

## Research Article

# Catalytic Allylic Chlorination of Natural Terpenic Olefins Using Supported and Nonsupported Lewis Acid Catalysts

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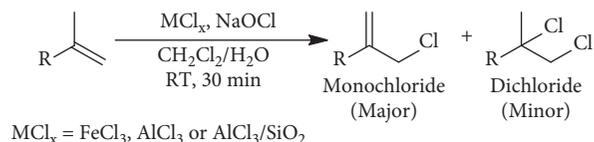
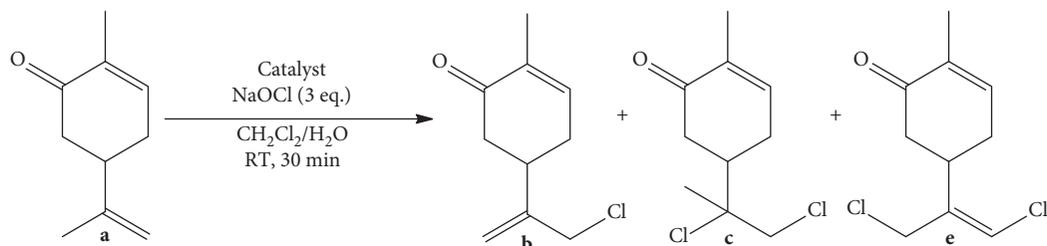
A mild and convenient method for the allylic chlorination of naturally occurring terpenic olefins was investigated in the presence of different supported and non-supported Lewis acid catalysts. The reaction has been tested on carvone as a model substrate in the presence of sodium hypochlorite as chlorine donor. The scope and limitations of transition metal-based Lewis acid catalysts, stoichiometry, and substrate structure were evaluated. Among the iron precursors used, FeCl<sub>3</sub> and FeCl<sub>2</sub> provide the promise of a general approach to allylic or vinylic chlorination reaction. Various terpenic olefins were examined in the presence of FeCl<sub>3</sub>/NaOCl combination system. The catalytic chlorination proceeds under mild conditions with short reaction time and shows a high selectivity affording the corresponding chlorides in good to excellent yields.

## 1. Introduction

Allyl, vinyl, or isopropenyl groups are present in different naturally occurring products as part of their structures [1–3]. They have been used repeatedly as starting materials to reach new natural products derivatives or more complex atomic arrangements [4, 5]. Among the versatile natural products bearing these groups, terpenes represent a sustainable supply of intermediates for several functionalization segments of the fine chemical industry, for example, the manufacture of flavors and fragrances [6–8]. Allylic chlorination represents a convenient way to functionalize terpenes bearing an allyl, vinyl, or isopropenyl group since further manipulation on the chloride may lead to several functional groups for the synthesis of natural products [9–13]. Previously, we have reported the allylic substitution of optically active natural terpenic allylic chloride derivatives in good yields [14, 15]. Moreover, isoprenoid chlorides are remarkably interesting

for the synthesis of  $\alpha$ -monoterpenes or vitamin A intermediates such as pseudoionone [16–18].

Allylic chlorination represents a convenient alternative method for allylic olefins functionalization. Various methodologies developing the allylic chlorination are reported in the literature, allowing the preparation of allyl chloride derivatives from the corresponding allylic alcohols using different reagents, such as thionyl chloride [19], hydrochloric acid [20], titanium (IV) chloride [21], N-chlorosuccinimide (NCS) [22], chlorosilanes [23, 24], methanesulfonyl chloride/lithium chloride [25], or iridium catalyst [26, 27]. Moreover, allylic chloride intermediates could be synthesized from aldehydes through olefination-reduction-halogenation sequences [28]. Torii et al. have reported electrochemical methods using sodium chloride as halogen source for the allylic chlorination of variety of isoprenoids [29, 30]. In organic synthesis, diselenides were also used as catalysts for the allylic chlorination of olefins

SCHEME 1: Allylic chlorination of terminal olefins catalyzed by  $\text{MCl}_x$ .

SCHEME 2: Allylic chlorination of carvone.

TABLE 1: Allylic chlorination of carvone using different Lewis acid catalysts.

