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## **Discovery of potent T-type calcium channel blocker**

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**Abstract**—The intensive SAR study of 3,4-dihydroquinazoline series led to the most potent compound **10** (**KYS05090**:  $IC_{50} = 41 \pm 1$  nM) against T-type calcium channel and its potency is nearly comparable to that of Kurtoxin. As a small organic molecule, this compound showed the highest blocking activity reported to date.

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Calcium channels are the primary route for translating electrical signals into the biochemical events underlying key processes such as neurotransmitter release, cell excitability, and gene expression.<sup>1</sup> Among calcium channels, T-type or low voltage activated (LVA) calcium channels are thought to contribute to neuronal excitability and also play crucial roles in the control of blood pressure.<sup>2</sup> Therefore, T-type calcium channels are important therapeutic targets for the treatment of epilepsy, neuropathic pain, and cardiovascular diseases such as hypertension and angina pectoris.<sup>3</sup> For example, a T-type calcium channel blocker, mibefradil (Posi $cor^{TM}$ ), was used in treatment of hypertension and stable angina.<sup>4</sup> Shortly following its introduction, mibefradil was withdrawn from the US market in May 1998 because of potential harmful interactions with other drugs.<sup>5</sup> Since the withdrawal of mibefradil, efforts aimed at discovery of new T-type calcium channel blockers have intensified.<sup>6</sup> We have recently reported that 3,4-dihydroquinazoline (KYS05001), as a new scaffold, showed T-type calcium channel blocking effect comparable to mibefradil.<sup>7</sup> In a development of this class, we have also reported that 2-aminopentyl-substituted 3,4-dihydroquinazoline (KYS05044) is more potent than KYS05001 (Fig. 1).<sup>7c</sup> As a result of further investigation of structure-activity relationships in

**KYS05044** series, we found that **KYS05090** is the most potent T-type calcium channel blocker reported to date. Herein, we report the synthesis and biological evaluation of **KYS05090** compound.

Compound KYS05090 was synthesized by the same procedure as described previously by our group as shown in Schemes 1 and 2. Amine nucleophile 4, N,N-dibenzyl-N'-methylpentane-1,5-diamine, was prepared from 1,5diaminopentane 1. Monoprotection of diamine 1 with di-tert-butyl dicarbonate afforded compound 2 and the other amine group was treated with 2 equivalents of benzyl bromide to provide dibenzyl compound 3. The reduction of Boc group with lithium aluminum hydride gave a secondary amine 4 (Scheme 1).<sup>8</sup> Azaphosphorane compound 6 was prepared from 2-nitrocinnamic acid 5 via 3 steps. The coupling of 6 with biphenylyl isocyanate gave the carbodiimide 7, which was treated with amine 4 to provide 3,4-dihydroquinazoline derivative 8 via tandem nucleophilic addition and intramolecular conjugate addition. Hydrolysis of compound 8 with LiOH afforded the free carboxylic acid 9. The coupling reaction of 9 with benzylamine, EDC, and HOBt followed by hydrogenation with 10% Pd/C in the presence of formaldehyde afforded the compound 10 (KYS05090). For comparison of biological activity, ester compound 8 was also treated with 10% Pd(C) under the same condition to provide 11 (KYS05089) as shown in Scheme 2. Both of N,N-dimethylated compounds (10 and 11) are formed via the tandem debenzylation-reductive amination: that is, debenzylated free amine group was further reacted with formaldehyde to convert into imine group,

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Scheme 1. Reagents and conditions: (a) ('Boc)<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, rt, 51%; (b) 2 equiv of benzyl bromide, Na<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, reflux, 49%; (c) LiAlH<sub>4</sub>, THF, 70 °C, 60%.

