Novel Arylalkenylpropargylamines as Neuroprotective, Potent, and Selective Monoamine Oxidase B Inhibitors for the Treatment of Parkinson's Disease

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Supporting Information



ABSTRACT: To develop novel neuroprotective agents, a library of novel arylalkenylpropargylamines was synthesized and tested for inhibitory activities against monoamine oxidases. From this, a number of highly potent and selective monoamine oxidase B inhibitors were identified. Selected compounds were also tested for neuroprotection in in vitro studies with PC-12 cells treated with 6-OHDA and rotenone, respectively. It was observed that some of the compounds tested yielded a marked increase in survival in PC-12 cells treated with the neurotoxins. This indicates that these propargylamines are able to confer protection against the effects of the toxins and may also be considered as novel disease-modifying anti-Parkinsonian agents, which are much needed for the therapy of Parkinson's disease.

INTRODUCTION

Parkinson's disease (PD) is a debilitating, progressive, neurodegenerative disorder that predominantly affects the elderly population. It is the second most common neurodegenerative disease after Alzheimer's disease and is characterized by the progressive loss of dopaminergic neurons in the substantia nigra pars compacta. In fact, most therapies are aimed at increasing the concentration of dopamine in the brain. L-DOPA (Figure 1), itself or in combination with a peripheral L-DOPA decarboxylase inhibitor, is utilized predominantly for dopamine replacement, being a prodrug that increases the concentration of dopamine. Other strategies include the use of inhibitors that reduce the metabolism of dopamine, such as catechol-O-methyl transferase (COMT) and monoamine oxidase (MAO) inhibitors. Despite the many advances in the treatment of PD





to date, these therapies are only symptomatic in nature. However, in addition to symptomatic relief, what is urgently lacking is a therapy that is able to halt disease progression. As the etiology and pathomechanism of PD are complex and not fully understood, such a therapy would necessarily require protection of the dopaminergic neurons from cell death. To illustrate some advances in this regard, selegiline, a selective MAO-B inhibitor that is used in early stage of PD, has been demonstrated to have antioxidant and protective effects in certain experimental models of PD.^{1,2} However, selegiline is metabolized to L-amphetamine and L-methamphetamine, which though less psychoactive than their D-enantiomers, can result in cardiovascular implications and, at higher concentrations $(>10^{-7} \text{ M})$, can increase the rate of apoptosis.³

A more recently FDA-approved drug for PD, rasagiline, is an even more selective MAO-B inhibitor⁴ as compared to selegiline, and as its lacks amphetamine-like metabolites, it may offer a therapeutic advantage over selegiline. In addition, several studies have shown that rasagiline, due also to its propargylamine moiety, is able to counteract various steps of

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the apoptotic cascade, which ultimately prevents cell death. These include activation of caspase 3 and poly(ADP-ribose) polymerase-1 (PARP-1), translocation of glyceraldehyde-3phosphate dehydrogenase (GADPH), and nucleosomal DNA fragmentation.⁵ Rasagiline is also able to induce the expression of antiapoptotic proteins Bcl-2 and Bcl-X_L, while pro-apoptotic Bad and Bax proteins are downregulated.⁶⁻¹⁰ Despite these promising findings and other studies on the neuroprotective properties of rasagiline, clinical trials have failed to show a clear neuroprotective action, and this has consequently led to the rejection of rasagiline by the FDA¹¹ as a disease-modifying drug for PD. The fate of rasagiline further exemplifies the urgent need for the development of an anti-Parkinsonian drug that is not only selective in MAO-B inhibition but also possesses clear, clinically proven neuroprotective action to address the unmet medical need in PD.

To this end, our strategy was to design multifunctional and multitargeted novel propargylamines with potent and selective MAO-B activity and neuroprotective effects. The design of our novel propargylamines is based on the knowledge that the propargylamino group is likely to be important in neuroprotection, while MAO-B selectivity and potency as well as overall neuroprotective profile can be tuned through judicious choice of the skeleton and its substituents. In addition, recent reports suggest that MAO-B inhibitors can have neuroprotective functions.¹² Herein we describe the synthesis of a library of novel compounds with the general structure shown in Figure 2. The test compounds were screened for MAO-A and



Figure 2. General structure of target compounds.

Scheme 1	. General	Synthetic	Routes to	o Target	Compounds	a

MAO-B inhibition, in both rat and human enzyme assays, as well as in neurotoxin-treated PC12 cells, a widely accepted model for neuroprotection.

RESULTS AND DISCUSSION

Synthetic Chemistry. The compounds synthesized and studied in the biological assays, comprising an (hetero)aryl moiety, a vinyl, and backbone substituents, can be conveniently divided into four groups. Groups 1 and 2 are comprised of compounds with a variety of unsubstituted aryl and heteroaryl groups, while in group 3, compounds bear mono substitution on the aryl ring and those in group 4 have disubstituted aryl rings. The target compounds 1-50 (except 15 and 18) were synthesized using the four general synthetic routes shown in Scheme 1. Compounds 15 and 18 were obtained using the synthetic routes shown in Schemes 2 and 3, respectively. The

Scheme 2. Synthesis of Compound 15^a



^aReagents and conditions: (a) Mg, I₂, ether, 1 h then propargyl alcohol, CuI, 40 °C, 16 h, 67%; (b) CBr₄, PPh₃, DCM, 16 h, 86%; (c) *N*-methylpropargylamine, DCM, 16 h, 55%; (d) oxalic acid, ⁱPrOH, 83%.

choice of synthetic routes was based on the availability of starting material/chemical reagents and the potential for formation of side products. The first route (route A) started from the commercially available aryl methyl ketone I, which



"Reagents and conditions: (a) $Br_{2\nu}$ AcOH, toluene, 0 °C to rt or benzyltrimethylammonium dichloroiodate, THF; (b) *N*-methylpropargylamine, ether, acetonitrile (ACN) or (dichloromethane) DCM; (c) TMSCHN₂, ⁱPrOH, (Ph₃P)₃RhCl, PPh₃, dioxane, 60 °C; (d) diphenyl diselenide, *N*chlorosuccinimide (NCS), DCM, or NBS, chlorobenzene, 160 °C in an oil bath or 180 °C in a microwave reactor; (e) *N*-methylpropargylamine, ⁱPrOH; (f) Pd(OAc)₂, XPhos, K₃PO₄·H₂O, EtOH/H₂O, 60 °C; (g) TFA/DCM then propargyl bromide, Na₂CO₃, DCM; (h) Buchwald's precatalyst (catalyst A or B), K₃PO₄·H₂O, THF/H₂O, 60 °C; (i) MsCl, NEt₃, DCM, then *N*-methylpropargylamine.



"Reagents and conditions: (a) LDA, CO₂, THF; (b) DCC, DMAP, 'BuOH, DCM, 16 h, 81% over two steps; (c) NaO'Bu, Freon 22 (ClCHF₂), 9 bar, THF, 16 h, 86%; (d) TFA, DCM, 40 min; (e) NaOH, 15 min, 68% over two steps; (f) DIBALH, THF, 1 h, 68%; (g) CBr₄, PPh₃, DCM, 16 h, 60%; (h) *N*-methylpropargylamine, ACN, 16 h, 60%; (i) oxalic acid, ⁱPrOH, 67%.

was converted to the aryl halo ketone II.¹³ Subsequently, the nucleophilic substitution was achieved using *N*-methylpropargylamine, followed by a modified Wittig olefination^{14,15} reaction to provide the desired tertiary amine IV. However, low yields were frequently obtained in the olefination protocol and the toxicity of TMSCHN₂ prohibited large-scale synthesis. As outlined in route B, an alternative procedure utilized the commercially available aryl methylstyrene V. Allylic bromination or chlorination followed by nucleophilic substitution of the halide with *N*-methylpropargylamine gave the desired compound IV. However, in some instances, allylic bromination using NBS alone or NCS and diphenyl diselenide¹⁶ led to a mixture of allylic and vinyl halides. The use of catalytic amount of Yb(OTf)₃ and TMSCl in this step alleviated these regioselectivity issues in some cases.¹⁷

In order to facilitate compound library synthesis, a modular approach was sought. Suzuki cross-coupling reactions (routes C and D) could be utilized depending on the availability of the precursors. In route C, the Suzuki coupling between the aryl boronate ester VII or aryl boronic acid VIII with tert-butyl (2bromoallyl)methylcarbamate proceeded in the presence of $Pd(OAc)_2$, XPhos, and K_3PO_4 as the base in a 2:1 EtOH/H₂O mixture.¹⁸ The resultant allylic amine IX was subjected to Boc deprotection followed by alkylation using propargyl bromide. In route D, Suzuki coupling was carried out between the aryl boronate ester VII or aryl boronic acid VIII with 2-bromoallyl alcohol in the presence of Buchwald's precatalyst (catalyst A or B) and K_3PO_4 as the base.^{19,20} The resultant alcohol X was subjected to mesylation and amination in a one-pot reaction to give the desired arylalkenyl N-methylpropargylamines. In cases where compounds bear an electron-donating group at the para position of the aryl group, the target compounds can only be accessed via route D. This is due to the formation of oxazolidinones during the Boc deprotection step of route C. All the propargyl amines IV (except compound 13, which was used as the free base) were converted to the final form, either the HCl salt or oxalate salt.

Scheme 2 depicts the synthetic route that was utilized for compound **15**. Using copper catalysis, phenylethylmagnesium bromide was added in a regioselective fashion to propargyl alcohol.²¹ The resultant allylic alcohol **51** underwent the Appel reaction²² followed by nucleophilic substitution to furnish the desired amine **53**. The free base was then converted to the oxalate salt **15**.

The synthetic route of the vinyl fluoride analog 18 is outlined in Scheme 3. The mixed malonate 54 was prepared in two steps from commercially available ethyl 2-phenylacetate. The introduction of a difluoromethyl group in **55** was achieved when a THF solution of **54** was stirred in the presence of NaO^tBu under an atmosphere of Freon 22 (ClCHF₂) at 9 bar for 16 h.²³ Decarboxylative fluoride elimination occurred in two steps from **55** to afford the vinyl fluoride **56**. The acrylate ester **56** was then reduced using DIBAL to provide the allylic alcohol **57**. Appel reaction²² of the resultant allylic alcohol **57**, followed by nucleophilic substitution, furnished the desired product **59**, which was converted to the oxalate salt **18**.

A summary of the synthesized compounds in groups 1-4 along with their characteristics and method of synthesis is shown in Table 1.

Biological Assays. Initial preliminary screening for MAO-A and MAO-B enzyme inhibitory activities was carried out for the majority of the compounds using rat MAO enzymes. Potent and selective inhibitors were subsequently screened against isolated human MAO-A and MAO-B enzymes. Selected compounds from groups 1–4 were also screened for neuroprotective properties in PC12 cells.

Rat MAO-A and MAO-B Inhibitory Activities. Rat MAO activities were determined on the rat brain homogenates with ¹⁴C-5-serotonin (MAO-A) or ¹⁴C-phenylethylamine (MAO-B) substrates (Table 2). Rasagiline displayed rMAO-B inhibition with an IC₅₀ of 1 nM (selectivity 3000). From group 1 compounds, compounds 1, 6, and 7, bearing phenyl, 3-furanyl, and 2-thiophenyl moieties, were potent and selective MAO-B inhibitors. However, substitution using pyridyl (2–4), 3-thiophenyl (8), and thiazolyl (9) moieties resulted in weak MAO-B inhibitors.

Next, in group 2 compounds, it was observed that both the *N*-desmethyl (10) and *N*-ethyl (11) analogues are micromolar inhibitors of rMAO-B. The introduction of a methyl group in either position of the methylene group (12 and 13) was generally detrimental to rMAO-B inhibition activity. Compound 14, with an increase in the distance (n = 3) between the alkene and the propargyl moiety of the structure as compared to compound 1, was inactive. Interestingly, increasing the distance between the aryl ring and the alkene moiety (m = 2 for 15, m = 1 for 16) gave comparable rMAO-B inhibitory activity as compared to the parent compound 1. Replacing the propargyl moiety with an allylic moiety (in 17) resulted in significant loss of activity. Masking the alkene moiety by a bioisosteric fluorine replacement (in 18) gave a micromolar selective rMAO-B inhibitor.

