

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Synthesis and BK channel-opening activity of 2-amino-1,3-thiazole derivatives

Xiao-Lei Qi^{a,1}, Heeji Jo^{b,1}, Xue-Ying Wang^a, Tong-Tong Ji^a, Hai-Xia Lin^a, Chul-Seung Park^{b,*}, Yong-Mei Cui^{a,*}

^a Department of Chemistry, Innovative Drug Research Center, College of Sciences, Shanghai University, Shanghai 200444, China

^b School of Life Sciences and National Leading Research Laboratory, Gwangju Institute of Science and Technology (GIST), Gwangju 61005, Republic of Korea

ARTICLE INFO	A B S T R A C T
Keywords: BK channels Openers Synthesis Thiazole	A series of 2-amino-5-arylmethyl- or 5-heteroarylmethyl-1,3-thiazole derivatives were synthesized and evaluated for BK channel-opening activities in cell-based fluorescence assay and electrophysiological recording. The assay results indicated that the activities of the investigated compounds were influenced by the physicochemical properties of the substituent at benzene ring.
SAR	

Large-conductance calcium-activated K⁺ channels (also called maxi-K or BK channels) are widely distributed in a number of organ systems, such as smooth muscle cells,¹ skeletal muscle cells,^{2,3} neuronal cells,⁴ and secretory epithelial cells,⁵ and participate in numerous physiological functions by coupling transmembrane K⁺ flux, changes in membrane potential, and intracellular Ca²⁺ concentration.^{6–8} The BK channel plays important physiological roles in modulating muscle contraction or neurotransmitter release and hormone secretion.^{9–12} The physiological role and widespread distribution of BK channels suggest that agents that open these channels could have profound impacts on diseases such as ischemic stroke, epilepsy, asthma, and bladder overactivity.^{13–17} During the past few years, various classes of BK channel openers such as the synthetic benzimidazolin-2-one derivative NS1619,¹⁸ the bisarylurea NS1608,¹⁹ the bisarylthiourea NS11021,^{20,21} the oxindole BMS-204352²², the benzofuroindole derivative CTBIC (4chloro-7-(trifluoromethyl)-10H-benzofuro[3,2-b]indole-1-carboxylic acid)²³ and the natural modulator dihydrosoyasaponin-1,²⁴ etc (Fig. 1), as well as their pharmacology have been described.²⁵ Wellcharacterized BK channel openers not only are expected to have therapeutic potential, but also should be of assistance in understanding the function, structure and role of BK channels.

NS19504, reported by Bernhard Nausch, ²⁶ represents a novel chemotype among BK activators. The structure of NS19504 is markedly different from that of the well-known BK activators, because of its low mol. wt. and absence of an acidic function. Therefore, NS19504 represents an interesting lead in the search for new BK channel modulators and provides a template from which more potent derivatives might be obtained by suitable substitution. In this letter, we survey the BK channel-opening properties of a series of 2-amino-1,3-thiazole derivatives of general structure **4** (Fig. 1) with the aim of studying the space around the terminal group and exploring the requirements for BK channel-opening activity.

Firstly, a series of 5-arylmethyl-2-amino-1,3-thiazole compounds **4au** were synthesized. As shown in Scheme 1, Meerwein reaction of the arenediazonium chlorides resulted from anilines, with acrolein gave 3aryl-2-chloropropanals, followed by cyclocondensation with thiourea, resulted in 2-amino-5-aryl-1,3-thiazoles **4a-s**. However, starting from pyridin-3-amine, using similar conditions failed to give the target compound **4t**. Therefore, α -bromine aldehyde was first prepared by Dess-Martin oxidation of the corresponding alcohol and subsequent bromination using Br₂ and HBr. Treatment of the α -bromine aldehydes **5c** and **6c** with thiourea in EtOH under heating at reflux afforded the corresponding target compounds **4t** and **4u**.

