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Selective Inhibition of the Immunoproteasome by Ligand-Induced Crosslinking of the Active Site **

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Abstract: The concept of proteasome inhibition ranks among the latest achievements in the treatment of blood cancer and represents a promising strategy for modulating autoimmune diseases. In this study, we describe peptidic sulfonyl fluoride inhibitors that selectively block the catalytic β 5 subunit of the immunoproteasome by inducing only marginal cytotoxic effects. Structural and mass spectrometric analyses revealed a novel reaction mechanism involving polarity inversion and irreversible crosslinking of the proteasomal active site. We thus identified the sulfonyl fluoride headgroup for the development and optimization of immunoproteasome selective compounds and their possible application in autoimmune disorders.

The 20S proteasome (core particle; CP) is a sensitive target for the clinically applied inhibitors bortezomib (Velcade) and carfilzomib (CFZ, Kyprolis; Figure 1). Its blockage in malignant cells emerged as an effective approach for the treatment of blood cancers such as multiple myeloma and mantle cell lymphoma.^[1,2] Moreover, the ongoing evaluation of the antiinflammatory immunoproteasome (iCP) inhibitor ONX 0914 (former PR-957; Figure 1) in preclinical studies has givenrise to a novel therapeutic strategy for modulating autoimmune disorders including rheumatoid arthritis and multiple sclerosis.^[3,4] A special attribute of ONX 0914 is the reduction of disease-associated immune responses by selectively blocking the $\beta 5$ subunit of the iCP ($\beta 5i$ or LMP7).^[3] However, the therapeutic window of iCP inhibitors like ONX 0914 entirely depends on their selectivity for \$5i over \$5c, in order to prevent cytotoxic effects that arise from undesired co-

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Figure 1. α',β' -Epoxyketones carfilzomib (CFZ) and ONX 0914 as well as their peptido sulfonyl fluoride (PSF) counterparts 1, 2, and compound 3.

inhibition of the constitutive proteasome (cCP).^[5,6] Thus, the decisive element of iCP inhibitor design is the capability to discriminate between the similar chymotrypsin-like (ChTL) activities of \$\beta5\$ i and \$\beta5\$c.^[7] CFZ as well as ONX 0914 feature an electrophilic α',β' -epoxyketone warhead that forms a covalent and irreversible adduct with both nucleophiles, Thr 10^{γ} and Thr1N, of the catalytically active threonine (Thr1) at the β 5 subunit (see Scheme S1 in the Supporting Information).^[8-10] Since CFZ and ONX 0914 have identical warheads, ONX 0914's favorable binding to 65i solely originates from its backbone architecture, which fulfills the individual binding requirements of β 5i as opposed to β 5c.^[11] Recently, extensive research on the optimization of peptide backbones by incorporation of unnatural amino acids resulted in ß5i- and ß5c-specific epoxyketones.^[5,12,13] However, studies on various functional reactive groups and peptido sulfonyl fluoride (PSF) proteasome inhibitors suggest that also warheads have a direct influence on the selectivity for individual active β -subunits.^[14-16] Notably, PSF compounds block the CP activity in the low nanomolar range,^[16] albeit they are the only peptidic CP inhibitors known so far whose electrophilic

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headgroup is shifted by a methylene unit, which demands an exceptional binding mode. Therefore, we set out to elucidate the PSF's mode of action, commencing with the synthesis of the CFZ and ONX 0914 PSF counterparts **1** and **2** (Figure 1). Next, we determined crystal structures of yeast CP (yCP):**1** with resolutions up to 2.1 Å by performing crystal soaking experiments with incubation times of 1–6 h prior to data collection (see Table ST1).

Based on the identical headgroups of the PSF and the common serine protease inhibitor phenylmethanesulfonyl fluoride (PMSF), we anticipated a similar inactivation mechanism through the formation of a covalent adduct on Thr1 upon attack of Thr1O^{γ} on the sulfur atom. Unexpectedly, after incubation of the crystal for 2 h, the yCP:**1** structure revealed empty substrate binding channels (2.1 Å resolution, $R_{\rm free} = 19.7$ %, PDB ID 4R17). Instead, the F_o-F_c map displayed negative electron density at the Thr1 side chain of subunit β 5, disclosing a chemical modification of the catalytic center (see Figure S1a), whereas β 1 and β 2 remained identical to the apo state. To identify a short-lived reaction intermediate of **1** at β 5 we conducted time-resolved intact protein mass spectrometry using LC-ESI-LTQ-FT-MS analysis on various CP types incubated with **1** and **2** (Figure 2).



