A Simple Synthesis of C-8 Modified 2-Keto-3-deoxy-D-*manno*-octulosonic Acid (KDO) Derivatives

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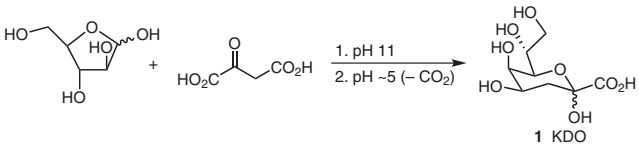
This paper is dedicated to Prof. Gerry Pattenden on the occasion of his 70th birthday – thank you for the wonderful insight into chemistry.

Abstract: This paper describes a simple and efficient method with which to prepare C-8 modified 2-keto-3-deoxy-D-*manno*-octulosonic acid (KDO) derivatives from C-5 modified arabinose derivatives.

Key words: carbohydrates, ulosonic acids, aldol condensation, benzylation, protecting groups

The eight-carbon acidic sugar 2-keto-3-deoxy-D-*manno*-octulosonic acid (KDO, **1**) is a member of the family of 3-deoxy-2-ulosonic acids. KDO is an essential component of the outer membrane lipopolysaccharide (LPS), or lipo-oligosaccharide (LOS), of Gram-negative bacteria, where it forms the link between the lipid A and polysaccharide components of the LPS.[1–3] KDO appears to be unique to Gram-negative bacteria and, since KDO biosynthetic pathway knockout mutants are no longer viable,[1b,4] it is reasonable to assume that small molecule inhibitors of KDO biosynthesis have the potential to act as a new class of antibacterial agent.[3] As part of a program aimed at exploring the binding specificity of enzymes involved in the biosynthesis of KDO and its incorporation into the LPS, we required an efficient and flexible route towards a range of structurally modified KDO derivatives. Specifically, we were interested in preparing C-8 modified KDO derivatives since, of those bacteria that have had their LPS structures elucidated, most have more than one KDO unit present in their core region, with the additional KDO residues usually attached to KDO through the C-4 or C-8 hydroxy groups.[2]

The chemical synthesis of KDO (**1**) was first reported by Ghalambor and Heath in 1963,[5] following a method developed by Cornforth for the synthesis of sialic acid.[6] The method involved a base-catalysed aldol condensation between D-arabinose and oxalacetic acid, followed by decarboxylation under mildly acidic conditions (Scheme 1). Since this first report there have been numerous papers describing the synthesis of KDO,[1a,7] often involving either the use of D-mannose and the addition of a two-carbon unit,[8] or starting from D-arabinose and adding a three-carbon unit.[9]



1 KDO

Scheme 1 Cornforth's method for the synthesis of KDO

Whilst there are several published procedures for the synthesis of KDO, many of these methods require the use of expensive reagents or involve multiple steps. Our aim was to find a single method that utilized relatively cheap reagents and would be flexible enough to allow the preparation of structural analogues of KDO. Towards this end, the method based on Cornforth's procedure, although not giving high yields, appeared attractive to us. In 1986 McNicholas et al.[9a] described a modification of the original procedure for the synthesis of KDO, wherein sodium carbonate was used to ensure that an optimal pH 11 was maintained for the aldol condensation. In this way KDO was consistently obtained in yields of ~35%. Subsequently, it was suggested[9b] that the modest yields for KDO may be due to the lack of optimised conditions for the decarboxylation step that follows the aldol condensation. By using a catalytic amount of $NiCl_2$ in the decarboxylation step, the yield of KDO was reported to increase to an acceptable 66%.[9b]

In using Cornforth's method to prepare our target C-8 modified KDO derivatives, we required ready access to C-5 modified arabinose derivatives, since carbon-5 of arabinose becomes carbon-8 of KDO after the aldol condensation. Our target compounds were those in which the normal 5-hydroxy group of arabinose was replaced by –OR, –SR or –NHR. It was reasoned that, after transformation into the corresponding C-8 modified KDO derivatives, such functional groups still have the potential to participate in hydrogen bonding interactions with KDO-recognising enzymes. In order to introduce these types of groups at C-5, we envisaged preparing a differentially protected arabinose derivative such as **2** (Scheme 2). Compounds such as **2** have been described before,[10–12] and the synthetic approaches generally rely on selectively introducing a temporary protecting group at the 5-hydroxy group, protecting the three remaining hydroxy groups, and then removal of the C-5 protecting group. However, as noted by others,[10] this proved more challeng-

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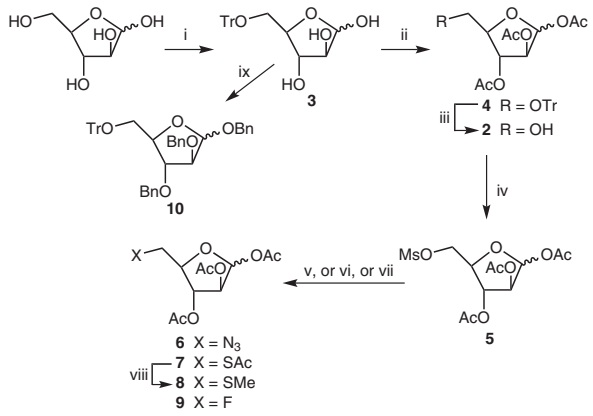
ing than could be inferred from previously reported procedures.

