$ER\beta$ Ligands. 3. Exploiting Two Binding Orientations of the 2-Phenylnaphthalene Scaffold To Achieve ER β Selectivity

Richard E. Mewshaw,*,[†] Richard J. Edsall, Jr.,[†] Cuijian Yang,[†] Eric S. Manas,[†] Zhang B. Xu,[‡] Ruth A. Henderson,[§] James C. Keith, Jr.,^{II} and Heather A. Harris[§]

Chemical and Screening Sciences and Women's Health Research Institute, Wyeth Research, 500 Arcola Road, Collegeville, Pennsylvania 19426, and Chemical and Screening Sciences and Cardiovascular and Metabolic Diseases, Wyeth Research, 200 Cambridge Park Dr., Cambridge Massachusetts 02140

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The 2-phenylnaphthalene scaffold was explored as a simplified version of genistein in order to identify ER selective ligands. With the aid of docking studies, positions 1, 4, and 8 of the 2-phenylnaphthalene template were predicted to be the most potentially influential positions to enhance ER selectivity using two different binding orientations. Both orientations have the phenol moiety mimicking the A-ring of genistein. Several compounds predicted to adopt orientations similar to that of genistein when bound to $ER\beta$ were observed to have slightly higher ER affinity and selectivity than genistein. The second orientation we exploited, which was different from that of genistein when bound to $\text{ER}\beta$, resulted in the discovery of several compounds that had superior ER selectivity and affinity versus genistein. X-ray structures of two ER selective compounds (i.e., 15 and 47) confirmed the alternate binding mode and suggested that substituents at positions 1 and 8 were responsible for inducing selectivity. One compound (i.e., 47, WAY-202196) was further examined and found to be effective in two models of inflammation, suggesting that targeting ER may be therapeutically useful in treating certain chronic inflammatory diseases.

Introduction

Estrogens are a family of naturally occurring steroid hormones that play a central role in the growth, development, and maintenance of a diverse range of tissues. Their actions are mediated by estrogen receptors (ER), which are members of a superfamily of nuclear receptors that function as ligand-activated gene transcription factors. Prior to 1996, estrogen-mediated events had been assumed to be regulated by only one estrogen receptor (now called ER α).¹ However, the discovery of a second estrogen receptor subtype (ER β) in 1996 led to an intense interest in elucidating $ER\beta$ function^{2,3} and determining which aspects of estrogen biology it mediated. Significant sequence homology is observed in the DNA and ligand binding domains (LBDs) of ER α and ER β ; however, the expression pattern of the two subtypes is different. Although widely found in many tissues, $ER\beta$ is predominately expressed in ovarian granulosa cells, lung, bladder, and prostate, whereas ERa is the predominant ER expressed in the uterus, kidney, and ovarian theca cells.4-6 Over the last several years our laboratories and others have been focused on identifying selective $ER\beta$ ligands from several classes of molecules, e.g., biphenyls,^{7,8} tetrahydrochrysenes,⁹ diarylpropionitriles,¹⁰ arylbenzthiophenes,¹¹ and triazines.¹² The challenge to identify highly selective compounds has stifled progress to probe the physiological function of ER β . Only a small group of molecules has been reported to have $ER\beta$ selectivities above 40fold.¹³ However, with the recent identification of the highly ER β selective agonist ERB-041, our company discovered that one function of ER β may be to modulate the inflammatory response.^{14,15}

In an effort to continue to identify novel nonsteroidal ER agonists, our group initially focused on the modestly selective structure of genistein (1) as a starting point.¹⁶ In the three-dimensional structure of human ER complexed with genistein (1),¹⁷ the phenolic hydroxyl group interacts with a Glu-Arg-water triad at one end of the cavity through hydrogen-bonding interactions, while the chromenone hydroxyl at C(7) forms a hydrogen bond with a His side chain at the other end of the cavity. The remaining hydroxyl at C(5) is believed to be hydrogen bonded to the adjacent carbonyl group¹⁸⁻²³ and does not interact with the protein. Our initial efforts focused on whether the isoflavone framework of genistein could be mimicked with an appropriately substituted biphenyl scaffold (Figure 1). Though we showed that incorporating only a single phenolic group (i.e. 2) within the 4-OHbiphenyl motif produced $ER\beta$ selective ligands, only modest affinity and selectivity could be achieved.⁸ To further mimic the key pharmacophoric groups of genistein, an oxime moiety was appended to the 4-OH-biphenyl scaffold, resulting in the identification of a 4-OH-

^{*} To whom correspondence should be addressed. Telephone: 484-865-2910. Fax: 484-865-9399. E-mail: mewshar@wyeth.com.

[†] Chemical and Screening Sciences, Collegeville, PA. [‡] Chemical and Screening Sciences, Cambridge, MA.

[§] Women's Health Research Institute, Collegeville, PA.

[&]quot;Cardiovascular and Metabolic Diseases, Cambridge, MA.

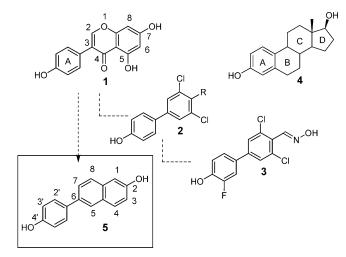


Figure 1. Compounds of interest.

biphenyl oxime derivative (i.e. 3) having affinity and selectivity similar to that of genistein (1).⁷ In this paper, we will disclose further efforts at creating a new template that can embrace similar spatial and chemical features of genistein (1) that fulfill the ER pharmacophoric requirements, while exploring how to enhance $ER\beta$ selectivity. The classical ER pharmacophore basically consists of an A-ring phenol linked by a hydrophobic core to a second hydroxyl group placed approximately 11 Å from the A-ring phenolic OH to mimic the 17hydroxy group of estradiol (4).²⁴ The phenol group appears to be the key pharmacophoric group needed to achieve ligand recognition and high affinity for ER. One such scaffold that conforms to this design strategy is the 6-phenylnaphthalene framework (i.e. 5). In fact, the 6-phenylnaphthalene motif has been previously employed in the anti-estrogen field.²⁵

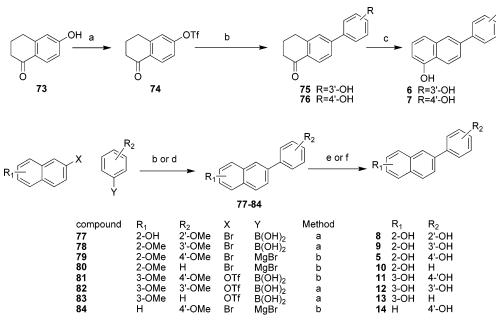
The 6-phenylnaphthalene scaffold can present similar spatial and conformational requirements as genistein (1), while at the same time provide the synthetic chemist a platform to expedite structural modifications. The 6-phenylnaphthalene moiety actually represents a simplified version of genistein, in which the heteroatoms in the B and C rings of genistein (1) have been removed, providing us with a further understanding of their role in ER β binding and selectivity. Furthermore, although the 6-phenylnaphthalene scaffold was initially intended to serve as a "stripped-down" version of genistein, we present evidence to support our hypothesis that two of the binding orientations, one genistein-like and another that is flipped relative to the genistein binding mode, provide opportunities to enhance ER β selectivity beyond that of genistein. We now report the design, synthesis, and structure-activity relationships (SAR) of a new class of $ER\beta$ selective ligands that embrace the 6-phenylnaphthalene scaffold to achieve superior ER affinity and selectivity to that of genistein (1).

Chemistry

Synthesis. All target molecules prepared as depicted in Schemes 1-10 are described in detail in the Experimental Section. Synthesis of the 1-hydroxy-6-phenylnaphthalenes (**6** and **7**) began with commercially available 6-hydroxytetralone, which was converted to the triflate and coupled to the appropriate phenyl derivative, followed by aromatization (Scheme 1). Compounds 5, 8–14 were prepared by coupling the bromo or triflate derivatives (i.e., 77-84) to the appropriately substituted boronic acid (method a) or Grignard reagent (method b), followed by demethylation using either boron tribromide or pyridinium hydrochloride (methods c or d, respectively). Chlorination or bromination led to 85 and 86, respectively (Scheme 2). Bromide 86 was converted to the 1-substituted derivatives (87–90), which were subsequently demethylated. Bromide 16 was converted to the corresponding methoxy derivative (21) using cuprous bromide and sodium methoxide. Nitration followed by Suzuki coupling and deprotection led to 22. Suzuki coupling of the appropriately substituted naphthalenes and phenyl derivatives followed by deprotection led to the 6-phenylnaphthalene derivatives (i.e., 23, **25**, **27**, **29**, **31**, **32**, **34**, **36**, **38**) as depicted in Scheme 3. Subsequent chlorination with NCS (method E) of the appropriate 6-(4-hydroxyphenyl)-2-naphthols led to the 1-chloro-6-phenylnaphthalenes (24, 26, 28, 30, 33, 35, **37**, **39**). Shown in Scheme 4 is the rather lengthy route used to prepare the 8-fluoro-6-phenylnaphthalenes 40-43, beginning with commercially available 1-amino-7-hydroxynaphthalene (106). A three-step sequence was used to methylate 106 to afford the methoxy derivative 109. Diazoniation and demethylation gave naphthol 111, which was dibrominated to afford 112. Selective debromination with SnCl₂ afforded **113**, which when reacted under Suzuki conditions led to 116 and 117. The methoxy derivatives (i.e., 116, 117) could either be demethylated with boron tribromide to afford 40 and 41 or further reacted with NCS, followed by demethylation, to produce 42 and 43. The 8-chloro-6-phenylnaphthalene derivatives 44 and 45 were prepared from intermediate 109 in a straightforward manner as depicted in Scheme 5. Target molecules 46-50 were prepared from commercially available tetralone 129 (Scheme 6). Briefly, tetralone 129 was converted to the cyano derivative 130, which was aromatized with palladium in cymene followed by demethylation with pyridinium hydrochloride to produce naphthol 132. Dibromination followed by selective debromination led to the key intermediate 133, which when coupled under Suzuki conditions led to the phenylnaphthalenes 134 and 135. Subsequent modifications of 134 and 135 in a straightforward manner afforded 46-50. As shown in Scheme 7, naphthol 133 was converted to 51 in four steps. Further elaboration of the 8-cyano group of 47 led to the corresponding aldehyde, vinyl, ethyl, and alkynyl analogues (52-56). Several fluoro-subsituted analogues (i.e., 57-60) were prepared using the appropriately substituted phenylboronic acids under Suzuki conditions (Scheme 8). The 4-cyano-6-phenylnaphthalenes (i.e., 61 and 62) were prepared in four steps (Scheme 9) from key intermediate 133. Subsequent modification of 61, 62, and 152 led to the 1,4-disubstituted-6-phenylnaphthalenes (63-67). Finally, the 4-cyano derivative 62 was modified (Scheme 10) to afford the corresponding 4-substituted derivatives **68–72** by employing similar chemistry as previous shown in Scheme 7.

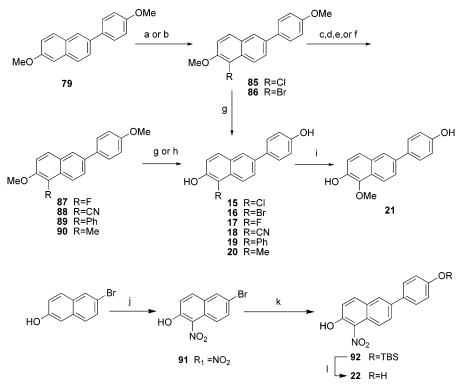
Crystallography. The overall structures of $\text{ER}\beta$ LBD complexed with 15 and with 47 are similar to those

Scheme 1^a



^{*a*} Reagents: (a) Tf₂O, C₆H₅N, CH₂Cl₂, 0 °C; (b) Pd[P(Ph)₃]₄, Na₂CO₃, DME/H₂O, 85 °C (method A); (c) 10% Pd/C, *p*-cymene, reflux; (d) ArMgBr, Pd[P(Ph)₃]₄, (method B); (e) C₆H₅N·HCl, 190 °C (method D); (f) BBr₃, CH₂Cl₂, 0 °C (method C).

