

Accepted Manuscript

Synthesis and evaluation of thieno[3,2-*d*]pyrimidine derivatives as novel FMS inhibitors

Yu-Yon Kim, Jaeyul Choi, Kyungjin Choi, Changhee Park, Young Hoon Kim, Kwee Hyun Suh, Young Jin Ham, Sun Young Jang, Kyu-Hang Lee, Kwang Woo Hwang

PII: S0960-894X(18)30904-1
DOI: <https://doi.org/10.1016/j.bmcl.2018.11.037>
Reference: BMCL 26148

To appear in: *Bioorganic & Medicinal Chemistry Letters*

Received Date: 19 September 2018
Revised Date: 16 November 2018
Accepted Date: 18 November 2018

Please cite this article as: Kim, Y.-Y., Choi, J., Choi, K., Park, C., Hoon Kim, Y., Suh, K.H., Ham, Y.J., Jang, S.Y., Lee, K.-H., Hwang, K.W., Synthesis and evaluation of thieno[3,2-*d*]pyrimidine derivatives as novel FMS inhibitors, *Bioorganic & Medicinal Chemistry Letters* (2018), doi: <https://doi.org/10.1016/j.bmcl.2018.11.037>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Short communication**Title**

Synthesis and evaluation of thieno[3,2-*d*]pyrimidine derivatives as novel FMS inhibitors

Yu-Yon Kim^{a,b}, Jaeyul Choi^b, Kyungjin Choi^b, Changhee Park^b, Young Hoon Kim^b, Kwee Hyun Suh^b, Young Jin Ham^b, Sun Young Jang^b, Kyu-Hang Lee^b, Kwang Woo Hwang^{a,1}

^aHost Defense Modulation Lab, Collage of Pharmacy, Chung-Ang University, 84 Heukseok-Ro, Dongjak-Gu, Seoul, 06974, Republic of Korea

^bHanmi Research Center, Hanmi Pharm. Co. Ltd., 550 Dongtangiheung-Ro, Hwaseong-Si, Gyeonggi-Do, 18469, Republic of Korea

E-mail addresses:

YYK, yykim@hanmi.co.kr; JYC, alchemistj@hanmi.co.kr; KJC, booim@hanmi.co.kr; CHP, cchange@hanmi.co.kr; YHK, yhkeem@hanmi.co.kr; KHS, khsuh@hanmi.co.kr; YJH, yjham@hanmi.co.kr; SYJ, jsy74@hanmi.co.kr; KHL, kyuhang@hanmi.co.kr; KWH, khwang@cau.ac.kr

¹Corresponding authors

Kwang Woo Hwang Ph.D., Host Defense Modulation Lab, College of Pharmacy, Chung-Ang University, 84 Heukseok-Ro, Dongjak-Gu, Seoul 06974, Republic of Korea.
khwang@cau.ac.kr

Abstract

Colony stimulating factor-1 receptor (CSF-1R or FMS) and its ligand, CSF-1, signaling regulates the differentiation and function of tumor-associated macrophages (TAMs) that play an important role in tumor progression. Derivatives of thieno[3,2-*d*]pyrimidine were synthesized and evaluated as kinase inhibitors of FMS. The most representative compound **21** showed strong activity ($IC_{50} = 2$ nM) against FMS kinase and served as candidate for proof of concept. Anti-tumor activity alone and/or in combination with paclitaxel was examined via a tumor cell growth inhibition assay and via an *in vitro* tumor invasion assay using human breast adenocarcinoma cells.