Entry	Catalyst	Catalyst/substrate*	Conversion		Selectivity		
			a (%)	b (%)	c (%)	e (%)	
1	$\text{AlCl}_3/\text{SiO}_2$ 10% wt%	40% wt./wt.	25	6	4, 7	0	
2	$\text{AlCl}_3/\text{SiO}_2$ 10% wt%	3 eq. of $\text{AlCl}_3$	74	53	20	0	
3	$\text{AlCl}_3$	3	74	60	14	0	
4	$\text{FeCl}_3$	3	84	68	6	0	
5	$\text{FeCl}_2$	3	99	4	12	84	
6	$\text{Fe}(\text{NO}_3)_3$	3	99	33	15	52	
7	$\text{Fe}(\text{acac})_3$	3	0	0	0	0	
8	$\text{MoCl}_5$	3	99	75	10	14	

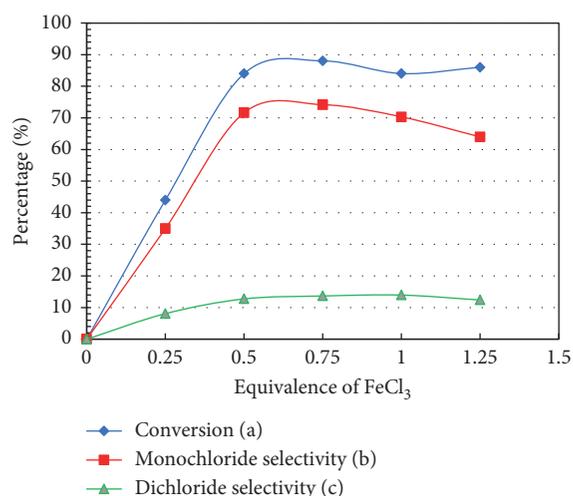
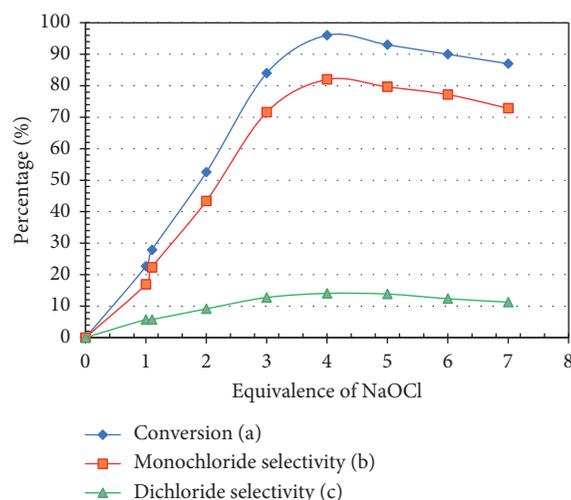
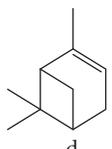
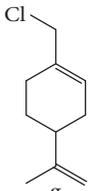
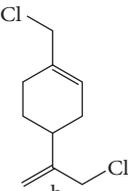
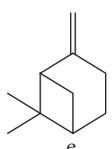
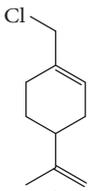
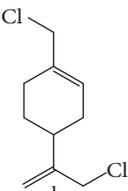
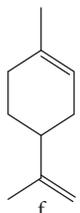
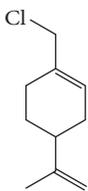
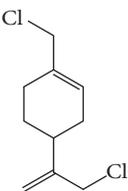
\*Reaction conditions: NaOCl (3 eq.),  $\text{CH}_2\text{Cl}_2/\text{H}_2\text{O}$  (1:1, 10 mL), RT, and 30 min.FIGURE 1: Effect of  $\text{FeCl}_3$  amount.

FIGURE 2: Effect of NaOCl amount.

[31–33]. Barrero et al. have reported a solid-phase selenium catalyst for the selective allylic chlorination of polyprenoids [34]. Recently, we have prepared an efficient new organoselenide for the allylic chlorination of various terpenic olefins [35]. In the presence of NCS, the allylic chloride can

be prepared from olefins using  $\text{Yb}(\text{OTf})_3\text{-TMSCl}$  or aniline catalyst [36, 37], while the allylic chlorination could be performed by a direct molecular chlorine bubbling through the reaction medium but suffer from disadvantages such as difficulties of handling chlorine gas [38]. On the other hand, reports are focused on the use of calcium hypochlorite

TABLE 2: Allylic chlorination of nonfunctionalized terpenic olefins<sup>a</sup>.

Entry	Substrate	Conversion <sup>b</sup> (%)	Product/isolated yield (%)	
1		99	 20%	 18%
2		99	 10%	 41%
3		99	 36%	 12%

<sup>a</sup>Reaction conditions: FeCl<sub>3</sub> (0.5 eq.), NaOCl (4 eq.), CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O (1:1, 10 mL), RT, and 30 min. <sup>b</sup>Conversion was determined by GC using dodecane as an internal standard.