In group 3, the effect of monosubstitution on the benzene ring was explored. Substitutions with a *p*-halogen resulted in the more active rMAO-B inhibitors (4-chloro, 4-bromo, and 4-

Table 1. Final Salt Form of the Test Compounds

Compound		Route	Salt	Molecular formula	Melting point (°C)
	GROUP 1				
	Aryl				
1	Aryl = phenyl	В	HC1	$C_{12}H_{16}ClN$	166 – 167
2	Aryl = 2-pyridyl	A	Oxalate	$C_{14}H_{16}N_2O_4$	113 – 114
3	Aryl = 3-pyridyl	A	Oxalate	$C_{16}H_{18}N_2O_8$	149 – 150
4	Aryl = 4-pyridyl	A	Oxalate	$C_{16}H_{18}N_2O_8$	158 – 160
5	Aryl = 2-furanyl	A	Oxalate	$C_{13}H_{15}NO_5$	121 – 122
6	Aryl = 3-furanyl	A	Oxalate	$C_{13}H_{15}NO_5$	116 - 118
7	Aryl = 2-thiophenyl	В	HC1	C ₁₁ H ₁₄ ClNS	155
8	Aryl = 3-thiophenyl	A	Oxalate	$C_{13}H_{15}NO_4S$	125 - 126
9	Aryl = 2-thiazolyl	Α	Oxalate	$C_{12}H_{14}N_2O_4S$	153 – 154
	GROUP 2				
10	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{R}_3 = \mathbf{H}$	В	HC1	C ₁₃ H ₁₄ ClN	135
11	$R_1 = Et, R_2 = R_3 = H$	C	Oxalate	$C_{16}H_{19}NO_4$	85 – 87
12	$R_1 = R_3 = Me, R_2 = H$	D	Oxalate	$C_{16}H_{19}NO_4$	109 - 110
13	$R_1 = R_3 = H, R_2 = Me$	A	Free base	$C_{14}H_{17}N$	n.a.
14	m = 0, n = 3, R = H	A	Oxalate	$C_{17}H_{21}NO_4$	96 – 97
15	m = 2, n = 1, R = H	Scheme 2	Oxalate	$C_{17}H_{21}NO_4$	117 – 118
16	m = 1, n = 1, R = 3-F, 4-F	A	Oxalate	$C_{16}H_{17}F_2NO_4$	97 – 99
17		В	HCl	C ₁₃ H ₁₈ ClN	127 – 128
18	F N	Scheme 3	Oxalate	C ₁₅ H ₁₆ FNO ₄	153 – 155

Compound		Route	Salt	Molecular formula	Melting point (°C)
	GROUP 3				
19	R = 3-Cl	A	Oxalate	C ₁₅ H ₁₆ ClNO ₄	142 – 144
20	$\mathbf{R} = 4\text{-}\mathbf{C}1$	В	Oxalate	C ₁₅ H ₁₆ ClNO ₄	160 – 161
21	R = 3-Br	A	Oxalate	C ₁₅ H ₁₆ BrNO ₄	130 - 132
22	R = 4-Br	В	HC1	C ₁₃ H ₁₅ BrClN	176 – 177
23	R = 2-F	В	HCl	C ₁₃ H ₁₅ FCIN	140
24	R = 3-F	C	Oxalate	C ₁₅ H ₁₆ FNO ₄	142 – 143
25	R = 4-F	В	HC1	C ₁₃ H ₁₅ FClN	182 – 183
26	R = 2-OMe	В	HC1	C ₁₄ H ₁₈ ClNO	135
27	R = 3-OMe	D	Oxalate	C ₁₆ H ₁₉ NO ₅	106 – 107
28	R = 4-OMe	В	HCl	C ₁₄ H ₁₈ ClNO	148 - 149
29	R = 3-OCH ₂ Ph	С	Oxalate	C ₂₂ H ₂₃ NO ₅	112 - 113
30	R = 4-OCH ₂ Ph	D	Oxalate	C ₂₂ H ₂₃ NO ₅	138 – 139
31	$R = 3-CH_2OPh$	С	Oxalate	C ₂₂ H ₂₃ NO ₅	126 – 127
32	$R = 4-CH_2OPh$	C	Oxalate	C ₂₂ H ₂₃ NO ₅	147 – 150
33	$R = 3-CH_2NHPh$	D	Oxalate	C ₂₂ H ₂₄ N ₂ O ₄	143 – 147
34	$R = 4-CH_2NHPh$	D	Oxalate	$C_{22}H_{24}N_2O_4$	140 - 141

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Table 1. continued

Compound		Route	Salt	Molecular formula	Melting point (°C)
35	R = 3-F	D	Oxalate	C ₂₂ H ₂₂ FNO ₅	100 - 102
36	R = 4-F	D	Oxalate	C ₂₂ H ₂₂ FNO ₅	118 - 119
37	R = 3-OMe	D	Oxalate	C ₂₃ H ₂₅ NO ₆	113 – 114
38	R = 4-OMe	D	Oxalate	C ₂₃ H ₂₅ NO ₆	104 - 105
39	R = 3-F	D	Oxalate	C ₂₂ H ₂₂ FNO ₅	126 – 127
40	R = 4-F	D	Oxalate	C ₂₂ H ₂₂ FNO ₅	111 – 112
41	R = 3-OMe	D	Oxalate	C ₂₃ H ₂₅ NO ₆	115 – 116
42	R = 4-OMe	C	Oxalate	C ₂₃ H ₂₅ NO ₆	118 – 119
43		D	Oxalate	$C_{21}H_{21}NO_5$	185 – 187
44	F C C C C C C C C C C C C C C C C C C C	D	Oxalate	$C_{21}H_{20}FNO_5$	182 - 184
	GROUP 4				
45	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{M}\mathbf{e}$	C	Oxalate	$C_{17}H_{21}NO_4$	136 – 137
46	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{F}$	A	Oxalate	$C_{15}H_{15}F_2NO_4$	144 – 145
47	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{C}1$	D	Oxalate	C ₁₅ H ₁₅ Cl ₂ NO ₄	153 – 156
48	$\mathbf{R}_1 = \mathbf{M}\mathbf{e}, \mathbf{R}_2 = \mathbf{F}$	C	Oxalate	C ₁₆ H ₁₈ FNO ₄	156 – 158
49	$R_1 = Me, R_2 = Cl$	D	Oxalate	C ₁₆ H ₁₈ ClNO ₄	157 – 158
50	$\mathbf{R}_1 = \mathbf{C}\mathbf{l}, \mathbf{R}_2 = \mathbf{F}$	D	Oxalate	C ₁₅ H ₁₅ ClFNO ₄	145 – 146

fluorobenzene compounds 20, 22, and 25), when compared to other halogen-substituted regioisomers. Next, we investigated the effect of the methoxy and benzyloxy substitution on the aromatic ring, synthesizing all possible regioisomers. The 2methoxyphenyl propargylamine 26 was found to be a less selective rMAO-B inhibitor. However, substitution of the methoxy and benzyloxy group at the meta- and para-positions (compounds 27-30) resulted in submicromolar rMAO-B inhibitors. Interestingly, the meta- and para-substituted phenoxymethylene compounds 31 and 32 are selective nanomolar inhibitors of rMAO-B, especially compound **31** (IC₅₀ = 1.5 nM). Substitution at ring B of compounds **29** and **31** afforded compounds **35–42**; further studies revealed that compound **35** (with a 3-fluoro substitution in ring B) is a potent and selective rMAO-B inhibitor (IC₅₀ = 2.0 nM and selectivity 2500, which are comparable to rasagiline).

For group 4 compounds, we investigated the effects of disubstitution of the aromatic ring, especially at the meta- and para-position for the methyl, fluoro, and chloro substituents. Interestingly, all the compounds (45-50) were found to be

Table 2. MAO-A and -B Inhibition Values of All Test Compounds Determined in the Rat Brain Assay

Cor	npound	rMAO-B, IC ₅₀ , nM	rMAO-A, IC ₅₀ , nM	Rat Selectivity, r(A/B)
	Rasagiline	1	3000	3000
	GROUP 1			
	Aryl			
1	Aryl = phenyl	50	2000	40
2	Aryl = 2-pyridyl	>100	N.D. ^a	
3	Aryl = 3-pyridyl	<1000	N.D.	
4	Aryl = 4-pyridyl	>1000	N.D.	
5	Aryl = 2-furanyl	100	3300	33
6	Aryl = 3-furanyl	33	800	24
7	Aryl = 2-thiophenyl	60	2500	41
8	Aryl = 3-thiophenyl	>100	N.D.	
9	Aryl = 2-thiazolyl	>100	N.D.	
	GROUP 2			
10	$R_1 = R_2 = R_3 = H$	1000	10000	10
11	$R_1 = Et, R_2 = R_3 = H$	>1000	~100000	<100
12	$R_1 = R_2 = H, R_3 = Me$	~100000	>1000	>0.01
13	$R_1 = R_3 = H, R_2 = Me$	~1000	N.D.	
14	m = 0, n = 3, R = H	>1000	700	<0.7
15	m = 2, n = 1, R = H	60	660	11
16	m = 1, n = 1, R = 3-F, 4-F	100	~1000	~10
17		~100000	N.D.	
18		800	66000	82

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Table 2. continued

Cor	npound	rMAO-B, IC ₅₀ , nM	rMAO-A, IC ₅₀ , nM	Rat Selectivity, r(A/B)
19	R = 3-C1	90	1000	11
20	$\mathbf{R} = 4 - \mathbf{C}1$	25	2000	80
21	R = 3-Br	500	>1000	>2
22	R = 4-Br	20	2500	125
23	R = 2-F	500	5000	10
24	R = 3-F	60	2000	33
25	R = 4-F	33	660	20
26	R = 2-OMe	5000	5000	1
27	R = 3-OMe	90	1500	16.6
28	R = 4-OMe	80	200	2.5
29	R = 3-OCH ₂ Ph	20	5000	250
30	R = 4-OCH ₂ Ph	400	700	1.7
31	R = 3-CH ₂ OPh	1.5	400	266
32	$R = 4$ - CH_2OPh	9	1500	16
33	$R = 3-CH_2NHPh$	60	>1000	>16.6
34	R = 4-CH ₂ NHPh	100	5000	50
	R O N N			
35	R = 3-F	2	5000	2500
37	R = 3-OMe	9	>1000	>111
42	R = 4-OMe	6	700	116

Table 2. continued

Cor	npound	rMAO-B, IC ₅₀ , nM	rMAO-A, IC ₅₀ , nM	Rat Selectivity, r(A/B)
	GROUP 4			
45	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{M}\mathbf{e}$	80	1000	12.5
46	$\mathbf{R}_1 = \mathbf{R}_2 = \mathbf{F}$	50	5000	100
47	$R_1 = R_2 = C1$	80	~1000	~12.5
48	$R_1 = Me, R_2 = F$	70	4000	57
49	$\mathbf{R}_1 = \mathbf{M}\mathbf{e}, \mathbf{R}_2 = \mathbf{C}\mathbf{l}$	80	~1000	~12.5
50	$R_1 = Cl, R_2 = F$	70	700	10

^{*a*}N.D. = not determined.

selective, submicromolar inhibitors of rMAO-B. In particular, the 3,4-difluoroaryl compound **46** displayed the best selectivity and most potent rMAO-B inhibition activity of this group of compounds.

Human MAO-A and MAO-B Inhibitory Activities of Selected Compounds. The human MAO-A and MAO-B inhibitory activities were measured for selected compounds chosen from the preliminary screening above (Table 3). From group 1 compounds, it was observed that the 3-furanyl analog **6** was the most selective, subnanomolar hMAO-B inhibitor (IC_{50} = 0.72 nM with >2500-fold selectivity) followed by the 2-thiophenyl compound 7 (IC_{50} = 3.4 nM).

Meanwhile, from group 2, the secondary amine 10 resulted in micromolar hMAO-B inhibition, while the amines 15 and 16 (m = 2 for 15 and m = 1 for 16) were submicromolar and poorly selective hMAO-B inhibitors, indicating the importance of methyl substitution at the amino group as well as the direct attachment of aryl group to the vinyl substituent.

From group 3 compounds, it was observed that parasubstitution appeared to be optimal for activity, consistent with observations from the rat data, as can be observed in the 4chloro, 4-bromo, and 4-fluorobenzene compounds (20, 22, and 25). Both 3-chloro- and 3-fluorobenzene compounds (19 and 24) were also potent hMAO-B inhibitors, although compound 19 was not a selective hMAO-B inhibitor. m-Benzyloxybenzene derivative **29** showed significant hMAO-B inhibition ($IC_{50} = 6$ nM, selectivity 350). Meanwhile, both meta- and parasubstituted phenoxymethylenebenzene derivatives (31 and 32) were submicromolar hMAO-B inhibitors. We further explored derivatives of 3-benzyloxybenzene and the 3phenoxymethylenebenzene compounds (35-42) and found that all meta- and para-substitution of fluoro and methoxy substituents in ring B resulted in moderate to excellent hMAO-B inhibitors. It was also interesting to note that the metabolite 44, which is the N-desmethyl analogue of 35, is a more potent

and selective hMAO-B inhibitor (IC₅₀ = 1.9 nM, selectivity >2200) as compared to the parent compound **35** (IC₅₀ = 50 nM, selectivity >20).

The effects of disubstitution with fluoro, chloro, and methyl groups on the aromatic ring resulted in moderate to excellent hMAO-B inhibitors. The 3,4-difluorobenzene and 3,4-dichlorobenzene compounds (46 and 47) were identified as the most active hMAO-B inhibitors among the compounds in the series, with 46 exhibiting >1300 times more selectivity for hMAO-B over hMAO-A inhibition. Interestingly, the 3,4-dimethylbenzene and 3-chloro-4-fluorobenzene derivatives (45 and 50) were potent but nonselective hMAO-B inhibitors.

Our studies with compound 1 (detailed in the Biology section within the Experimental Section) suggest that it behaves as an irreversible inhibitor to the hMAO enzyme.