Figure 2. Deconvoluted intact-protein mass spectra of the β 5c and β 5i subunits following treatment of cCP with 1 (left panel) and iCP with 2 (right panel) (25 μ M) after incubation for 12 h. The species labeled " $-H_2O$ " represent either the aziridine or the crosslinked state (expected mass difference: 18.0 Da). The insets feature enlargements of the major species. The species labeled "+2–HF" represents the covalently modified intermediate of the β 5 subunits prior to dehydroxylation (expected mass difference: 572.7 Da). See Figure S5 for spectra of β 5 of untreated cCP and iCP.

Spectra with incubation times up to 2 h confirmed the formation of a covalent adduct on the β 5 subunit of all applied CP types with an observed mass increase corresponding to the ligand upon fluorine release (see Figure 2 and Figure S2). Furthermore, we observed a formal loss of a water molecule (-18 Da) at the β 5 subunit which was validated by multiple experiments and different mass spectra deconvolution algorithms (see Figures S3 and S4 and Tables ST2 and ST3).

These findings suggest either an addition–elimination reaction as described for PMSF, which for example converts the active Ser195 of thrombin into dehydroalanine,^[17] or an addition–displacement mechanism comprising sulfonylation of Thr1O^{γ}, followed by an intramolecular substitution by Thr1N to yield an aziridine. While the PMSF-induced elimination reaction requires strong alkaline conditions, our experiments were carried out at pH 6.8–7.5, indicating that

dehydroxylation of Thr1 is rather an integral part of the inhibition mechanism than an artificially base-induced event. Consistently, the F_o-F_c electron density map clearly depicts the intramolecular displacement product (*S*,*S*)-aziridine-T1' (Figure 3b), thus excluding the elimination product (*E*)-dehydrobutyrine (Dhb) (Figure S1b, Supporting Information). Moreover, cyclization initiated by Thr1N is confirmed



Figure 3. Comparative X-ray analysis of the β5 active site after timedependent soaking experiments of yCP crystals with **1**. The 2*F*_o-*F*_c electron density maps (blue mesh, contoured at 1σ) show distances in Å as black dashed lines. The active site triad Thr1 (T1), Asp17 (D17), and Lys33 (K33) has been excluded prior to phasing. Stereoviews of (a)-(d) are depicted in Figure S6. a) Subunit β5 of the apo structure with unmodified T1 (yellow).^[18] b) Aziridine-T1' (yellow) formation on β5 after 2 h soaking time with **1**. The trajectory of the nucleophilic attack of K33 (green) is shown as a pink dashed arrow (PDB ID 4R17). c) K33-T1'-crosslink (yellow) formation on the apo structure with unmodified T1 (light gray), the aziridine-T1' intermediate (I, yellow), and the K33-T1'-crosslink (II, yellow). The structural rearrangement of T1 upon its conversion into intermediate I and crosslink II is illustrated with a pink dashed arrow.

by the inverted stereoconfiguration of the methyl group in (S,S)-aziridine-Thr1', implying an S_N2-like displacement. To analyze the stability of the aziridine ring we conducted further soaking experiments with extended incubation times up to 6 h at pH 6.8. The $2F_o-F_c$ electron density map of the yCP:**1** structure revealed a S_N2-type ring-opening of the aziridine-Thr1' by attack of the amino group of Lys33 (Lys33N^e), yielding an intramolecular crosslink in β 5 (Figure 3 c; 2.4 Å resolution, R_{free} = 19.5 %, PDB ID 4R18). This Lys33-Thr1' bond proves the presence of a polarity-inversed Thr1 intermediate and is certainly surprising, since the function of Lys33N^e is to maintain the pK_a of Thr1O^{γ}, and hence

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Scheme 1. Proposed three-step inactivation mechanism of the PSF compounds at the proteasomal active site of subunit β 5. Hypothetically, the sulfonylation of Thr1O^{γ} is conceivable by two different mechanisms: a direct nucleophilic attack of Thr1O^{γ} on the electrophilic sulfur center (upper left corner), or by elimination and sulfene formation (lower left corner). The substituent R indicates the rest of the peptidic backbone and P¹ refers to the amino acid side chain of the inhibitor protruding into the S1 specificity pocket.