Scheme 2^a



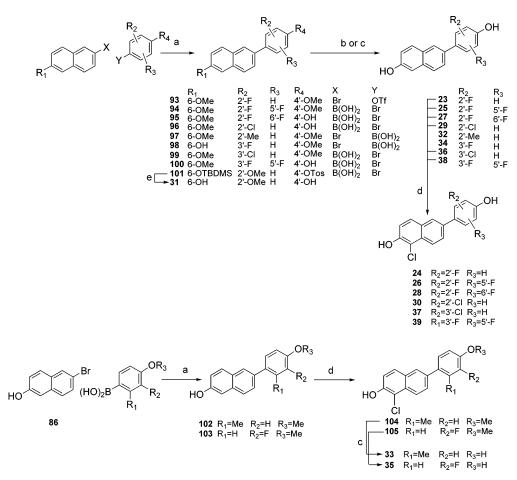
^a Reagents: (a) NCS (method E) (b) Br₂/AcOH; (c) n-BuLi, NFSI; (d) CuCN, DMF, 120 °C; (e) PhMgBr, Pd[P(Ph)₃]₄ (method A); (f) n-BuLi, MeI; (g) C₆H₅N·HCl, 190 °C (method D); (h) BBr₃, CH₂Cl₂, 0 °C (method C); (i) CuBr, NaOMe, DMF; (j) 4-NO₂-4-Me-2,3,5,6-Br₄-2,5-cyclohexadien-1-one; (k) Pd[P(Ph)₃]₄, Na₂CO₃, DME/H₂O, 85 °C (method A); (l) TBAF.

previously reported by our group^{14,26} and by others^{17,27–29} and thus they will not be described in detail here. We point out that in both cases the helix-12 conformation corresponds to that observed for other bound agonists, consistent with the observed functional activity within this series (see below). When helix-12 adopts an "agonist-like" conformation, the ligand becomes completely encapsulated by the protein, as a result of the helix "closing" over the binding pocket. The closing of helix-

12 also forms a cavity on the surface of the ER LBD, formed by helices H3, H4, H5, and H12, and the turn between H3 and H4. In our structures, a coactivator fragment binds to this cavity in a α -helical conformation, in a manner similar to what has been previously described for other ER α and ER β LBD complexes (see Figure 2).

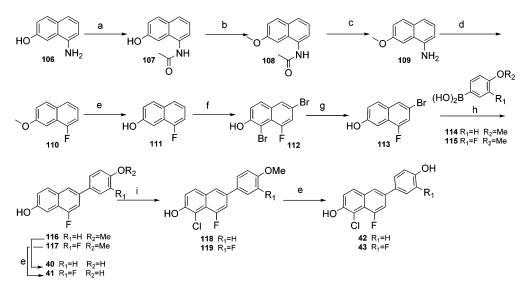
Crystal structures of the ER α and ER β binding pockets have been reported to have relatively consistent

Scheme 3^a



^{*a*} Reagents: (a) $Pd[P(Ph)_3]_4$, Na_2CO_3 , DME/H_2O , 85 °C (method A); (b) C_6H_5N ·HCl, 190 °C (method D); (c) BBr₃, CH_2Cl_2 , 0 °C (method C); (d) NCS (method E); (e) KOH/H₂O/EtOH, 90 °C.

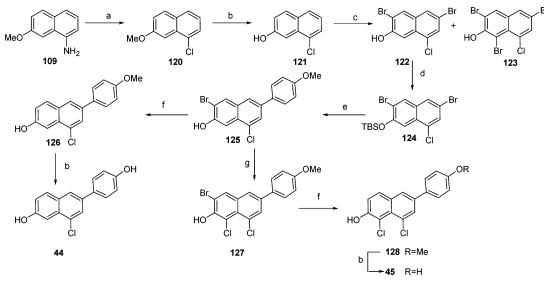
Scheme 4^a



^{*a*} Reagents: (a) Ac₂O (b) MeI, K₂CO₃; (c) 1N HCl; (d) (i) NaNO₂ (ii) HBF₄; (iii) xylenes, reflux; (e) BBr₃, CH₂Cl₂, 0° (method C); (f) Br₂/AcOH; (g) SnCl₂; (h) Pd[P(Ph)₃]₄, Na₂CO₃, DME/H₂O, 85 °C (method A); (i) NCS (method E).

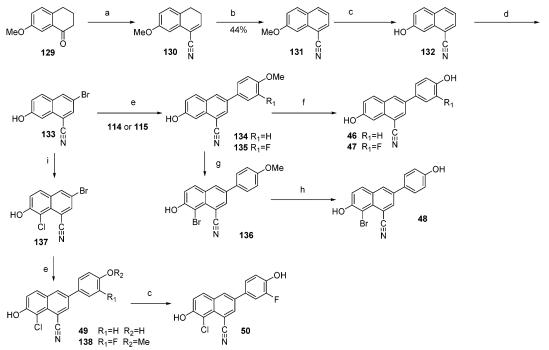
tertiary architecture. Comparing X-ray cocrystal structures of ER α and ER β complexed with agonists^{14,17,26–29} reveals only two residue substitutions within 5 Å of the bound ligand. The leucine present at position 384 in ER α is replaced by a methionine in ER β (i.e., Met₃₃₆), and the methionine at position 421 in ER α is replaced by an isoleucine in ER β (i.e. Ile₃₇₃). It has been hypothesized from modeling studies in our labs^{7,8,14,26} and others¹² that there exist an opportunity to achieve enhanced selectivity for ER β by identifying a ligand that could differentiate the substitution of ER α Met₄₂₁ to ER β Ile₃₇₃, which lies at the α -face of ER-bound estradiol. Briefly, the substitution of Met₄₂₁ by Ile₃₇₃ may impart the ability of certain appropriately placed func-

Scheme 5^a



^{*a*} Reagents: (a) CuCl, *tert*-butyl nitrite, MeCN; (b) BBr₃, CH₂Cl₂, 0 °C (method C); (c) Br₂/AcOH; (d) TBSCl; (e) ArMgBr, Pd[P(Ph)₃]₄, (method B); (f) (i) t-BuLi, (ii) H₂O; (g) NCS (method E).

Scheme 6^a



^a Reagents: (a) (i) TMSCN/ZnI₂ (ii) POCl₃, AcOH (b) 10% Pd/C, *p*-cymene, reflux; (c) C_6H_5N ·HCl, 190 °C (method D); (d) (i) $Br_2/AcOH$ (ii) $SnCl_2$; (e) $Pd[P(Ph)_3]_4$, Na_2CO_3 , DME/H_2O , 85 °C (method A); (f) C_6H_5N ·HCl, 190 °C (method D); (g) NBS; (h) BBr_3, CH_2Cl_2, 0° (method C); (i) NCS (method E).

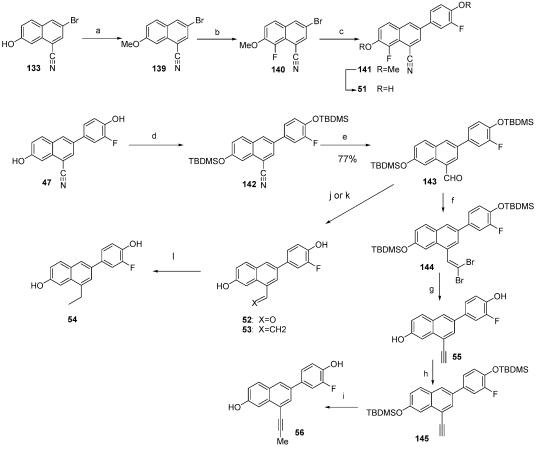
tional groups to achieve stereo electronic differentiation between these two residue side chains, leading to enhanced $\text{ER}\beta$ selectivity.

$ER\beta$ Selectivity

Optimization of Key Hydroxyl Groups. Shown in Tables 1–6 are the affinities (as measured by IC₅₀s) and selectivities for human ER α or ER β of all compounds of interest using a competitive radioligand binding assay.³⁰ Since the ER has been reported to have some tolerance for the oxygen–oxygen distance²⁴ (D_1) (e.g., genistein $D_1 = 12.2$ Å; estradiol $D_1 = 10.9$ Å), a better understanding of the roles of the crucial hydroxyl groups, with respect to ER β affinity and

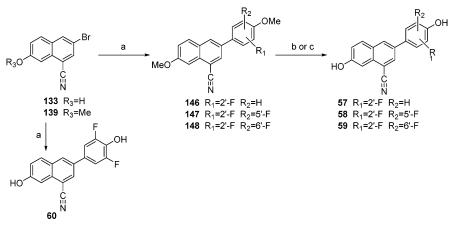
selectivity, needed to be addressed within the phenylnaphthalene scaffold. As shown in Table 1, compound 5 ($D_1 = 12.0$ Å), having its hydroxyl groups in comparable positions to that of genistein (1), was also observed to have similar affinity. However, the ER β selectivity of 5 decreased 3-fold versus 1, suggesting that one or more of the other three oxygens of 1, not present in 5, plays a significant role in determining ER β selectivity. Naphthalene 7 ($D_1 = 10.7$ Å) was observed to have the highest affinity for ER β ; however, selectivity decreased 10-fold versus 1. The 4'-OH phenyl group was always observed to be the most potent derivative within its respective group of 1-, 2- or 3-OH substituted naphthalenes (i.e., 6 vs 7; 8 vs 5; 9 vs 5; 11 vs 12). Removal of





^{*a*} Reagents: (a) MeI, K₂CO₃; (b) Selectfluor; (c) Pd[P(Ph)₃]₄, Na₂CO₃, DME/H₂O, 85 °C (method A); (d) TBSCl; (e) DIBAL; (f) CBr₄, Zn, P(Ph)₃; (g) (i) n-BuLi, (ii) TBAF; (h) TBDSCl, (i) n-BuLi, MeI; (j) P(Ph₃)CH₃Br, n-BuLi; (k) TBAF; (l) H₂ 10% Pd/C.

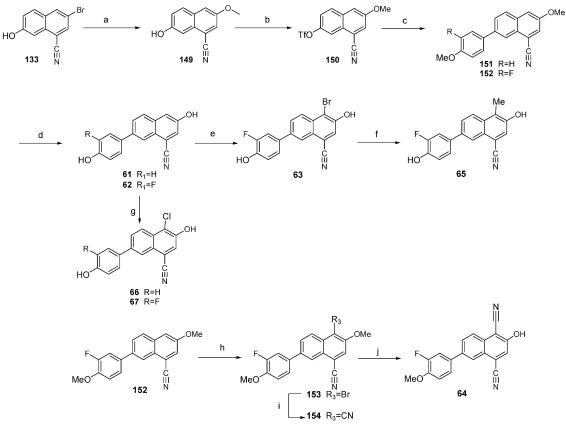
Scheme 8^a



^{*a*} Reagents: (a) $Pd[P(Ph)_3]_4$, Na_2CO_3 , DME/H_2O , 85 °C (method A); (b) C_6H_5N ·HCl, 190 °C (method D); (c) BBr₃, CH_2Cl_2 , 0 °C (method C).