In Vitro Neuroprotective Action in the PC12 Assay. Cellbased cell survival assays are relatively simplified systems and, therefore, widely used for in vitro high-throughput screening of potential anti-Parkinsonian drugs. We chose PC12 cells for this purpose, as they are catecholaminergic in origin, similar to dopaminergic neurons, and we used different types of neurotoxins to increase the predictive value of the model for clinical neuroprotection. In the first series of experiments, 6hydroxydopamine (6-OHDA), a catecholaminergic neurotoxin, was used to elicit cell death, whereas in the second series, rotenone, a mitochondrial toxin implicated in the etiology of PD, was used. In these experiments, rasagiline was used as reference compound, and the procedures described by Zheng et al.²⁴ were followed, with slight modification. Cells were incubated with 6-OHDA (200 μ M) for 24 h, and test compounds and rasagiline were applied in a single 10⁻⁵ M concentration for 1 h prior to 6-OHDA application. The MTT assay was used to assess cell death. In a similar manner, rotenone was used to assess the ability of the target compounds

Table 3. MAO-A and -B Inhibition Values of All Group 1-4 Compounds Determined in the Human Recombinant Assay

Comp	ound	hMAO-B, IC ₅₀ , nM	hMAO-A, IC ₅₀ , nM	Selectivity, (A/B)
1		60	3500	58
6		0.72	1900	2638
7		3.4	2500	735
10		1500	20000	13
16		34	130	4
19	GROUP 3	30	83	3
20	CI N	3.9	120	31
22	Br	0.85	60	70
24	FN	7.6	160	21
25	F N	3.7	280	74
27		65	200	3
28		370	56	0

Table 3. continued

Comp	ound	hMAO-B, IC ₅₀ , nM	hMAO-A, IC ₅₀ , nM	Selectivity, (A/B)
29		6	2100	350
31		20	400	20
32		50	620	12
33		7.4	1400	185
34	H. H. H.	80	850	11
35	F O N	50	1200	24
36		13	63	5
37		6.6	590	88
38		50	2500	50
39	F O N	14	71	5
40	F O O N	1.4	5.5	4
41		20	350	17

Table 3. continued

Compound		hMAO-B, IC ₅₀ , nM	hMAO-A, IC ₅₀ , nM	Selectivity, (A/B)
42		9	440	48
43	K K	6.3	250	40
44	F C C C C C C C C C C C C C C C C C C C	1.9	4200	2211
	GROUP 4			
45		18	91	5
46	F N F	2.3	3100	1347
47		1.0	61	61
48	F N	17	230	14
49		4.1	100	24
50		6.3	26	4

to facilitate cell survival.²⁵ The results are summarized in Table 4.

In line with literature data,^{24,26} rasagiline exerted significant, 48% protective effect in 10 μ M concentration (Table 4). Of the compounds tested, compounds 1, 10, 15, 18, 20, 24, 31, 46, and 48 are protective against 6-OHDA-induced cell death (>40% protection), while only compounds 15, 21, and 45 show >20% protection against rotenone-induced cell death. Of these, compound 24 showed the highest increase of 117% in the survival of 6-OHDA-treated cells, while the best compound for rotenone-induced cell death is less spectacular at 33% increase (shown by compound 15). The latter is, however, also a very important finding, since 15 is still much more effective than rasagiline (6% increase) on rotenone treatment of PC-12 cells. It is interesting to note that, among those compounds which were tested in both assays, compound **15**, a selective hMAO-A inhibitor, is the only compound that shows neuroprotective activities in both 6-OHDA- and rotenone-treated PC12 cells.

Cytotoxicity of Selected Compounds and Their Putative Metabolites. The cytotoxicities of selected compounds (1, 20, 40, 43, and 46) and their principal metabolites (a-c, Scheme 4) were also investigated using a metabolically competent immortalized liver cell line, TAMH (TGF α overexpressing mouse hepatocytes). This assay provides a glimpse of the safety index of the compounds, i.e., the margin of activity vs toxicity, to support further development. Viability at 100 and 10 μ M (with 24 and 72 h exposures) was used as a preliminary indicator of cytotoxicity.

From Table 5, it is noted that rasagiline and its metabolite, aminoindane, along with compounds 1, 20, and 46 and their

Table 4. Effect of Compounds (10 μ M) on the Survival of PC12 Cells after 6-OHDA and Rotenone Treatment^{*a*}

	% survi	val
compd	6-OHDA	rotenone
control (no test compd)	100 ± 2.31	100 ± 1.88
rasagiline	$147.82 \pm 7.23^{***}$	106.05 ± 1.28
1	$166.71 \pm 7.17^{***}$	116.8 ± 2.1
2	N.D.	121.01 ± 4.20
3	N.D.	106.05 ± 3.15
4	N.D.	110 ± 4.15
5	N.D.	113.11 ± 2.35
6	132.9 ± 7.3	115.7 ± 4.2
7	127.94 ± 8.87	104.57 ± 7.3
9	N.D.	115.10 ± 3.0
10	$151.781 \pm 8.79^{***}$	111.68 ± 4.6
11	110.61 ± 1.59	97.45 ± 3.24
12	108.95 ± 2.13	100.79 ± 1.91
14	102.83 ± 1.00	100.05 ± 5.39
15	163.99 ± 9.7***	$133.40 \pm 2.4^{***}$
17	110.14 ± 1.69	113.06 ± 7.0
18	$167.06 \pm 11.74^{***}$	101.86 ± 1.99
19	108.81 ± 5.335	107.92 ± 3.54
20	$147.17 \pm 6.76^{**}$	112.03 ± 4.8
21	110.24 ± 3.52	131.64 ± 6.76***
22	119.23 ± 2.62	106.25 ± 5.03
23	117.26 ± 2.95	109.17 ± 4.6
24	$217.27 \pm 14.92^{***}$	101.92 ± 3.24
25	119.89 ± 4.81	102.12 ± 1.2
26	105.03 ± 1.52	111.07 ± 5.3
27	112.02 ± 3.56	109.19 ± 1.17
28	115.66 ± 2.52	109.72 ± 4.6
29	95.39 ± 7.89	89.67 ± 2.18
31	$174.24 \pm 6.27^{***}$	106.90 ± 0.99
44	109.50 ± 1.94	109.51 ± 2.13
45	119.89 ± 2.16	$125.96 \pm 7.50^*$
46	174.46 ± 14.07**	114.48 ± 4.68
47	81.27 ± 6.87	89.31 ± 3.18
48	156.54 ± 9.13**	88.61 ± 5.12
49	106.09 ± 9.79	104.46 ± 4.16
50	111.49 ± 1.18	93.95 ± 2.65

"Survival data are expressed as the percentage of 6-OHDA- and rotenone-treated cells. Symbols represent significant changes from 6-OHDA- and rotenone-treated (*P < 0.05; **P < 0.01; ***P < 0.001) cells, respectively. Statistical analysis: one-way ANOVA followed by the Tukey test. All data are the means \pm SEM of at least eight values measured in two independent plates. N.D. not determined.

Scheme 4. Principal Metabolites of Propargylamines



respective metabolites are generally not toxic, even at suprapharmacological concentrations (100 μ M) after 24 and 72 h, respectively. However, for compound 46, high toxicities were observed after 72 h at both 100 and 10 μ M. For compounds 40 and 40a, cell viability is less than 50% when administered at 100 μ M concentration after 24 h. This loss of cell viability is more prominent for 40 and all its metabolites at the concentration of 100 μ M. Similar observations were also made for the metabolites of 31, with cell viabilities ranging between 15 and 40% after 72 h with treatment of 100 μ M of test compound. These compounds with measurable toxicities prompted IC_{50} determination, which revealed that 40 is the most toxic (28.3 μ M) followed by the desmethyl metabolite of 31 (i.e., 43, IC₅₀) = 33.7 μ M) that is followed by the desmethyl metabolite of 40, i.e., **40a** (IC₅₀ = 43.9 μ M). Nonetheless, these studies show that most of the novel compounds and their metabolites can be considered to be non-cytotoxic at the projected concentrations used for MAO B inhibition and therapeutic effect.

Finally, ADMET properties of a selected set of compounds (1, 20, 31, 35, 46) were characterized by QikProp parameters (see the Supporting Information, Table 5) obtained with the Schrodinger Suite in silico modeling package. No violation of druglikeness scores was observed.

CONCLUSIONS

The studies above have identified a number of new compounds possessing selective and potent human MAO B inhibitory activities that also possess in vitro neuroprotective (as measured by survival of 6-OHDA- or rotenone-treated cells) properties. The most promising of these compounds are identified as compound 1, the *m*-fluorophenyl compound 24, the *m*-benzyloxyphenyl compound 31, 3,4-dimethylphenyl compound 45, 3,4-difluorophenyl compound 46, and the 3-methyl-4-fluorophenyl analogue 48. On the basis of these highly promising results, further studies on these compounds are now in progress to confirm their roles in the development of disease-modifying anti-Parkinsonian drugs.

EXPERIMENTAL SECTION

General. All reaction solvents were dried by the PureSolv MD 7 Solvent Purification System (Innovative Technology) prior to use. All reagents were used as purchased without further purification. All reactions were conducted under a slight positive pressure of argon using oven-dried (110 $^{\circ}$ C) glassware. Room temperature corresponds to a temperature interval from 20 to 25 $^{\circ}$ C. The solvents were removed under reduced pressure using standard rotary evaporators.

Analytical thin layer chromatography (TLC) separations were performed on Merck's silica gel 60 F₂₅₄-precoated aluminum sheets. Visualization was accomplished with UV light (254 or 365 nm) and/or ninhydrin (200.0 mmol in ethanol) stain followed by heating. Solvent mixtures used for chromatography are always given as a vol/vol ratio. Flash column chromatography was generally performed using silica gel 60 (spherical, 40–100 μ m or 40–63 μ m), purchased from Sigma-Aldrich, or the Biotage SP1 purification system by gradient. Melting points were determined on a Büchi-540 capillary melting point apparatus and are uncorrected. Fourier-transform infrared spectroscopy was conducted using Spectrum 100 software (PerkinElmer). High-resolution mass spectrometry was run by the electrospray ionization time-of-flight (ESI-TOF) mode on an Agilent 6210 mass spectrometer.

High-performance liquid chromatography (HPLC) analysis was performed with the following systems: (a) a JASCO PU-2089 pump and a JASCO MD-2010 diode array detector, with manual injection of 20 μ L on a column kept at ambient temperature (25 °C) at a MP flow rate of 0.5 mL/min or (b) Shimadzu LC-20AD parallel double micro

Table 5. Cytotoxicity of Selected Compounds/Principal Metabolites

		viab	ility		
	24	h	72	h h	
compd	100 µM	10 µM	100 µM	10 µM	IC_{50} (μM)
rasagilline mesylate	88.8 ± 1.7	90.5 ± 2.8	84.1 ± 1.7	91.6 ± 3.8	
aminoindane	87.5 ± 2.9	96.4 ± 1.7	81.2 ± 2.4	82.7 ± 1.6	
1	83.3 ± 4.1	76.6 ± 7.0	88.6 ± 1.7	92.6 ± 2.2	
10 (=1a)	85.2 ± 2.9	93.2 ± 3.3	71.9 ± 2.8	90.3 ± 3.4	
1b	93.3 ± 2.5	89.8 ± 8.4	95.0 ± 4.1	92.8 ± 7.7	
1c	80.9 ± 2.7	89.8 ± 2.7	78.3 ± 3.5	89.1 ± 4.4	
20	63.1 ± 3.5	72.7 ± 5.0	96.2 ± 4.7	94.0 ± 2.8	
20a	71.8 ± 2.5	80.9 ± 4.2	84.3 ± 1.1	96.6 ± 3.5	
20b	74.2 ± 6.0	75.6 ± 1.6	81.2 ± 4.3	86.0 ± 10.4	
20c	58.5 ± 10.9	71.7 ± 3.9	62.5 ± 6.4	67.5 ± 14.7	
31	66.9 ± 5.3	90.2 ± 4.5	71.3 ± 8.7	92.6 ± 8.6	
43 (=31a)	38.2 ± 11.2	84.1 ± 5.5	15.8 ± 5.4	90.3 ± 7.4	33.7
31b	65.6 ± 1.5	82.6 ± 9.2	39.4 ± 7.7	87.1 ± 3.9	80.7
31c	50.1 ± 6.9	78.8 ± 2.6	22.4 ± 5.9	83.0 ± 2.2	86.4
40	47.1 ± 8.7	70.6 ± 7.5	19.9 ± 19.8	59.1 ± 6.1	28.3
40a	31.8 ± 6.8	69.3 ± 2.3	6.5 ± 11.8	70.8 ± 10.6	43.9
40b	61.6 ± 7.2	89.8 ± 5.8	27.0 ± 9.9	87.1 ± 4.1	
40c	55.2 ± 4.8	89.6 ± 2.2	29.8 ± 8.4	66.4 ± 0.7	
46	107.3 ± 1.4	113.3 ± 9.4	3.95 ± 15.5	5.1 ± 25.4	
46a	82.3 ± 2.8	93.6 ± 4.5	77.1 ± 3.5	81.8 ± 0.9	
46b	103.6 ± 0.6	113.3 ± 3.5	83.1 ± 4.4	84.4 ± 1.0	
46c	110.3 ± 0.8	112.2 ± 1.6	79.2 ± 0.9	84.4 ± 1.5	

plunger type chromatograph, Shimadzu SPD-M20A Prominence HPLC photo diode array detector, and Shimadzu SIL-20A Prominence HPLC autosampler, with autoinjection of 10–30 μ L, a column kept at 40 °C, and a MP flow rate of 1 mL/min. The following are the conditions for system a: condition A used a Phenomenex Gemini NX C18 250 × 4.6 mm, 5 μ m analytical column and ACN/H₂O 65:35 as eluent, and condition B used a Phenomenex Gemini NX C18 250 × 4.6 mm, 5 μ m analytical column and ACN/H₂O 80:20 as eluent. For system b, condition C used a Kinetex 2.6 μ m C18 100 Å, LC column 150 × 4.6 mm analytical column, with the following gradient: 35%–95% solvent B in gradient flow from 0 to 10 min, followed by 2 min of 95% solvent B and 3 min of 35% solvent B, where solvent A is water and solvent B is ACN spiked with 0.1% triethylamine.

