Chiral Auxiliaries as Docking/Protecting Groups in Biohydroxylation: (S)-Specific Hydroxylation of Enantiopure *tert*-Butyl-Substituted Spirooxazolidines Derived From Cyclopentanone

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Dedicated to the memory of Professor Herbert Holland^[‡]

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An enantiopure *tert*-butyl-substituted derivative of cyclopentanone, which is a vital member of the chiral docking/ protecting group series, is employed, for the first time, to stereoselectively (90 % *de*) introduce an (*S*)-configured hydroxyl group onto an unactivated carbon atom present in the cyclo-

pentane ring using the fungus *Beauveria bassiana* ATCC 7159.

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Introduction

The stereoselective introduction of hydroxyl groups onto unactivated carbon atoms present in organic compounds is, generally, still a very daunting task in modern synthetic chemistry.^[1,2] For this reason, we have been investigating the docking/protecting (d/p) group concept as a means to easily employ biohydroxylation in preparative chemistry.^[3,4] We have found that a range of organic compounds, such as carboxylic acids, alcohols, aldehydes and ketones, can be easily hydroxylated following this concept.

Chiral d/p groups have also been investigated^[5,6] and employed for the hydroxylation of ketones. In this manner, the stereoselective functionalisation of these compounds was achieved. Indeed, depending on the nature of the chiral d/ p group used, the configuration of the introduced hydroxyl moiety could be readily determined.

The d/p concept is a three-step process (Scheme 1). Using parent ketone 1 and a chiral d/p group as an example, derivatisation gives the enantiopure biohydroxylation substrate 2 in the first step. Subsequent exposure of this spirooxazolid-

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- ^[1] Not only a leader in the field of biohydroxylation but a most valued friend and colleague



Scheme 1. The d/p concept as exemplified by a chiral d/p group and model ketone 1: step 1: (2R)-2-amino-1-propanol, K₂CO₃, CH₂Cl₂, 20 °C, 24 h, followed by BzCl, 20 °C, 24 h; step 2: *Beauveria bassiana* ATCC 7159; step 3: BnBr, NaH, THF/DMF, 20 °C followed by IR 120 (H⁺, cat), CH₃CN, 20 °C

ine derivative to a suitable microorganism, for example *Beauveria bassiana* ATCC 7159, yields hydroxylated product **3**. The third and final step of this approach is to remove the d/p group to furnish the desired hydroxylated product **4**. In this particular example, an atypical protection step (benzylation) was also needed to prevent elimination of the introduced group.

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Despite the fact that a wide range of chiral d/p groups has been investigated, while the (*R*)-configured product (3) could be prepared in 90% de – recrystallisation increased this value to over 99% – the (*S*)-configured counterpart could not be obtained with more than 20% de from other substrates.

In this account we wish to disclose results obtained from the last member of this chiral d/p group series, namely compounds containing *tert*-butyl substituents. In this manner, the synthesis of the (S)-configured biohydroxylation product could be accomplished with 90% de.

Results and Discussion

The commercially available, but relatively expensive, *tert*butyl amino alcohols **5** and **6** required for substrate synthesis could be easily prepared from the corresponding cheaper amino acids in high yield (over 80%) by the reduction method reported by Drauz and coworkers.^[7] Subsequent reaction with cyclopentanone afforded the desired, enantiopure substrates **7** and **8**, respectively (Scheme 2). In contrast to previous observations, these compounds were obtained in only modest yields under standard, unoptimized conditions^[5,8] (22% and 17%, respectively).



Scheme 2. Preparation of enantiopure *tert*-butyl biohydroxylation substrates 7 and 8: step 1: NaBH₄, I₂, THF; step 2: cyclopentanone, K₂CO₃, CH₂Cl₂, 20 °C, 24 h followed by BzCl, 20 °C, 24 h