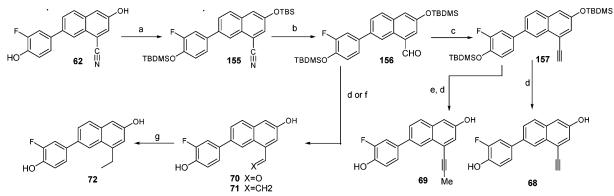
one of the hydroxyl groups of **5** led to a 19–29-fold loss in affinity (i.e., **5** vs **10**, and **5** vs **14**), confirming the importance of having two hydrogen bonding groups within the 6-phenylnaphthalene scaffold. However, the monohydroxyl derivatives **10**, **13**, and **14** were still more potent than the dihydroxyl analogue **8**, suggesting that the 2'-OH phenyl group was playing no role in hydrogen bonding ($D_1 = 8-8.4$ Å) and was actually having a detrimental effect. Also worth mentioning are that monohydroxyl derivatives **10** and **14** had similar ER β affinities. Since it is known that the phenolic A-ring of estrogens participates in key hydrogen bonds with the receptor,²⁴ this suggests that the hydroxyl groups of **10** and **14** are involved in the essential hydrogenbond network that mimics the A-ring of estradiol using two completely different binding orientations of the phenylnaphthalene scaffold. In the case of **10**, the naphthol moiety is mimicking the A-ring of estradiol, whereas in **14** the phenolic ring mimics the A-ring of estradiol. This supports the notion that the phenylnaphthalene scaffold is capable of what we term "A-C-ring flipping" or "horizontal flipping," due prima-

Scheme 9^a



^{*a*} Reagents: (a) NaOMe, CuBr (method F); (b) Tf₂O, C₆H₅N, CH₂Cl₂, 0 °C; (c) Pd[P(Ph)₃]₄, Na₂CO₃, DME/H₂O, 85 °C (method A); (d) C₆H₅N·HCl, 190 °C (method D); (e) NBS; (f) SnMe₄, PdCl₂P(*o*-tolyl)₃; (g) NCS (method E); (h) Br₂/AcOH; (i) Pd(0), Zn(CN)₂; (j) C₆H₅N·HCl, 190 °C (method D).





^{*a*} Reagents: (a) TBDSCl; (b) DIBAL; (c) (i) CBr₄, Zn, P(Ph)₃; (ii) n-BuLi; (d) TBAF; (e) n-BuLi, MeI; (f) P(Ph)₃CH₃Br, n-BuLi; (g) H₂ 10% Pd/C.

rily to the pseudosymmetry of this simplified scaffold (see below).

Opportunities To Enhance Selectivity. To be able to fully understand how to exploit the residue substitutions in ER β relative to ER α , it is necessary to have an understanding of the binding orientation that is favored by the chosen template. As discussed above in the case of estradiol, the phenol ring is involved in a hydrogenbonding interaction with Glu₃₀₅ and Arg₃₄₆, and the 17 β -OH group forms a hydrogen bond to His₄₇₅. In this case, one of the primary determinants of the ligand-binding mode appears to be the shape and asymmetry of the estradiol steroidal scaffold. Similarly, in the case of genistein the phenol ring mimics the A-ring of estradiol,

while the second phenolic hydroxyl group on the chromenone ring mimics the 17β -OH group of estradiol. Again, the ability of ER to differentiate the two phenolic groups and ultimately determine the true binding mode must rely on the structural (i.e. geometric) features of the genistein framework. On the other hand, a simplified template, such as phenylnaphthalene **5**, may have no obvious means to allow ER to differentiate one potential binding mode from another. In fact, one could argue that **5** possesses pseudo D_{2h} symmetry, and thus there appear to be essentially four reasonable binding orientations that **5** can adapt when bound to ER (Figure 3). While these orientations all seem to yield similar interactions with the receptor, docking studies also sug-

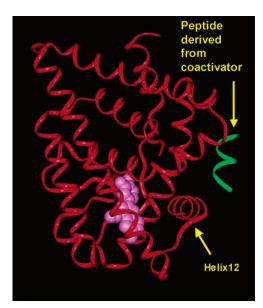
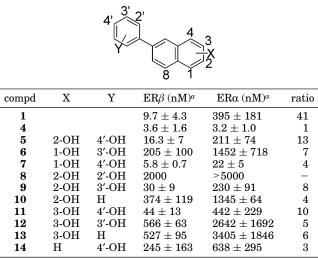


Figure 2. Ribbon diagram of the X-ray structure of **47** complexed to $\text{ER}\beta$.

Table 1. Optimizing Hydroxyl Groups



 a IC₅₀ values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate). Values without SDs are for a single determination only.

gest that orientations a ("genistein-like") and b ("vertically flipped", i.e., about the longest axis of the molecule) may provide the greatest opportunity to take advantage of the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution, by introducing functional groups that interact less favorably with ER α Met₄₂₁ relative to ER β Ile₃₇₃. In contrast, the two "horizontally flipped" orientations c and d may be less desirable, because the phenyl ring can easily rotate to achieve more favorable interactions with ER α Met₄₂₁. In addition, interactions between the B-ring and $\text{ER}\beta$ Met₃₃₆, which previous studies suggest may contribute to ER β selectivity,⁸ appear to be more difficult to maintain in these orientations. In the light of this discussion, the most likely positions to influence $ER\beta$ selectivity on the phenylnaphthalene template would be the 4 position, using the genistein-like orientation, and positions 1 and 8, using the vertically flipped orientation. Interestingly, even though we initially intended to mimic the genistein scaffold, we were most successful at enhancing ER β selectivity beyond that of genistein by taking advantage of the vertically flipped orientation. It is to these results that we now turn our attention.

Selectivity Enhancements Obtained Utilizing the Vertically Flipped Orientation. As shown in Table 2, attaching a chlorine substituent (i.e. 15) to the 1 position led to a 6.5-fold increase in ER β affinity versus 5 and 36-fold selectivity for ER β . Docking studies predicted 15 to adopt a vertically flipped binding mode, which was verified by an X-ray cocrystal structure of 15 bound to the ER β LBD (see Figure 4a). As predicted, the 1-chloro substituent is directed at the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution, consistent with the selectivity improvement seen for 15 relative to 5. We also point out that the chloro group was found to be an effective substituent in our earlier work within the biphenyl series.⁸

Shown in Table 3 is a summary of the substituent effects on the phenolic ring with respect to 5 and 15. In general, with the exception of 31, all modifications of the phenolic ring of **5** led to an improvement in $ER\beta$ affinity, with very little effect on $ER\beta$ selectivity. However, this trend was not observed when attaching substituents to the phenolic ring of 15. Only in the case of attaching a 2'-fluoro group to 15 (i.e. 24) did a slight increase in both $ER\beta$ affinity and selectivity occur. Attachment of a second chloro group ortho to the phenol hydroxyl of **15** (i.e. **37**) led to over a 12-fold loss in ER β affinity. The fact that this trend was not observed when comparing 5 to 36, in terms of a loss in $\text{ER}\beta$ affinity, suggests that **36** may be binding in a different orientation than 15. Since it is known that the A-ring of estradiol is not very tolerant toward o-chloro groups,²⁴ we hypothesize that **36** may be maintaining its affinity using the naphthol moiety as the A-ring mimic (i.e. horizontally flipped binding mode), whereas dichloro 37 does not have this alternative orientation to exploit, since both of its hydroxyl groups are flanked by chloro groups, which thus results in a diminished ER affinity. Addition of a single 2'-F or 3'-F group to the phenolic ring of 15 either maintained or slightly improved $\text{ER}\beta$ affinity and selectivity (i.e. 24 and 35). Attaching two o-fluoro groups to the phenol of 15 (i.e. 39) led to a slight increase in ER β selectivity, with the concurrent loss in $ER\beta$ affinity. This loss in affinity can be explained by electrostatic repulsion between one of these fluoro groups and the side chain of Glu₃₀₅.¹⁴

As mentioned above, docking calculations predicted that position 8 might have a potentially beneficial effect on ER β selectivity. As shown in Table 4, several compounds were indeed identified to possess enhanced $ER\beta$ selectivity when exploiting this position. For example, the cyano derivative **46** was found to be 46fold selective for $\text{ER}\beta$ and could be further improved to achieve 78-fold selectivity by the addition of a 3'-fluoro group (i.e. 47). An X-ray cocrystal structure of 47 complexed to $\text{ER}\beta$ was obtained (Figures 2 and 4b) in order to confirm the predicted binding mode (Figure 3b). In particular, the 8-cyano group is directed toward $\text{ER}\beta$ Ile₃₇₃, which may represent a more favorable interaction than with the corresponding ERa Met₄₂₁.²⁶ However, this difference is a subtle one, given that the 3.5-fold difference in selectivity observed when comparing 5 to **46** corresponds to only a 0.75 kcal/mol binding free

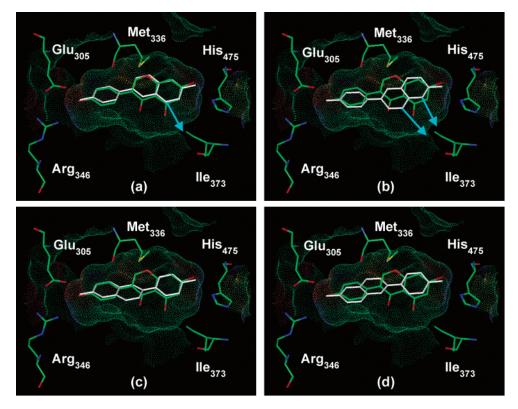
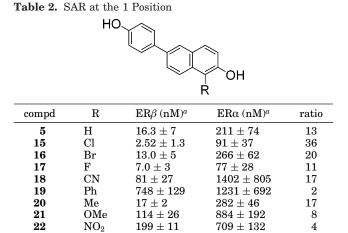


Figure 3. Four potential binding motifs (orientations a-d) identified by docking compound 5 to the ER β LBD.

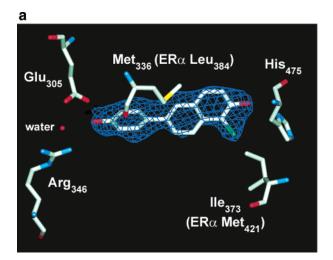


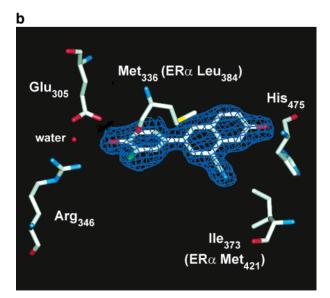
 a IC_{50} values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate).

energy difference at room temperature. We also point out that, in addition to direct interaction with ERa Met₄₂₁/ER β Ile₃₇₃, 8-position substituents may indirectly modulate the electronic interaction between the B-ring and ER β Met₃₃₆ relative to ERa Leu₃₈₄. The even more subtle selectivity effect induced by the 3'-fluoro substituent was also observed within the 4-OH-biphenyl series reported in parts 1 and 2,⁸ as well as within a 2-phenyl benzoxazole series reported previously.^{14,26} A structure-based explanation for this effect has been provided elsewhere.²⁶

Several other analogues were also observed to be more selective than genistein. In particular, the ethyl analogue **54** was 98-fold selective for ER β . Docking calculations on **54** suggest a vertically flipped binding mode, similar to that observed for **47** complexed to ER β , placing the ethyl group near ER α Met₄₂₁/ER β Ile₃₇₃. Thus, the selectivity enhancement due to the ethyl moiety can be explained by a favorable dispersive interaction with $\text{ER}\beta$ Ile₃₇₃, relative to a less favorable or unfavorable (i.e. steric) interaction with $ER\alpha$ Met₄₂₁ (which may translate to increased strain of either the ethyl group, the protein, or both). However, the docking calculations also suggest the possibility that 54 might adopt a genistein-like binding mode when bound to ERa and/or ER β . In this case, the enhanced selectivity of 54 relative to 5 can also be rationalized by more favorable contact (and thus more favorable dispersive interactions) with $ER\beta$ Met₃₃₆ relative to $ER\alpha$ Leu₃₈₄, resulting from the ethyl projecting $\sim 90^\circ$ out of the naphthalene plane. As with the 8-cyano substituent, we cannot discount the possibility that the 8-ethyl group modulates the differential interaction between the Bring and ER β Met₃₃₆ relative to ER α Leu₃₈₄ in either binding mode.