Tandem liquid chromatography–mass spectrometry (LCMS) was performed on a Waters 2996 separations module and 3100 mass detector using a Luna-5u-C18(2) ($150 \times 4.6 \text{ mm}$) reverse-phase analytical column at a flow rate of 1.0 mL/min. The eluting system, consisting of A (water) and B (acetonitrile), was used under a linear gradient. The performance was monitored by photodiode array detection with a wavelength of 254 nm. Generally, the eluting program started at 80% of A and 20% of B for the first 2 min, followed by changing the concentration of B from 20% to 80% in 20 min. This condition was maintained for another 5 min.

Elemental analyses were performed on an Elementar VarioEL III apparatus. Microwave (mw) irradiation experiments were carried out in a monomode CEM-Discover mw reactor, using the standard configuration as delivered, including proprietary software. The experiments were executed in open vessel mode with control of the temperature by infrared detection. After completion of the reaction, the vial was cooled to 50 $^\circ$ C by air jet cooling.

¹H and ¹³C nuclear magnetic resonance (NMR) spectra were recorded at ambient temperature (if not indicated otherwise), in the solvent indicated, on a Bruker 400 Ultra Shield spectrometer operating at 400 and 101 MHz, a Varian Mercury Plus 400 spectrometer at frequency of 400 and 100 MHz, a Varian Unity 600 spectrometer at frequency at 600 and 150 MHz, or on a Bruker Avance III 500 spectrometer at frequency of 500 or 125 MHz, respectively. Chemical shifts for ¹H NMR were recorded in parts per million (ppm)

downfield from tetramethylsilane or the proton signal of residual nondeuterated solvent (δ 7.26 for CHCl₃, δ 4.87 for CH₃OH, δ 4.79 for H₂O, δ 2.50 for DMSO, and δ 2.05 for acetone) as the internal signal. Coupling constants are indicated in hertz (Hz). Chemical shifts are given using the δ -scale relative to tetramethylsilane or the residual solvent signal as an internal reference. The following abbreviations are used for spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad, dd = doublet doublet, dm = doublet multiplet, and tm = triplet multiplet.

Chemistry. Detailed experimental procedures for the syntheses of the base form of the target compounds and the respective intermediates are described in the Supporting Information. The data below are for the final compounds used in the biological studies.

General Synthetic Procedure for Salt Formation from the Free Base (Method A). To a solution of the appropriate base in 96% EtOH or dioxane or EtOAc, 37% HCl was added dropwise or HCl gas was bubbled into an ice-cooled solution of the amine in ether until pH <3. After 30 min of stirring at room temperature, the residue was filtered or the mixture was evaporated to dryness. The residue obtained was recrystallized to afford the salts in 15–93% yields (unoptimized), depending on the scale.

General Synthetic Procedure for Salt Formation from the Free Base (Method B). A stock solution of oxalic acid (2.6 mmol, MW = 126.07, oxalic acid dihydrate) in abs *i*-PrOH (5.0 mL) was prepared and used immediately. Stock solutions should not be kept since ester formation has previously been observed. To a solution of the appropriate base (0.9 mmol) in abs *i*-PrOH (0.5 mL) was added a 0.512 M solution of oxalic acid dihydrate in *i*-PrOH (1.8 mL, 0.9 mmol). The resulting solution was allowed to stir at rt for 30 min, and then it was kept at 0 °C for 10 min. Then, the solid was filtered, washed with ether (2.0 mL), and dried under high vacuum to afford the products in 50–95% unoptimized yields.

N-Methyl-2-phenyl-N-(prop-2-yn-1-yl)prop-2-en-1-amine Hydrochloride (1). Following method A, the title compound was isolated (87%): mp 166–167 °C; ¹H NMR (400 MHz, D₂O) δ 7.58–7.44 (m, 5H), 5.84 (s, 1H), 5.70 (br s, 1H), 4.42 (br s, 2H), 4.01 (d, *J* = 2.4 Hz, 2H), 3.17 (t, *J* = 2.4 Hz, 1H), 2.90 (s, 3H); ¹³C NMR (D₂O, 100 MHz) δ 140.3, 139.2, 132.0, 131.9, 129.2, 127.0, 83.1, 74.0, 61.3, 47.6,

42.4. Anal. Calcd for $C_{12}H_{15}N$ ·HCl (MW = 221.73): C, 70.42%; H, 7.27%; N, 6.32%. Found: C, 70.50%; H, 7.51%; N, 6.30%.

N-*Methyl*-*N*-(*prop*-2-*yn*-1-*yl*)-2-(*pyridin*-2-*yl*)*prop*-2-*en*-1-*aminium Carboxyformate* (2). Following method B, the title compound was isolated (60%): mp 113–114 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 8.55 (ddd, *J* = 4.8, 1.6, 0.8 Hz, 1H), 7.78 (td, *J* = 8.0, 1.6 Hz, 1H), 7.69 (dt, *J* = 8.0, 0.8 Hz, 1H), 7.30 (ddd, *J* = 8.0, 4.8, 1.2 Hz, 1H), 6.09 (d, *J* = 1.6 Hz, 1H), 5.50 (d, *J* = 0.4 Hz, 1H), 3.67 (s, 2H), 3.47 (d, *J* = 2.0 Hz, 2H), 3.35 (t, *J* = 2.0 Hz, 1H), 2.32 (s, 3H); ¹³C NMR (DMSO- d_6 , 100 MHz) δ 161.7, 155.8, 148.7, 142.2, 136.8, 122.8, 120.8, 119.4, 77.6, 77.2, 57.8, 44.7, 40.7; HRMS (ESI+) *m*/*z* calcd for C₁₂H₁₄N₂⁺ [M + H]⁺ 187.1230, found 187.1230. Anal. Calcd for C₁₂H₁₄N₂·(COOH)₂ (MW = 276.29): C, 60.86%; H, 5.84%; N, 10.14%.

3-(3-(Methyl(prop-2-yn-1-yl)ammonio)prop-1-en-2-yl)pyridin-1ium Carboxyformate (**3**). Following method B, the title compound was isolated (55%): mp 149–150 °C; ¹H NMR (DMSO- d_6) δ 8.74 (dd, *J* = 2.0, 0.4 Hz, 1H), 8.48 (dd, *J* = 4.8, 1.6 Hz, 1H), 7.93 (ddd, *J* = 8.0, 2.0, 1.6 Hz, 1H), 7.37 (ddd, *J* = 8.0, 4.8, 0.8 Hz, 1H), 5.69 (d, 1H, *J* = 0.8 Hz), 5.40 (d, 1H, *J* = 0.8 Hz), 3.55 (s, 2H), 3.39 (d, 2H, *J* = 2.4 Hz), 3.29 (t, 1H, *J* = 2.4 Hz), 2.25 (s, 3H). ¹³C NMR (DMSO- d_6) δ 161.2, 148.4, 147.3, 141.1, 134.4, 133.9, 123.3, 117.9, 78.1, 76.8, 58.9, 44.3, 40.5; HRMS (ESI+) *m*/*z* calcd for C₁₂H₁₄N₂⁺ [M + H]⁺ 187.1230, found 187.1229. Anal. Calcd for C₁₂H₁₄N₂·2(COOH)₂ (MW = 366.32): C, 52.46%; H, 4.95%; N, 7.65%. Found: C, 51.86%; H, 5.04%; N, 7.64%.

4-(3-(Methyl(prop-2-yn-1-yl)ammonio)prop-1-en-2-yl)pyridin-1ium Carboxyformate (4). Following method B, the title compound was isolated (50%): mp 158–160 °C; ¹H NMR (DMSO- d_6) δ 8.53 (d, J = 6.0 Hz, 1H), 7.54 (d, J = 6.0 Hz, 1H), 5.80 (s, 1H), 5.45 (s, 1H), 3.46 (s, 2H), 3.32 (d, J = 2.4 Hz, 2H), 3.23 (t, J = 2.4 Hz, 1H), 2.19 (s, 3H); ¹³C NMR (DMSO- d_6) δ 161.3, 149.4, 146.4, 141.9, 121.1, 119.4, 78.3, 76.5, 58.5, 44.3, 40.6; HRMS (ESI+) m/z calcd for C₁₂H₁₄N₂⁺ [M + H]⁺ 187.1230, found 187.1227. Anal. Calcd for C₁₂H₁₄N₂. 2(COOH)₂ (MW = 366.32): C, 52.46%; H, 4.95%; N, 7.65%. Found: C, 51.18%; H, 5.13%; N, 7.68%.

2-(Furan-2-yl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**5**). Following method B, the title compound was isolated (50%): mp 121–122 °C; ¹H NMR (DMSO-*d*₆) δ 7.63 (dd, *J* = 2.0, 0.8 Hz, 1H), 6.59 (dd, *J* = 3.2, 0.8 Hz, 1H), 6.49 (dd, *J* = 3.2, 2.0 Hz, 1H), 5.60 (d, *J* = 1.2 Hz, 1H), 5.18 (d, *J* = 1.2 Hz, 1H), 3.42 (d, *J* = 2.4 Hz, 2H), 3.35 (s, 2H), 3.27 (t, *J* = 2.4 Hz, 1H), 2.28 (s, 3H); ¹³C NMR (DMSO-*d*₆) δ 161.5, 152.7, 142.7, 133.3, 112.2, 111.5, 107.9, 78.2, 76.8, 57.4, 44.6, 40.8; HRMS (ESI+) *m*/*z* calcd for C₁₁H₁₄NO⁺ [M + H]⁺ 176.1070, found 176.1069. Anal. Calcd for C₁₁H₁₃NO· (COOH)₂ (MW = 265.26): C, 58.86%; H, 5.70%; N, 5.28%. Found: C, 58.44%; H, 5.88%; N, 5.22%.

2-(Furan-3-yl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (6). Following method B, the title compound was isolated (77%): mp 116–118 °C; ¹H NMR (DMSO- d_6) δ 7.84 (s, 1H), 7.61 (t, *J* = 1.6 Hz, 1H), 6.74 (dd, *J* = 1.6, 0.8 Hz, 1H), 5.46 (s, 1H), 5.12 (s, 1H), 3.38 (s, 2H), 3.30 (s, 2H), 3.25 (s, 1H), 2.26 (s, 3H); ¹³C NMR (DMSO- d_6) δ 161.3, 143.3, 140.2, 135.4, 124.7, 113.9, 108.5, 78.2, 76.6, 59.3, 44.5, 40.7; HRMS (ESI+) *m/z* calcd for C₁₁H₁₄NO⁺ [M + H]⁺ 176.1070, found 176.1072. Anal. Calcd for C₁₁H₁₃NO·(COOH)₂ (MW = 265.26): C, 58.86%; H, 5.70%; N, 5.28%. Found: C, 57.84%; H, 5.81%; N, 5.26%.

N-*Methyl*-*N*-(*prop*-2-*yn*-1-*yl*)-2-(*thiophen*-2-*yl*)*prop*-2-*en*-1amine Hydrochloride (7). Following method A, the title compound was isolated (36%): mp 155 °C (dec); ¹H NMR (400 MHz, DMSO d_6) δ 10.85 (br s, 1H), 7.55 (m, 1H), 7.39 (m, 1H), 7.10 (dd, *J* = 5.1, 3.6 Hz, 1H), 5.80 (br s, 1H), 5.60 (br s, 1H), 4.16 (br s, 2H), 4.08 (br s, 2H), 3.87 (br s, 1H), 2.75 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 141.4, 131.8, 128.0, 126.7, 126.1, 120.3, 81.5, 73.3, 56.3, 45.3, 44.4. Anal. Calcd for C₁₁H₁₃NS·HCl (MW = 227.75): C, 58.01%; H, 6.20%; N, 6.15%. Found: C, 57.98%; H, 6.22%; N, 5.97%.

N-Methyl-N-(prop-2-yn-1-yl)-2-(thiophen-3-yl)prop-2-en-1-aminium Carboxyformate (8). Following method B, the title compound was isolated (55%): mp 125–126 °C; ¹H NMR (DMSO- d_6) δ 7.78 (dd, *J* = 2.8, 1.2 Hz, 1H), 7.64 (dd, *J* = 4.8, 2.8 Hz, 1H), 7.52 (dd, *J* =

4.8, 1.2 Hz, 1H), 5.74 (s, 1H), 5.34 (s, 1H), 3.55 (bs, 4H), 3.40 (s, 1H), 2.41 (s, 3H); 13 C NMR (DMSO- d_6) δ 161.3, 141.3, 140.3, 126.1, 125.9, 122.0, 115.0, 78.0, 76.9, 59.5, 44.5, 40.7; HRMS (ESI+) m/z calcd for C₁₁H₁₄NS⁺ [M + H]⁺ 192.0841, found 192.0844. Purity by HPLC (condition A) 100%.

N-Methyl-N-(prop-2-yn-1-yl)-2-(thiazol-2-yl)prop-2-en-1-aminium Carboxyformate (9). Following method B, the title compound was isolated (60%): mp 153–154 °C; ¹H NMR (DMSO-*d*₆) δ 7.82 (d, *J* = 3.2 Hz, 1H), 7.69 (d, *J* = 3.2 Hz, 1H), 6.04 (s, 1H), 5.47 (s, 1H), 3.49 (s, 2H), 3.41 (d, *J* = 2.4 Hz, 2H), 3.25 (t, *J* = 2.4 Hz, 1H), 2.28 (s, 3H); ¹³C NMR (DMSO-*d*₆) δ 166.1, 161.4, 142.7, 137.7, 120.7, 118.5, 78.1, 76.7, 58.3, 44.7, 40.7; HRMS (ESI+) *m*/*z* calcd for C₁₀H₁₃N₂S⁺ [M + H]⁺ 193.0794, found 193.0795. Purity by HPLC (condition A) 97%.