Keywords

FMS; CSF-1R; Colony stimulating factor-1; Tumor associated macrophages (TAMs); Anti-tumor activity; Human breast adenocarcinoma cells

Macrophages are characterized as classical M1- and alternative M2-subtypes depending on how they influence the immune response.¹ M1 macrophages influence the inflammatory response and anti-tumor immunity, whereas M2 macrophages are involved in immunosuppressive and tumor promoting activities. Tumor-associated macrophages (TAMs), which are classified as M2-polarized macrophages, are recruited by tumor cells and infiltrate into the stromal environment of malignant tumors. TAMs in the tumor microenvironment are known to play a crucial role in tumor progression, including tumor initiation and development, matrix remodeling and metastasis, and immune suppression.²⁻³

Colony stimulating factor-1 (known as CSF-1 or macrophage-colony stimulating factor, M-CSF) is a growth factor for monocyte and macrophage-derived cells. CSF-1 and its receptor, colony stimulating factor-1 receptor (CSF-1R or M-CSFR, also known as FMS) have been reported as having important roles in regulating the production, migration, differentiation, survival, and function of macrophages and their precursors.⁴ Therefore, over-activation of CSF-1/CSF-1R signaling in macrophages has been implicated in many disease states including osteoclast proliferation in bone osteolysis, and a number of inflammatory disorders as well as the growth/metastasis of cancer.⁵⁻¹⁴ Especially, a number of CSF1/CSF1R-targeting agents such as Pexidartinib and ARRY-382 are undergoing clinical trials in several cancer including breast cancer, prostate cancer, and melanoma.¹⁵

A previous report showed that the transfer of the CSF1 gene into tumor cells induced TAM infiltration into syngeneic BALB/c mice.¹⁶ High expressions of CSF-1 and CSF-1R in tumor and stromal cells and a high TAM density are known to be correlated with poor prognoses in many cancer types.¹⁷ CSF-1 promotes metastases via the regulation of TAMs in the tumor microenvironment.¹⁸ In human metastatic breast cancer, elevated CSF-1 levels were correlated with marked CSF-1R-positive macrophage infiltration into tumors and were associated with poor outcomes.¹⁹ Another study also showed that macrophage-deficient/CSF-1-deficient mice rarely developed pulmonary metastases, despite the rapid growth of primary mammary tumors.²⁰

Furthermore, some chemotherapies stimulate tumor cells to release CSF-1, which then recruits TAMs expressing CSF-1R. These TAMs in the microenvironment stimulate tumor progression by enhancing tumor resistance to chemotherapy. In combination therapy with paclitaxel and CSF-1R antagonists on primary and metastatic tumors of mammary tumor-bearing mice, inhibition of TAM infiltration via CSF-1R antagonists increased the anti-tumor efficacy of cytotoxic agents.²¹ For these reasons, modulation of the CSF-1 signaling pathway in the microenvironment has been of great interest in chemotherapy research.

In this paper, we describe the identification of the potent FMS inhibitor compound **21** and evaluate its anti-tumor activities against breast cancer. By computational screening compounds from the internal small molecule compound bank of Hanmi Pharmaceutical, we identified 1*H*-indazol-4-yl)thieno[3,2-*d*]pyrimidine derivative **7** (Fig. 1) as a highly potent (IC₅₀ = 13 nM) FMS inhibitor.