Interestingly, the chloro group, which induced modest selectivity from the 1 position (i.e. 15), has virtually no effect on ER β selectivity relative to 5 when placed at the 8-position (i.e. 44; 13-fold selective), demonstrating that certain substituents can only induce $\text{ER}\beta$ selectivity from appropriate positions of the phenylnaphthalene scaffold, even though they may have a similar trajectory within the ligand binding cavity. Also, no synergistic effect was observed by having both the 1-chloro and 8-cyano substituents (i.e. 49) placed onto the phenylnaphthalene template, which actually had a detrimental effect on ER β affinity and selectivity. This appears to be a consequence of the fact that any given functional group can modulate the interactions between the rest of the ligand and the protein. For example, the interaction of the 8-cyano of 49 with nearby residues can





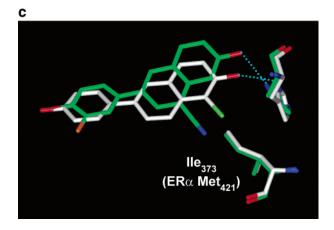
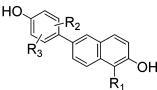


Figure 4. Unbiased $2F_{\circ} - F_{c}$ maps for **15** and **47** complexed to the ER β LBD unambiguously define the ligand binding modes of (a) **15** and (b) **47**. For both ligands, the phenol mimics the estradiol A-ring, with the phenolic hydroxyl (4'-OH) involved in a hydrogen-bonding network between ER β residues Glu₃₀₅ and Arg₃₄₆ (ER α residues Glu₃₅₃ and Arg₃₉₄) and a highly ordered water molecule. Another hydrogen bond is formed between the naphthalene hydroxyl (2-OH) and N₆₁ of ER β His₄₇₅ (ER α His₅₂₄). The core scaffold, consisting of the A-ring phenyl and the naphthalene B and C rings, fills the remainder of the primarily hydrophobic pocket. (c) An overlay of **15** and **47** in the ligand binding pocket.

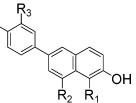
Table 3. A-Ring Substituent Effects



compd	\mathbf{R}_{1}	R_2	\mathbf{R}_3	$\mathrm{ER}\beta(\mathrm{nM})^a$	$ER\alpha(nM)^a$	ratio
5	Н	Н	н	16.3 ± 7	211 ± 74	13
15	Cl	н	Н	2.52 ± 1.3	91 ± 37	36
23	Η	2'-F	Н	2 ± 0.3	24 ± 0.8	12
24	Cl	2'-F	Н	1.2 ± 0.3	58 ± 6	48
25	Η	2'-F	5'-F	2.8 ± 0.1	27 ± 13	10
26	Cl	2'-F	5'-F	8.5 ± 4	118 ± 40	14
27	Η	2'-F	6′-F	2.3 ± 0.1	10.3 ± 2.4	4
28	Cl	2'-F	6'-F	3.4 ± 1.3	35 ± 5	10
29	Η	2'-Cl	Н	1.4 ± 0.6	10 ± 5	7
30	Cl	2'-Cl	Н	3.3 ± 2	36 ± 23	11
31	Η	2'-OMe	Н	27 ± 6	174 ± 26	6
32	н	2'-Me	Н	10 ± 1	40 ± 1	4
33	Cl	2'-Me	Н	13	40	11
34	н	3'-F	Η	5 ± 1	92 ± 25	17
35	Cl	3'-F	Н	4.0 ± 1	143 ± 28	36
36	Η	3'-Cl	Н	11 ± 1.4	107 ± 30	10
37	Cl	3'-Cl	Η	32 ± 3	356 ± 65	11
38	Η	3'-F	5'-F	8.4 ± 3.6	92 ± 52	11
39	Cl	3'-F	5'-F	10.6 ± 0	519 ± 126	47

 $^a\,IC_{50}$ values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate). Values without SDs are for a single determination.

Table 4. SAR at the 1, 8, and 3' Positions

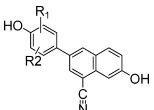


compd	\mathbf{R}_{1}	$ m R_2$	R_3	$\mathrm{ER}\beta(\mathrm{nM})^a$	$ER\alpha(nM)^a$	ratio
5	Η	Н	Η	16.3 ± 7	211 ± 74	13
40	Η	F	Η	1.6 ± 0.7	22 ± 1	14
41	н	F	\mathbf{F}	1.5 ± 0.6	21 ± 6	14
42	Cl	F	Η	1.1 ± 0.5	40 ± 11	36
43	Cl	F	\mathbf{F}	2.5 ± 1	125 ± 24	50
44	Η	Cl	Η	2.3 ± 1.7	30 ± 1	13
45	Cl	Cl	Η	9	43	5
46	Η	CN	Η	2.3 ± 1.3	105 ± 47	46
47	Η	CN	\mathbf{F}	2.7 ± 1.9	210 ± 122	78
48	\mathbf{Br}	CN	Η	12 ± 4	131 ± 50	11
49	Cl	CN	Η	6.0 ± 3.3	109 ± 29	18
50	Cl	CN	\mathbf{F}	11 ± 6	299 ± 147	27
51	\mathbf{F}	CN	\mathbf{F}	5.6 ± 2.7	312 ± 75	56
52	\mathbf{H}	CHO	\mathbf{F}	3.4 ± 0.5	231 ± 91	68
53	Η	$CH=CH_2$	\mathbf{F}	4.4 ± 3.7	254 ± 112	58
54	Η	ethyl	\mathbf{F}	2.4 ± 1	235 ± 31	98
55	Η	C≡CH	\mathbf{F}	6.3 ± 1.3	247 ± 190	40
56	Н	$C \equiv CCH_3$	\mathbf{F}	10 ± 6	343 ± 61	34

 a IC_{50} values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate). Values without SDs are for a single determination.

modify the trajectory groups at the 1-position (e.g. chloro), leading to reduced affinity and selectivity. This can be understood by overlaying the X-ray structures of ER β complexed with **15** and **47**, which clearly demonstrates that steric interactions between the 8-nitrile of **46** and the protein contribute to a shift of the 1-position relative to **15** (see Figure 4c).

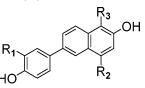
Table 5. A-Ring Substituent Effect Based on 46



compd	R_1	R_2	$\mathrm{ER}\beta(\mathrm{nM})^a$	ERa $(nM)^a$	ratio
46	Н	Н	2.3 ± 1.3	105 ± 47	46
47	3'-F	Η	2.7 ± 1.9	210 ± 122	78
57	2'-F	Η	0.5 ± 0.5	26 ± 3	55
58	2'-F	5-F	6.9 ± 1	186 ± 16	27
59	2'-F	6-F	2.2 ± 1.1	45 ± 17	20
60	3'-F	5-F	58 ± 45	548 ± 225	9

 a IC₅₀ values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate).

Table 6. SAR at the 1, 4, and 3' Positions

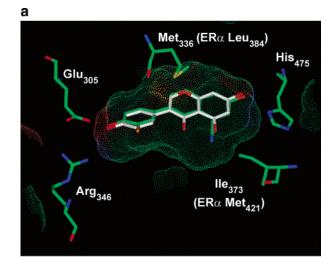


compd	\mathbf{R}_1	$ m R_2$	R_3	$\mathrm{ER}\beta(\mathrm{nM})^a$	$ER\alpha(nM)^a$	ratio
5	Н	Н	Н	16 ± 7	211 ± 74	13
61	Η	CN	н	2.0 ± 1.5	84 ± 33	42
62	\mathbf{F}	CN	н	2.1 ± 0.9	96 ± 42	46
63	\mathbf{F}	CN	\mathbf{Br}	4.5 ± 1.8	116 ± 47	26
64	\mathbf{F}	CN	CN	28 ± 5	877 ± 48	31
65	\mathbf{F}	CN	Me	6.0 ± 0.1	406 ± 136	68
66	Η	CN	Cl	3.0 ± 2.6	98 ± 12	32
67	\mathbf{F}	CN	Cl	2.6 ± 2.7	55 ± 49	21
68	\mathbf{F}	CCH	н	1.2 ± 0.7	73 ± 73	61
69	\mathbf{F}	CCMe	н	5.5 ± 4.0	180 ± 91	33
70	\mathbf{F}	CHO	н	1.1 ± 0.2	73 ± 23	67
71	\mathbf{F}	$CH=CH_2$	н	7.3 ± 7.5	527 ± 607	72
72	\mathbf{F}	ethyl	Н	2.5 ± 1.2	113 ± 82	45

 a IC₅₀ values are the means of at least two experiments \pm SD, determined from eight concentrations (performed in triplicate).

Attempts to further optimize cyano analogue **46** by various fluorine substitutions showed again that the 2'-fluoro group (i.e., **57**) resulted in an increase ER β affinity at the expense of a slight loss in ER β selectivity (Table 5). In contrast to the 3',5'-difluoro analogue **39** (Table 3), the 3',5'-difluoro substitutents pattern resulted in a dramatic loss in ER β affinity and selectivity when incorporated with the 8-cyano group (**46** vs **60**). As previously discussed,¹⁴ this loss in affinity can be explained by electrostatic repulsion between one of these fluoro groups and the side chain of Glu₃₀₅.

Selectivity Enhancements Obtained Utilizing a Genistein-Like Orientation. To attempt to exploit a second "genistein-like" binding mode (i.e., orientation a in Figure 2), the 4-position of the naphthalene scaffold was focused on. As shown in Table 6, several 4-substituted naphthalene derivatives (i.e., **61**, **62**, **65**, **68**, **70**, **71**, and **72**) were all observed to have higher affinity and selectivity for ER β than that of genistein (1). Shown in Figure 5a,b is **62** docked into the binding site of ER β , showing how the cyano group might enhance ER β selectivity by accessing the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution. The overlay with genistein in Figure 5a illustrates that this compound



b

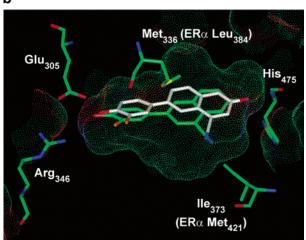


Figure 5. (a) **62** overlayed with genistein and docked to the ER β /genistein binding site and (b) **62** docked to the ER β /**47** binding site, colored by atom type. For **62**, carbon atoms were colored white for clarity. A Connolly surface of the binding site is also shown.

may be taking advantage of the genistein-like binding mode, while Figure 5b shows how the docked binding mode of **62** compares to that of **47** bound to $\text{ER}\beta$. Clearly, **47** and **62** appear to use two different orientations to enhance $\text{ER}\beta$ selectivity, by directing the common cyano functional group toward the $\text{ER}\beta$ Ile_{373} \rightarrow $\text{ER}\alpha$ Met_{421} residue substitution but at different angles.