To investigate the importance of the methylene linker, 5-aroyl-2amino-1,3-thiazole compounds **9a-b** were synthesized. Thiourea was firstly activated as bis-thiazadiene **7** with excess of commercially available N,N-dimethylformamide dimethylacetal in methanol. The thiazole ring was formed after addition of the corresponding

* Corresponding authors.

https://doi.org/10.1016/j.bmcl.2021.128083

Received 12 March 2021; Received in revised form 23 April 2021; Accepted 1 May 2021 Available online 6 May 2021 0960-894X/© 2021 Published by Elsevier Ltd.

E-mail addresses: cspark@gist.ac.kr (C.-S. Park), ymcui@shu.edu.cn (Y.-M. Cui).

¹ These authors contributed equally to this work.



Figure 1. The structures of representative BK channel openers and 4.

 α -bromoketones **7a** and **7b** in THF. Intermediates **8a-b**, which were not isolated, were deprotonated in situ by addition of triethylamine. The imine was subsequently removed in situ with HCl to form the expected products **9a** and **9b**. Meanwhile, **10a-b** was synthesized by direct acylation of NS19504 with acetyl chloride or 2-chloroacetyl chloride under basic conditions (Scheme 2). All the compounds prepared in this study are compiled in Table 1.

First of all, the BK channel-modulating activities of all the target compounds in this study were evaluated using the FluxOR potassium channel assay (Invitrogen, U.S.A.) with AD-293 cells expressing hyperactive mutant BK channels (G803D/N806K).²⁷ Fluorescence signals were measured and normalized against the basal level of each trace to give normalized values in relative fluorescence units (RFU) and the relative fluorescence value in the presence of a test compound (Δ RFU, 5 μ M) was expressed as versus of the drug-free control. The values represent an average of data obtained from at least three separate

measurements. In the assays, DMSO was used as a negative control and CTBIC at a fixed concentration as a positive control. Also, the results for NS19504 are included for comparison.

From the results presented in Table 1 and Fig. 2, most of the 21 compounds **4a-u** showed moderate BK channel-opening activity. At a final concentration of 5 μ M, two compounds **4a** and **4 h** were found to show higher BK channel-opening activity than NS19504. Among the aryl-group containing derivatives **4a-s**, the activity is quite sensitive to the location and properties of the aromatic substituents. Substitution with an electron-donating group, such as a methoxy group (**4o-q**) resulted in inactivity, and the methyl derivatives **41-n** were only marginally active. Substitution of an electron-withdrawing bromo or trifluoromethyl group on the aromatic ring resulted in increases in the channel activity. Among the three regioisomers **4f-h** of the trifluoromethyl substituent, the *para* isomer **4h** (Δ RFU = 2.10 ± 0.16 of control at 5 μ M) was more potent than the *ortho* and *meta* CF₃-



Scheme 1. Regents and conditions: (a) NaNO₂, HCl, 0 °C, H₂O; (b) NaHCO₃, MgO, CuCl₂·2H₂O, acrolein, r.t, acetone, 21–43%; (c) thiourea, r.t, EtOH, 21–51%; (d) DMP, r.t, CH₂Cl₂; (e) HBr, Br₂, r.t, Ac₂O.



Scheme 2. Reagents and conditions: (a) (Me)₂NCH(OMe)₂, MeOH, reflux; (b) (Et)₃N, MeOH, reflux, 53–64%; (c)10% HCl, reflux, 4 hrs, 23–31%; (d) (Et)₃N, DMAP, 0 °C, 88–91%.

Table 1 (continued)

substituted isomers (4f, 1.07 \pm 0.04; 4g, 1.22 \pm 0.05). However, among the three regioisomers of the bromo substituent, the ortho isomer 4a was more potent than the para and meta Br-substituted isomers (NS19504, 4b). In particular, the ortho-bromo isomer 4a exhibited the highest potency for channel activity with ΔRFU 2.67 \pm 0.22 of control at 5 μM among the synthesized compounds. Compounds 4d and 4e bearing a Cl atom substituent in the 3- or 4-position, with similar activities, were found to be more potent than the 2-Cl isomer 4c. All the three