Β'n

which was hydrogenated to afford *N*,*N*-dimethylated compound.<sup>9</sup>

The biological activity of new synthesized compounds (10 and 11) was evaluated against HEK293 cells which stably express both T-type calcium channel  $Ca_v 3.1$  with  $\alpha_{1G}$  subunit and potassium channel Kir2.1.10 The % inhibition of Ca^{2+} current was measured at 10  $\mu M$  concentration of the synthesized compounds. For this purpose, two assay methods were employed: highthroughput screening (FDSS6000 system)<sup>11</sup> and patchclamp assay using a single cell.<sup>10</sup> High-throughput screening is applied for the preliminary screening. On the other hand, patch-clamp assay is more accurate and sensitive but measures Ca<sup>2+</sup> current one by one. Compounds were primary-screened by high-throughput screening and secondary-screened by patch-clamp assay to obtain IC<sub>50</sub> values. Their in vitro blocking data are summarized in Table 1 and the data of KYS05001 and KYS05044 were also inserted for comparison.

With respect to % inhibition at 10  $\mu$ M concentration, both the new synthetic compounds (10 and 11) exhibited lower % inhibitory activity than mibefradil under highthroughput screening (FDSS6000 system). However, accurate patch–clamp assay showed that both compounds were more or less active than mibefradil but superior to **KYS05044**. Compound 10 exhibited the most potency among them. With respect to IC<sub>50</sub> value, both compounds (10 and 11) were found to be more potent than the other compounds. In accordance with % inhibition data, compound 10 exhibited the most potency (IC<sub>50</sub> = 41  $\pm$  1 nM). This potency is nearly comparable to that of Kurtoxin ( $IC_{50} = 15 \text{ nM}$ ), a peptide compound from the venom of the south African scorpion.<sup>12</sup> To the best of our knowledge, 10 (KYS05090) shows the most powerful activity compared to small organic molecules reported to date. With respect to channel selectivity (T/N-type calcium channel), both compounds (10 and 11) showed less selectivity than parent compound KYS05044 based on % inhibition at 10 µM concentration. To determine the exact channel selectivity of 10 (KYS05090), however, we obtained its IC<sub>50</sub> value (4.9  $\mu$ M) against N-type channel ( $\alpha_{1B}$ ) by patch-clamp assay. As a result, compound 10 (KYS05090) showed higher selectivity for T-type over N-type calcium channel than based on % inhibition [Selectivity index:  $119.5 = 4.9 \,\mu\text{M}$  (N-type)/0.041  $\mu\text{M}$ (T-type)].

In summary, new compounds 10 and 11 (KYS05090 and KYS05089) were synthesized as a result of further structure–activity relationship study in KYS05044 series.<sup>13</sup> Compound 10 (KYS05090) showed the comparable potency to Kurtoxin as well as high selectivity for T-type calcium channel. The detailed result of SAR study in KYS05044 series will be reported in due course.



Scheme 2. Reagents and conditions: (a) (i)  $H_2SO_4$ , MeOH, 70 °C; (ii)  $SnCl_2 \cdot 2H_2O$ , EtOAc, 70 °C; (iii) PPh<sub>3</sub>,  $C_2Cl_6$ , Et<sub>3</sub>N, toluene, reflux, 60% (3 steps); (b) 4-biphenylyl isocyanate, toluene, rt, 80%; (c) compound 4, toluene, rt, >99%; (d) LiOH, THF-H<sub>2</sub>O (1:1), 70 °C, >99%; (e) PhCH<sub>2</sub>NH<sub>2</sub>, HOBt, EDC, THF-CH<sub>2</sub>Cl<sub>2</sub> (1:1), 0 °C to rt, >79%; (f) HCHO, H<sub>2</sub>, 10% Pd/C, CH<sub>3</sub>OH, 4 days, rt, 62% (for 10) and 60% (for 11).