Several compounds were prepared with substitutions at both the 1 and 4 positions (i.e. **63–67**). Interestingly, these compounds have the potential of adopting either the genistein-like or vertically flipped orientations. For example, if compounds **63–67** exploit a genistein-like orientation, the 4-cyano group would be directed toward the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution. In contrast, if **63–67** exploit a vertically flipped orientation, the 1-bromo, cyano, methyl, and chloro groups would be directed toward the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution. Since we have learned that placing a cyano group at the 1 position had no significant effect on ER β selectivity (i.e. **5** vs **18**, Table 2), this suggests that **64** may be using a genistein-like orientation to achieve its 30-fold selectivity. Since it was also observed

Table 7. Regulation of IGFBP-4 mRNA in SAOS-2 Cells

$\operatorname{compound}^a$	% activity relative to 10 nM 17 β -estradiol
15 (WAY-169122)	106
20	100
35	90
42	88
43	100
46	103
47 (WAY-202196)	120
49	143
50	100
39	100

 a Compounds were tested at 1 $\mu M,$ except for 15, which was tested at 0.2 $\mu M.$

 Table 8. Uterine Weights of Immature Rats after 3 days of Treatment

compound	$\begin{array}{c} mean \; uterine \\ wt \; (mg) \pm SEM \end{array}$
$ \begin{array}{l} \mbox{vehicle (50\% DMSO/50\% 1\times Dulbecco's PBS)} \\ 17\alpha\mbox{-ethinyl-}17\beta\mbox{-estradiol} (0.06\ \mu\mbox{g/rat/day}) \\ \mbox{47} (2\ \mbox{mg/rat/day}) \\ \mbox{47} + 17\alpha\mbox{-ethinyl-}17\beta\mbox{-estradiol} \end{array} $	$\begin{array}{c} 21.4 \pm 1.6 \\ 85.5 \pm 3.1^a \\ 23.3 \pm 1.3 \\ 81.9 \pm 4.2^b \end{array}$

^a Significantly greater than vehicle, p < 0.05. ^b Not significantly different from 17α -ethinyl- 17β -estradiol alone, p > 0.05.

that attaching a methyl group to the 1 position (i.e. **20**) had no effect of affinity or selectivity, this suggests that **65** may also be achieving its 68-fold ER β selectivity by exploiting a genistein-like orientation. The methyl group of **65** may be introducing some steric bulk, thus redirecting the 4-cyano group more optimally toward the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution, leading to an improvement in ER β selectivity with respect to **62**. No improvement in ER β selectivity was observed when the 3'-fluoro group was attached to the phenolic moiety (i.e., **61** vs **62**, and **66** vs **67**), which suggests the enhanced ER β selectivity from a 3'-fluoro group is only manifested via the vertically flipped binding orientation.

Biological Evaluation

Cell-Based Transcriptional Assay. To assess whether compounds were ER β agonists, a cell-based transcriptional assay was conducted. Insulin-like growth factor binding protein-4 (IGFBP4) is upregulated by 17 β -estradiol in human osteosarcoma cells expressing ER β .³¹ Ten compounds were evaluated for their ability to upregulate this mRNA at a single dose (typically 1 μ M). As shown in Table 7, each of the 10 compounds tested was essentially a full agonist in this assay.

Rat Uterotrophic Assay. The uterus of the sexually immature rat is highly responsive to estrogens, and this model is commonly used as an estrogenic bioassay.^{32,33} Compounds such as 17α -ethynyl- 17β -estradiol (e.g. 0.06 $\mu g/\sim 50$ g rat, given subcutaneously) increase organ weight by about 4-fold. Because the rat uterus expresses essentially only ER α and little or no ER β , this model can be used to assess the in vivo ER selectivity of compounds. Cyano derivative 47 was evaluated in this model at high doses given subcutaneously each day for three consecutive days. Specifically, 47 was dosed at 2 $mg/\sim 50$ g rat, a > 30 000-fold excess over the reference compound. No increase in organ weight was seen (Table 8). Moreover, when given in combination, organ weights were not reduced, indicating no antagonistic effect. The ER α -selective ligand propylpyrazoletriol (PPT) is fully

uterotrophic,³¹ suggesting ER α activation is necessary and sufficient for a full uterine response to nonreceptor subtype selective estrogens. **47** was also tested in an analogous model using sexually immature mice. Doses as high as 50 mg/kg were tested subcutaneously for 4 days. Consistent with the data from the rat model, organ weight did not increase (data not shown). Finally, **47** was also nonuterotrophic following 28 days of oral dosing, and this regimen achieved blood exposure substantially higher than those needed for efficacy in the Lewis rat adjuvant-induced model discussed below (data not shown).

HLA-B27 Transgenic Rat Model. The HLA-B27 transgenic rat expresses two human proteins (HLA-B27 and $\beta 2$ microglobulin) that, over time, provoke a misdirected immune response.³⁴ One of the first phenotypes these rats manifest is chronic intestinal inflammation, and thus they provide a model of inflammatory bowel disease. A later phenotype to appear is joint inflammation, and they are also used as a model of arthritis. Daily oral administration of 47 rapidly converted the chronic diarrhea these rats experienced to a normal stool. Efficacy was seen as low as 0.1 mg/kg, dosed daily, but the response was slower at this dose compared to higher doses. A composite graph of these data is shown in Figure 6. Upon euthanasia, the colons were examined histologically for quantification of four disease parameters, which were then summed to calculate a total disease score. As shown in Table 9, all doses of 47 significantly reduced total disease score. However, as seen with the stool scores, the 0.1 mg/kg dose was slightly less effective than the higher doses. As mentioned above, the joints of the HLA-B27 transgenic rat become inflamed as they age. One of the studies looking at intestinal endpoints was extended to cover the period of time when swelling occurred in the vehicle-treated rats. Thus, this was a preventative study. Joint swelling was evaluated by visual inspection each day, with a possible maximum score of 12. As shown in Figure 7, rats treated with 47 (10 mg/kg, po) had less swelling than those treated with vehicle. The difference in the two curves is significantly different (p < 0.05).

Lewis Rat Adjuvant-Induced Arthritis Model. Because positive joint data were seen in the HLA-B27 transgenic rat, 47 was evaluated in a second model of arthritis. When immunized with Complete Freund's Adjuvant, the joints of Lewis rats swell dramatically over a period of 8 days. After maximal swelling occurred, rats received an oral daily dose of 47. Thus, this was a full treatment regimen. Joint swelling was evaluated every day, and upon euthanasia, histological analysis was performed. As shown in the Figure 8, a composite graph of two studies, joint swelling was rapidly and dramatically reduced in rats treated with 47. The first study evaluated doses of 10, 5, and 1 mg/kg and the second study tested doses of 1, 0.3, and 0.1 mg/kg. The 1 mg/kg dose was tested in both studies to illustrate the consistency of the data. Efficacy on this endpoint is seen with the lowest dose tested, 0.1 mg/kg. Serum from some of these rats was analyzed for haptoglobin, an acute phase protein that is elevated in many inflammatory conditions. 47, at doses of 1 and 0.3 mg/kg, reduced haptoglobin levels by >70% (Table 10). Joints of the rats from this study were assessed for histological evidence

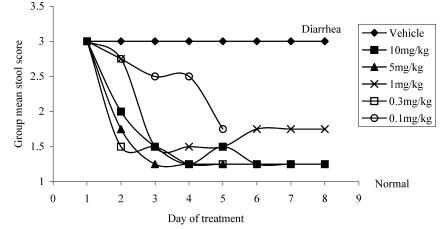


Figure 6. Effect of **47** on stool character of HLA-B27 transgenic rats. Fully diseased male rats were treated with oral daily doses of compound and stool character assessed daily according to following scale: diarrhea = 3; soft stool = 2; normal = 1.

Table 9. Effect of 47 on Colon Histology of HLA-B27 Transgenic Rats

group	ulceration (0-2)	inflammation (0-3)	$\begin{array}{c} \text{lesion depth} \\ (0-3) \end{array}$	fibrosis (0-2)	total score $(0-10)$
vehicle	1.81 ± 0.24	2.75 ± 0.20	2.38 ± 0.32	1.69 ± 0.32	8.63 ± 0.85
47					
10 mg/kg	0.25 ± 0.35^a	1.19 ± 0.52^a	0.20 ± 0.28^a	0.06 ± 0.13^a	1.75 ± 1.10^{a}
5 mg/kg	0.75 ± 0.41^a	1.13 ± 0.72^a	0.50 ± 0.12^a	0.31 ± 0.32^a	2.69 ± 1.52^a
1 mg/kg	0.31 ± 0.38^a	1.00 ± 0.29^a	0.19 ± 0.24^a	0.13 ± 0.14^a	1.63 ± 1.00^a
vehicle	1.81 ± 0.24	2.81 ± 0.24	1.81 ± 0.24	1.81 ± 0.24	8.25 ± 0.74
47					
0.3 mg/kg	1.00 ± 0.50	0.83 ± 0.29^a	0.58 ± 0.14^a	0.17 ± 0.29^a	2.58 ± 0.52^a
0.1 mg/kg	1.65 ± 0.55	1.85 ± 0.60^a	0.95 ± 0.11^a	0.45 ± 0.45^a	4.90 ± 1.63^a

^{*a*} Significantly less than vehicle (p < 0.05).

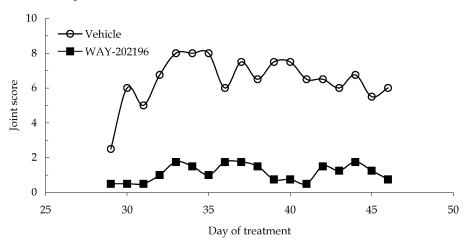


Figure 7. Effect of **47** on preventing arthropathy in HLA-B27 transgenic rats. Eight to 10 week old rats were treated with oral daily doses of 10 mg/kg, and joint redness and swelling were assessed on days 28–46 of treatment.

of disease (synovitis and Mankin scores). Each category of these disease indices was significantly reduced for the 10, 5, 1, and 0.3 mg/kg dose of **47**. At 0.1 mg/kg, although total synovitis scores were reduced, there was no difference in the Mankin scores (Table 11).

Receptor Selectivity Assays. To assess the receptor selectivity of **47**, several assays were conducted. **47** was tested in three cell-based assays to evaluate its ability to direct transcription of genes via the progesterone (PR), glucocorticoid receptors (GR), or androgen receptors (AR). First, progestins upregulate alkaline phosphatase activity in T47D cells, and this in vitro assay has been used to characterize new progestins.³⁵ When tested alone, **47** (8 nM to 5 μ M) did not upregulate alkaline phosphatase activity, indicating that the com-

pound does not have PR agonist activity in this assay. When tested in combination with 1 nM progesterone, a decrease in alkaline phosphatase activity was seen at 5μ M, and the ED₅₀ was estimated to be between 1 and 5μ M. Second, A549 lung carcinoma cells express GR, and a hormone response element luciferase reporter system was used to test for cross-reactivity on this receptor. **47** (0.3 nM to 10 μ M) had little to no effect ($\leq 5\%$ of the positive control) on reporter gene activity when tested alone. In combination with 10 nM dexamethasone, no inhibitory effect was seen. Third, L929 cells express AR, and a hormone response element luciferase reporter system was used to test for crossreactivity with this receptor. When tested alone, **47** (0.1 nM to 10 μ M) did not upregulate reporter gene activity,

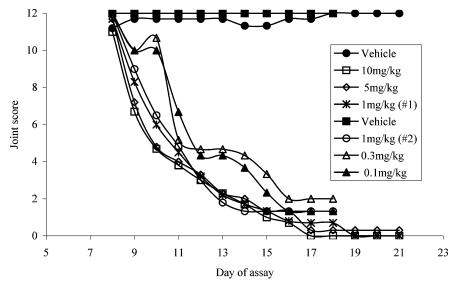


Figure 8. Effect of 47 on gross joint scores in the Lewis rat adjuvant-induced arthritis model. Data are a composite of two studies, both of which included a 1 mg/kg dose. Treatment began 8 days after adjuvant injection, when maximal redness and swelling developed in all rats.