[39, 40] or by a combination of Vilsmeier reagent and H<sub>2</sub>O<sub>2</sub>, but this excludes the use of acid-sensitive substrates [41]. In addition, sodium hypochlorite and acetic acid were used for the chlorine generation with limitation to nonsensitive substrates [42].

In the last two decades, the use of Brønsted acid (acetic acid) instead of Lewis acid has gained considerable importance. The method has the advantage of mild reaction conditions when the reaction occurs in a smoothly two-phase system (CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O) [38]. Different metallic Lewis acids in combination with NaOCl were studied for the allylic chlorination of terpenic olefins such as CeCl<sub>3</sub>, InCl<sub>3</sub>, or MoCl<sub>5</sub> [38, 43, 44]. The optimum solvent was reported to be the biphasic system of dichloromethane and water with addition of sodium hypochlorite under vigorous stirring to ensure the homogeneous distribution of the in situ generated electrophilic chlorine [38]. Moreover, Lewis acids such as NbCl<sub>5</sub> and NbBr<sub>5</sub> are reported to be efficient for the allylic and allenic chlorination via a mediated alkoxide rearrangements [28].

As part of our studies directed towards the valorization of natural terpenes via new catalytic systems [45–47], herein we report an efficient and convenient method for the allylic chlorination of terpenes using a combination of sodium hypochlorite and Lewis acid catalyst. The allylic chlorination

was achieved in a high degree of efficiency and selectivity. Among the Lewis acids used, aluminum and iron salts exhibit multiple interesting features such as their high abundance, low environmental impact, high chemoselectivity, and tolerance to aqueous media. The method represents a good choice for the preparation of new functionalized compounds derived from natural products under mild conditions (Scheme 1).

## 2. Results and Discussion

The scope and limitation of the allylic chlorination was examined first using carvone, chosen as model substrate, in the presence of supported and nonsupported Lewis acid catalysts (Scheme 2). The results are summarized in Table 1.

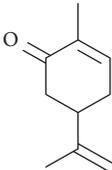
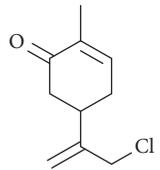
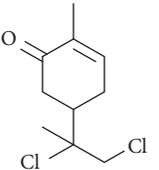
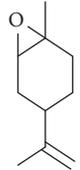
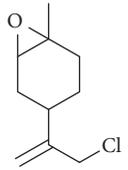
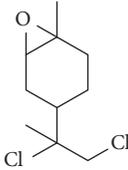
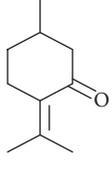
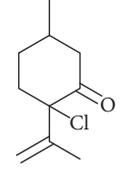
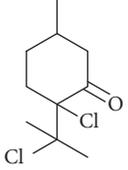
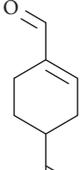
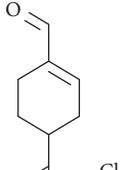
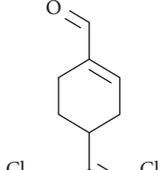
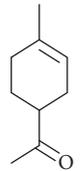
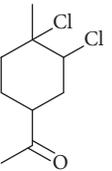
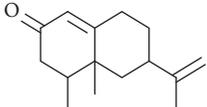
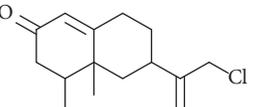
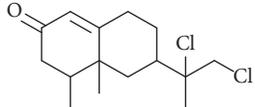
First, we have checked the reaction in the presence of a prepared supported catalyst AlCl<sub>3</sub>/SiO<sub>2</sub> 10% wt% (Table 1, entries 1-2). A slight conversion is observed when the reaction is carried out with a catalytic amount of a prepared supported catalyst AlCl<sub>3</sub>/SiO<sub>2</sub> 10% wt% (entry 1). However, a conversion of 74% with selectivity of 53% (b) and 10% (c) were obtained with a stoichiometric amount of AlCl<sub>3</sub>/SiO<sub>2</sub> 10% wt% (entry 2). From entries 1 and 2, we can assume that supported catalyst is less efficient for the allylic chlorination of terminal olefins due to the need of a stoichiometric amount of the Lewis acid.