Table 1

Structure and BK-opening properties of 2-amino-1,3-thiazole derivatives based

Ar N NHa					
4a-u					
Compound	Ar	ΔRFU (5 $\mu M,n=3)$			
DMSO	-	1.00 ± 0.01			
CTBIC	-	3.30 ± 0.13			
NS19504	The second se	1.85 ± 0.17			
4a	Br er	2.67 ± 0.22			
4b	, as Br	1.37 ± 0.14			
4c	CI	1.13 ± 0.17			
4d	2 ² CI	1.51 ± 0.18			
4e	And a second sec	1.59 ± 0.05			
4f	CI CF3	1.07 ± 0.04			
4g	CF3	1.22 ± 0.05			
4h	×	$\textbf{2.10} \pm \textbf{0.16}$			
4i	CN CN	1.06 ± 0.05			
4j	\sim	1.00 ± 0.01			

Compound	Ar	ΔRFU (5 $\mu M,n=3$)
	² ^ℓ CN	
		1.01 0.00
4K		1.21 ± 0.02
41	<pre></pre>	1.37 ± 0.27
	*	
4		1.07 0.15
4m	P CIN	1.27 ± 0.15
4n	× Š	1.11 ± 0.01
	CH3	
40	OCH3	0.94 ± 0.03
4p	JCH3	1.02 ± 0.03
		1.00 + 0.00
4q		1.02 ± 0.02
4r	ž	0.94 ± 0.04
	NO	
4s	JCH3	0.97 ± 0.03
	\forall	
<i>A</i> +	ĊF ₃	1.08 ± 0.13
41	T)	1.00 ± 0.15
4u	N N	1.11 ± 0.03
0	, N	0.04 + 0.04
9a 9h	-	0.94 ± 0.04 1 08 + 0 16
10a	_	1.14 ± 0.15
10b	_	0.94 ± 0.02



Figure 2. Effects of the synthesized 2-amino-1,3-thiazole derivatives on BK_{Ca} channel activity. (For statistical analyses, Student T-test was used,*P < 0.05, **P < 0.01, ***P < 0.005).

regioisomers **4i-k** with CN substituent and the *para*-nitro isomer **4r** were inactive, except for compounds **4k**, which was only marginally active. Moreover, the bis-substituted derivative **4s** with *meta*-CF₃ and *meta*-OMe, as well as the pyridine derivatives **4t-u** exhibited no BK-channel opening activity.

As mentioned above, we also synthesized the ketone derivatives to confirm the importance of the methylene linker for BK channel-opening activity. Both the ketone derivatives (**9a** and **9b**, respectively) of NS19504 showed no activity. Meanwhile, it's noteworthy that acylation of the amino group of NS19504 resulted in decrease in activity, and compounds **10a-b** showed no channel-opening activity, indicating the importance of the methylene linker and the amino functionality in thiazole ring for the activity.

The potentiating effects of 4a and 4h were further investigated at

different concentrations $(1-30 \ \mu\text{M})$. Both compounds progressively increased the fluorescence signal in a dose-dependent manner. The fluorescence signal evoked by each compound was completely blocked by co-treatment with 1 μ M paxilline, a known BK_{Ca} channel inhibitor, confirming that the Tl⁺ fluorescence induced by the compounds was due to the activation of BK_{Ca} channels (Fig. 3).

We further validated the effects of 4a and 4h electrophysiologically. Electrophysiological recording was performed using the α -subunit of the rat BK_{Ca} channel (Slo1) expressed in Xenopus laevis oocytes. The Gigaohm-seal patch-clamp method was used for current recordings in an outside-out configuration as previously described.²⁸ Macroscopic BK_{Ca} channel currents were evoked by voltage pulses from -80 to 200 mV in the absence and presence of 4a or 4h. Both outward and inward tail currents were increased by both compounds at a concentration of $10 \,\mu M$ (Fig. 4A). The effects of each compound were further quantified by plotting the G-V relationship. Both compounds shifted the G-V curve toward more negative voltage (Fig. 4B). In the presence of 10 μ M compound, the shift in $V_{1/2}$ was 23.4 \pm 3.40 mV for 4a, and 24.5 \pm 1.53 mV for 4h (Fig. 4C). These two compounds also increased the maximum conductance (G_{max}) by 1.4-fold for 4a, and 1.3-fold for 4h compared to vehicle trace (Fig. 4D). Thus, the results indicate that 4a and 4h can potentiate BK_{Ca} channel activation with almost the same degrees of potency.