possesses only a moderate intrinsic nucleophilicity.^[19] From an organic chemical point of view this aziridine ring-opening is also unexpected, since normally the presence of electron-withdrawing groups on the aziridine nitrogen is required.^[20]

Intriguingly, superposition of the apo and yCP:1 structures illustrates that the flexibility of the aziridine-Thr1'containing β 5 chain solely accounts for closing the 3.3 Å gap between Lys33N^e and the electrophile, while all remaining residues retain their position (Figure 3d). Consequently, the decline of structural integrity upon dehydroxylation of Thr1 presents a precondition to perform the crosslinkage. Based on these distinct snapshots of the reaction intermediates, we propose a three-step mechanism of the PSF compounds resulting in the crosslinking of the proteasomal active site (Scheme 1).

Our next goal was to investigate whether this complex mechanism could contribute to increased β 5i selectivity and thus reduced cytotoxicity. Therefore, we compared the potency and β 5 subunit selectivity of **1** and **2** with that of their α',β' -epoxyketone originals by performing in vitro IC₅₀ assays with purified human iCP, cCP, and yeast CP (yCP) (Table 1, Figure 4). Remarkably, **2** (IC₅₀ β 5c/ β 5i: 25) turned out to be roughly three times more selective for β 5i than ONX 0914 (IC₅₀ β 5c/ β 5i: 9), despite identical backbone architecture. However, the improved β 5i selectivity of **2** was accompanied by a 20-fold decreased potency (IC₅₀ (β 5i): 1134 nM) compared to ONX 0914 (IC₅₀ (β 5i): 57 nM), indicat-

Table 1: In vitro IC_{50} values [nm] against the ChTL activity of various purified CP types.

Compound	IC ₅₀ yβ5	IC ₅₀ β5c	IC ₅₀ β5i	IC ₅₀ β5c/β5i ^{[a}
CFZ	3 ± 1	5 ± 1	21 ± 3	0.2
1	21 ± 2	28 ± 2	54 ± 10	0.5
ONX 0914	$145\pm\!15$	513 ± 30	57 ± 10	9
2	15420 ± 635	$28460{\pm}1305$	1134 ± 146	25
3	1775 ± 476	3927 ± 550	139 ± 34	28

[a] A high IC₅₀ β 5c/ β 5i ratio indicates selectivity for β 5i.

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100 CFZ yβ5 CFZ B5c residual activity [% 75 CFZ 85 1 v 85 50 1 B5c 1 B5 25 0 0.00001 0.0001 0.001 0.01 0.1 10 100 c(inhibitor) [µM] 100 ONX 0914 v65 ONX 0914 85c ONX 0914 β5i residual activity [%] 75 **2** yβ5 2 ß5c 50 **2** β5i **3** yβ5 25 3 B5c **3** β5i 0.0001 0.001 0.01 0.1 10 1000 100 c(inhibitor) [µM]

Figure 4. In vitro IC_{50} assays against the ChTL activity of various purified CP types after 1 h incubation at various concentrations of CFZ and 1 (left panel) as well as ONX 0914, 2, and 3 (right panel) using a fluorogenic substrate assay. Data of three repetitions were normalized to DMSO-treated controls and are presented as relative activity with standard deviation (Table 1).

ing that PSF ligands require at least capped tetrapeptidic backbones for sufficient stabilization during proteasomal binding. Thus, we extended 2 with a homophenylalanine in P4 to generate compound 3 (Figure 1).