The use of nonsupported catalyst AlCl<sub>3</sub> leads to similar result (entry 3). Table 1 shows that FeCl<sub>3</sub> was the most reactive and selective catalyst towards monochloride (b) (entry 4). The catalytic activity of different iron Lewis acid catalysts has been performed (entries 4–7). While the presence of FeCl<sub>3</sub> orients the reaction towards the formation of the allylic monochloride (b), FeCl<sub>2</sub> gave mainly the vinyl allyl dichloride (e) with 86% of yield (entry 5). The examination of iron(III) Lewis catalyst ligands effect indicates that with acetylacetonate practically no reaction took place which could be due to the ligand steric hindrance effect (entry 7). When the reaction was carried out with MoCl<sub>5</sub>, the corresponding monochloride (b) is formed as the major product (entry 8). It appeared that the molybdenum catalyst promotes the formation of allylic monochloride (b) with a moderate selectivity of dichloride (c) and vinyl allyl dichloride (e) contrarily to what was reported previously [44].

The catalytic chlorination of carvone (a) was performed by varying the amount of FeCl<sub>3</sub> (Figure 1). In the absence of Lewis acid catalyst, no reaction was observed even after stirring for a long reaction time. The increase of the FeCl<sub>3</sub> equivalence resulted in the increase of the conversion of carvone (a) with the formation of both the mono- and dichloride product. The best result was obtained with 0.5 equivalence of catalyst, and a conversion of 84% was obtained. The 0.75 equivalence amount reached a slight increase of conversion with no significant increase of selectivity.

The effect of NaOCl on the allylic chlorination reaction was evaluated using 0.5 eq. of FeCl<sub>3</sub> (Figure 2). In the absence of NaOCl as a chlorination agent, no reaction took place even in presence of Lewis acid catalyst. An excess of

TABLE 3: Allylic chlorination of functionalized terpenes<sup>a</sup>.

Entry	Substrate	Conversion <sup>b</sup> (%)	Product/isolated yield (%)	
1		96	 82%	 14%
2		93	 70%	 8%
3		76	 50%	 8%
4		99	 68%	 30%
5		85	 80%	
6		99	 80%	 17%

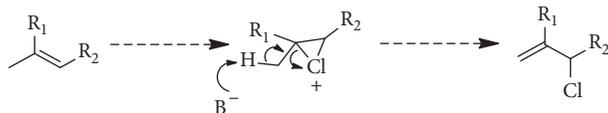
<sup>a</sup>Reaction conditions: FeCl<sub>3</sub> (0.5 eq.), NaOCl (4 eq.), CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O (1 : 1, 10 mL), RT, and 30 min. <sup>b</sup>Conversion was determined by GC using dodecane as an internal standard.

NaOCl up to 4 equivalence resulted in a maximum of conversion (96%) and a maximum of selectivity of (b) (82%).

According to the literature [38, 43, 44], the modest selectivity of dichlorinated product proved its presence for the first time in Lewis acid catalytic system. In order to confirm the formation of dichloride compound, a chlorination reaction starting from monochloride (b) was carried

out and no conversion was observed. Based on this result, we can predict that the dichlorinated product is formed by direct addition on the double bond.

Under the optimized conditions with FeCl<sub>3</sub> as a catalyst, allylic chlorination of various nonfunctionalized olefins has been carried out (Table 2). All the proposed terpenic olefins ( $\alpha$ -pinene d,  $\beta$ -pinene e, and limonene f) were converted



SCHEME 3: Proposed mechanism of allylic chlorination of terpenic olefins.

totally to the corresponding monochlorinated g and the dichlorinated h. At lower temperature of 0°C, no improvement on the selectivity is detected. In addition, a complex mixture was obtained.

To shed more light on the activity of  $\text{FeCl}_3$ , we have extended the allylic chlorination to a much more demanding functionalized terpenic olefins such as limonene oxide, pulegone, perillyl aldehyde, limona ketone and nootkatone (Table 3). The results depicted in Table 3 demonstrate that functionalized terpenic olefins were found to be more reactive than the nonfunctionalized ones. Except for limona ketone (entry 5), all substrates were converted to the corresponding allylic monochlorides as a major product (entries 1–4 and 6). It is noteworthy that perillyl aldehyde and carvone lead to the formation of a new vinyl allyl dichlorides in moderate yield (entries 5–6 (Table 1) and entry 4 (Table 3)). As vinyl chloride derivatives represent great interest in organic synthesis and biological activity [38, 48, 49], this procedure may serve as a tool for their synthesis in a simple one step. All isolated pure products were fully characterized by  $^1\text{H}$ ,  $^{13}\text{C}$  NMR, and mass spectroscopy (Supporting Information, Figures S1–S41).