Table 1. In vitro calcium channel blocking effects of 3,4-dihydroquinazoline derivatives

Compounds	HEK293 cell (T-type: $\alpha_{1G}$ )		HEK293 cell (N-type: $\alpha_{1B}$ )	Selectivity (T/N-type) at 10 µM
	% Inhibition $(10 \ \mu M)^a$	$IC_{50} \left(\mu M\right)^{b}$	% Inhibition $(10 \ \mu M)^a$	
KYS05001 KYS05044 KYS05089 (11) KYS05090 (10)	90.1 $\pm$ 2.3 82.5 $\pm$ 0.7 90.1 $\pm$ 2.3 (57.9) <sup>d</sup> 08.0 $\pm$ 1.6 (42.0) <sup>d</sup>	$1.16 \pm 0.04 \\ 0.56 \pm 0.10 \\ 0.23 \pm 0.03 \\ 0.041 \pm 0.001$	28.1 ± 1.7 No blocking <sup>c</sup> 24.4 ± 2.8 70.6 ± 2.1	3.2 >100 3.7
Mibefradil	$98.0 \pm 1.6 (42.9)^{\circ}$ $95.9 \pm 1.7 (77.6)^{\circ}$	$0.041 \pm 0.001$ $1.34 \pm 0.49$	$70.6 \pm 3.1$ $67.6 \pm 1.2$	1.4 (119.5) 1.4

<sup>a</sup> % Inhibition value ( $\pm$ SE) was obtained by repeated procedures ( $n \ge 4$ ) under patch–clamp assay.

<sup>b</sup> IC<sub>50</sub> value was determined from the dose-response curve.

<sup>c</sup> 'No blocking' means the inhibition was less than 1%.

<sup>d</sup> Data by HTS (FDSS6000 system).

<sup>e</sup> Selectivity index based on IC<sub>50</sub> values.

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## Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl. 2007.08.070.

## **References and notes**

- (a) Barclay, J. W.; Morgan, A.; Burgoyne, R. D. Cell Calcium 2005, 38, 343; (b) Zheng, X.; Bobich, J. A. Brain Res. Bull. 1998, 47, 117; (c) Himpens, B.; Missiaen, L.; Casteels, R. J. Vasc. Res. 1995, 32, 207; (d) Levi, A. J.; Brooksby, P.; Hancox, J. C. Cardiovasc. Res. 1993, 27, 1743.
- (a) Opie, L. H.; Frishman, W. H.; Thadani, U. Calcium channel antagonists (calcium entry blockers). In *Drugs for the Heart*; Opie, L. H., Ed., 4th ed.; W. B Saunders: Philadelphia, 1994; p 50; (b) Abernethy, D. R.; Schwartz, J. B. N. *Engl. J. Med.* **1999**, *341*, 1447; (c) Thadani, U. *Curr. Opin. Cardiol.* **1999**, *14*, 349.