Our initial synthetic approach to compounds such as **2** involved attempts at introducing a silyl protecting group onto the C-5 hydroxy group of arabinose. However, we found it difficult to achieve consistent outcomes using this approach, a finding comparable with the observations of others.[10] Ultimately, we settled on a synthetic sequence to **2** involving the use of a trityl ether at C-5 and acetate groups on the other hydroxyls. Accordingly, treatment of D-arabinose with trityl chloride in pyridine gave the known[12] 5-*O*-trityl ether **3** which, without purification, was acetylated (pyridine and acetic anhydride) to give the fully protected compound **4**[12] (Scheme 2). Removal of the *O*-trityl group of **4** was achieved smoothly and efficiently by treatment with 80% acetic acid at 65 °C for one hour.[12] In this way the key 5-hydroxy arabinose derivative **2** was obtained in 49% yield from arabinose. Importantly, we found that when working on a large scale (>5 g of arabinose), the tritylation, acetylation, de-*O*-tritylation sequence could be carried out without any purification of intermediates.



Scheme 2 *Reagents and conditions*: (i) Ph_3CCl, pyridine, r.t., 48 h, 58%; (ii) Ac_2O, pyridine, r.t., 16 h, 96%; (iii) 80% aq AcOH, 65 °C, 1 h, 88%; (iv) MsCl, CH_2Cl_2, DIPEA, 0 °C to r.t., 1 h, 97%; (v) NaN_3, DMF, 60 °C, 16 h, 73%; (vi) KSAc, acetone, 40 °C, 72 h, 82%; (vii) TBAF or DAST; (viii) $H_2NNH_2{\cdot}HOAc$, DMF, $(CH_3O)_2SO_2$, r.t., 4 h, 61%; (ix) NaH, BnBr, TBAI, DMF, 0 °C to r.t.

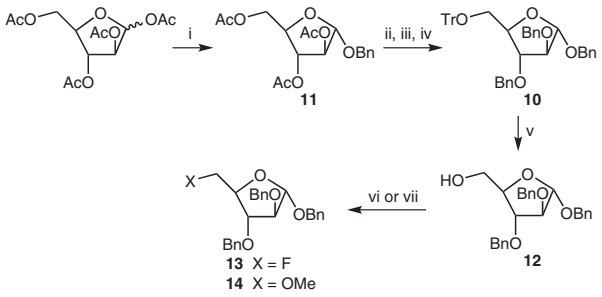
With the key alcohol **2** in hand, we turned our attention to introducing alternative functionality at C-5, and opted for a strategy involving displacement of a good leaving group. Thus, exposure of **2** to methanesulfonyl chloride gave the mesylate **5** (97%) which, upon treatment with sodium azide in DMF, afforded the known[10,11] 5-azido derivative **6** (Scheme 2). We felt that an azide would provide maximum flexibility for the preparation of a range of KDO derivatives with nitrogen-based functionality, since the azide group could be reduced after the aldol condensation, allowing the introduction of both amine and amide functionality at C-8 in KDO.

For the introduction of sulfur functionality at C-5, the mesylate **5** was treated with potassium thioacetate in acetone to give the 5-thioacetyl derivative **7**[12] in 82% yield. Selective unmasking of the thioacetate group in **7** was achieved by treatment with hydrazine acetate;[13] subsequent exposure to dimethyl sulfate in situ gave the thiomethyl derivative **8** in 61% yield.