2-Phenyl-N-(prop-2-yn-1-yl)prop-2-en-1-amine Hydrochloride (10). Following method A, the title compound was isolated (14%): mp 135 °C; ¹H NMR (400 MHz, D₂O) δ 7.56–7.43 (m, 5H), 5.77 (br s, 1H), 5.59 (br s, 1H), 4.29 (br s, 2H), 3.93 (d, J = 2.6 Hz, 2H), 3.03 (t, J = 2.6 Hz, 1H). Anal. Calcd for C₁₂H₁₃N·HCl (MW = 207.70): C, 69.39%; H, 6.97%; N, 6.74%. Found: C, 69.10%; H, 6.80%; N, 6.73%.

N-Ethyl-2-phenyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (11). To the amine (89.6 mg, 0.449 mmol) in *i*-PrOH (0.39 mL) was added a solution of oxalic acid (40.4 mg, 0.448 mmol) in ⁱPrOH (0.39 mL). A white solid did not form, even after the reaction was left at 0 °C overnight. The oil was left stirring in ether for 3 d. Eventually, a white solid which formed was filtered, washed with cold ether, and dried to give the title compound (86.8 mg, 67%): mp 85–87 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.53–7.51 (m, 2H), 7.35–7.26 (m, 3H), 5.52 (d, *J* = 1.2 Hz, 1H), 5.29 (d, *J* = 1.2 Hz, 1H), 3.53 (s, 2H), 3.38 (d, *J* = 2.4 Hz, 2H), 3.17 (t, *J* = 2.4 Hz, 1H), 2.57 (q, *J* = 7.2 Hz, 2H), 1.00 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 161.8, 144.3, 139.6, 128.6, 128.1, 126.7, 116.5, 78.5, 76.8, 57.5, 47.1, 40.9, 12.5; HRMS (ESI+) *m/z* calcd for C₁₆H₁₉NNaO₄⁺ [M + Na]⁺ 312.1206, found 312.1202. Purity by HPLC (condition C) 99%.

N-Methyl-N-(2-phenylallyl)but-3-yn-2-aminium Carboxyformate (12). Following method B, the title compound was isolated as a white solid (63%): mp 108.5–110.3 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.52–7.50 (m, 2H), 7.35–7.28 (m, 3H), 5.50 (d, *J* = 1.2 Hz, 1H), 5.26 (d, *J* = 1.2 Hz, 1H), 3.67 (qd, *J* = 6.8, 2.0 Hz, 1H), 3.55 (d, *J* = 13.6 Hz, 1H), 3.23 (d, *J* = 2.0 Hz, 1H), 2.14 (s, 3H), 1.23 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 161.2, 144.1, 139.3, 128.1, 127.6, 126.2, 115.8, 81.7, 75.8, 58.2, 49.7, 36.5, 19.2; HRMS (ESI+) *m*/*z* calcd for C₁₄H₁₈N ⁺ [M + H]⁺ 200.1434, found 200.1430. Purity by HPLC (condition A) 99%.

(*rac*)-*N*-*Methyl*-3-*phenyl*-*N*-(*prop*-2-*ynyl*)*but*-3-*en*-2-*amine* (13). This was submitted as a free base. Please refer to the Supporting Information for the method to synthesize the free base. ¹H NMR (400 MHz, CDCl₃) δ 7.45–7.42 (m, 2H), 7.26–7.10 (m, 3H), 5.25–5.23 (m, 2H), 3.46 (q, *J* = 6.8 Hz, 1H), 3.42 (t, *J* = 2.4 Hz, 2H), 2.30 (s, 3H), 2.16 (t, *J* = 2.4 Hz, 1H), 1.12 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 144.6, 141.1, 128.0, 127.3, 127.1, 114.5, 79.2, 73.0, 61.9, 43.4, 39.1, 18.1. EI MS: *m*/*z* (relative abundance) 200 (M⁺⁺, 25%); 131 (100%). Purity by HPLC (conditions C) 95%.

N-*Methyl*-4-*phenyl*-*N*-(*prop*-2-*yn*-1-*yl*)*pent*-4-*en*-1-*aminium Carboxyformate* (14). Following method B, the title compound was isolated (77%): mp 96–97 °C; ¹H NMR (400 MHz, acetone- d_6) δ 7.48–7.45 (m, 2H), 7.36–7.26 (m, 3H), 5.33 (d, *J* = 1.2 Hz, 1H), 5.14 (d, *J* = 1.2 Hz, 1H), 3.83 (d, *J* = 2.4 Hz, 2H), 3.04 (t, *J* = 2.4 Hz, 1H), 3.00 (t, *J* = 7.6 Hz, 2H), 2.67 (s, 3H), 2.63 (t, *J* = 7.6 Hz, 2H), 1.83 (m, 2H); ¹³C NMR (101 MHz, acetone- d_6) δ 163.3, 148.2, 141.5, 129.3, 128.4, 126.9, 113.2, 79.0, 55.1, 45.1, 40.5, 32.9, 24.2; HRMS (ESI+) *m*/*z* calcd for C₁₅H₂₀N ⁺ [M + H]⁺ 214.1590, found 214.1596. Anal. Calcd for C₁₅H₁₉N·(COOH)₂ (MW = 303.36): C, 67.31%; H, 6.98%; N, 4.62%. Found: C, 66.62%; H, 6.83%; N, 4.54%.

N-Methyl-2-methylene-4-phenyl-N-(prop-2-yn-1-yl)butan-1-aminium Carboxyformate (15). Following method B, the title compound was isolated as a white solid (38%): mp 116–118 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.29–7.25 (m, 2H), 7.22–7.14 (m, 3H), 4.96 (s,

0

1H), 4.93 (s, 1H), 3.36 (s, 2H), 3.23 (s, 1H), 3.05 (s, 2H), 2.72 (t, J = 8.0 Hz, 2H), 2.32 (t, J = 8.0 Hz, 2H), 2.24 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.4, 144.9, 141.7, 128.19, 128.16, 125.68, 113.66, 78.18, 76.56, 60.33, 44.64, 40.75, 35.11, 33.17; HRMS (ESI+) m/z calcd for C₁₅H₂₀N⁺ [M + H]⁺ 214.1596, found 214.1595. Anal. Calcd for C₁₅H₁₉N·(COOH)₂ (MW = 303.35): C, 67.31%; H, 6.98%; N, 4.62%. Found: C, 67.38%; H, 6.80%; N, 4.59%.

2-(3,4-Difluorobenzyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**16**). Following method B, the title compound was isolated as a white solid (78%): mp 98–99 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.23–7.12 (m, 2H), 7.05–7.01 (m, 1H), 5.35 (d, *J* = 1.2 Hz, 1H), 5.19 (d, *J* = 1.2 Hz, 1H), 3.86 (d, *J* = 2.4 Hz, 2H), 3.51 (s, 2H), 3.46 (s, 2H), 3.12 (t, *J* = 2.4 Hz, 1H), 2.73 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.8, 151.5 (dd, *J* = 248.4, 12.8 Hz), 150.5 (dd, *J* = 248.4, 12.8 Hz), 142.0, 137.0 (dd, *J* = 5.7, 4.0 Hz), 126.7 (dd, *J* = 6.3, 3.5 Hz), 121.4, 119.0 (d, *J* = 17.3 Hz), 118.3 (d, *J* = 17.2 Hz), 79.5, 74.8, 60.2, 45.9, 41.0, 40.6 (d, *J* = 1.4 Hz); HRMS (ESI+) *m*/z calcd for C₁₄H₁₆F₂N⁺ [M + H]⁺ 236.1245, found 236.1249. Anal. Calcd for C₁₄H₁₅F₂N·(COOH)₂ (MW = 325.31): C, 59.07%; H, 5.27%; N, 4.31%. Found: C, 59.02%; H, 5.31%; N, 4.15%.

N-Methyl-2-phenyl-*N*-(prop-2-en-1-yl)prop-2-en-1-amine Hydrochloride (**17**).²⁷ Following method A, the title compound was isolated (64%): mp 127–128 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 10.87 (br s, 1H), 7.55 (m, 2H), 7.44–7.34 (m, 3H), 6.05 (m, 1H), 5.83 (br s, 1H), 5.81 (br s, 1H), 5.51–5.43 (m, 2H), 4.30 (m, 1H) and 4.16 (m, 1H), 3.78 (m, 1H) and 3.76 (m, 1H), 2.60 (br s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 141.7, 138.1, 128.7, 128.5, 127.6, 126.3, 124.8, 122.7, 57.3, 56.5, 38.8. Anal. Calcd for C₁₃H₁₇N·HCl (MW = 223.74): C, 69.79%; H, 8.11%; N, 6.26%. Found: C, 69.54%; H, 8.22%; N, 6.40%.

(*E*)-3-*Fluoro-N-methyl-2-phenyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate* (**18**). Following method B, the title compound was isolated as a white solid (67%): mp 153–155 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.48 (d, *J* = 7.6 Hz, 2H), 7.36 (t, *J* = 7.6 Hz, 2H), 7.28 (d, *J* = 7.6 Hz, 1H), 7.03 (d, *J* = 84.0 Hz, 1H), 3.32 (d, *J* = 2.0 Hz, 2H), 3.28 (d, *J* = 3.2 Hz, 2H), 3.20 (brs, 1H), 2.19 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 161.2, 147.3 (d, *J* = 265.3 Hz), 133.7, 128.1 (d, *J* = 4.5 Hz), 128.1, 127.5, 119.0, 78.3, 76.5, 54.5 (d, *J* = 9.2 Hz), 44.3, 40.3; HRMS (ESI+) *m/z* calcd for C₁₃H₁₅FN⁺ [M + H]⁺ 204.1183, found 204.1184. Purity by HPLC (condition A) >99%.

2-(3-Chlorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**19**). Following method B, the title compound was isolated (78%): mp 139–140 °C; ¹H NMR (400 MHz, acetone-*d*₆) δ 7.58 (t, *J* = 2.0 Hz, 1H), 7.50 (dt, *J* = 6.8, 2.0 Hz, 1H), 7.39–7.33 (m, 2H), 5.60 (d, *J* = 1.2 Hz, 1H), 5.31 (d, *J* = 1.2 Hz, 1H), 3.45 (s, 2H), 3.33 (d, *J* = 2.0 Hz, 2H), 3.21 (t, *J* = 2.0 Hz, 1H), 2.21 (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 161.2, 142.5, 141.3, 133.0, 130.0, 127.4, 126.1, 125.0, 117.6, 78.2, 76.5, 59.1, 44.3; HRMS (ESI+) *m/z* calcd for C₁₃H₁₅ClN⁺ [M + H]⁺ 220.0888, found 220.0888. Anal. Calcd for C₁₃H₁₄ClN·(COOH)₂ (MW = 309.74): C, 58.16%; H, 5.21%; N, 4.52%; Found: C, 57.88%; H, 5.15%; N, 4.43%.

2-(4-Chlorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**20**). Following method B, the title compound was isolated (88%): mp 160–161 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.55 (m, 2H), 7.38 (m, 2H), 5.57 (d, J = 1.1 Hz, 1H), 5.31 (q, J = 0.9 Hz, 1H), 3.50 (br s, 2H), 3.37 (d, J = 2.4 Hz, 2H), 3.25 (t, J = 2.4 Hz, 1H), 2.24 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 161.6, 142.3, 137.7, 132.4, 128.2, 128.2, 117.4, 77.9, 76.9, 59.0, 44.3, 40.6. Anal. Calcd for C₁₃H₁₄CIN·(COOH)₂ (MW = 309.74): C, 58.16%; H, 5.21%; N, 4.52%. Found: C, 58.04%; H, 5.13%; N, 4.46%.

2-(3-Bromophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (21). Following method B, the title compound was isolated as a white solid (56%): mp 130–132 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.71 (s, 1H), 7.53 (d, *J* = 7.6 Hz, 1H), 7.48 (d, *J* = 7.6 Hz, 1H), 7.30 (t, *J* = 7.6 Hz, 1H), 5.59 (s, 1H), 5.31 (s, 1H), 3.45 (s, 2H), 3.34 (d, *J* = 2.0 Hz, 2H), 3.21 (t, *J* = 2.0 Hz, 1H), 2.22 (br, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.3, 142.5, 141.5, 130.3, 130.3, 128.9, 125.4, 121.7, 117.7, 78.2, 76.5, 59.1, 44.3, 40.6; HRMS (ESI+) *m*/z calcd for C₁₃H₁₅BrN⁺ [M + H]⁺ 264.0382, found 264.0381. Anal. Calcd for $C_{13}H_{14}BrN \cdot (COOH)_2$ (MW = 354.20): C, 50.86%; H, 4.55%; N, 3.95%. Found: C, 50.48%; H, 4.51%; N, 3.90%.

2-(4-Bromophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1amine Hydrochloride (22). Following method A, the title compound was isolated (49%): mp 176–177 °C; ¹H NMR (400 MHz DMSO- d_6) δ 11.10 (br s, 1H), 7.61–7.51 (m, 4H), 5.85 (br s, 1H), 5.78 (br s, 1H), 4.24 (br s, 2H), 4.03 (br s, 2H), 3.86 (br s, 1H), 2.69 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 137.0, 136.8, 131.4, 128.6, 123.8, 121.8, 81.4, 73.2, 56.4, 44.1, 39.2. Anal. Calcd for C₁₃H₁₄BrN·HCl (MW = 300.62): C, 51.94%; H, 5.03%; N, 4.66%. Found: C, 51.84%; H, 5.11%; N, 4.52%.