Since the S4 specificity pockets of iCP and cCP are identically shaped by the $\beta6$ subunit, **3** exhibited up to eightfold improved potency against $\beta5i$ (IC₅₀ ($\beta5i$): 139 nM) and $\beta5c$ (IC₅₀ ($\beta5c$): 3927 nM), respectively. Importantly, **3**

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Compound	IC ₅₀ HeLa	IC ₅₀ THP-1	LC ₅₀ HeLa	LC ₅₀ THP-1	LC ₅₀ /IC ₅₀ ^[a] HeLa	LC ₅₀ /IC ₅₀ ^[a] THP-1		
CFZ	6±1	7 ± 1	28 ± 6	12±1	5	2		
1	29 ± 3	35 ± 4	$353{\pm}63$	156 ± 16	12	4		
ONX 0914	78 ± 10	22 ± 2	333 ± 48	110 ± 14	4	5		
2	5032 ± 577	1789 ± 378	>100000	>100000	>19	>56		
3	550 ± 59	146 ± 40	>100000	5746 ± 1436	>182	39		

Table 2: In vivo IC_{50} values [nm] against the ChTL activity and LC_{50} values [nm] against HeLa and THP-1 cells.

[a] A high LC₅₀/IC₅₀ ratio indicates low cytotoxicity relative to the corresponding IC₅₀ value.

displayed an even enhanced β 5i selectivity (IC₅₀ β 5c/ β 5i: 28) compared to **2**. By contrast, **1** and CFZ inhibited the β 5 subunits of all analyzed CP types in the single- and double-digit nanomolar range without any preference for β 5i. This suggests that the PSF headgroup emphasizes the influence of the peptidic backbone to a greater extent than the α',β' -epoxyketone warhead.

The in vitro findings were confirmed by determining the in vivo IC₅₀ values of **1**, **2**, their α',β' -epoxyketone counterparts as well as **3** in cell culture assays (Table 2, Figure 5 a). We used either the THP-1 cell line, which is derived from acute monocytic leukemia and expresses high levels of iCP, or the HeLa cell line, which primarily contains cCP.^[21] ONX 0914, **2**, and **3** turned out to be three to four times more selective for THP-1 cells over HeLa cells, while **1** and CFZ displayed virtually similar potencies on both cell lines in accordance with the in vitro IC₅₀ assays.

To evaluate the cytotoxic profiles of the compounds we determined the LC₅₀ values against THP-1 and HeLa cells in viability assays (Table 2, Figure 5b). Notably, **2** and **3** hardly affected the viability of either cell line, arguably reflecting their high β 5i selectivity and low off-target binding. The in vivo IC₅₀ values of **1** (IC₅₀ (THP-1): 35 nm, IC₅₀ (HeLa): 29 nm) exclude that the low cytotoxicity is caused by

a rapid hydrolysis of the PSF headgroup.

In summary, the presented comparative study of iCPselective α',β' -epoxyketone and PSF counterparts highlights the sulfonyl fluoride headgroup as a promising motif for $\beta5i$ targeting. In contrast to all analyzed CP inhibitors to date, PSFs manipulate the proteasomal activity by a previously unobserved mode of action through polarity inversion and intramolecular crosslinking of the active site. In contrast to the unspecific serine protease inhibitor PMSF, the additional peptidic backbone accounts for site-selective proteasome inhibition. Thus, target-specific iCP blockage of the PSF in the nanomolar range along with low cytotoxicity broadens the therapeutic window of PSF as potential future anti-inflammatory inhibitors.

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Figure 5. a) In vivo IC₅₀ assays against the ChTL activity in HeLa and THP-1 cells after 105 min incubation at various concentrations of CFZ and 1 (left panel) as well as ONX 0914, **2**, and **3** (right panel) using a luminogenic substrate assay. b) LC_{50} against HeLa (left panel) and THP-1 (right panel) cells after 48 h incubation at concentrations of CFZ, **1**, ONX 0914, **2**, and **3** between 1 nm and 100 μ m using an AlamarBlue-based cell viability assay. Data of three repetitions were normalized to DMSO-treated controls and are presented as relative activity with standard deviation (Table 2).

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Immunoproteasome Inhibition

C. Dubiella, H. Cui, M. Gersch, A. J. Brouwer, S. A. Sieber, A. Krüger, R. M. J. Liskamp,* M. Groll*

Selective Inhibition of the Immunoproteasome by Ligand-Induced Crosslinking of the Active Site



The current benchmark for proteasome inhibition, immunoproteasome inhibitors featuring α',β' -epoxyketones, were compared to peptido sulfonyl fluorides (PSF) in terms of their mechanism of action,

selectivity, and cytotoxicity. PSFs were found to remove the catalytically active nucleophile and then crosslink the active site. Cell-based activity and viability assays designate this warhead for selective immunoproteasome blockage.

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