At this stage, we felt that introducing fluorine at C-5 would be beneficial for our studies, since fluorine is detectable by NMR and would therefore provide us with an excellent 'spectroscopic handle' for studies involving interactions between our KDO derivatives and KDO-recognising enzymes. Additionally, fluorine is known to have similar electronic properties to hydroxy groups but is unable to participate in hydrogen bonding interactions;[14] it is therefore an excellent functionality for use in studies aimed at exploring ligand–protein interactions. Unfortunately, we found the introduction of fluorine to be quite problematic. Attempted fluoride displacement of the mesylate group in **5** using tetrabutylammonium fluoride (TBAF),[15] in our hands gave a complex mixture of products from which the desired 5-fluoro-arabinose derivative **9** was obtained in poor yield. A common reagent for converting alcohols directly into fluorides is diethylamino sulfur trifluoride (DAST). Unfortunately however, treatment of **2** with DAST in dichloromethane,[16] gave poor (~15%) yields of the desired product. It has been reported that changing the solvent to diglyme can improve the outcome of this reaction[10] but, in our hands, this was not the case. We reasoned that the attempted introduction of fluoride failed because the acetate protecting groups were too labile for this type of transformation, since some of the components we isolated from these reactions contained fewer acetate groups than expected.

Dissatisfied with this approach, we investigated an alternative strategy involving the use of the more stable benzyl ether protecting groups. Accordingly, the 5-*O*-trityl ether **3** was treated with sodium hydride and benzyl bromide in DMF. Unfortunately, but perhaps not unexpectedly, this resulted in a highly complex reaction mixture from which the desired tri-*O*-benzyl ether **10** was obtained in modest (<40%) and highly variable yield. Interestingly, one of the products from this benzylation reaction appeared, by inspection of the ^1H NMR spectrum, to contain four benzyl ether groups in addition to the 5-*O*-trityl ether moiety, which was possibly due to sodium hydride mediated reduction of the acyclic aldehyde form of the sugar.[17] After considerable experimentation, we opted for a longer sequence of reactions that proved far more reliable and efficient overall. Accordingly, acetylation of arabinose (Ac_2O/pyridine) followed by treatment with $SnCl_4$ and benzyl alcohol[18] gave the α-glycoside **11** in 86% yield (Scheme 3). Removal of the acetate groups, tritylation of the C-5 hydroxy group, and then benzylation of the remaining two hydroxy groups gave the desired arabinoside **10** in 30% yield from **11**. Removal of the trityl group from **10** proceeded as expected to give the target alcohol **12** in 72% yield (Scheme 3).

To see if changing the protecting groups would indeed result in an improved outcome, the alcohol **12** was exposed to DAST. To our delight, the desired 5-fluoro derivative



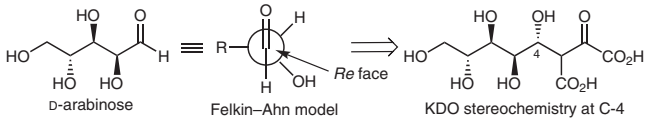
Scheme 3 *Reagents and conditions*: (i) $SnCl_4$, BnOH, MeCN, 1 h, 86%; (ii) aq NaOH (1 M), MeOH, r.t., 16 h, 72%; (iii) Ph_3CCl, pyridine, r.t., 48 h, 52%; (iv) NaH, BnBr, TBAI, DMF, 0 °C to r.t., 16 h, 81%; (v) 80% aq AcOH, 65 °C, 1 h, 72%; (vi) DAST, CH_2Cl_2, 0 °C to r.t., 3 h, 58%; (vii) NaH, MeI, DMF, 16 h, 81%.

13 was obtained in a modest 58% yield, which was a significant improvement over our results with substrates bearing acetate protecting groups. Attempts to increase this yield by altering the reaction temperature and the number of equivalents of DAST failed to significantly improve the outcome. Having prepared the alcohol **12**, we felt we should exploit the robust nature of the benzyl ethers by introducing a methyl ether at C-5. Thus, treatment of **12** with sodium hydride and methyl iodide smoothly gave the methyl ether **14** in 81% yield (Scheme 3).

Each of the C-5 modified arabinose derivatives was deprotected in the standard way. Acetate protecting groups were removed by treatment with dilute sodium hydroxide (1 M) in methanol, whilst benzyl ethers were removed by hydrogenolysis. Interestingly, deprotection of the 5-thioacetyl derivative **7** gave the known 5-thio-D-arabinose,[12] as a mixture of the pyranose and furanose forms in a 3:1 ratio, respectively. Attempted deprotection of the ester groups of the 5-thiomethyl derivative **8** resulted in extensive decomposition, even when dilute aqueous lithium hydroxide and shorter reaction times were employed.

Having successfully prepared a range of C-5 modified arabinose derivatives, we next explored their condensation with oxalacetic acid to give KDO derivatives. Initially we followed the method reported by McNicholas et al.[9a] using arabinose itself as a model compound, and obtained the ammonium salt of KDO in ~30% yield after ion-exchange chromatography on Amberlite CG-400 (HCO_3^-) resin. As expected, the product obtained in this way was a complex mixture of components due to both the lack of stereoselectivity of the aldol condensation, giving a mixture of epimers at C-4, as well as the fact that reducing sugars exist as a mixture of α- and β-anomers in both pyranose and furanose forms. Since this aldol condensation is conducted in the absence of chelation control, the stereochemical outcome of the enolate addition to the aldehyde should fit the Felkin–Ahn model,[19] where the incoming nucleophile preferentially attacks from the sterically less-hindered (*Re*) face of the aldehyde (Scheme 4).

