cultivated for 72 h, 17 °C, 5% CO₂. A quantity of survived cells was determined using crystal violet (CV) or 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT). The quantity of live cells on control plates was taken in calculations for 100% [17, 18]. Concentration of NO was determined according to Gryess method (by NO₂ level in cultural medium). Sodium nitrite standard solution was used for the calibration curve [17].

5.3. *In vivo* activity assay

The compounds were tested *in vivo* against sarcoma S-180 cells. Sarcoma S-180 (5 × 10⁶ cells) was inoculated s.c. to male ICR mice (6 weeks old, 18–20 g) on day 0. Drugs were administrated i.p., and the treatment was started 4 h after tumour transplantation. The number of

mice used in each group was between six and ten. The efficacy of the treatment was estimated by the ellipsoid formula, and *V* of control group was taken in calculations for 100%.

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References

- [1] Spasov A.A., Iezhitsa I.N., Bugaeva L.I., Anisimova V.A., *Khim.-Farm. Zhurn.* 33 (5) (1999) 6–17.
- [2] Ülküseven B., Tavman A., Ötük G., *Metal-Based Drugs* 6 (1999) 163–167.
- [3] Garuti L., Roberti M., Cermelli C., *Bioorg. Med. Chem. Lett.* 9 (1999) 2525–2530.
- [4] Zarrinmayeh H., Nunes A.M., Ornstein P.L., Zimmerman D.M., Arnold M.B., Schober D.A., Gackenheimer S.L., Bruns R.F., Hipskind P.A., Britton T.C., Cantrell B.E., Gehlert D.R., *J. Med. Chem.* 41 (1998) 2709–2719.
- [5] Zimmerman D.M., Cantrell B.E., Smith E.C.R., Nixon J.A., Bruns R.F., Gitter B., Hipskind P.A., Ornstein P.L., Zarrinmayeh H., Britton T.C., Schober D.A., Gehlert D.R., *Bioorg. Med. Chem. Lett.* 8 (1998) 473–476.
- [6] Zarrinmayeh H., Zimmerman D.M., Cantrell B.E., Schober D.A., Bruns R.F., Gackenheimer S.L., Ornstein P.L., Hipskind P.A., Britton T.C., Gehlert D.R., *Bioorg. Med. Chem. Lett.* 9 (1999) 647–652.
- [7] McNally J.J., Youngman M.A., Lovenberg T.W., Nepomuceno D., Wilson S., Dax S.L., *Bioorg. Med. Chem. Lett.* 10 (2000) 1641–1643.
- [8] Hamaguchi T., Takahashi A., Kagamizono T., Manaka A., Sato M., Osada H., *Med. Chem. Lett.* 10 (2000) 2657–2660.
- [9] David W.M., Kumar D., Kerwin S.M., *Med. Chem. Lett.* 10 (2000) 2509–2512.
- [10] Ahn C.N., Tak J.A., Choi S.J., *Arch. Pharm. Res.* 23 (2000) 288–301.
- [11] Novelli F., Tasso B., Sparatore F., Sparatore A., *Farmaco* 52 (1997) 499–507.
- [12] Ahn C.M., Kim S.K., Han J.L., *Arch. Pharm. Res.* 21 (1998) 599–609.
- [13] el-Hawash S.A., Badawey S.A., Kappe T., *Pharmazie* 54 (1999) 341–346.
- [14] Craigo W.A., LeSueur B.W., Skibo E.B., *J. Med. Chem.* 42 (1999) 3324–3333.
- [15] Grimaudo S., Tolomeo M., Chimirri A., Zappala M., Gancitano R.A., D'Alessandro N., *Eur. J. Cancer* 34 (1998) 1756–1763.
- [16] Sturkovich R., Goldberg Yu., Verovsky V., Augustane I., Prodanchuk N., Deineka S., Lukevics E., *Appl. Organomet. Chem.* 3 (1989) 393–399.
- [17] Fast D.J., Lynch R.C., Leu R.W., *J. Leukocyt. Biol.* 52 (1992) 255–261.
- [18] Freshney P.J., *Culture of Animal Cells (A Manual of Basic Technique)*, Wiley-Liss, New York, 1994, pp. 296–297.