In summary, a series of new 2-amino-1,3-thiazole derivatives were synthesized and characterized in approach of BK channel openers. The assay results indicated that the activities of the investigated compounds were influenced by the physicochemical properties of the substituent at benzene ring. Further modifications of these lead structures with the aim of improving the potency as well as the specificity *in vitro* and the efficacy *in vivo* are in progress.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.



Figure 3. Effects of 4a and 4 h on BK_{Ca} channel activity based on cell-based fluorescence (For statistical analyses, Student T-test was used:*P < 0.05, ***P < 0.005).



Figure 4. Effects of 4a and 4 h on BK_{Ca} channel macroscopic currents. For statistical analyses, Student T-test was used, *p < 0.05, ***p < 0.001.

Acknowledgements

The authors are grateful for support from Science and Technology Commission of Shanghai Municipality (No. 19ZR1419700) and a grant supported by the Korea Institute of Planning and Evaluation for Technology in Food, Agriculture and Forestry (IPET) through the Agri-Bio Industry Technology Development Program, funded by the Ministry of Agriculture, Food and Rural Affairs (MAFRA), Korea (317070-4) to CSP.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.bmcl.2021.128083.

References

- 1 Wang Y, Zhang HT, Su XL, et al. Experimental diabetes mellitus down-regulates large-conductance Ca²⁺-activated K⁺ channels in cerebral artery smooth muscle and alters functional conductance. *Curr Neurovasc Res.* 2010;7(2):75–84.
- 2 Foppen RJG, Van Heukelom JS. Isoprenaline-stimulated differential adrenergic response of K⁺ channels in skeletal muscle under hypokalaemic conditions. *Pflugers Arch.* 2003;446(2):239–247.
- 3 Maqoud F, Cetrone M, Mele A, Tricarico D. Molecular structure and function of big calcium-activated potassium channels in skeletal muscle: pharmacological perspectives. *Physiol Genomics*. 2017;49(6):306–317.

- 4 Kimm T, Khaliq ZM, Bean BP. Differential regulation of action potential shape and burst-frequency firing by BK and Kv2 channels in substantia nigra dopaminergic neurons. J Neurosci. 2015;35(50):16404–16417.
- 5 Yang C, Gonzalez-Perez V, Mukaibo T, Melvin JE, Xia X-M, Lingle CJ. Knockout of the LRRC26 subunit reveals a primary role of LRRC26-containing BK channels in secretory epithelial cells. *Proc Natl Acad Sci.* 2017;114(18):E3739–E3747.
- 6 Zhou Y, Zeng X-H, Lingle CJ. Barium ions selectively activate BK channels via the Ca² ⁺-bowl site. Proc Natl Acad Sci. 2012;109(28):11413–11418.
- 7 Meera P, Wallner M, Jiang Z, et al. A calcium switch for the functional coupling between α (hslo) and β subunits (KV, Ca β) of maxi K channels. *FEBS Lett.* 1996;382 (1–2):84–88.
- 8 Xu JW, Slaughter MM. Large-conductance calcium-activated potassium channels facilitate transmitter release in salamander rod synapse. J Neurosci. 2005;25(33): 7660–7668.
- 9 Griguoli M, Sgritta M, Cherubini E. Presynaptic BK channels control transmitter release: physiological relevance and potential therapeutic implications. J Physiol. 2016;594(13):3489–3500.
- 10 Sun X, Zhou D, Zhang P, et al. Beta-subunit-dependent modulation of hSlo BK current by arachidonic acid. *J Neurophysiol.* 2007;97:62–69.
- 11 Garcia ML, Shen D-M, Kaczorowski GJ. High-conductance calcium-activated potassium channels: validated targets for smooth muscle relaxants? *Expert Opin Ther Pat.* 2007;17(7):831–842.
- 12 JrJE S, Dworetzky SI, Gribko VK. Modulators of large-conductance calcium-activated potassium (BK) channels as potential therapeutic targets. *Curr Pharm Des.* 1996;2(4): 413–428.
- 13 Cox DH. The BK_{Ca} channel's Ca²⁺-binding sites, multiple sites, multiple ions. J Gen Physiol. 2005;125(3):253–255.
- 14 Hristov KL, Parajuli SP, Soder RP, Cheng Q, Rovner ES, Petkov GV. Suppression of human detrusor smooth muscle excitability and contractility via pharmacological activation of large conductance Ca²⁺-activated K⁺ channels. *Am J Physiol Cell Physiol.* 2012;302(11):C1632–C1641.