For the biohydroxylation step, the fungus Beauveria bassiana ATCC 7159 was then employed in the usual manner to afford crystalline products 9 and 10 in modest yields (50% and 22%, respectively; Scheme 3). We believe that this outcome could be improved by optimising the fermentation parameters if required.^[9] More importantly, however, the de and configuration of the introduced alcohol were deemed to be very interesting. While the *de* of product 10 was found to be rather modest (18%) by HPLC, compound 9 exhibited a high de of 90%. After subsequent benzylation and d/ p group removal, the configuration of the newly introduced chiral center could be determined by GC comparison of the resulting ketones 4 and 11 with model compounds. Satisfyingly, although ketone 4 was found to be (R)-configured (19% ee), ketone 11 was found to be (S)-configured and with a high ee (89%). It should be mentioned at this

point that, based on published experiments^[5] with product **3**, this *ee* value should increase after simple recrystallisation of compound **9**. This result nicely complements those obtained from the (*R*)-methyl d/p used in previous studies (Scheme 1) where product **3** could be obtained in 90% *de* in a single biohydroxylation step.



Scheme 3. Biohydroxylation of *tert*-butyl substrates 7 and 8, subsequent benzylation and d/p removal: step 1: *Beauveria bassiana* ATCC 7159; step 2: BnBr, NaH, THF/DMF, 20 °C followed by IR 120 (H⁺, cat), CH₃CN, 20 °C

Conclusions

In conclusion, it has been shown that, for the biohydroxylation step, the appropriate choice of chiral d/p group can be exploited to obtain either configuration of the newly introduced alcohol in 90% *de*. Simple recrystallisation of this biohydroxylation product could improve this value to over 99% *de* if desired. Simple removal of the chiral d/p group yields the corresponding ketone derivative.

Experimental Section

General Remarks: Please refer to ref.^[3] for all general methods unless stated otherwise. NMR: signals from the minor isomer are given in *italics*. The ¹³C NMR spectra for spirooxazolidine derivatives 7 to **13** suggested that a number of different conformations were present in the NMR sample (see Figure 1 for numbering scheme).



Figure 1. Numbering scheme for the spirooxazolidine derivatives

Amino Alcohols 5 and 6:¹H and ¹³C NMR spectroscopic data, optical rotations and melting points were in agreement with published values.^[10]

Spirooxazolidine Derivative 7: Employing the standard procedure for preparing spirooxazolidine derivatives,^[5] compound 7 was prepared in 22% yield as a pale-yellow, crystalline solid. M.p. 97.0–

97.5 °C. [α]_D²⁰ = +65.3 (c = 1.11 in CH₂Cl₂). ¹H NMR (CDCl₃): δ = 0.72 (br. s, 9 H, 11-H, 12-H, 13-H), 1.40–2.10 (br. m, 6 H, 6^a-H, 7-H, 8-H, 9^a-H), 2.28, 2.73 (2×br. m, 2×1 H, 6^b-H, 9^b-H), 3.95 (m, 3 H, 2-H, 3-H), 7.38 (s, 5 H, CO*Ph*) ppm. ¹³C NMR (CDCl₃): δ = 24.7, 25.2 (2×br. s, C-7, C-8), 27.4 (C-11, C-12, C-13), 35.2, 37.5 (2×br. s, C-6, C-9), 35.7 (C-10), 65.5, 66.2 (2×br. s, C-2, C-3), 105.7 (C-5), 127.7, 128.4, 129.9, 138.8 (CO*Ph*), 170.2 (*C*OPh) ppm. MS (70 eV): *mlz* (%) = 287 (15) [M]⁺, 258 (13) [M – C₂H₅]⁺, 230 (43) [M – C₄H₉]⁺, 105 (100) [Bz]⁺, 77 (16) [Ph]⁺. HRMS: calcd. 287.1885; found 287.1887.