2-(2-Fluorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1amine Hydrochloride (23). Following method A, the title compound was isolated (73%): mp 140 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.50 (m, 1H), 7.42 (m, 1H), 7.27–7.21 (m, 2H), 6.00 (br s, 1H), 5.72 (br s, 1H), 4.22 (br s, 2H), 4.15 (br s, 2H), 3.84 (br s, 1H), 2.70 (br s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 159.2 (d, *J* = 243 Hz), 133.7, 130.5 (d, *J* = 8 Hz), 130.2 (d, *J* = 3 Hz), 126.9, 126.2 (d, *J* = 13 Hz), 124.8 (d, *J* = 3 Hz), 115.9 (d, *J* = 22 Hz), 81.4, 73.0, 57.2, 44.1, 39.0. Anal. Calcd for C₁₃H₁₄FN·HCl (MW = 239.71): C, 65.13%; H, 6.31%; N, 5.84%. Found: C, 65.09%; H, 6.18%; N, 5.78%.

2-(3-Fluorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (24). Following method B, the title compound was isolated (60%): mp 142.2–142.9 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.45–7.40 (m, 1H), 7.37–7.30 (m, 2H), 7.14– 7.09 (m, 1H), 5.81 (s, 1H), 5.68 (s, 1H), 4.22 (s, 2H), 3.94 (d, *J* = 2.5 Hz, 2H), 3.26 (t, *J* = 2.5 Hz, 1H), 2.79 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.6, 164.5 (d, *J* = 245.2 Hz), 141.4 (d, *J* = 7.7 Hz), 140.1 (d, *J* = 2.4 Hz), 131.8 (d, *J* = 8.4 Hz), 124.5, 123.6 (d, *J* = 2.9 Hz), 116.6 (d, *J* = 21.4 Hz), 114.6 (d, *J* = 22.7 Hz), 80.6, 73.9, 59.4, 45.9, 40.8. Anal. Calcd for C₁₃H₁₄FN·(COOH)₂ (MW = 293.29): C, 61.43%; H, 5.50%; N, 4.78%. Found: C, 61.42%; H, 5.43%; N, 4.64%.

2-(4-Fluorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1amine Hydrochloride (**25**). Following method A, the title compound was isolated (62%): mp 182–183 °C; ¹H NMR (400 MHz, DMSO d_6) δ 11.20 (br s, 1H), 7.60 (m, 2H), 7.21 (m, 2H), 5.77 (br s, 1H), 5.74 (br s, 1H), 4.36–4.16 (br m, 2H), 4.02 (br s, 2H), 3.85 (br s, 1H), 2.68 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 163.0 (d, J = 245 Hz), 137.8, 134.9 (d, J = 3 Hz), 129.4 (d, J = 8 Hz), 123.8, 116.1 (d, J = 22 Hz), 82.2, 73.9, 57.3, 44.8, 40.0. Anal. Calcd for C₁₃H₁₄FN-HCl (MW = 239.72): C, 65.13%; H, 6.31%; N, 5.84%. Found: C, 65.26%; H, 6.11%; N, 5.84%.

2-(2-Methoxyphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1amine Hydrochloride (**26**). Following method A, the title compound was isolated (44%): mp 135 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 11.03 (br s, 1H), 7.36 (ddd, J = 8.0, 7.6, 1.8 Hz, 1H), 7.27 (dd, J = 7.6, 1.8 Hz, 1H), 7.04 (dm, J = 8.0 Hz, 1H), 6.97 (td, J = 7.2, 1.0 Hz, 1H), 5.82 (br s, 1H), 5.52 (br s, 1H), 4.36–4.05 (m, 2H), 4.00 (br s, 2H), 3.85 (br s, 1H), 3.81 (s, 3H), 2.68 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 156.3, 137.7, 129.9, 129.9, 127.5, 124.6, 120.7, 111.4, 81.2, 73.1, 57.2, 55.4, 44.1, 39.1. Anal. Calcd for C₁₄H₁₇NO·HCl (MW = 251.75): C, 66.79%; H, 7.21%; N, 5.56%. Found: C, 66.70%; H, 7.11%; N, 5.49%.

2-(3-Methoxyphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (27). Following method B, the title compound was isolated as a white solid (86%): mp 106–107 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.25 (t, J = 8.0 Hz, 1H), 7.12–7.08 (m, 1H), 7.06 (t, J = 2.4 Hz, 1H), 6.87 (ddd, J = 8.0, 2.4, 0.4 Hz, 1H), 5.56 (d, J = 1.2 Hz, 1H), 5.29 (d, J = 1.2 Hz, 1H), 3.76 (s, 3H), 3.53 (s, 2H), 3.41 (d, J = 2.4 Hz, 2H), 3.27 (t, J = 2.4 Hz, 1H), 2.28 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 161.7, 159.2, 143.1, 140.4, 129.2, 118.6, 117.0, 113.1, 112.1, 77.8, 77.0, 59.1, 55.0, 44.4, 40.6; HRMS (ESI+) *m*/z calcd for C₁₄H₁₈NO⁺ [M + H]⁺ 216.1383, found 216.1391. Anal. Calcd for C₁₄H₁₇NO·(COOH)₂ (MW = 305.33): C, 62.94%; H, 6.27%; N, 4.59%. Found: C, 62.87%; H, 6.14%; N, 4.56%.

2-(4-Methoxyphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1amine Hydrochloride (**28**). Following method A, the title compound was isolated (93%): mp 148–149 °C; ¹H NMR (400 MHz, DMSO d_6) δ 7.50 (m, 2H), 6.96 (m, 2H), 5.71 (br s, 1H), 5.62 (br s, 1H), 4.23 (br s, 2H), 4.04 (br s, 2H), 3.88 (br s, 1H), 3.77 (s, 3H), 2.70 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6) δ 159.6, 137.4, 129.8, 127.7, 121.0, 114.0, 81.5, 73.2, 56.7, 55.2, 44.2, 39.3. Anal. Calcd for C₁₄H₁₇NO·HCl (MW = 251.75): C, 66.79%; H, 7.21%; N, 5.56%. Found: C, 66.36%; H, 7.09%; N, 5.51%.

2-(3-(Benzyloxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**29**). Following method B, the title compound was isolated (73%): mp 112–113 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.48–7.31 (m, 6H), 7.16–7.15 (m, 1H), 7.12–7.09 (m, 1H), 7.03 (ddd, *J* = 8.0, 2.4, 0.8 Hz, 1H), 5.75 (d, *J* = 0.8 Hz, 1H), 5.61 (d, *J* = 0.8 Hz, 1H), 5.13 (s, 2H), 4.22 (s, 2H), 3.92 (d, *J* = 2.4 Hz, 2H), 3.27 (t, *J* = 2.4 Hz, 1H), 2.77 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.9, 160.7, 140.8, 140.2, 138.6, 131.2, 129.5, 129.0, 128.6, 123.7, 120.2, 116.5, 114.5, 80.7, 73.7, 71.1, 59.6, 45.9, 40.7; HRMS (ESI+) *m/z* calcd for C₂₀H₂₂NO⁺ [M + H]⁺ 292.1696, found 292.1689. Anal. Calcd for C₂₀H₂₁NO·(COOH)₂ (MW = 381.42): C, 69.28%; H, 6.08%; N, 3.67%. Found: C, 69.25%; H, 6.07%; N, 3.52%.

2-(4-(Benzyloxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**30**). Following method B, the title compound was isolated (66%): mp 138–139 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.49–7.30 (m, 7H), 6.97 (d, *J* = 8.8 Hz, 2H), 5.45 (d, *J* = 1.2 Hz, 1H), 5.16 (d, *J* = 0.4 Hz, 1H), 5.11 (s, 2H), 3.46 (s, 2H), 3.37 (d, *J* = 2.4 Hz, 2H), 3.24 (t, *J* = 2.4 Hz, 1H), 2.24 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.4, 158.0, 142.7, 137.1, 131.4, 128.4, 127.8, 127.6, 127.4, 114.6, 114.4, 78.0, 76.7, 69.2, 59.3, 44.3, 40.6; HRMS (ESI+) *m*/*z* calcd for C₂₀H₂₂NO⁺ [M + H]⁺ 292.1696, found 292.1697. Purity by HPLC (condition A) >99%.

N-Methyl-2-(3-(phenoxymethyl)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (31). Following method B, the title compound was isolated as a white solid (85%): mp 125–126 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.60 (br, 1H), 7.50–7.47 (m, 1H), 7.37–7.35 (m, 2H), 7.31–7.27 (m, 2H), 7.03–7.00 (m, 2H), 6.96–6.92 (m, 1H), 5.55 (d, *J* = 1.2 Hz, 1H), 5.29 (d, *J* = 1.2 Hz, 1H), 5.10 (s, 2H), 3.50 (s, 2H), 3.37 (d, *J* = 2.3 Hz, 2H), 3.23 (t, *J* = 2.3 Hz, 1H), 2.25 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.3, 158.3, 143.6, 139.2, 137.0, 129.5, 128.3, 127.0, 125.7, 125.6, 120.7, 116.3, 114.8, 78.2, 76.5, 69.1, 59.2, 44.4, 40.7. Anal. Calcd for C₂₀H₂₁NO-(COOH)₂ (MW = 381.42): C, 69.28%; H, 6.08%; N, 3.67%. Found: C, 69.21%; H, 5.72%; N, 3.74%.

N-Methyl-2-(4-(phenoxymethyl)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**32**). Following method B, the title compound was isolated as a white solid (75%): mp 150–151 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.54 (d, *J* = 8.3 Hz, 2H), 7.41 (d, *J* = 8.3 Hz, 2H), 7.32–7.26 (m, 2H), 7.00 (dd, *J* = 8.7, 0.9 Hz, 2H), 6.93 (t, *J* = 7.3 Hz, 1H), 5.55 (d, *J* = 1.6 Hz, 1H), 5.27 (d, *J* = 1.6 Hz, 1H), 5.09 (s, 2H), 3.49 (s, 2H), 3.36 (d, *J* = 2.3 Hz, 2H), 3.23 (t, *J* = 2.3 Hz, 1H), 2.24 (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 161.3, 158.3, 143.3, 138.5, 136.5, 129.5, 127.5, 126.3, 120.7, 116.3, 114.7, 78.1, 76.6, 68.7, 59.2, 44.3, 40.6; HRMS (ESI+) *m/z* calcd for C₂₀H₂₁NO⁺ [M + H]⁺ 292.1696, found 292.1691. Anal. Calcd for C₂₀H₂₁NO·(COOH)₂ (MW = 381.42): C, 69.28%; H, 6.08%; N, 3.67%. Found: C, 68.52%; H, 6.16%; N, 3.29%.

N-Methyl-2-(3-((phenylamino)methyl)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (33). Following method B, the title compound was isolated (80%): mp 152–154 °C; ¹H NMR (400 MHz, methanol-*d*₄) δ 7.54 (s, 1H), 7.45–7.33 (m, 3H), 7.09 (dd, J = 8.4, 7.6 Hz, 2H), 6.73–6.59 (m, 3H), 5.77 (s, 1H), 5.63 (s, 1H), 4.38 (s, 2H), 4.28 (s, 2H), 3.94 (d, J = 2.4 Hz, 2H), 3.29 (t, J = 2.4 Hz, 1H), 2.78 (s, 3H); ¹³C NMR (101 MHz, methanol-*d*₄) δ 165.0, 149.4, 142.4, 140.5, 138.7, 130.3, 130.1, 129.3, 126.7, 126.2, 124.1, 118.7, 114.7, 81.2, 73.2, 59.5, 46.0, 40.7; HRMS (ESI+) *m/z* calcd for C₂₀H₂₃N₂⁺ [M + H]⁺ 291.1856, found 291.1861. Purity by HPLC (condition C) 96%.

N-Methyl-2-(4-((phenylamino)methyl)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (34). Following method B, the title compound was isolated (85%): mp 139–140 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.49 (d, *J* = 8.4 Hz, 2H), 7.43 (d, *J* = 8.4 Hz, 2H), 7.06 (dd, *J* = 8.8, 7.2 Hz, 2H), 6.64–6.55 (m, 3H), 5.74 (s, 1H), 5.58 (s, 1H), 4.35 (s, 2H), 4.21 (s, 2H), 3.91 (d, *J* = 2.4 Hz, 2H), 3.26 (t, *J* = 2.4 Hz, 1H), 2.76 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.8, 149.7, 142.9, 140.8, 137.2, 130.0, 129.0, 127.7, 122.9,

118.2, 114.2, 80.6, 73.8, 59.7, 48.3, 45.9, 40.8; HRMS (ESI+) m/z calcd for $C_{20}H_{23}N_2^+$ [M + H]⁺ 291.1856, found 291.1856. Purity by HPLC (condition C) 97%.

2-(3-((3-Fluorobenzyl)oxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**35**). Following method B, the title compound was isolated as a white solid (87%): mp 107–109 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.47–7.41 (m, 1H), 7.31–7.23 (m, 3H), 7.18–7.10 (m, 3H), 6.95–6.93 (m, 1H), 5.53 (d, *J* = 1.6 Hz, 1H), 5.26 (d, *J* = 1.6 Hz, 1H), 5.14 (s, 2H), 3.45 (s, 2H), 3.34 (d, *J* = 2.4 Hz, 2H), 3.22 (t, *J* = 2.4 Hz, 1H), 2.22 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 162.2 (d, *J* = 243.6 Hz), 161.3, 158.0, 143.5, 140.7, 140.1 (d, *J* = 7.4 Hz), 130.4 (d, *J* = 8.3 Hz), 129.2, 123.5 (d, *J* = 2.7 Hz), 119.0, 116.5, 114.5 (d, *J* = 20.9 Hz), 114.2 (d, *J* = 21.8 Hz), 113.9, 112.9, 78.2, 76.6, 68.3 (d, *J* = 1.7 Hz), 59.3, 44.4, 40.6. Anal. Calcd for C₂₀H₂₀FNO·(COOH)₂ (MW = 399.41): C, 66.16%; H, 5.55%; N, 3.51%. Found: C, 65.97%; H, 5.21%; N, 3.59%.