 Table 10. Effect of 47 on Colon Histology of HLA-B27

 Transgenic Rats

	mean [haptoglobin] (mg/mL) ± SD
vehicle	1.55 ± 0.74
47 (1 mg/kg)	0.45 ± 0.16^a
47 (0.3 mg/kg)	0.35 ± 0.09^a

^{*a*} Significantly less than vehicle; p < 0.05.

indicating that the compound does not have AR agonist activity in this assay. When tested in combination with 3 nM 5 α -dihydrotestosterone, reporter gene activity was decreased by 42% at 10 μ M, thus the IC₅₀ of this compound in this assay is predicted to be $\geq 10 \ \mu$ M. Finally, **47** was tested for binding to human sex hormone binding globulin (SHBG) in a competition binding assay using [³H]-5 α -dihydrotestosterone (DHT) as the tracer. **47** (10 nM to 10 μ M) did compete for binding at concentrations above 1 μ M, and the IC₅₀ was estimated to be $\sim 10 \ \mu$ M. For comparison, radioinert DHT had an IC₅₀ value of 1.3 nM.

Conclusion

Our goal was to design a selective ligand for $ER\beta$ because we expected that such a compound would exhibit a radically different pharmacologic profile from nonselective estrogens, such as 17β -estradiol. We have shown that, though the possibility of multiple binding modes may present a greater challenge toward the rational design of ER β selective ligands, introducing the proper substituent at the appropriate position of the phenylnaphthalene scaffold can allow a single orientation to predominate, while at the same time allowing us to exploit the ER β Ile₃₇₃ \rightarrow ER α Met₄₂₁ residue substitution to afford $ER\beta$ selective compounds. Within the phenylnaphthalene scaffold, we learned that both hydroxyl groups were optimized when placed at either the 1 or 2 positions of the naphthalene framework and 4' position of the phenyl ring (Table 1). ER β selectivity could be achieved by exploiting two different orientations using the appropriate substituents attached to the 1, 4, or 8 position of the 6-(4'-hydroxyphenyl)naphthalen-2-ol template (5). An *o*-fluoro substituent slightly improved ER β selectivity when the phenylnaphthalene scaffold exploited orientation b, an effect which was not observed for orientation a. One of the most selective compounds (i.e., **47**) showed no evidence of activity via ER α in vivo and had little or no effect on GR, PR, or AR activity in vitro. However, **47** was observed to be active in two inflammation models (i.e., HLA-B27 transgenic rat and Lewis rat adjuvant-induced arthritis models), which suggests it may have utility in treating chronic inflammatory diseases such as inflammatory bowel disease or rheumatoid arthritis. Efforts to further improve ER β selectivity by exploiting multiple binding modes as well as evolving from the phenylnaphthalene motif to other new scaffolds will be reported shortly.

Experimental Section

Melting points were measured on a Mel-Temp II (Laboratory Device Inc.) melting point apparatus and are uncorrected. ¹H NMR spectra were recorded on a Bruker DPX300, Varian INOVA 400, or Varian INOVA 500 instrument. Chemical shifts are reported in δ values (parts per million, ppm) relative to an internal standard of tetramethylsilane in CDCl₃, acetone d_6 , or DMSO- d_6 . Electrospray (ESI) mass spectra were recorded using a Hewlett-Packard 5989B MS engine or Waters Alliance-ZMD mass spectrometer. Electron impact ionization (EI, EE = 70 eV) mass spectra were recorded on a Finnigan Trace mass spectrometer. Elemental analyses were carried out on a modified Perkin-Elmer model 2400 series II CHN analyzer or sent to Robertson Microlit. Analytical thin-layer chromatography (TLC) was carried out on precoated plates (silica gel, 60 F-254), and spots were visualized with UV light and stained in iodine. Preparative HPLC purifications were performed on a preparative Gilson HPLC system using a CombiPrep Pro C18 column with acetonitrile (0.1% TFA) and water (0.1% TFA) as solvents at a flow rate of 20 mL/min.

6-Trifluoromethanesulfonic Acid 5-Oxo-5,6,7,8-tetrahydronaphthalen-2-yl Ester (74). A solution of commercially available 6-hydroxy-1-tetralone (4.8 g, 29.6 mmol, Aldrich) in anhydrous CH_2Cl_2 (220 mL) was azeotroped with xylenes to remove any residual water. The solution was then cooled to 0 °C and anhydrous pyridine (3.35 mL, 41.4 mmol) was added followed by triflic anhydride (10.0 g, 35.5 mmol). After 0.5 h at 0 °C the reaction was quenched with saturated aqueous bicarbonate and washed with water. The organic layer was passed through a silica plug and concentrated to 8.39 g (96%)

Table 11.	Effect of WAY-202196	on Histology Scores	of Lewis Rats with Adjuva	nt-Induced Arthritis
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Synovitis Scores						
group	synovial structure (0-3)	fibroplasia (0-3)	inflammatory cells $(0-3)$	pannus (0-2)	total score $(0-11)$	
vehicle	2.75(0.27)	2.67 ± 0.26	3.00 ± 0.00	2.00 ± 0.00	10.5 ± 0.45	
47						
10 mg/kg	1.50 ± 0.32^a	1.08 ± 0.67^a	1.08 ± 0.59^a	0.50 ± 0.55^a	4.00 ± 1.41^a	
5 mg/kg	2.08 ± 0.38^a	1.42 ± 0.49^a	1.33 ± 0.41^a	0.83 ± 0.75^a	5.67 ± 1.72^a	
1 mg/kg	2.25 ± 0.52	1.50 ± 0.45^a	1.33 ± 0.41^a	1.00 ± 0.89^a	6.08 ± 2.18^a	
vehicle	2.92 ± 0.2	2.58 ± 0.38	2.92 ± 0.2	2 ± 0	10.42 ± 0.59	
47						
1 mg/kg	1.25 ± 0.52^a	0.75 ± 0.61^a	0.75 ± 0.42^a	0 ± 0^a	2.75 ± 1.41^a	
0.3 mg	1.67 ± 0.41^a	1.42 ± 0.38^a	1.25 ± 0.42^a	0.5 ± 0.55^a	4.83 ± 1.47^a	
0.1 mg	2.25 ± 0.69^a	1.92 ± 0.59^a	1.67 ± 0.41^a	0.67 ± 0.82^a	6.5 ± 2.15^a	
		Mar	kin Scores			
	cartilage structure	cartilage cells	Safranin-o/Fast Green	tidemark integrity	total score	
group	(0-6)	(0-3)	staining $(0-4)$	(0-1)	(0-14)	
vehicle	2.83 ± 0.26	2.58 ± 0.38	2.83 ± 0.26	0	8.25 ± 0.42	
47						
10 mg/kg	1.83 ± 0.68^a	1.58 ± 0.49^a	1.75 ± 0.82^a	0	5.17 ± 1.81^a	
5 mg/kg	2.33 ± 0.26^a	1.58 ± 0.20^a	2.00 ± 0.45^a	0	5.92 ± 0.74^a	
1 mg/kg	2.08 ± 0.38^a	1.75 ± 0.42^a	1.83 ± 0.4^a	0	5.67 ± 0.98^a	
vehicle	3.58 ± 0.59	2.75 ± 0.42	3 ± 0	0	9.33 ± 0.88	
47						
1 mg/kg	1.08 ± 0.2^a	0.92 ± 0.49^a	1.67 ± 0.41^a	0	3.67 ± 0.88^a	
0.3 mg	2.17 ± 0.68^a	1.58 ± 0.49^a	2.25 ± 0.42^a	0	6.0 ± 1.23^a	
0.1 mg	3.31 ± 0.41	2.58 ± 0.49	2.58 ± 0.49	0	8.5 ± 1.27	

Supervitia Secret

^{*a*} Significantly less than vehicle (p < 0.05).

product as a pale yellow oil: ¹H NMR (DMSO- d_6) δ 2.07 (2H, m), 2.65 (2H, t, J = 6.5 Hz), 3.03 (2H, t, J = 6.0 Hz), 7.47 (1H, dd, J = 8.7 Hz, 2.3 Hz), 7.57 (1H, d, J = 1.9 Hz), 8.03 (1H, d, J = 8.7 Hz).

6-(3-Hydroxyphenyl)-3,4-dihydronaphthalen-1(2H)one (75). Method A. A solution of 74 (1.01 g, 3.43 mmol), 3-hydroxyphenyl boronic acid (0.56 g, 4.1 mmol), sodium carbonate (7 mL of 2 N aqueous, 14 mmol), and tetrakis-(triphenylphosphine)palladium (0.20 g, 0.17 mmol) in DME $(20\ mL)$ were stirred at reflux overnight. The reaction mixture was cooled to room temperature, poured into 100 mL of 1 N NH₄Cl, and extracted with ethyl acetate (2 \times 100 mL). The combined organic layers were washed with water, washed with brine, dried over sodium sulfate, concentrated, and purified on silica to yield 0.68 g (83%) of a white solid: mp 156-160 °C; ¹H NMR (DMSO- d_6) δ 1.99–2.10 (2H, m), 2.62 (2H, t, J =5.95 Hz), 3.01 (2H, t, J = 6.08 Hz), 6.81–6.89 (1H, m), 7.07– 7.08 (1H, m), 7.12-7.15 (1H, m), 7.56-7.58 (2H, m), 7.92 (1H, d, J = 8.15 Hz), 9.60 (1H, bs); MS (ESI) m/z 239; MS (ESI) m/z 237.

6-(4-Hydroxyphenyl)-1-tetralone (76). A mixture of **74** (5.0 g, 17.0 mmol), 4-[[*tert*-butyl(dimethyl)silyl]oxy]phenylboronic acid^{7,8} (5.45 g, 20.4 mmol), sodium carbonate (4.56 g as 2 N aqueous, 42.5 mmol), and tetrakis(triphenylphosphine)palladium (0.98 g, 0.85 mmol) in DME (200 mL) was reacted according to method A to yield 3.68 g (92%) of a tan solid: mp 208–210 °C; ¹H NMR (DMSO-*d*₆) δ 2.06 (2H, m), 2.60 (2H, t, J = 6.5 Hz), 2.99 (2H, t, J = 5.9 Hz), 6.87 (2H, d, J = 8.6 Hz), 7.57 (4H, m), 7.86 (1H, d, J = 8.7 Hz), 9.74 (1H, s); MS (ESI) *m/z* 237 (M + H⁺)⁺. Anal. (C₁₆H₁₄O₂•0.25H₂O) C, H.

6-(3-Hydroxyphenyl)-1-naphthol (6). A mixture of **75** (0.51 g) and 10% Pd/C (0.61 g) in *p*-cymene (50 mL) was heated to reflux overnight. The reaction was cooled to room temperature and filtered through Celite. The Celite was rinsed with ethyl acetate, and the combined organics were removed under vacuum and purified on silica to yield 0.38 g (75%) of a white solid: mp 137–140 °C; ¹H NMR (DMSO-*d*₆) δ 6.79–6.82 (1H, m), 6.87 (1H, dd, *J* = 7.44 Hz, *J* = 0.97 Hz), 7.15–7.16 (1H, m), 7.19–7.22 (1H, m), 7.28–7.34 (2H, m), 7.42 (1H, d, *J* = 8.28 Hz), 7.69 (1H, dd, *J* = 8.79 Hz, *J* = 1.94 Hz), 8.04 (1H, d, *J* = 1.81 Hz), 8.19 (1H, d, *J* = 8.79 Hz), 9.55 (1H, bs), 10.11 (1H, bs); MS (ESI) *m*/*z* 237 (M + H⁺)⁺. Anal. (C₁₆H₁₂O₂•0.3H₂O) C, H.

6-(4-Hydroxyphenyl)-1-naphthol. (7) A mixture of **76** (500 mg, 2.12 mmol), 10% palladium on carbon (510 mg), and *p*-cymene (15 mL) was reacted according to the procedure used to prepare **6** to afford 260 mg (53%) of product as a brown solid: mp above 200 °C (dec); ¹H NMR (DMSO- d_6) δ 6.82 (1H, d, J = 7.5 Hz), 6.87 (2H, d, J = 8.1 Hz), 7.29 (1H, t, J = 7.5 Hz), 7.38 (1H, d, J = 7.5 Hz), 7.63 (2H, d, J = 8.2 Hz), 7.70 (1H, d, J = 8.5 Hz), 7.99 (1H, s) 8.14 (1H, d, J = 8.5 Hz), 9.59 (1H, s), 10.09 (1H, s); MS (ESI) m/z 235 (M - H⁺)⁻. Anal. (C₁₆H₁₂O₂) C, H.