Despite no reaction took place in the absence of Lewis acid catalyst, it has been reported that chlorine is usually generated from  $\text{NaOCl}$  [38, 43, 44]. The allylic chlorination reaction is probably based on the mild generation of electrophilic chlorine from  $\text{NaOCl}$  to a chlorination of the corresponding alkene. The loss of proton of the cationic intermediate leads to the formation of the major allylic chlorinated product (Scheme 3).

### 3. Conclusion

An efficient methodology for the catalytic allylic chlorination of naturally occurring terpenes using inexpensive and readily available Lewis acid catalysts combined with  $\text{NaOCl}$  has been investigated. The reaction was performed with a high degree of efficiency and selectivity. All the proposed terpenic olefins exhibit marked activity under mild conditions and lead to the corresponding mono- or dichlorides derivatives in good yields. Various supported and non-supported Lewis acid catalysts were studied. Different iron precursors have been checked and interesting results have been obtained with  $\text{FeCl}_3$  and  $\text{FeCl}_2$ . The reaction provides a useful entry to new functionalized terpenic olefin products.

### 4. General Procedure

In a typical procedure, terpenic olefin (1 mmol) in 10 mL of  $\text{CH}_2\text{Cl}_2$  is added to a vigorously stirred solution of  $\text{FeCl}_3$  (0.5 eq.) in 10 mL of  $\text{H}_2\text{O}$ . The mixture is vigorously stirred and a diluted  $\text{NaOCl}$  (4 eq.) is added dropwise for 5 min. After

30 min, a saturated aqueous  $\text{Na}_2\text{SO}_3$  solution is added and the mixture is extracted with  $\text{CH}_2\text{Cl}_2$  ( $3 \times 10$  mL). The organic layer is dried over anhydrous  $\text{Na}_2\text{SO}_4$ . Then, the solvent was removed under reduced pressure. The pure chlorinated products were obtained by column chromatography over silica gel using hexane and ethyl acetate as eluents. The isolated pure products were fully characterized by  $^1\text{H}$ ,  $^{13}\text{C}$  NMR, and GC-MS.

#### Carvone monochloride b

$^1\text{H}$  NMR (300 MHz)  $\delta$  6.69 (m, 1H, =CH), 5.19 (s, 1H, =CH<sub>2</sub>), 4.99 (s, 1H, =CH<sub>2</sub>), 4.03 (s, 2H, -CH<sub>2</sub>Cl), 2.9 (m, 1H, CH), 2.61 (m, 2H, CH<sub>2</sub>), 2.4 (m, 2H, CH<sub>2</sub>), 1.75 (s, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR (75 MHz)  $\delta$  198.8 (C=O), 146.5 (=C-), 144.1 (=CH-), 135.5 (=C-), 115.0 (=CH<sub>2</sub>), 46.9 (CH<sub>2</sub>Cl), 43.0 (CH<sub>2</sub>), 37.8 (CH), 31.3 (CH<sub>2</sub>), 15.6 (CH<sub>3</sub>). MS (EI):  $m/z$  = 184.0176 [M]<sup>+</sup>.

#### Carvone dichloride c

$^1\text{H}$  NMR (300 MHz)  $\delta$  7.21 (m, 1H, =CH), 3.45 (m, 2H, -CH<sub>2</sub>Cl), 2.5 (m, 2H, CH<sub>2</sub>), 2.3 (m, 1H, CH), 2.15 (m, 2H, CH<sub>2</sub>), 1.71 (s, 3H, -CH<sub>3</sub>), 1.19 (s, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR (75 MHz)  $\delta$  198.0 (C=O), 144.15 (=CH-), 135.30 (=C-), 72.69 (-C-Cl), 52.60 (CH<sub>2</sub>Cl), 41.64 (CH), 39.36 (CH<sub>2</sub>), 26.28 (CH<sub>2</sub>), 21.80 (CH<sub>3</sub>), 15.62 (CH<sub>3</sub>). MS (EI):  $m/z$  = 220.0758 [M]<sup>+</sup>.

#### Carvone vinyl allyl dichloride e

$^1\text{H}$  NMR (300 MHz)  $\delta$  6.74 (m, 1H, =CH), 6.12 (s, 1H, =CHCl), 4.20 (s, 2H, -CH<sub>2</sub>Cl), 3.00 (m, 1H, CH), 2.60 (m, 2H, CH<sub>2</sub>), 2.49 (m, 2H, CH<sub>2</sub>), 1.70 (s, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR (75 MHz)  $\delta$  197.97 (C=O), 143.69 (=CH-), 139.70 (=C-), 135.77 (=C-), 119.91 (=CHCl), 42.40 (-CH<sub>2</sub>Cl), 42.0 (CH<sub>2</sub>), 39.56 (CH), 31.15 (CH<sub>2</sub>), 15.62 (CH<sub>3</sub>). MS (EI):  $m/z$  = 218 [M]<sup>+</sup>.