2-(3-((4-Fluorobenzyl)oxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**36**). Following method B, the title compound was isolated as a white solid (68%): mp 117.6– 118.9 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.50–7.45 (m, 2H), 7.34 (t, *J* = 8.0 Hz, 1H), 7.15 (t, *J* = 1.8 Hz, 1H), 7.13–7.07 (m, 3H), 7.04–7.01 (m, 1H), 5.75 (s, 1H), 5.61 (s, 1H), 5.11 (s, 2H), 4.20 (s, 2H), 3.91 (d, *J* = 2.8 Hz, 2H), 3.27 (t, *J* = 2.8 Hz, 1H), 2.76 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.8, 163.9 (d, *J* = 244.7 Hz), 160.6, 141.0, 140.3, 134.6 (d, *J* = 3.1 Hz), 131.2, 130.7 (d, *J* = 8.3 Hz), 123.4, 120.3, 116.4, 116.2 (d, *J* = 21.7 Hz), 114.5, 80.5, 73.9, 70.4, 59.7, 45.9, 40.8; HRMS (ESI+) *m*/*z* calcd for C₂₀H₂₁FNO⁺ [M + H]⁺ 310.1602, found 310.1589. Anal. Calcd for C₂₀H₂₀FNO·(COOH)₂ (MW = 399.41): C, 66.16%; H, 5.55%; N, 3.76%. Found: C, 66.16%; H, 5.31%; N, 3.61%.

2-(3-((3-Methoxybenzyl)oxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**37**). Following method B, the title compound was isolated as a white solid (68%): mp 113.9–114.3 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.30 (app t, 1H), 7.24 (app t, 1H), 7.16–7.15 (m, 1H), 7.11–7.09 (m, 1H), 7.03–7.01 (m, 2H), 6.94–6.92 (m, 1H), 6.90–6.88 (m, 1H), 5.52 (d, *J* = 1.2 Hz, 1H), 5.25 (d, *J* = 1.2 Hz, 1H), 5.08 (s, 2H), 3.76 (s, 3H), 3.44 (s, 2H), 3.34 (d, *J* = 2.4 Hz, 2H), 3.21 (t, *J* = 2.4 Hz, 1H), 2.22 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.3, 159.3, 158.2, 143.6, 140.6, 138.7, 129.5, 129.2, 119.8, 118.8, 116.4, 113.9, 113.2, 113.0, 78.3, 76.5, 69.0, 59.3, 55.0, 44.4, 40.7. Anal. Calcd for C₂₁H₂₃NO₂·(COOH)₂ (MW = 411.45): C, 67.14%; H, 6.12%; N, 3.40%. Found: C, 66.98%; H, 6.21%; N, 3.07%.

2-(3-((4-Methoxybenzyl)oxy)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**38**). Following method B, the title compound was isolated as a white solid (57%): mp 104–105 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.38–7.31 (m, 3H), 7.13 (s, 1H), 7.10 (d, *J* = 7.7 Hz, 1H), 7.02 (dd, *J* = 8.2, 2.1 Hz, 1H), 6.94–6.91 (m, 2H), 5.74 (s, 1H), 5.60 (br d, 1H), 5.05 (s, 2H), 4.19 (br, 2H), 3.90 (br, 2H), 3.80 (s, 3H), 3.26 (t, *J* = 2.4 Hz, 1H), 2.75 (s, 3H); ¹³C NMR (100 MHz, methanol- d_4) δ 165.8, 161.0, 160.7, 141.4, 140.3, 131.2, 130.5, 130.4, 122.9, 120.1, 116.4, 114.9, 114.5, 80.2, 74.2, 70.9, 59.9, 55.7, 45.9, 40.9; HRMS (ESI+) *m*/*z* calcd for C₂₁H₂₄NO₂+ [M + H]⁺ 322.1802, found 322.1801. Anal. Calcd for C₂₁H₂₃NO₂. (COOH)₂ (MW = 411.45): C, 67.14%; H, 6.12%; N, 3.40%. Found: C, 66.96%; H, 5.89%; N, 3.39%.

2-(3-((3-Fluorophenoxy)methyl)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**39**). Following method B, the title compound was isolated as a white solid (65%): mp 126.1–127.0 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.62 (br, 1H), 7.52–7.43 (m, 3H), 7.30–7.24 (m, 1H), 6.84–6.81 (m, 1H), 6.77 (dt, *J* = 11.1, 2.4 Hz, 1H), 6.70–6.65 (m, 1H), 5.78 (d, *J* = 0.8 Hz, 1H), 5.63 (d, *J* = 0.8 Hz, 1H), 5.13 (s, 2H), 4.23 (s, 2H), 3.91 (d, *J* = 2.8 Hz, 2H), 3.24 (t, *J* = 2.8 Hz, 1H), 2.77 (s, 3H); ¹³C NMR (100 MHz, methanol- d_4) δ 164.8 (d, *J* = 183.0 Hz), 161.6 (d, *J* = 11.0 Hz), 141.3, 139.3, 139.2, 131.6 (d, *J* = 10.1 Hz), 130.3, 129.1, 127.3, 126.8, 123.2, 112.0 (d, *J* = 3.0 Hz), 108.6 (d, *J* = 21.5 Hz), 103.5 (d, *J* = 25.2 Hz), 80.2, 74.2, 71.0, 59.8, 46.0, 40.9; HRMS (ESI+) *m/z* calcd for C₂₀H₂₁FNO⁺ [M + H]⁺ 310.1602, found 310.1599. Anal. Calcd for

 $C_{20}H_{20}FNO\cdot(COOH)_2$ (MW = 399.41): C, 66.16%; H, 5.55%; N, 3.51%. Found: C, 66.05%; H, 5.35%; N, 3.61%.

2-(3-((4-Fluorophenoxy)methyl)phenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (40). Following method B, the title compound was isolated as a white solid 59%): mp 111.0–112.2 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.61 (brs, 1H), 7.51–7.42 (m, 3H), 7.02–6.97 (m, 4H), 5.76 (s, 1H), 5.61 (s, 1H), 5.10 (s, 2H), 4.19 (s, 2H), 3.88 (s, 2H), 3.22 (s, 1H), 2.74 (s, 3H); ¹³C NMR (100 MHz, methanol- d_4) δ 165.6, 158.8 (d, *J* = 237.2 Hz), 156.3 (d, *J* = 2.1 Hz), 141.3, 139.6, 139.2, 130.3, 129.1, 127.2, 126.8, 123.2, 117.2 (d, *J* = 8.0 Hz), 116.7 (d, *J* = 23.4 Hz), 80.2, 74.2, 71.4, 59.8, 45.9, 40.9; HRMS (ESI+) *m*/*z* calcd for C₂₀H₂₁FNO⁺ [M + H]⁺ 310.1602, found 310.1599. Anal. Calcd for C₂₀H₂₀NO·(COOH)₂ (MW = 399.41): C, 66.16%; H, 5.55%; N, 3.51%. Found: C, 65.66%; H, 5.59%; N, 3.56%.

2-(3-((3-Methoxyphenoxy)methyl)phenyl)-N-methyl-N-(prop-2yn-1-yl)prop-2-en-1-aminium Carboxyformate (41). Following method B, the title compound was isolated as a white solid (55%): mp 114.7–116.0 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.62 (s, 1H), 7.52–7.41 (m, 3H), 7.16 (t, *J* = 8.0 Hz, 1H), 6.60–6.51 (m, 3H), 5.77 (s, 1H), 5.62 (s, 1H), 5.11 (s, 2H), 4.21 (s, 2H), 3.89 (s, 2H), 3.76 (s, 3H), 3.23 (s, 1H), 2.75 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 165.6, 162.4, 161.3, 141.3, 139.7, 139.2, 131.0, 130.3, 129.1, 127.1, 126.7, 123.2, 108.3, 107.5, 102.5, 80.3, 74.2, 70.7, 59.8, 55.7, 45.9, 40.9; HRMS (ESI+) *m*/*z* calcd for C₂₁H₂₄NO₂+ [M + H]+ 322.1802, found 322.1805. Anal. Calcd for C₂₁H₂₃NO₂·(COOH)₂ (MW = 411.45): C, 67.14%; H, 6.12%; N, 3.40%. Found: C, 67.16%; H, 5.90%; N, 3.53%.

2-(3-((4-Methoxyphenoxy)methyl)phenyl)-N-methyl-N-(prop-2yn-1-yl)prop-2-en-1-aminium Carboxyformate (42). Following method B, the title compound was isolated as a white solid (47%): mp 117.8–180 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.58 (brs, 1H), 7.49–7.46 (m, 1H), 7.35–7.34 (m, 2H), 6.97–6.93 (m, 2H), 6.88– 6.83 (m, 2H), 5.53 (d, *J* = 1.2 Hz, 1H), 5.28 (d, *J* = 1.2 Hz, 1H), 5.03 (s, 2H), 3.69 (s, 3H), 3.47 (s, 2H), 3.35 (d, *J* = 2.4 Hz, 2H), 3.21 (t, *J* = 2.4 Hz, 1H), 2.23 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 161.2, 153.5, 152.3, 143.6, 139.2, 137.2, 128.3, 127.0, 125.6, 125.5, 116.2, 115.8, 114.6, 78.3, 76.5, 69.7, 59.2, 55.3, 44.4, 40.7; HRMS (ESI+) *m*/ *z* calcd for C₂₁H₂₄NO₂⁺ [M + H]⁺ 322.1802, found 322.1804. Purity by HPLC (condition B) 99%.

2-(3-(Phenoxymethyl)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**43**). Following method B, the title compound was isolated as a white solid (81%): mp 188–190 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.60 (s, 1H), 7.50–7.43 (m, 3H), 7.27 (dd, *J* = 8.8, 7.6 Hz, 2H), 6.99 (dd, *J* = 8.8, 0.8 Hz, 2H), 6.94 (m, 1H), 5.75 (s, 1H), 5.55 (s, 1H), 5.13 (s, 2H), 4.23 (d, *J* = 0.4 Hz, 2H), 3.91 (d, *J* = 2.4 Hz, 2H), 3.21 (t, *J* = 2.4 Hz, 1H); ¹³C NMR (101 MHz, methanol- d_4) δ 166.4, 160.1, 141.1, 139.8, 138.8, 130.5, 130.3, 129.2, 126.8, 126.5, 122.1, 120.5, 116.0, 79.3, 74.8, 70.6, 50.7, 37.2; HRMS (ESI+) *m*/z calcd for C₁₉H₂₀NO⁺ [M + H]⁺ 278.1539, found 278.1542. Purity by HPLC (condition C) 99%.

2-(3-((3-Fluorobenzyl)oxy)phenyl)-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (44). Following method B, the title compound was isolated as a white solid (84%): mp 182–184 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.42–7.33 (m, 2H), 7.27–7.25 (m, 1H), 7.22–7.19 (m, 1H), 7.14–7.13 (m, 1H), 7.12–7.09 (m, 1H), 7.07–7.02 (m, 2H), 5.72 (s, 1H), 5.53 (s, 1H), 5.16 (s, 2H), 4.19 (s, 2H), 3.90 (d, *J* = 2.4 Hz, 2H), 3.22 (t, *J* = 2.4 Hz, 1H); ¹³C NMR (101 MHz, methanol- d_4) δ 166.5, 164.4 (d, *J* = 244.6 Hz), 160.4, 141.5 (d, *J* = 7.3 Hz), 141.1, 140.1, 131.4 (d, *J* = 8.2 Hz), 131.2, 124.1 (d, *J* = 2.9 Hz), 120.4, 120.1, 116.3, 115.5 (d, *J* = 21.4 Hz), 115.0 (d, *J* = 22.3 Hz), 114.4, 79.2, 74.8, 70.2 (d, *J* = 2.0 Hz), 50.8, 37.2; HRMS (ESI+) *m/z* calcd for C₁₉H₁₉FNO⁺ [M + H]⁺ 296.1445, found 296.1460. Anal. Calcd for C₁₉H₁₈FNO·(COOH)₂ (MW = 385.39): C, 65.45%; H, 5.23%; N, 3.63%. Found: C, 65.21%; H, 5.06%; N, 3.82%.

2-(3,4-Dimethylphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**45**). Following method B, the title compound was isolated (71%): mp 136–137 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.30 (s, 1H), 7.24 (dd, *J* = 7.6, 2.0 Hz, 1H), 7.17 (d, *J* = 7.6 Hz, 1H), 5.71 (d, *J* = 0.4 Hz, 1H), 5.55 (d, *J* = 0.4 Hz, 1H), 4.24 (s, 2H), 3.94 (d, J = 2.4 Hz, 2H), 3.26 (s, 1H), 2.80 (s, 3H), 2.30 (s, 3H), 2.28 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 166.0, 140.7, 138.9, 138.4, 136.1, 131.3, 128.7, 125.1, 122.5, 80.9, 73.6, 59.6, 45.9, 40.7, 19.9, 19.5. Anal. Calcd for C₁₅H₁₉N·(COOH)₂ (MW = 303.35): C, 67.31%; H, 6.98%; N, 4.62%. Found: C, 67.32%; H, 6.93%; N, 4.30%.