Spirooxazolidine Derivative 8: Employing the standard procedure for preparing spirooxazolidine derivatives,^[5] compound **8** was prepared in 17% yield as a pale-yellow, crystalline solid. M.p. 92.0–95.0 °C. [α]_D²⁰ = -57.8 (*c* = 1.51 in CH₂Cl₂). ¹H NMR (CDCl₃): δ = 0.72 (br. s, 9 H, 11-H, 12-H, 13-H), 1.40–2.10 (br. m, 6 H, 6^a-H, 7-H, 8-H, 9^a-H), 2.28, 2.73 (2×br. m, 2×1 H, 6^b-H, 9^b-H), 3.95 (m, 3 H, 2-H, 3-H), 7.38 (s, 5 H, CO*Ph*) ppm. ¹³C NMR (CDCl₃): δ = 24.7, 25.2 (2×br. s, C-7, C-8), 27.4 (C-11, C-12, C-13), 35.2, 37.5 (2×br. s, C-6, C-9), 35.7 (C-10), 65.5, 66.2 (2×br. s, C-2, C-3), 127.7, 128.4, 129.9, 138.8 (CO*Ph*) ppm. MS (70 eV): *m/z* (%) = 287 (15) [M]⁺, 258 (12) [M – C₂H₅]⁺, 230 (43) [M – C₄H₅]⁺, 105 (100) [Bz]⁺, 77 (17) [Ph]⁺. HRMS: calcd. 287.1885; found 287.1875.

Biohydroxylation Product 9: Employing published methods,^[5] treatment of compound 7 (1.277 g) with Beauveria bassiana ATCC 7159 gave the title compound (586.6 mg), together with unreacted starting material (97.1 mg), as a white solid (50% yield, taking into account unreacted starting material). M.p. 89.0–104.0 °C. $[\alpha]_{D}^{20}$ = +79.4 (c = 1.06 in CH₂Cl₂); 90% de (HPLC: CHIRALCEL AD, T = 10 °C, 0.5 mL min⁻¹, *n*-heptane/IPA = 4:1, measured at 238 nm), retention time of $(3S, 5\Xi, 7R)$ -9 = 17.6 min, retention time of $(3S,5\Xi,7S)$ -9 = 20.8 min. ¹H NMR (CDCl₃): δ = 0.72 (br. s, 9 H, 11-H, 12-H, 13-H), 1.65–2.90 (3×br. m, 7 H, 6-H, 8-H, 9-H, OH), 4.00 (br. m, 3 H, 2-H, 3-H), 4.39 (br. s, 1 H, 7-H), 7.39 (s, 5 H, COPh) ppm. ¹³C NMR (CDCl₃): δ = 27.4 (C-11, C-12, C-13), 34.3, 35.3 (2×br. s, C-6, C-9), 35.6 (C-10), 42.7 (br. s, C-8), 65.8 (br. s, C-2, C-3), 73.4 (br. s, C-7), 105.0 (br. s, C-5), 127.6, 128.4, 130.0, 138.4 (COPh) ppm. MS (70 eV): m/z (%) = 303 (12) [M]⁺, 286 (2) $[M - OH]^+$, 274 (10) $[M - C_2H_5]^+$, 258 (16), 246 (28) $[M - C_4H_9]^+$, 124 (42), 105 (100) [Bz]⁺, 77 (27) [Ph]⁺. HRMS: calcd. 303.1834; found 303.1831.

Biohydroxylation Product 10: Employing published methods,^[5] treatment of compound 8 (759.6 mg) with Beauveria bassiana ATCC 7159 gave the title compound (160.6 mg), together with unreacted starting material (65.8 mg), as a white solid (22% yield, taking into account unreacted starting material). M.p. 85.0-105.0 °C; 18% de (HPLC, CHIRALCEL OD-H, T = 10 °C, 0.5 mL/min, n-heptane/IPA = 7:2, measured at 238 nm), retention time of $(3R,5\Xi,7S)$ -9 = 16.0 min, retention time of $(3R,5\Xi,7R)$ -9 = 22.1 min. ¹H NMR (CDCl₃): $\delta = 0.72$ (br. s, 9 H, 11-H, 12-H, 13-H), 1.65–2.90, 3.12 (4×m, 7 H, 6-H, 8-H, 9-H, OH), 4.00 (m, 3 H, 2-H, 3-H), 4.39 (br. m, 1 H, 7-H), 7.39 (s, 5 H, COPh) ppm. ¹³C NMR (CDCl₃): δ = 27.4 (C-11, C-12, C-13), 34.4, 34.8, 35.3 (3×br. s, C-6, C-9), 35.7 (C-10), 42.7, 46.0 (2×br. s, C-8), 66.0 (br. s, C-2, C-3), 73.5 (br. s, C-7), 127.6, 128.5, 130.1, 138.4 (COPh) ppm. MS $(70 \text{ eV}): m/z \ (\%) = 303 \ (11) \ [M]^+, 286 \ (2) \ [M - OH]^+, 274 \ (10) \ [M - OH]^+,$ C_2H_5 ⁺, 258 (13), 246 (27) [M - C_4H_9]⁺, 124 (40), 105 (100) [Bz]⁺, 77 (26) [Ph]⁺. HRMS: calcd. 303.1834; found 303.1841.