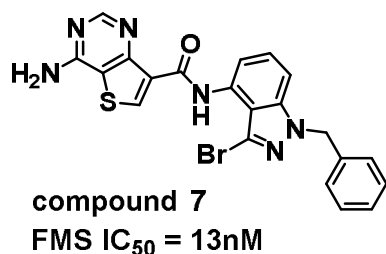








Figure 1. Hit compound 7

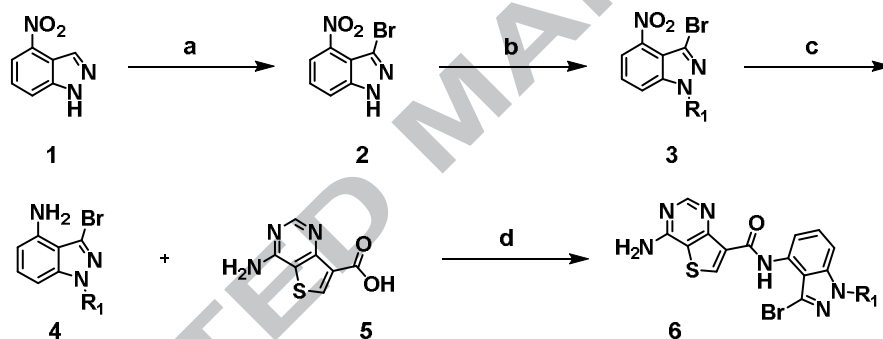
To improve its enzyme activity and cellular activities on NFS-60 cells that express endogenous FMS,²²⁻²³ structure-activity relationship (SAR) studies probed substituent effects at the N-1 position (R¹) and C-3 position of the indazole moiety (R²). Initially, analogues were synthesized with substitutions at R₁ (Table 1). The replacement of the phenyl moiety with methoxy phenyl moiety (**8**) led to an almost two-fold loss of enzyme and cellular activity. In contrast, the replacement of the phenyl moiety with a pyridine moiety (**9-11**) showed good enzyme activity (IC₅₀ < 10 nM). To further improve enzyme and cellular activity, we added an adopted substituent to the pyridine moiety. Halogen substituents (**12-13**) in the pyridine moiety had similar enzyme activities compared to regular pyridine moieties (**9-11**), and cellular activity did not dramatically improve. In contrast, alkyl substituents (**14-15**) on the pyridine moiety also showed improved cellular activity. In particular, a methyl substituent (**14**) on the pyridine moiety showed good cellular activity (GI₅₀ = 27.5 nM) in an NFS-60 cell line whose growth depends on FMS.²²⁻²³ Next, we tried to attach an alicyclic, heterocyclic, or alkynyl substituent (**16-18**) to the indazole moiety. Although all the resultant compounds showed good enzyme activities, they did not show sufficiently strong cellular activity.

Table 1. FMS enzyme and cellular activity of compounds 7-18

Compound	R ¹	FMS IC ₅₀ (nM)	NFS-60 GI ₅₀ (nM)
GW-2580	-	3	94
7		13	1005
8		37	2390
9		9	2502
10		6	77
11		7	135
12		13	233

13		12	243
14		23	28
15		10	34
16		30	254
17		19	62
18		26	>10000

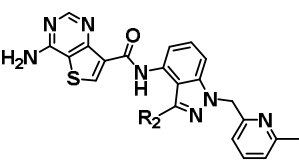
The synthetic route of R¹-changed derivatives is shown in **Scheme 1**. Compounds **7-18** were prepared from commercially available 4-nitro-1*H*-indazole (**1**). Bromination of **1** was performed using bromine gas, and gave an excellent yield (94%). The 1-substituted indazole of **2** was easily derived from an SN2 reaction using an alkyl-halide. Reduction of **3** was performed using Pd/C, and the amide coupling of **4** and **5** using commercially available HATU. Each step was isolated via column chromatography.



Scheme 1. Preparation of compounds **7-18**. Reagents and conditions: (a) Br₂, NaOAc, AcOH/CHCl₃(1:1), 20°C; (b) R¹CH₂Cl, K₂CO₃, DMF, rt; (c) H₂, Pd/C, THF/MeOH(1:1), rt; (d) HATU, DIPEA, DMF, rt.

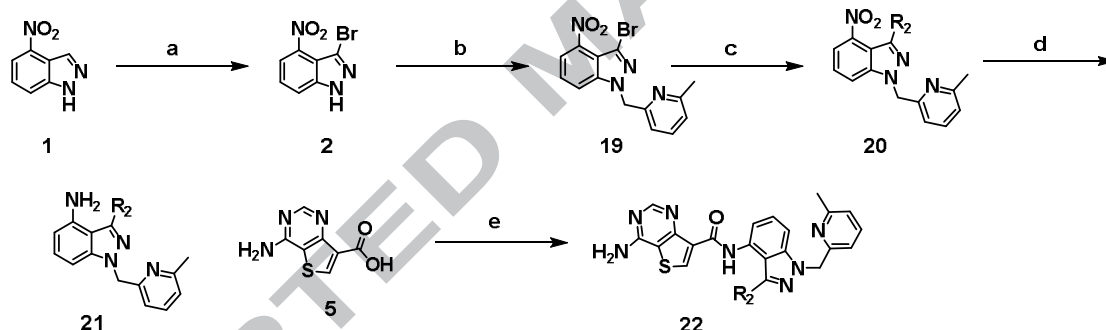
Compound **14** was the most potent compound of the R¹ changed-derivatives, based on cellular activity. This compound also showed good oral bioavailability (37%) (Supplementary data).