2-(3,4-Difluorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**46**). Following method B, the title compound was isolated as a white solid (60%): mp 147–148 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.66–7.56 (m, 1H), 7.38 (m, 2H), 5.59 (s, 1H), 5.30 (s, 1H), 3.44 (s, 2H), 3.34 (d, *J* = 2.1 Hz, 2H), 3.21 (t, *J* = 2.1 Hz, 1H), 2.21 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.2, 150.2, 147.8, 141.7, 136.7, 123.2 (dd, *J* = 3.3, 6.3 Hz), 117.5, 117.1 (d, *J* = 17.0 Hz), 115.4 (d, *J* = 17.7 Hz), 78.2, 76.5, 59.2, 44.3, 40.5; HRMS (ESI+) *m*/*z* calcd for C₁₃H₁₃F₂NNa⁺ [M + Na]⁺ 244.0908, found 244.0913. Anal. Calcd for C₁₃H₁₃F₂N·(COOH)₂ (MW = 311.28): C, 57.88%; H, 4.86%; N, 4.50%. Found: C, 57.38%; H, 4.82%; N, 4.40%.

2-(3,4-Dichlorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1aminium Carboxyformate (**47**). Following method B, the title compound was isolated as a white solid (81%): mp 139–140 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.79 (d, J = 2.0 Hz, 1H), 7.58 (d, J = 8.4 Hz, 1H), 7.53 (dd, J = 8.4, 2.0 Hz, 1H), 5.65 (d, J = 0.8 Hz, 1H), 5.34 (d, J = 0.8 Hz, 1H), 3.45 (s, 2H), 3.33 (d, J = 2.4 Hz, 2H), 3.22 (t, J = 2.4 Hz, 1H), 2.21 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.2, 141.6, 139.7, 130.9, 130.2, 130.1, 128.2, 126.6, 118.2, 78.2, 76.5, 59.0, 44.3, 40.5; HRMS (ESI+) m/z calcd for C₁₃H₁₃Cl₂N·(COOH)₂ (MW = 344.19): C, 52.34%; H, 4.39%; N, 4.07%. Found: C, 53.12%; H, 4.44%; N, 4.12%.

2-(4-Fluoro-3-methylphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2en-1-aminium Carboxyformate (**48**). Following method B, the title compound was isolated as a white solid (76%): mp 158–159 °C; ¹H NMR (400 MHz, methanol- d_4) δ 7.51–7.28 (m, 2H), 7.05 (app t, 1H), 5.71 (s, 1H), 5.60 (s, 1H), 4.23 (s, 2H), 3.96 (s, 2H), 3.28 (s, 1H), 2.81 (s, 3H), 2.29 (s, 3H); ¹³C NMR (101 MHz, methanol- d_4) δ 164.5, 161.7 (d, *J* = 246.0 Hz), 138.6, 133.3 (d, *J* = 3.5 Hz), 129.7 (d, *J* = 5.2 Hz), 125.7 (d, *J* = 8.3 Hz), 125.1 (d, *J* = 17.6 Hz), 122.0, 115.0 (d, *J* = 22.9 Hz), 79.4, 72.2, 58.2, 44.5, 39.3, 13.1 (d, *J* = 3.5 Hz). Anal. Calcd for C₁₄H₁₆FN·(COOH)₂ (MW = 307.32): C, 62.53%; H, 5.90%; N, 4.56%. Found: C, 62.53%; H, 6.15%; N, 4.34%.

2-(4-Chloro-3-methylphenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2-en-1-aminium Carboxyformate (**49**). Following method B, the title compound was isolated as a white solid (73%): mp 171–173 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.50 (br, 1H), 7.36 (br, 2H), 5.54 (d, *J* = 1.2 Hz, 1H), 5.27 (d, *J* = 1.2 Hz, 1H), 3.45 (s, 2H), 3.34 (d, *J* = 2.4 Hz, 2H), 3.22 (t, *J* = 2.4 Hz, 1H), 2.33 (s, 3H), 2.22 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.3, 142.7, 137.9, 135.0, 132.5, 129.0, 128.5, 125.5, 116.6, 78.2, 76.5, 59.1, 44.3, 40.6, 19.6; HRMS (ESI+) *m*/*z* calcd for C₁₄H₁₇ClN⁺ [M + H]⁺ 234.1044, found 234.1044. Anal. Calcd for C₁₄H₁₆ClN·(COOH)₂ (MW = 323.77): C, 59.35%; H, 5.60%; N, 4.33%. Found: C, 60.07%; H, 5.60%; N, 4.25%.

2-(3-Chloro-4-fluorophenyl)-N-methyl-N-(prop-2-yn-1-yl)prop-2en-1-aminium Carboxyformate (**50**). Following method B, the title compound was isolated as a white solid (77%): mp 144–145 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 7.75 (dd, J = 7.2, 2.4 Hz, 1H), 7.57– 7.53 (m, 1H), 7.37 (app t, 1H), 5.59 (d, J = 0.6 Hz, 1H), 5.30 (d, J = 0.6 Hz, 1H), 3.45 (s, 2H), 3.34 (d, J = 2.3 Hz, 2H), 3.22 (t, J = 2.3 Hz, 1H), 2.21 (s, 3H); ¹³C NMR (101 MHz, DMSO- d_6) δ 161.2, 156.7 (d, J = 247.1 Hz), 141.6, 136.8 (d, J = 3.9 Hz), 128.3, 127.0 (d, J = 7.3 Hz), 119.2 (d, J = 17.6 Hz), 117.6, 116.5 (d, J = 20.8 Hz), 78.2, 76.5, 59.2, 44.3, 40.5. Anal. Calcd for C₁₃H₁₃CIFN·(COOH)₂ (MW = 327.74): C, 54.97%; H, 4.61%; N, 4.27%. Found: C, 55.55%; H, 4.63%; N, 4.22%.

Biology. *Rat Monoamine Oxidase B and A Enzyme Assays.* MAO-A and MAO-B enzyme inhibitory activities were determined on rat brain homogenates with ¹⁴C-5-HT (MAO-A) or ¹⁴C-PEA (MAO-B) substrates and liquid scintillation method.

1. Tissue Preparation. MAO-A and MAO-B enzyme activities were determined on nuclei-free rat brain homogenates. Following

decapitation, the brain (excluding cerebellum) was removed, the exact weight being determined. On 1.0 g of brain tissue was layered 2.0 mL of 0.25 M freshly dissolved saccharose. The sample was homogenized (blade homogenizer) in an ice–water bath (33.3% homogenate) and centrifuged (1000 rpm, 10 min, 4 $^{\circ}$ C). For the assays the diluted supernatants were used (MAO-B, 8.325%; MAO-A, 16.5%).

2. Radiometric Assay of MAO-B/MAO-A Enzyme Activity. MAO-B/MAO-A enzyme activity was determined by a radiometric method.²⁸ From ¹⁴C-phenylethylamine/¹⁴C-5-hydroxytriptamine substrate MAO-B/MAO-A forms ¹⁴C-phenylacetaldehyde/¹⁴C-5-hydroxyindole-3-acetaldehyde, which could be extracted under acidic conditions and quantitated. The incubation mixtures contained phosphate buffer (100 µL, 0.1 M, pH 7.4), EDTA (20 µL, 0.01 M), distilled (deionized) water/inhibitor (20 μ L), and ¹⁴C-phenylethylamine (MAO-B; 20 μ L, 0.2 mM)/¹⁴C-hydroxytriptamine (MAO-A, 20 μ L, 0.5 mM), and 40 μ L of tissue (nuclei-free homogenate). Mixtures were preincubated with the inhibitors for 10 min (37.5 °C), followed by subsequent addition of the substrate and incubation for 20 min at 37 °C. The reaction was stopped by adding 200 μ L of 2 M citric acid. The samples were extracted with 1 mL of ethyl acetate by vortexing and then centrifuged (2500 rpm, 5 min). The enzyme activity was determined by liquid scintillation counting (600 μ L of the organic phase + 5 mL of scintillation liquid).

Human Monoamine Oxidase B and A Enzyme Assays. MAO-A and MAO-B enzyme inhibitory activities were determined on brain human recombinant MAO-A and MAO-B enzymes, and results are summarized in Tables 3 and 4.

Human recombinant MAO-A and MAO-B were purchased in the form of supersomes (BD Biosciences) expressed in Sf9. The enzymatic activity was measured in a fluorescent coupled reaction. MAO-A and MAO-B oxidize their substrates (5-hydroxytryptamine and phenylethylamine respectively) to produce hydrogen peroxide, which produces the oxidized form of Amplex UltraRed (Invitrogen) that can be readily measured in a fluorimetric plate reader at 540/590 nm (excitation/emission). Measurements were conducted in a 384-well format in the final volume of 40 μ L. The enzymes were preincubated on ice for 1 h prior to tests. The drug candidates were incubated for 10 min at room temperature with the enzyme, and then substrate was added to initialize the reaction at 30 °C. Fluorescence was read at 1 h of reaction and corrected with the value read before substrate addition. Dose-response curves were measured using at least seven dilution points with 5-fold dilution steps. Duplicate points were determined for each concentration. IC₅₀ values were calculated from the remaining activity, and the graphs were fitted using Origin 5.0 software.

Irreversible Inhibition of Monoamine Oxidase B. To investigate whether compound 1 inhibited MAO-B reversibly or irreversibly, we designed an equilibrium dialysis experiment. First, we measured the dose–response curve of 1 for MAO-B inhibition. For the dialysis experiment, a mixture of the enzyme MAO-B and 1 was prepared, where the concentration of compound 1 was 10× higher than its IC_{50} value, and the inhibition (86%) was measured. Then, the enzyme–inhibitor mixture was dialyzed overnight against 1000× volume of buffer containing no inhibitor; the percentage of inhibition was determined on the next day. The inhibition we obtained was 99%, indicating that the inhibitor could not be washed out. In a control experiment under similar conditions except that there was no inhibitor present, the enzyme activity did not change, indicating that the enzyme was intact after the overnight dialysis.

As a validation, we performed another study in which known reversible (safinamide) and irreversible inhibitors (rasagiline) were used. It was found that extent of inhibition caused by the irreversible inhibitor did not change by dialysis, while the extent of inhibition achieved by the reversible inhibitor was significantly decreased after the dialysis, indicating that the enzyme—inhibitor complex formed reversibly and dissociation occurred during the dialysis.

In the next experiment, the influence of preincubation of the enzyme–inhibitor mixture was investigated. We found that the longer the preincubation period, the lower the virtual IC_{50} value of compound 1. These results could be interpreted as follows. Compound 1 first binds to MAO B with a K_i value of 60 nM, and then it irreversibly

inactivates the enzyme; preincubation for 90 min at room temperature results in a virtual IC_{50} of 50 pM.

From the above experiments, it could be concluded that **1** inhibited the MAO-B enzyme in an irreversible fashion.

Cell Culture. PC12 cells were grown in Dulbecco's modified Eagle's medium (DMEM 4500) supplemented with 10% horse serum, 5% fetal bovine serum (FBS), and L-glutamine. The cultures were maintained in a humidified atmosphere containing 5% CO₂ at 37 °C. For experiments assessing the effect of drugs on 6-OHDA/ rotenone-induced cell death, PC12 cells were seeded on 96-well plates in a density of 10 000 cells/well. The data are expressed as the percentage of the survival of 6-OHDA/rotenone-treated cells of the same plate. Every measurement was performed at least in n = 8 number and repeated on at least two independent plates.

MTT Assay. Cell viability was assessed using the colorimetric reagent 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT). A stock solution of the dye was prepared, filter-sterilized, and stored at -20 °C. After 6-OHDA/rotenone treatment in 96-well plates, 5 mg/mL MTT was added to each well, and the incubation was continued for another 4 h. The converted dye was solubilized with acidic isopropyl alcohol (0.04 M HCl in anhydrous isopropyl alcohol). Reduced MTT was measured at a wavelength of 570 nm.

Cytotoxicity Assay. Transforming growth factor- α overexpressing mouse hepatocytes (TAMH) were used as the metabolically competent immortalized cell line to access the cytoxicity of compounds arising from both the administered ones and the metabolites. These cells were maintained in accordance to established conditions.²⁹ TAMH cells were seeded at a cell density of 12 000 cells/ well in a 96-well plate for the cytotoxicity assays. Cells were first tested at three different concentrations, 100, 10, and 1 μ M, at both 24 and 72 h of incubation, as a preliminary determinant of toxicity. Compounds exhibiting >50% reduction in viability were subjected to subsequent IC₅₀ determination. These assays were performed with the CellTiter-Glo cell viability assay (Promega Corp.) as per the manufacturer's instructions. Resultant luminescence readings (as an indication of intracellullar ATP levels as a surrogate for viability) were measured with an integration time of 0.25 s on a Tecan Infinite M200 microplate reader.

ASSOCIATED CONTENT

S Supporting Information

Tables summarizing the intermediates synthesized by routes A–D, detailed synthetic procedures and characterization data, and calculated QikProp parameters. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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ABBREVIATIONS USED

PD, Parkinson's disease; MAO, monoamine oxidase; rMAO, rat monoamine oxidase; hMAO, human monoamine oxidase; 6-OHDA, 6-hydroxydopamine; COMT, catechol-*O*-methyl transferase; X-Phos, 2-dicyclohexylphosphino-2',4',6'-triisopropylbiphenyl; MTT, 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide; TAMH, transforming growth factor- α overexpressing mouse hepatocytes.

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