Compound 12: Standard benzylation^[5] of **9** (135.0 mg) furnished derivative **12** (146.2 mg; Figure 2) in 83% yield as a pale-yellow syrup. $[a]_D^{20} = +65.5$ (c = 1.90 in CH₂Cl₂). ¹H NMR (CDCl₃): $\delta = 0.65$ (br. s, 9 H, 11-H, 12-H, 13-H), 1.72, 2.13, 2.55 (3×m, 6 H, 6-H, 8-H, 9-H, OH), 3.60, 3.95 (2×m, 3 H, 2-H, 3-H), 4.18 (m, 1 H,

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7-H), 4.45 (br. s, 2 H, CH_2 Ph), 7.24 (m, 10 H, COPh) ppm. ¹³C NMR (CDCl₃): δ = 27.4 (C-11, C-12, C-13), 31.3, 36.0 (2×br. s, C-6, C-9), 35.6 (C-10), 42.4 (br. s, C-8) 65.8, 66.3 (2×br. s, C-2, C-3), 70.7 (CH_2 Ph), 79.8 (br. s, C-7) 104.6 (br. s, C-5), 127.0, 127.5, 127.6, 127.8, 128.4, 128.7 129.9, 131.4, 138.6, 139.0 (COPh) ppm. MS (70 eV): m/z (%) = 393 (3) [M]⁺, 364 (1) [M – C_2H_5]⁺, 336 (3) [M – C_4H_9]⁺, 287 (5), 258 (9), 245 (7), 214 (5), 182 (4), 146 (4), 124 (5), 105 (100) [Bz]⁺, 91 (37) [Bn]⁺, 77 (28) [Ph]⁺. HRMS: calcd. 393.23039; found 393.23040.



Figure 2. Compounds 12 and 13

Compound 13: Standard benzylation^[5] of **10** (73 mg) furnished derivative **13** (83.9 mg; Figure 2) in 89% yield as a pale-yellow syrup. ¹H NMR (CDCl₃): $\delta = 0.65$ (br. s, 9 H, 11-H, 12-H, 13-H), 1.72, 2.13, 2.55, *3.08* (4×m, 6 H, 6-H, 8-H, 9-H, OH), *3.60*, 3.95 (2×m, 3 H, 2-H, 3-H), 4.18 (m, 1 H, 7-H), 4.45 (br. s, 2 H, CH₂Ph), 7.24 (m, 10 H, COPh) ppm. ¹³C NMR (CDCl₃): $\delta = 27.4$ (C-11, C-12, C-13), 31.3, 36.0 (2×br. s, C-6, C-9), 35.6 (C-10), 42.5, *43.9* (2×br. s, C-8) 65.8, 66.3 (2×br. s, C-2, C-3), 70.6, *71.3* (CH₂Ph), *79.8* (2×br. s, C-7) 104.6 (br. s, C-5), 127.0, 127.5, 127.8, 127.8, 128.4, 128.7 130.0, 131.5, 138.6, 139.0 (COPh) ppm. MS (70 eV): *mlz* (%) = 393 (2) [M]⁺, 364 (1) [M – C₂H₃]⁺, 336 (2) [M – C₄H₉] ⁺, 287 (3), 258 (5), 245 (4), 214 (3), 182 (2), 146 (5), 124 (2), 105 (100) [Bz]⁺, 91 (49) [Bn]⁺, 77 (25) [Ph]⁺. HRMS: calcd. 393.23039; found 393.22975.

Ketones 4 and 11: The known^[5] title compounds were obtained for chiral GC experiments from the corresponding benzylated derivatives **13** and **12**, employing published conditions.^[3] These compounds were then compared with reference substances available from previous studies.

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