Next, we fixed R¹ as 2-methylpyridine and replaced R² with the C-3 position of an indazole moiety (compound **14**: Br). We then tried halogen (**23-24**) and alkyl (**25-27**) substituents at R² (Table 2). Compound **23** with a halogen moiety showed more potent cellular activity (3.4 fold) than compound **14**, and compound **25** with an alkyl moiety had the most potent enzyme activity (IC₅₀ = 2 nM). Compound **25** showed more stable liver metabolism than compound **23** in microsomal stability test using human hepatocytes. Remaining value of parent molecule at 60 minutes of compound **25** and **23** were 87% and 66%, respectively.

Table 2. FMS enzyme and cellular activity of compounds **23-27**


Compound	R ²	FMS IC ₅₀ (nM)	NFS-60 GI ₅₀ (nM)
23	Cl	20	8
24	I	9	30
25	Me	2	80
26	Et	9	>10000
27	Vinyl	12	4458

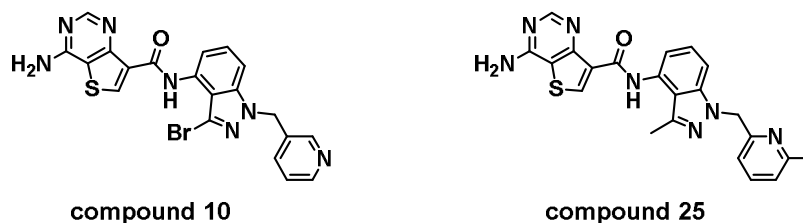
The synthetic route of R²-changed derivatives is shown in **Scheme 2**. Compounds **25-27** were prepared from commercially available 4-nitro-1H-indazole (**1**). After same two steps with scheme 1, the N-substituted indazole of **19** was produced using the Suzuki reaction in a good yield (85%). Reduction of **20** was performed using Pd/C, and the amide coupling of **21** and **5** using commercially available HATU. Each step was isolated via column chromatography.



Scheme 2. Preparation of compounds **25-27**. Reagents and conditions: (a) R₂B(OH)₂, K₃PO₄, Pd(OAc)₂, PhMe/H₂O, 100°C; (b) H₂, Pd/C, THF/MeOH(1:1), rt; (c) HATU, DIPEA, DMF, rt.

Having successfully achieved good enzyme and/or cellular activities, the drug metabolism and pharmacokinetic profiles for the more potent compounds (**10** and **25**) were evaluated. To understand the potential drug-drug interaction liabilities of these compounds, a CYP (Cytochromes P450) reversible inhibition assay using human liver microsomes was conducted. In this test, compound **25** showed weak inhibition of five major CYP enzymes, whereas compound **10** inhibited four CYP enzymes either strongly or moderately (IC₅₀ range from < 0.25 to 2.02 μM) except CYP2D6 (Table 3). These results indicate that compound **25** has a lower risk of drug-drug interactions than compound **10**. Moreover, compound **25** showed both good oral bioavailability (F = 36%) in mice (Table 4) and also an IC₅₀ value > 50 μM for the binding capacity to the hERG (Human ether-a-go-go-related gene) membrane. Based on these results, compound **25** was chosen for further study.

Table 3. CYP enzyme inhibition assay for compounds **10** and **25**



IC ₅₀ (μM)					
Compound	CYP1A2	CYP2C9	CYP2C19	CYP2D6	CYP3A4
10	< 0.25	1.38	2.02	> 20.00	0.31
25	> 20.00	> 20.00	> 20.00	> 20.00	> 20.00

CYP enzyme inhibition assay used the HLM with LC/MS/MS analysis method.