In this way we, as well as others,[1a,9a,b] have observed that the stereochemical outcome for the condensation between

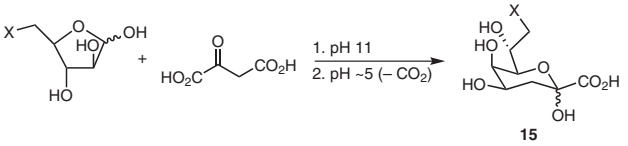


Scheme 4 Diagram showing the preferential *Re*-face attack on D-arabinose, giving the desired KDO stereochemistry at C-4

arabinose and oxalacetic acid appears to favour the formation of KDO over 4-*epi*-KDO, with a ratio of KDO to 4-*epi*-KDO typically around 4:1. Purification of our KDO product using Dowex 1 × 8 (200–400 mesh) anion-exchange chromatography following the method reported by Kragl et al.,[20] resulted in separation of KDO from 4-*epi*-KDO, allowing complete assignment of their respective ^{1}H NMR spectra (see the Supporting Information).

Although satisfied that we could prepare and isolate pure KDO, the poor yield was a significant limitation of this approach, especially given that many of our arabinose derivatives were not trivial to prepare. In an attempt to optimise this reaction, we repeated the condensation between arabinose and oxalacetic acid using the $NiCl_2$ modification reported by Shirai and Ogura,[9b] and obtained a 60% yield of KDO as a ~5:1 epimeric mixture at C-4. Whilst delighted with this improved chemical yield, a limitation with this method (as indeed with all previous reports on the preparation of KDO using this approach) is the use of a molar excess of arabinose. Since, in our strategy, the arabinose derivative is the more difficult coupling partner to prepare, the aldol condensation was repeated using a molar excess of oxalacetic acid. In this way, KDO could be routinely obtained in ~65% yield based on arabinose. Having established appropriate reaction conditions[21] for the synthesis of KDO, we turned our attention to using our C-5 modified arabinose derivatives in the aldol condensation with oxalacetic acid. The results are summarised in Table 1, and clearly show that this approach represents an efficient and flexible synthesis of C-8 modified KDO derivatives **15**. The KDO derivatives **15** were all obtained together with their 4-*epi* analogues, with the ratio of **15** to 4-*epi*-**15** typically better than 5:1 (as determined by ^{1}H NMR). Reduction of the azide group in **15** (X = N_3) to an amine (**15**, X = NH_2) was accomplished by hydrogenation.

Table 1 Synthesis of C-8 Modified KDO Derivatives



X	Yield of **15** (%)
OH	65
OMe	84
N_3	87
SH	67

In conclusion, we have shown that C-8 modified KDO derivatives can be prepared efficiently from readily available starting materials. We are currently investigating simple ways to improve the stereochemical outcome of the aldol condensation between structurally modified arabinose derivatives and oxalacetic acid. We are also using the C-8 modified KDO derivatives described herein as probes for KDO-utilizing proteins. These investigations will be described in due course.

Supporting Information for this article is available online at http://www.thieme-connect.com/ejournals/toc/synlett.

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(21) General procedure for the synthesis of KDO and C-8 modified derivatives: D-Arabinose (500 mg, 3.3 mmol) was added to a solution of Na_2CO_3 (860 mg, 8.1 mmol) in H_2O (8 mL). Oxalacetic acid (525 mg, 4.0 mmol) was added portionwise over 5 min, and the solution was adjusted to pH 11 using NaOH (10 M). After stirring for 2 h at r.t. the solution was acidified to pH 5 using AcOH, $NiCl_2$ (7.5 mg, 0.03 mmol) added, and the mixture was heated at 50 °C for 1 h. After cooling to r.t., the reaction was neutralised to pH 8 with ammonia and the KDO product was isolated by column chromatography using CG-400 (HCO_3^-) resin, washing first with H_2O and then eluting with ammonium hydrogen carbonate (0.5 M). The eluant was concentrated under reduced pressure, and then freeze-dried. The lyophilised residue was purified using reversed-phase (C_{18}) silica gel with H_2O as the mobile phase. Fractions containing KDO can be visualised as bluish-grey spots on silica gel TLC plates ($CHCl_3$–MeOH–H_2O, 5:5:1) by staining with anisaldehyde–sulfuric acid dip.