#### Limonene oxide monochloride

$^1\text{H}$  NMR (300 MHz)  $\delta$  4.96 (s, 1H, =CH<sub>2</sub>), 4.95 (s, 1H, =CH<sub>2</sub>), 4.02 (s, 2H, -CH<sub>2</sub>Cl), 2.99 (m, 1H, -O-CH-), 2.36 (m, 1H, CH), 1.35–1.93 (m, 6H), 1.30 (s, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR (75 MHz)  $\delta$  148.76 (=C-), 113.65 (=CH<sub>2</sub>), 58.97 (O-CH), 57.12 (O-C), 47.37 (CH<sub>2</sub>Cl), 32.42 (CH), 30.70 (CH<sub>2</sub>), 28.46 (CH<sub>2</sub>), 24.66 (CH<sub>2</sub>), 22.96 (CH<sub>3</sub>). MS (EI):  $m/z$  = 185.0668 [M]<sup>+</sup>.

#### Limonene oxide dichloride

$^1\text{H}$  NMR (300 MHz)  $\delta$  3.55 (s, 2H, -CH<sub>2</sub>Cl), 3.02 (m, 1H, -O-CH-), 2.05 (m, 1H, CH), 2.01–1.50 (m, 6H), 1.33 (s, 3H, -CH<sub>3</sub>), 1.13 (s, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR (75 MHz)  $\delta$  73.37 (-C-Cl), 58.78 (O-CH), 57.66 (O-C), 53.23 (CH<sub>2</sub>Cl), 39.78 (CH), 30.58 (CH<sub>2</sub>), 24.89 (CH<sub>2</sub>), 22.85 (CH<sub>3</sub>), 22.96 (CH<sub>3</sub>), 20.45 (CH<sub>2</sub>). MS (EI):  $m/z$  = 220.9909 [M]<sup>+</sup>.

#### Pulegone monochloride

$^1\text{H}$  NMR (300 MHz)  $\delta$  5.20 (s, 1H, =CH<sub>2</sub>), 5.14 (s, 1H, =CH<sub>2</sub>), 2.35–2.80 (m, 2H, CH<sub>2</sub>), 2.30 (m, 2H, CH<sub>2</sub>), 1.96 (m, 1H, CH), 1.90 (m, 3H, -CH<sub>3</sub>), 1.84 (m, 2H, CH<sub>2</sub>), 1.06 (d, 3H, -CH<sub>3</sub>).  $^{13}\text{C}$  NMR

(75 MHz) d 203.76 (C=O), 143.44 (=C-), 115.25 (=CH<sub>2</sub>), 76.09 (-C-Cl), 45.54 (CH<sub>2</sub>), 37.97 (CH<sub>2</sub>), 34.39 (CH), 29.64 (CH<sub>2</sub>), 21.48 (CH<sub>3</sub>), 20.58 (CH<sub>3</sub>). MS (EI): m/z = 186.1188 [M]<sup>+</sup>.

#### Pulegone dichloride

<sup>1</sup>H NMR (300 MHz) d 2.38–2.86 (m, 2H, CH<sub>2</sub>), 2.15 (m, 1H, CH), 1.79–2.38 (m, 2H, CH<sub>2</sub>), 1.79 (m, 2H, CH<sub>2</sub>), 1.44 (s, 3H, -CH<sub>3</sub>), 1.35 (s, 3H, -CH<sub>3</sub>), 1.05 (d, 3H, -CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz) d 208.21 (C=O), 75.13 (-C-Cl), 74.96 (-C-Cl), 46.63 (CH<sub>2</sub>), 35.22 (CH<sub>2</sub>), 29.49 (CH<sub>2</sub>), 25.52 (CH), 21.99 (2CH<sub>3</sub>), 18.15 (CH<sub>3</sub>). MS (EI): m/z = 222.9984 [M]<sup>+</sup>.