Table 4. Pharmacokinetic parameters for compound **25**

Compound	Route	t _{1/2} (hr)	C _{max} (ng/ml)	T _{max} (h)	AUC _{0-7h} (ng·hr/ml)	V _d (L/kg)	Cl (L/hr/kg)	BA (%)
25^a	IV	1.6	2546.5	NA	3367.2	2.0	0.9	NA
	PO	2.6	900	2.0	4037.8	7.5	2.0	36

Male CD-1 mice treated with 3 mg/kg IV and 10 mg/kg PO.

^a 5 % DMSO in 30 % PEG400 solution for IV and 0.5 % HPMC in 0.05 % Tween80 solution for PO

To determine the biochemical selectivity for compound **25**, twenty eight kinases were tested.²⁴ As summarized in Table 5, compound **25** was highly selective against FMS and inhibited FMS with an IC₅₀ of 2 nM. It also inhibited the receptor tyrosine kinases c-RAF and DDR2 (IC₅₀ < 100 nM for both) and FGFR1, KDR, MEKL, and Tie2 (IC₅₀ 100 ~ 600 nM). However, the other twenty one kinases were inhibited only at much higher concentrations of **25**.

Table 5. Kinase selectivity of compound **25**

Kinase	IC ₅₀ (nM)
FMS	2
DDR2	64
c-RAF	80
FGFR1	145

KDR	527
MELK	258
Tie2	208

IC₅₀ values were measured at the respective ATP Km for each kinase.

The inhibition potential of compound **25** on the migration of tumor cells by TAMs was evaluated via an invasion assay using MDA-MB-231 cells (human breast adenocarcinoma cells) and RAW264.7 cells (mouse macrophage cells). Compound **25** inhibited the migration of MDA-MB-231 cells in a dose-dependent manner; the inhibitory effect after treatment with 1 μ M was 67%. (Fig. 2)

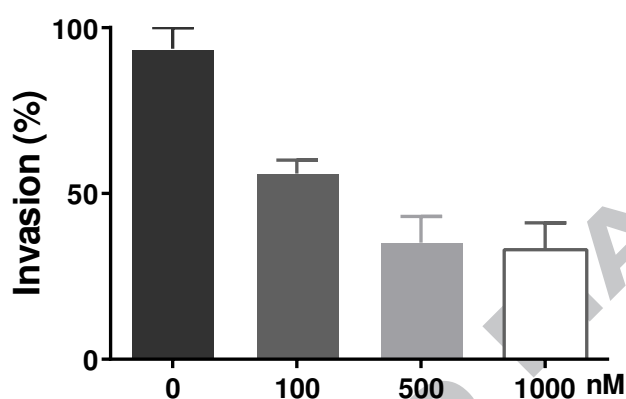
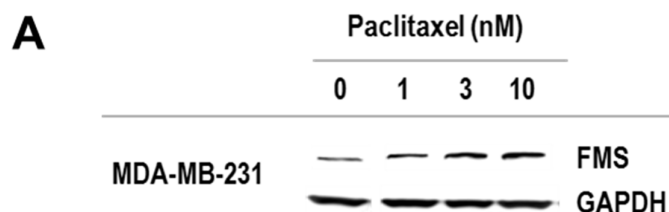


Figure 2. Tumor invasion assay in MDA-MB-231/Luc cells

The anti-tumor effects of compound **25** against breast cancer in combination with cytotoxic agents was evaluated using an *in vitro* tumor cell growth inhibition assay. As shown in Figure 3, paclitaxel dose-dependently induced FMS expression in MDA-MB-231 cells, and compound **25** effectively reduced paclitaxel-induced FMS expression.