2-Methoxy-6-(4-methoxyphenyl)naphthalene (79). Method B. To a mixture of 2-bromo-6-methoxynaphthalene (23.79 g, 100.3 mmol, Aldrich) and tetrakis(triphenylphosphine)palladium (5.8 g, 5 mmol) was added a solution of 4-methoxyphenylmagnesium bromide in THF (400 mL of 0.5 N solution, 200 mmol). The solution was heated to reflux for 3 h, cooled to room temperature, and poured into 200 mL of 1 $\,$ N HCl. The mixture was extracted with dichloromethane and filtered through a short silica plug with dichloromethane. The solvent was removed to yield a crude yellow solid that was further purified by silica chromatography (20-50% ethyl acetate-hexanes) to yield 25.7 g (97%) of the title compound as a white solid: mp 190 °C; ¹H NMR (CDCl₃): δ 3.86 (3H, s), 3.93 (3H, s), 7.01 (2H, d, J = 8.57 Hz), 7.14-7.18 (2H, m), 7.63 (2H, d, J = 8.50 Hz), 7.67 (1H, dd, J = 1.68 Hz, J = 8.73Hz), 7.76-7.80 (2H, m), 7.91 (1H, d, J = 0.94 Hz); MS (ESI) m/z 265 (M + H)⁺. Anal. (C₁₈H₁₆O₂) C, H.

7-Methoxy-2-naphthyl Trifluoromethanesulfonate. Treatment of commercially available 7-methoxy-2-naphthol (4.75 g, 27.27 mmol) with trifluoromethanesulfonic anhydride (10.0 g, 35 mmol) according to the procedure used to prepare **74** afforded 8.08 g (97%) of 7-methoxy-2-naphthyl trifluoromethanesulfonate as a clear colorless oil: ¹H NMR (DMSO- d_6) δ 3.92 (3H, s), 7.30 (1H, dd, J = 2.58 Hz, J = 8.93 Hz), 7.42 (1H, dd, J = 2.59 Hz, J = 8.93 Hz), 7.52 (1H, d, J = 2.38 Hz), 7.96 (1H, d, J = 9.12 Hz), 8.00 (1H, d, J = 2.38 Hz), 8.06 (1H, d, J = 8.73 Hz); MS (EI) *m/z* 306 (M•)⁺. Anal. (C₁₂H₉F₃O₄S) C, H.

2-Methoxy-7-(4-methoxyphenyl)naphthalene (81). The title compound was prepared by reacting 7-methoxy-2-naph-thyl trifluoromethanesulfonate (3.15 g, 10.3 mmol), prepared above, with 4-methoxyphenylboronic acid (2.2 g, 14 mmol) according to method A to afford 2.17 g (79%) of **81** as a white

solid: mp 154 °C; ¹H NMR (CDCl₃) δ 3.87 (3H, s), 3.94 (3H, s), 7.02 (2H, d, J = 8.72 Hz), 7.13 (1H, dd, J = 2.54 Hz, J = 9.09 Hz), 7.18 (1H, d, J = 2.55 Hz), 7.56 (1H, dd, J = 1.82 Hz, J = 8.36 Hz), 7.65 (2H, d, J = 8.72 Hz), 7.74 (1H, d, J = 9.09 Hz), 7.81 (1H, d, J = 8.36 Hz), 7.89 (1H, d, J = 1.09 Hz); MS (EI) m/z 264 (M·). Anal. (C18H16O2) C, H.

2-Methoxy-7-phenylnaphthalene (83). The title compound was prepared by reacting 7-methoxy-2-naphthyl trifluoromethanesulfonate (3.01 g, 9.83 mmol) with phenylboronic acid (1.4 g, 12 mmol) according to method A to yield 1.95 g (85%) of a white solid: mp 62–64 °C; ¹H NMR (CDCl₃) δ 3.95 (3H, s), 7.15 (1H, dd, J = 2.56 Hz, J = 8.79 Hz), 7.20 (1H, d, J = 2.56 Hz), 7.36–7.39 (1H, m), 7.46–7.50 (2H, m), 7.60 (1H, dd, J = 1.83 Hz, J = 8.42 Hz), 7.70–7.73 (2H, m), 7.76 (1H, d, J = 8.79 Hz), 7.84 (1H, d, J = 8.42 Hz), 7.94 (1H, d, J = 1.46 Hz); MS (EI) *m/z* 234 (M•)⁺. Anal. (C₁₇H₁₄O·0.1H₂O) C, H.

6-(2-Hydroxyphenyl)-2-naphthol (8). Method C. To a solution of 77 (0.38 g, 1.52 mmol) in dichloromethane (20 mL) at 0 °C was slowly added BBr₃ (4.5 mL of 1 N, 4.5 mmol). The solution was allowed to warm to room temperature overnight with stirring and was poured into 100 mL of water. The mixture was extracted with 100 mL of ethyl acetate and the organic layer was washed with water and with brine, dried over sodium sulfate, filtered, and concentrated, and the product was purified on silica (25% ethyl acetate/hexanes) to yield 0.29 g (81%) of an off-white solid: mp 179–180 °C; ^{1}H NMR (DMSO- d_6) δ 6.87–6.91 (1H, m), 6.96 (1H, dd, J = 8.14Hz, J = 1.04 Hz), 7.08 (1H, dd, J = 8.72 Hz, J = 2.40 Hz), 7.12–7.18 (2H, m), 7.33 (1H, dd, J = 7.63 Hz, J = 1.68 Hz), 7.61 (1H, dd, J = 8.48 Hz, J = 1.74 Hz), 7.68 (1H, d, J = 8.80Hz), 7.78 (1H, d, J = 8.92 Hz), 7.90 (1H, D, J = 0.91 Hz), 9.48 (1H, bs), 9.70 (1H, bs); MS (ESI) m/z 235 (M - H)⁻. Anal. $(C_{16}H_{12}O_2{\boldsymbol{\cdot}}0.1H_2O)\ C,\ H,\ N.$

6-(4-Hydroxyphenyl)-2-naphthol (5). Method D. Compound 79 (5.61 g, 21.2 mmol) was added to pyridinium HCl (30 g) at 190 °C for 2 h, and the mixture cooled to room temperature and was stirred with 200 mL of 1 N HCl. The resulting suspension was filtered and dissolved ethyl acetate (500 mL). The combined organic layers were washed with water (200 mL), dried over anhydrous magnesium sulfate, and filtered; the solvent was removed under vacuum; and the product was purified by silica chromatography (40% ethyl acetate-hexanes) to yield 4.88 g (98%) of a white solid: mp >220 °C(discolors above 200 °C); ¹H NMR (DMSO- d_6) δ 6.87 (2H, d, J = 8.54 Hz), 7.08 (1H, dd, J = 2.34 Hz, J = 8.76 Hz),7.11 (1H, d, J = 2.14 Hz), 7.58 (2H, d, J = 8.54 Hz), 7.65 (1H, dd, J = 1.92 Hz, J = 8.76 Hz), 7.71 (1H, d, J = 8.54 Hz), 7.79 (1H, d, *J* = 8.97 Hz), 7.95 (1H, s), 9.56 (1H, bs), 9.70 (1H, bs); MS (ESI) m/z 235 (M – H)⁻. Anal. (C₁₆H₁₂O₂) C, H.

1-Chloro-2-methoxy-6-(4-methoxyphenyl)naphthalene (85). Method E. A suspension of **79** (0.51 g, 1.93 mmol) and NCS (0.28 g, 2.13 mmol) in acetonitrile (20 mL) was heated to reflux for 3 h. The resulting solution was coolected by filtration and rinsed with acetonitrile to yield 0.41 g (72%) of a white solid: mp 158–164 °C; ¹H NMR (CDCl₃) δ 3.87 (3H, s), 4.05 (3H, s), 7.03 (2H, d, J = 8.62 Hz), 7.32 (1H, d, J =9.02 Hz), 7.65 (2H, d, J = 8.55 Hz), 7.82 (2H, d, J = 9.01 Hz), 7.95 (1H, d, J = 1.36 Hz), 8.27 (1H, d, J = 8.81 Hz); MS (EI) m/z 298 (M·)⁺. Anal. (C₁₈H₁₅ClO₂) C, H.

1-Bromo-2-methoxy-6-(4-methoxyphenyl)naphthalene (86). To a mixture of **79** (9.68 g, 36.6 mmol) and glacial acetic acid (150 mL) was slowly added a solution of bromine (5.85 g, 36.6 mmol) in glacial acetic acid (20 mL). The mixture was stirred for 1 h and the resulting suspension was poured into water (200 mL) and the solid product was collected by filtration. The solid was triturated with water and then with ethyl acetate to yield 11.25 g (90%) of a white solid: mp 172– 174 °C; ¹H NMR (CDCl₃) δ 3.88 (3H, s), 4.05 (3H, s), 7.04 (2H, d, J = 8.56 Hz), 7.30 (1H, d, J = 9.01 Hz), 7.67 (2H, d, J =8.65 Hz), 7.81 (1H, dd, J = 1.61 Hz, J = 8.84 Hz), 7.87 (1H, d, J = 9.04 Hz), 7.94 (1H, d, J = 1.47 Hz), 8.27 (1H, d, J = 8.92Hz); MS (EI) *m/z* 343 (M[•])⁺. Anal. (C₁₈H₁₅BrO₂) C, H.

2-Methoxy-6-(4-methoxyphenyl)-1-fluoronaphthalene (87). To a solution of 86 (1.28 g, 3.72 mmol) in THF (25 mL) at -78 °C was slowly added *n*-butyllithium (1.9 mL of 2.5 N in hexanes). The resulting solution was stirred at -78°C for 0.5 h and a solution of N-fluorobenzenesulfonimide (1.40 g, 4.5 mmol) in THF (10 mL) was added. After an additional 2 h at -78 °C, the reaction was warmed to room temperature, poured into water, and extracted with ethyl acetate (2×250) mL). The combined organic layers were washed with water, dried over sodium sulfate, filtered, and concentrated, and the product was purified by silica chromatography (5% THFhexanes) to yield 0.66 g (63%) of a white solid: mp 150–155 °C; ¹H NMR (CDCl₃) δ 3.88 (3H, s), 4.04 (3H, s), 7.02 (2H, d, J = 8.69 Hz), 7.28–7.34 (1H, m), 7.62–7.67 (3H, m), 7.74 (1H, dd, J = 1.60 Hz, J = 8.79 Hz), 7.93 (1H, s), 8.09 (1H, d, J =8.77 Hz). Anal. (C₁₈H₁₅O₂F) C, H.

2-Methoxy-6-(4-methoxyphenyl)-1-naphthonitrile (88). A mixture of **86** (0.76 g, 2.21 g), CuCN (0.24 g, 2.66 mmol), and DMF (10 mL) was stirred at 120 °C for 4 h. The reaction mixture was cooled to room temperature, triturated with ethyl acetate, filtered through silica, and concentrated, and the product was purified by silica column chromatography (20% ethyl acetate—hexanes) to yield 0.19 g (30%) of a pale yellow solid: mp 182–185 °C; ¹H NMR (CDCl₃) δ 3.88 (3H, s), 4.09 (3H, s), 7.03 (2H, d, J = 8.73 Hz), 7.29 (1H, d, J = 9.12 Hz), 7.63 (2H, d, J = 8.73 Hz), 7.88 (1H, dd, J = 1.98 Hz, J = 8.73 Hz), 7.97 (1H, d, J = 1.98 Hz), 8.09 (1H, d, J = 9.12 Hz), 8.14 (1H, d, J = 8.73 Hz).