X.-L. Qi et al.

Bioorganic & Medicinal Chemistry Letters 43 (2021) 128083

- 15 Meredith AL, Thorneloe KS, Werner ME, Nelson MT, Aldrich RW. Overactive bladder and incontinence in the absence of the BK large conductance Ca2⁺-activated K⁺ channel. J Biol Chem. 2004;279(35):36746–36752.
- 16 Sausbier M, Hu H, Arntz C, et al. Cerebellar ataxia and Purkinje cell dysfunction caused by Ca²⁺-activated K⁺ channel deficiency. *Proc Natl Acad Sci.* 2004;101(25): 9474–9478.
- 17 Olesen S-P, Munch E, Moldt P, Drejer J. Selective activation of Ca²⁺-dependent K⁺ channels by novel benzimidazolone. *Eur J Pharmacol.* 1994;251(1):53–59.
- 18 Han X, Xi L, Wang H, et al. The potassium ion channel opener NS1619 inhibits proliferation and induces apoptosis in A2780 ovarian cancer cells. *Biochem Biophys Res Commun.* 2008;375(2):205–209.
- 19 Siemer C, Bushfield M, Newgreen D, et al. Effects of NS1608 on MaxiK channels in smooth muscle cells from urinary bladder. J Membr Biol. 2000;173(1):57–66.
- 20 Borchert GH, Hlaváčková M, Kolář F. Pharmacological activation of mitochondrial BK_{Ca} channels protects isolated cardiomyocytes against simulated reperfusioninduced injury. *Exper Biol Med.* 2013;238(2):233–241.
- 21 Layne JJ, Nausch B, Olesen S-P, Nelson MT. BK channel activation by NS11021 decreases excitability and contractility of urinary bladder smooth muscle. Am J Physiol Regul Integr Comp Physiol. 2010;298(2):R378–R384.

- 22 Hewawasam P, Gribkoff VK, Pendri Y, et al. The synthesis and characterization of BMS-204352 (MaxiPostTM) and related 3-fluorooxindoles as openers of maxi-K potassium channels. *Bioorg Med Chem Lett.* 2002;12(7):1023–1026.
- **23** Gormemis AE, Ha TS, Im I, et al. Benzofuroindole analogues as potent BK(Ca) channel openers. *ChemBioChem.* 2005;6(10):1745–1748.
- 24 McManus OB, Helms LM, Pallanck L, et al. Functional role of the beta subunit of high conductance calcium-activated potassium channels. *Neuron*. 1995;14:645–650.
- 25 Bukiya AN, Patil SA, Li W, Miller DD, Dopico AM. Calcium- and Voltage- Gated Potassium (BK) Channel Activators in the 5β-Cholanic Acid-3α-ol Analogue Series with Modifications in the Lateral Chain. *ChemMedChem*. 2012;7(10):1784–1792.
- 26 Nausch B, Rode F, Jørgensen S, et al. NS19504: a novel BK channel activator with relaxing effect on bladder smooth muscle spontaneous phasic contractions. *J Pharmacol Exp Ther.* 2014;350(3):520–530.
- 27 Lee B-C, Kim H-J, Park SH, Phuong TTT, Kang TM, Park C-S. Development of cellbased assay system that utilizes a hyperactive channel mutant for high-throughput screening of BKCa channel modulators. J Biotechnol. 2013;167(1):41–46.
- 28 Lee N, Lim BH, Lee K-S, et al. Identification and Characterization of a Novel Large-Conductance Calcium-Activated Potassium Channel Activator, CTIBD, and Its Relaxation Effect on Urinary Bladder Smooth Muscle. *Mol Pharmacol.* 2021;99(2): 114–124.