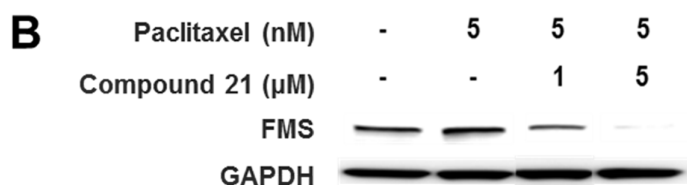


Figure 3. (A) FMS expression levels after treatment with paclitaxel in MDA-MB-231 cells was increased. (B) Compound **25** dose-dependently inhibited overexpressed FMS by paclitaxel treatment in MDA-MB-231 cells. GAPDH, Glyceraldehyde 3-phosphate dehydrogenase

In a combination study with paclitaxel and compound **25** in MDA-MB-231 cells, treatment with compound **25** (5 μM) alone showed a reduction in cell growth by 20%, whereas treatment with paclitaxel in combination with compound **25** showed a synergistic growth inhibition effect (CI < 0.1) compared with treatment with paclitaxel alone. (Fig. 4)

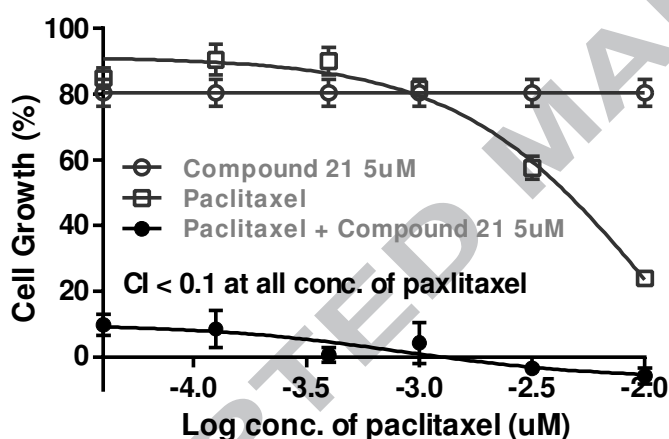


Figure 4. *In vitro* combination study of compound **25** and/or paclitaxel on cell growth inhibition in MDA-MB-231 cells. The synergistic effect was determined by calculating the combination index (CI) using the Calcsyn program (Cambridge.UK). The CI value was considered statistically significant when it was less than 1.0.

In summary, we identified and optimized a novel and potent class of FMS kinase inhibitors based on a thieno[3,2-*d*]pyrimidine core. One of these, compound **25**, was employed in an *in vitro* efficacy study to demonstrate its potential benefit in the treatment of cancer. The inhibition of FMS signaling by compound **25** was efficacious in reducing the migration of cancer cells and showed a synergistic anti-tumor activity against breast cancer in combination with a cytotoxic agent.

Declarations of interest: none.

Funding: not applicable

A. Supplementary data

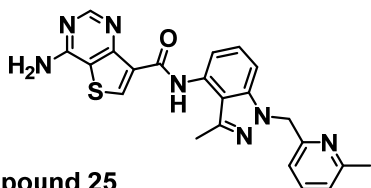
Supplementary data associated with this article can be found in the online version.

References

1. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000Prime Rep.* 2014; 6: 13.
2. Cannarile MA, Weisser M, Jacob W, et al. Colony-stimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy. *J Immunother Cancer.* 2017; 5(1): 53-66.
3. Ao JY, Zhu XD, Chai ZT, et al. Colony-stimulating factor 1 receptor blockade inhibits tumor growth by altering the polarization of tumor-associated macrophages in hepatocellular carcinoma. *Mol Cancer Ther.* 2017; 16(8):1544-1554.
4. Chanmee T, Ontong P, Konno K, Itano N. Tumor-associated macrophages as major players in the tumor microenvironment. *Cancers.* 2014; 6(3): 1670-1690.
5. Szekanecz Z, Koch AE. Macrophages and their products in rheumatoid arthritis. *Curr Opin Rheumatol.* 2007; 19(3): 289-295.
6. Conway JG, Pink H, Bergquist ML, et al. Effects of the cFMS kinase inhibitor 5-(3-methoxy-4-((4-methoxybenzyl)oxy)benzyl)pyrimidine-2,4-diamine (GW2580) in normal and arthritic rats. *J Pharmacol Exp Ther.* 2008; 326(1): 41-50.
7. Sarah O. Antibody against CSF-1 receptor protects bone and cartilage. *Nat Rev Rheumatol.* 2014; 10: 260.
8. Fleisch H, Hofstetter W, Felix R, et al. The role of macrophage stimulating factor M-CSF in bone resorption. *Osteoporos Int.* 1993; 3(1):108-110.
9. Escamilla J, Schokrpur S, Liu C, et al. CSF1 receptor targeting in prostate cancer reverses macrophage-mediated resistance to androgen blockade therapy. *Cancer Res.* 2015; 75(6): 950-962.
10. Woo HH, László CF, Greco S, et al. Regulation of colony stimulating factor-1 expression and ovarian cancer cell behavior in vitro by miR-128 and miR-152. *Mol Cancer.* 2012; 11: 58.
11. Toy EP, Azodi M, Folk NL, et al. Enhanced ovarian cancer tumorigenesis and metastasis by the macrophage colony-stimulating factor. *Neoplasia.* 2009; 11(2): 136-144.
12. Richardsen E, Uglehus RD, Johnsen SH, et al. Macrophage-colony stimulating factor