#### Perillyl aldehyde monochloride

<sup>1</sup>H NMR (300 MHz) d 9.43 (s, 1H, HC=O), 6.83 (m, 1H, =CH), 5.23 (s, 1H, =CH<sub>2</sub>), 5.02 (s, 1H, =CH<sub>2</sub>), 4.09 (s, 2H, -CH<sub>2</sub>Cl), 2.02–2.53 (m, 2H, CH<sub>2</sub>), 2.25 (m, 1H, CH), 1.42–1.72 (m, 2H, CH<sub>2</sub>), 0.87–1.24 (m, 2H, -CH<sub>2</sub>). <sup>13</sup>C NMR (75 MHz) d 193.74 (HC=O), 149.82 (=CH-), 148.25 (=C-), 141.15 (=C-), 114.20 (=CH<sub>2</sub>), 47.56 (CH<sub>2</sub>Cl), 36.21 (CH), 32.13 (CH<sub>2</sub>), 26.51 (CH<sub>2</sub>), 21.46 (CH<sub>2</sub>). MS (EI): m/z = 184.0441 [M]<sup>+</sup>.

#### Perillyl aldehyde vinyl allyl dichloride

<sup>1</sup>H NMR (300 MHz) d 9.45 (s, 1H, HC=O), 6.82 (m, 1H, =CH), 6.12 (m, 1H, =CHCl), 4.29 (s, 2H, -CH<sub>2</sub>Cl), 1.58–2.59 (m, 2H, CH<sub>2</sub>), 1.99 (m, 1H, CH), 1.26 (m, 2H, -CH<sub>2</sub>), 0.87–2.30 (m, 2H, CH<sub>2</sub>). <sup>13</sup>C NMR (75 MHz) d 193.51 (HC=O), 148.79 (=CH-), 141.26 (=C-), 141.10 (=C-), 119.00 (=CHCl), 40.11 (CH<sub>2</sub>Cl), 37.86 (CH), 31.81 (CH<sub>2</sub>), 26.28 (CH<sub>2</sub>), 21.40 (CH<sub>2</sub>). MS (EI): m/z = 218.0015 [M]<sup>+</sup>.

#### Limona ketone dichloride

<sup>1</sup>H NMR (300 MHz) d 4.04 (m, 1H, -CHCl), 2.82 (m, 1H, CH), 1.89–2.35 (m, 2H, CH<sub>2</sub>), 2.17 (s, 3H, -CH<sub>3</sub>), 1.76–1.95 (m, 2H, CH<sub>2</sub>), 1.56–1.76 (m, 2H, CH<sub>2</sub>), 1.35 (s, 3H, -CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz) d 211.13 (C=O), 71.46 (-CCl), 64.75 (-CHCl), 44.64 (CH), 32.62 (CH<sub>2</sub>), 31.80 (CH<sub>2</sub>), 28.14 (CH<sub>3</sub>), 27.32 (CH<sub>3</sub>), 23.28 (CH<sub>2</sub>). MS (EI): m/z = 206.9165 [M]<sup>+</sup>.

#### Nootkatone monochloride

<sup>1</sup>H NMR (300 MHz) d 5.66 (m, 1H, =CH), 5.09 (s, 1H, =CH<sub>2</sub>), 4.90 (s, 1H, =CH<sub>2</sub>), 4.01 (s, 2H, -CH<sub>2</sub>Cl), 2.47 (m, 2H, CH<sub>2</sub>), 2.29 (m, 1H, CH), 2.15 (m, 2H, CH<sub>2</sub>), 1.91 (m, 2H, CH<sub>2</sub>), 1.27 (m, 1H, CH), 1.04–1.97 (m, 2H, CH<sub>2</sub>), 1.04 (s, 3H, -CH<sub>3</sub>), 0.87 (d, 3H, -CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz) d 199.08 (C=O), 169.58 (=C-), 148.67 (=C-), 124.74 (=CH-), 113.80 (=CH<sub>2</sub>), 47.70 (CH<sub>2</sub>Cl), 44.17 (CH<sub>2</sub>), 41.95 (CH<sub>2</sub>), 40.30 (CH), 39.35 (-C-), 35.72 (CH), 32.85 (CH<sub>2</sub>), 31.91 (CH<sub>2</sub>), 16.67 (CH<sub>3</sub>), 14.87 (CH<sub>3</sub>). MS (EI): m/z = 252.0591 [M]<sup>+</sup>.