- (a) Flatters, S. J. L. Drugs Future 2005, 30, 573; (b) Nelson, M. T.; Todorovic, S. M. Curr. Pharm. Des. 2006, 12, 2189; (c) Perez-Reyes, E. Physiol. Rev. 2003, 83, 117; (d) Vassort, G.; Alvarez, J. J. Cardiovasc. Electrophysiol. 1994, 5, 376.
- (a) Clozel, J.; Ertel, E.; Ertel, S. J. Hypertens. Suppl. 1997, 15, S17; (b) Hermsmeyer, K.; Mishra, S.; Miyagawa, K.; Minshall, R. Clin. Ther. 1997, 19, 18; (c) Van der Vring, J.; Cleophas, T.; Van der Wall, E.; Niemeyer, M. Am. J. Ther. 1999, 6, 229.
- 5. Asirvatham, S.; Sebastian, C.; Thadani, U. Drug Saf. 1998, 19, 23.
- 6. (a) Park, J. H.; Choi, J. K.; Lee, E.; Lee, J. K.; Rhim, H.; Seo, S. H.; Kim, Y.; Doddareddy, M. R.; Pae, A. N.; Kang, J.; Roh, E. J. Bioorg. Med. Chem. 2007, 15, 1409; (b) Doddareddy, M. R.; Choo, H.; Cho, Y. S.; Rhim, H.; Koh, H. Y.; Lee, J.-H.; Jeong, S.-W.; Pae, A. N. Bioorg. Med. Chem. 2007, 15, 1091; (c) Jo, M. N.; Seo, H. J.; Kim, Y.; Seo, S. H.; Rhim, H.; Cho, Y. S.; Cha, J. H.; Koh, H. Y.; Choo, H.; Pae, A. N. Bioorg. Med. Chem. 2007, 15, 365; (d) Ku, I. W.; Cho, S.; Doddareddy, M. R.; Jang, M. S.; Keum, G.; Lee, J.-H.; Chung, B. Y.; Kim, Y.; Rhim, H.; Kang, S. B. Bioorg. Med. Chem. Lett. 2006, 16, 5244; (e) Furukawa, T.; Yamada, O.; Matsumoto, H.; Yamashita, T. WO 2005051402, 2005; (f) McCalmont, W. F.; Heady, T. N.; Patterson, J. R.; Lindenmuth, M. A.; Haverstick, D. M.; Gray, L. S.; MacDonald, T. L. Bioorg. Med. Chem. Lett. 2004, 14, 3691; (g) Jung, H. K.; Doddareddy, M. R.; Cha, J. H.; Rhim, H.; Cho, Y. S.; Koh, H. Y.; Jung, B. Y.; Pae, A. N. Bioorg. Med. Chem. 2004, 12, 3965; (h) Kumar, P. P.; Stotz, S. C.; Paramashivappa, R.; Beedle, A. M.; Zamponi, G. W.; Rao, A. S. Mol. Pharmacol. 2002, 61, 649.
- (a) Choi, J. Y.; Seo, H. N.; Lee, M. J.; Park, S. J.; Park, S. J.; Jeon, J. Y.; Kang, J. H.; Pae, A. N.; Rhim, H.; Lee, J. Y. Bioorg. Med. Chem. Lett. 2007, 17, 471; (b) Park, S. J.; Park, S. J.; Lee, M. J.; Rhim, H.; Kim, Y.; Lee, J.-H.; Chung, B. Y.; Lee, J. Y. Bioorg. Med. Chem. 2006, 14, 3502; (c) Rhim, H.; Lee, Y. S.; Park, S. J.; Chung, B. Y.; Lee, J. Y. Bioorg. Med. Chem. Lett. 2005, 15, 283; (d) Lee, Y. S.; Lee, B. H.; Park, S. J.; Kang, S. B.; Rhim, H.; Park, J.-Y.; Lee, J.-H.; Jeong, S.-W.; Lee, J. Y. Bioorg. Med. Chem. Lett. 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005, 16, 2005,
- Phuan, P.-W.; Kozlowski, M. C. J. Org. Chem. 2002, 67, 6339.
- (a) Fattori, D.; Rossi, C.; Fincham, C. I.; Caciagli, V.; Catrambone, F.; D'Andrea, P.; Felicetti, P.; Gensini, M.; Marastoni, E.; Nannicini, R.; Paris, M.; Terracciano, R.; Bressan, A.; Giuliani, S.; Maggi, C. A.; Meini, S.; Valenti, C.; Quartara, L. J. Med. Chem. 2007, 50, 550; (b) Ramamohan, R. D.; Sankara, R. C.; Pamujula, S. WO 02090339, 2002.