- (CSF1) predicts breast cancer progression and mortality. *Anticancer Res.* 2015; 35(2): 865-874.
13. Swierczak A, Cook AD, Lenzo JC, et al. The promotion of breast cancer metastasis caused by inhibition of CSF-1R/CSF-1 signaling is blocked by targeting the G-CSF receptor. *Cancer Immunol Res.* 2014; 2(8): 765-776.
 14. Ding J, Guo C, Hu P, et al. CSF1 is involved in breast cancer progression through inducing monocyte differentiation and homing. *Int J Oncol.* 2016; 49(5):2064-2074.
 15. Cannarile MA, Weisser M, Jacob W, et al. Colony-stimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy. *J Immunother Cancer.* 2017; 5(1): 53.
 16. Dorsch M, Hock H, Kunzendorf U, et al. Macrophage colony-stimulating factor gene transfer into tumor cells induces macrophage infiltration but not tumor suppression. *Eur J Immunol.* 1993; 23(1): 186-190.
 17. Richardsen E, Uglehus RD, Johnsen SH, Busund LT. Macrophage-colony stimulating factor (CSF1) predicts breast cancer progression and mortality. *Anticancer Res.* 2015; 35(2):865-874.
 18. Dwyer AR, Greenland EL, Pixley FJ. Promotion of tumor invasion by tumor-associated macrophages: the role of CSF-1-activated phosphatidylinositol 3 kinase and Src family kinase motility signaling. *Cancers.* 2017; 9(6): 68.
 19. Scholl SM, Pallud C, Beuvon F, et al. Anti-colony-stimulating factor-1 antibody staining in primary breast adenocarcinomas correlates with marked inflammatory cell infiltrates and prognosis. *J Natl Cancer Inst.* 1994; 19; 86(2):120-126.
 20. Lin EY, Nguyen AV, Russell RG, Pollard JW. Colony-stimulating factor 1 promotes progression of mammary tumors to malignancy. *J Exp Med.* 2001; 19; 193(6):727-740.
 21. DeNardo DG, Brennan DJ, Rexhepaj E, et al. Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy. *Cancer Discov.* 2011; 1: 54-67.
 22. Nakoinz I, Lee MT, Weaver JF, Ralph P. Differentiation of the IL-3-dependent NFS-60 cell line and adaption to growth in macrophage colony-stimulating factor. *J Immunol.* 1990; 145(3): 860-864.
 23. Zhang C, Ibrahim PN, Zhang J, et al. Design and pharmacology of a highly specific dual FMS and KIT kinase inhibitor. *Proc Natl Acad Sci U S A.* 2013; 110(14):5689-5694.
 24. ProbeChem®. [http://www.probechem.com/target_cFms\(CSF1R\).aspx](http://www.probechem.com/target_cFms(CSF1R).aspx); 2018 Accessed 15 November 2018

Graphic Abstract

**compound 25**FMS (IC₅₀): 2nMNFS-60 (GI₅₀): 79.7nMAnti-tumor activity in tumor cell inhibition assay
and in vitro tumor invasion assay