#### Nootkatone dichloride

<sup>1</sup>H NMR (300 MHz) d 5.76 (m, 1H, =CH), 3.57 (m, 1H, CH<sub>2</sub>Cl), 3.64 (m, 1H, CH<sub>2</sub>Cl), 2.40 (m, 1H, CH), 2.23–2.50 (m, 2H, CH<sub>2</sub>), 2.06 (m, 2H, CH<sub>2</sub>), 1.99–2.29

(m, 2H, CH<sub>2</sub>), 1.87–2.12 (m, 2H, CH<sub>2</sub>), 1.25 (m, 1H, CH), 1.19 (s, 3H, -CH<sub>3</sub>), 1.10 (m, 3H, -CH<sub>3</sub>), 0.98 (m, 3H, -CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz) d 199.50 (C=O), 170.08 (=C-), 124.62 (=CH-), 73.33 (-CCl), 53.47 (CH<sub>2</sub>Cl), 42.04 (CH<sub>2</sub>), 40.53 (CH), 39.96 (CH), 39.68 (CH), 32.72 (CH<sub>2</sub>), 27.70 (CH<sub>2</sub>), 26.67 (CH<sub>2</sub>), 21.22 (CH<sub>3</sub>), 16.75 (CH<sub>3</sub>), 14.96 (CH<sub>3</sub>). MS (EI): m/z = 288.0343 [M]<sup>+</sup>.

### Data Availability

<sup>1</sup>H and <sup>13</sup>C NMR and mass spectroscopy spectra used to support the findings of this study are available free of charge via the Internet in the Electronic Supporting Information (ESI).

### Disclosure

The present research work is a part of a thesis work of a Ph.D. student.

### Conflicts of Interest

The authors declare that they have no conflicts of interest.

### Supplementary Materials

Figure S1: <sup>1</sup>H NMR spectrum of product b. Figure S2: APT spectrum of product b. Figure S3: MS spectrum of product b. Figure S4: <sup>1</sup>H NMR spectrum of product c. Figure S5: APT spectrum of product c. Figure S6: MS spectrum of product c. Figure S7: <sup>1</sup>H NMR spectrum of product e. Figure S8: APT spectrum of product e. Figure S9: MS spectrum of product e. Figure S10: <sup>1</sup>H NMR spectrum of the limonene oxide monochloride. Figure S11: <sup>13</sup>C spectrum of the limonene oxide monochloride. Figure S12: DET 135 spectrum of the limonene oxide monochloride. Figure S13: MS spectrum of the limonene oxide monochloride. Figure S14: <sup>1</sup>H NMR spectrum of the limonene oxide dichloride. Figure S15: <sup>13</sup>C spectrum of the limonene oxide dichloride. Figure S16: DET 135 spectrum of the limonene oxide dichloride. Figure S17: MS spectrum of the limonene oxide dichloride. Figure S18: <sup>1</sup>H NMR spectrum of the pulegone monochloride. Figure S19: <sup>13</sup>C spectrum of the pulegone monochloride. Figure S20: DET 135 spectrum of the pulegone monochloride. Figure S21: MS spectrum of the pulegone monochloride. Figure S18: <sup>1</sup>H NMR spectrum of the pulegone dichloride. Figure S19: <sup>13</sup>C spectrum of the pulegone dichloride. Figure S20: DET 135 spectrum of the pulegone dichloride. Figure S21: MS spectrum of the pulegone dichloride. Figure S22: <sup>1</sup>H NMR spectrum of the perillyl aldehyde monochloride. Figure S23: APT spectrum of the perillyl aldehyde monochloride. Figure S24: DET 135 spectrum of the perillyl aldehyde monochloride. Figure S25: MS spectrum of the perillyl aldehyde monochloride. Figure S26: <sup>1</sup>H NMR spectrum of the perillyl aldehyde vinyl allyl chloride. Figure S27: APT spectrum of the perillyl aldehyde vinyl allyl chloride. Figure S28: DET 135 spectrum of the perillyl aldehyde vinyl allyl chloride. Figure S29: MS spectrum of the perillyl aldehyde vinyl allyl chloride. Figure S30: <sup>1</sup>H NMR

spectrum of the limona ketone dichloride. Figure S31: <sup>13</sup>C spectrum of the limona ketone dichloride. Figure S32: DET 135 spectrum of the limona ketone dichloride. Figure S33: MS spectrum of the limona ketone dichloride. Figure S34: <sup>1</sup>H NMR spectrum of the nootkatone monochloride. Figure S35: <sup>13</sup>C spectrum of the nootkatone monochloride. Figure S36: DET 135 spectrum of the nootkatone monochloride. Figure S37: MS spectrum of the nootkatone monochloride. Figure S38: <sup>1</sup>H NMR spectrum of the nootkatone dichloride. Figure S39: <sup>13</sup>C spectrum of the nootkatone dichloride. Figure S40: DET 135 spectrum of the nootkatone dichloride. Figure S41: MS spectrum of the nootkatone dichloride. (Supplementary Materials)

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