- 10. (a) Kim, T.; Choi, J.; Kim, S.; Kwon, O.; Nah, S. Y.; Han, Y. S.; Rhim, H. Biochem. Biophys. Res. Commun. 2004, 324, 401; (b) For the recordings of  $\alpha_{1G}$  T-type Ca<sup>2</sup> currents, the standard whole-cell patch-clamp method was utilized. Briefly, borosilicate glass electrodes with a resistance of 3–4 M $\Omega$  were pulled and filled with the internal solution containing (in mM): 130 KCl, 11 EGTA, 5 Mg-ATP, and 10 Hepes (pH 7.4). The external solution contained (in mM): 140 NaCl, 2 CaCl<sub>2</sub>, 10 Hepes, and 10 glucose (pH 7.4).  $\alpha_{IG}$  T-type Ca<sup>2+</sup> currents were evoked every 15 s by a 50 ms depolarizing voltage step from -100 mV to -30 mV. The molar concentrations of test compounds required to produce 50% inhibition of peak currents (IC<sub>50</sub>) were determined from fitting raw data into dose-response curves. The current recordings were obtained using an EPC-9 amplifier and Pulse/Pulsefit software program (HEKA, Germany).
- Kim, Y.; Seo, S.; Kim, D.; Rhim, H. The 11th Annual Conference and Exhibition, Geneva, Switzerland, September 11–15, 2005; Society for Biomolecular Screening: Danbury, CT, 2005; P07016.
- 12. Chuang, R. S.; Jaffe, H.; Cribbs, L.; Perez-Reyes, E.; Swartz, K. J. Nat. Neurosci. 1998, 1, 668.
- 13. Spectral data of the compounds 10 and 11: for compound 10 (KYS05090), <sup>1</sup>H NMR (400 MHz,  $CDCl_3\delta$  7.72–6.98 (18H, m, Ph), 5.32 (1H, dd, J = 9.1and 4.6 Hz, COCH<sub>2</sub>CH), 4.50 (2 H, d, J = 5.8 Hz, PhCH<sub>2</sub>-), 3.50-3.20 (2 H, m, CH<sub>3</sub>N-CH<sub>2</sub>), 2.71-2.33 (4 H,  $CH_3$ -N and COCH), 2.44 (1H, dd, J = 14.5 and 5.2 Hz, COCH), 2.30-2.29 (2 H, m, -NCH<sub>2</sub>), 2.21 (6 H, s, 2×N-CH<sub>3</sub>), 1.65-1.35 (4 H, m, 2×-CH<sub>2</sub>), 1.25-1.10  $(2 \text{ H}, \text{ m}, CH_2)$ ; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) $\delta$  170.1, 153.9, 145.3, 143.8, 140.4, 138.4, 136.7, 128.8, 128.6, 35.3, 27.1, 27.0, 24.7; MS (FAB+), m/z (relative intensity, %) 596.7([M+Na]<sup>+</sup>, 100), 574.7 ([M+H]<sup>+</sup>, 30); MS (FAB-), *m/z* (relative intensity, %) 572.7([M-H]<sup>+</sup>, 100); HRMS (FAB+) calcd for  $C_{37}H_{44}N_5O$ :  $[M+H]^+ =$ found = 574.3516: 574.3546. for compound 11 (KYS05089), <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.52–6.88 (13 H, m, Ph), 5.11 (1H, dd, J = 10.4 and 4.5 Hz, COCH<sub>2</sub>CH), 3.75 (3H, s, OCH<sub>3</sub>), 3.50 (1H, m, CH<sub>3</sub>N-CH), 3.15 (1H, m, CH<sub>3</sub>N-CH), 2.88-2.82 (4H, m, CH<sub>3</sub>-N and COCH), 2.51 (1H, dd, J = 15.5 and 4.6 Hz, COCH), 2.25-2.23 (2H, m, -NCH<sub>2</sub>), 2.20 (6H, s, 2×N- $CH_3$ ), 1.52–1.45 (4H, m, 2×– $CH_2$ ), 1.24–1.23 (2H, m, CH<sub>2</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)δ 171.9, 153.4, 145.4, 144.2, 140.3, 136.8, 128.8, 128.4, 127.9, 127.1, 126.8, 125.4, 124.8, 122.9, 122.6, 122.1, 61.2, 59.7, 49.5, 45.2, 43.8, 41.8, 35.3, 27.1, 27.0, 24.7; HRMS (FAB+) calcd for  $C_{31}H_{39}N_4O_2$ :  $[M+H]^+ = 499.3073$ , found = 499.3060.