2-Methoxy-6-(4-methoxyphenyl)-1-phenylnaphthalene (89). The title compound was prepared by reacting **86** (0.80 g, 2.33 mmol) with phenylmagnesium bromide (2.2 mL of 3 N in ether) according to method B to yield 0.42 g (53%) of a gray solid: mp 157–160 °C; ¹H NMR (CDCl₃) δ 3.85 (3H, s), 3.87 (3H, s), 7.01 (2H, d, J = 8.64 Hz), 7.38–7.46 (4H, m), 7.49–7.57 (4H, m), 7.64 (2H, d, J = 8.64 Hz), 7.92 (1H, d, J = 9.05 Hz), 7.97 (1H, s); MS (ESI) *m/z* 341 (M + H)⁺. Anal. (C₂₄H₂₀O₂•0.5H₂O) C, H.

2-Methoxy-6-(4-methoxyphenyl)-1-methylnaphthalene (90). To a solution of 86 in THF (25 mL) at 0 °C were slowly added *n*-butyllithium (2 mL of 2.5N) and TMEDA (0.60 g, 5.13 mmol, freshly distilled from KOH). After 0.5 h at 0 °C, iodomethane (7.3 g, 51.3 mmol, passed through basic alumina) was added and the solution was allowed to warm to room temperature overnight. The reaction was poured into water (100 mL) and extracted with ethyl acetate (3 \times 200 mL). The combined organic layers were washed with water, dried over magnesium sulfate, filtered, and concentrated, and the product purified by silica column chromatography (5% THF-hexanes) to yield 0.63 g (88%) of a white solid: mp 136–138 °C; ¹H NMR $(CDCl_3) \delta 3.87 (3H, s), 3.96 (3H, s), 7.02 (2H, d, J = 8.74 Hz),$ 7.29 (1H, d, J = 9.01 Hz), 7.60–7.80 (4H, m), 7.95 (1H, d, J =1.76 Hz), 8.00 (1H, d, J = 8.86 Hz); MS (ESI) m/z 279 (M + $H)^+$. Anal. (C₁₉ $H_{18}O_2 \cdot 0.3H_2O)$ C, H.

1-Chloro-6-(4-hydroxyphenyl)-2-naphthol (15). The title compound was prepared by reacting **85** (0.32 g, 1.07 mmol) with pyridinium HCl (7 g) at 190 °C according to method D to yield 0.12 g (41%) of an off-white solid: mp 222–224 °C (dec); ¹H NMR (DMSO- d_6) δ 6.88 (2H, d, J = 8.46 Hz), 7.29 (1H, d, J = 8.88 Hz), 7.63 (2H, d, J = 8.50 Hz), 7.81–7.88 (2H, m), 8.02–8.08 (2H, m), 9.59 (1H, s), 10.43(1H, s); MS (ESI) *m/z* 269/271 (M – H)⁻. Anal. (C₁₆H₁₁ClO₂) C, H.

1-Bromo-6-(4-hydroxyphenyl)-2-naphthol (16). To a solution of **5** (4.02 g, 17.0 mmol) in acetonitrile (150 mL) at 0 °C was added NBS (3.03 g, 17.0 mmol). The reaction was stirred at 0 °C for 3 h and then poured into water (200 mL) and extracted with ethyl acetate (3×300 mL). The combined organic layers were washed with water, dried over sodium sulfate, filtered, and concentrated, and the product was purified by silica column chromatography (20% ethyl acetate–hexanes) to yield 5.33 g (100%) of a tan solid. Further purification by reverse phase HPLC yielded a white solid: mp 208–210 °C; ¹H NMR (DMSO- d_6) δ 6.89 (2H, d, J = 8.47 Hz), 7.28 (1H, d, J = 8.80 Hz), 7.63 (2H, d, J = 8.50 Hz), 7.85

(2H, d, $J=8.84~{\rm Hz}),\, 8.02-8.06$ (2H, m), 9.60 (1H, s), 10.54 (1H, s); MS (ESI) m/z 313/315 (M - H)^-. Anal. (C_{16}H_{11}BrO_2) C, H.

1-Fluoro-6-(4-hydroxyphenyl)-2-naphthol (17). The title compound was prepared by reacting **87** (0.250 g, 0.886 mmol) with boron tribromide (2.7 mL of 1 N solution, 2.7 mmol) according to method C to yield 0.13 g (15%) of a white solid: mp 219–224 °C; ¹H NMR (DMSO-*d*₆) δ 6.89 (2H, d, *J* = 8.49 Hz), 7.22–7.28 (1H, m), 7.61 (2H, d, *J* = 8.53 Hz), 7.65 (1H, d, *J* = 9.12 Hz), 7.79 (1H, d, *J* = 8.77 Hz), 7.92 (1H, d, *J* = 8.73 Hz), 8.05 (1H, s), 9.63 (1H, bs), 10.00 (1H, bs); MS (ESI) *m/z* 253 (M - H)⁻. Anal. (C₁₆H₁₁FO₂) C, H: calcd, 75.58; found, 75.13.

2-Hydroxy-6-(4-hydroxyphenyl)-1-naphthonitrile (18). The title compound was prepared by reacting **88** (0.115 g, 0.397 mmol) with pyridinium HCl (4 g) at 190 °C according to method D to yield 0.025 g (24%) of a tan solid: mp >220 °C; ¹H NMR (DMSO-*d*₆) δ 6.89 (2H, d, J = 8.37 Hz), 7.28 (1H, d, J = 9.07 Hz), 7.63 (2H, d, J = 8.42 Hz), 7.88–7.98 (2H, m), 8.12–8.16 (2H, m), 9.63 (1H, s), 11.65 (1H, bs); MS (ESI) *m/z* 260 (M – H)⁻. Anal. (C₁₇H₁₁NO₂·0.4H₂O) C, H, N.

6-(4-Hydroxyphenyl)-1-phenyl-2-naphthol (19). The title compound was prepared by reacting **89** (0.36 g, 1.06 mmol) with boron tribromide (3.2 mL of 1 N in CH₂Cl₂) according to method C to yield 0.14 g (42%) of a white solid: mp 142–146 °C; ¹H NMR (DMSO-*d*₆) δ 6.86 (2H, d, J = 8.50 Hz), 7.26–7.42 (5H, m), 7.48–7.61 (5H, m), 7.84 (1H, d, J = 8.96 Hz), 8.02 (1H, d, J = 1.46 Hz), 9.52 (2H, s); MS (ESI) *m/z* 311 (M – H)⁻. Anal. (C₂₂H₁₆O₂·0.1H₂O) C. H.

6-(4-Hydroxyphenyl)-1-methyl-2-naphthol (20). The title compound was prepared by reacting **90** (0.35 g, 1.26 mmol) with boron tribromide (3.8 mL of 1 N solution, 3.8 mmol) according to method C to yield 0.15 g (48%) of a white solid: mp >170 °C (dec); ¹H NMR (DMSO- d_6) δ 2.42 (3H, s), 6.87 (2H,k d, J = 8.33 Hz), 7.15 (1H, d, J = 8.81 Hz), 7.59 (2H, d, J = 8.35 Hz), 7.65 (1H, d, J = 8.93 Hz), 7.71 (1H, dd, J = 1.24 Hz, J = 8.90 Hz), 7.88 (1H, d, J = 8.87 Hz), 7.95 (1H, s), 9.49 (1H, s), 9.52 (1H, s); MS (ESI) 249 *m/z* (M - H)⁻. Anal. (C₁₇H₁₄O₂) C, H.

6-(4-Hydroxyphenyl)-1-methoxy-2-naphthol (21).Method F. A mixture of 16 (0.41 g, 1.30 mmol), CuBr (0.19 g, 0.13 mmol), sodium methoxide (3 mL of 4.4 N in methanol), and DMF (6 mL) was heated to reflux for 3 h. The reaction mixture was cooled to room temperature, poured into HCl (50 mL of 1 N aqueous solution), and extracted with ethyl acetate $(3 \times 100 \text{ mL})$. The combined organic layers were washed with sodium bicarbonate solution, dried over sodium sulfate, and filtered; the solvent was removed under vacuum; and the product was purified by silica column chromatgraphy (40% ethyl acetate-hexanes) to yield 0.31 g (89%) of the title compound as a white solid: mp 204-206 °C; ¹H NMR (DMSO d_6) δ 6.88 (2H, d, J = 8.47 Hz), 7.19 (1H, d, J = 8.81 Hz), 7.57– 7.60 (3H, m), 7.71 (1H, dd, J = 1.41 Hz, J = 8.78 Hz), 7.94– 7.98 (2H, m), 9.54 (2H, s); MS (ESI) *m/z* 265 (M – H)⁻. Anal. $(C_{17}H_{14}O_3)$ C, H.

6-Bromo-1-nitro-2-naphthol (91). Commercially available 4-nitro-4-methyl-2,3,5,6-tetrabromo-2,5-cyclohexadien-1-one (2.53 g, 4.95 mmol) was added to a solution of 6-bromo-2naphthol (1 g, 4.5 mmol) in of dry ether (40 mL). The mixture was allowed to react for 1.5 h at room temperature. The solid byproduct, 2,3,5,6-tetrabromo-4-methylphenol (144 mg), was removed by filtration. The solution was then evaporated under vacuum and the crude mixture was dissolved in ethyl acetate and washed with water. The organic layer was dried over anhydrous sodium sulfate and filtered, and the solvent was removed under vacuum. An additional batch of 2,3,5,6-tetrabromo-4-methylphenol (1.2 g) was separated by recrystallization of the crude product with ethyl acetate-hexane. The mother liquid was concentrated onto Florosil and purified on a silica column (15-20% ethyl acetate-hexane) to yield 0.731 g (61%) of the desired product as a yellow solid: mp 111–113 °C; ¹H NMR (DMSO- d_6) δ 7.39 (1H, d, J = 9.07 Hz), 7.54 (1H, d, J = 9.05 Hz), 7.75 (1H, dd, J = 9.05 Hz, J = 1.70 Hz), 8.03

(1H, d, J = 9.14 Hz), 8.29 (1H, d, J = 1.84 Hz), 11.65 (1H, s); MS (ESI) m/z 266/268 (M - H)⁻; IR 1350 cm⁻¹, 1490 cm⁻¹. Anal. (C₁₀H₆BrNO₃·0.2H₂O) C, H.

6-[4-(tert-Butyldimethylsilanyloxy)phenyl]-1-nitro-2-naphthol (92). The title compound was prepared by reacting **91** (560 mg, 2.1 mmol) with 4-*tert*-butyl-dimethylsilyoxyphenylboronic acid⁸ (688 mg, 2.73 mmol) according to method A to yield 347 mg (42%) of yellowish solid: ¹H NMR (DMSO- d_6) δ 0.23 (6H, s), 0.98 (9H, s), 6.98 (2H, d, J = 8.47 Hz), 7.35 (1H, d, J = 9.05 Hz), 7.64 (1H, d, J = 8.86 Hz), 7.70 (2H, d, J = 8.51 Hz), 7.94 (1H, dd, J = 6.62 Hz, J = 1.25 Hz), 8.08 (1H, d, J = 9.16 Hz), 8.22(1H, s), 11.45 (1H, s); MS (ESI) *m/z* 394 (M - H)⁻.

6-(4-Hydroxyphenyl)-1-nitro-2-naphthol (22). Method G. To a solution of 92 (302 mg, 0.764 mmol) in THF (12 mL) was added TBAF (0.92 mL, 0.917 mmol, 1.0 M solution in THF). The solution was stirred for 10 min at room temperature and poured into water, and the mixture was extracted with ethyl acetate. The combined organic layers were washed with brine, dried with anhydrous sodium sulfate, and filtered; the solvent was removed; and the product was purified on a silica column (20-40% ethyl acetate-hexane) to yield 130 mg (61%) of a orange solid. An analytical sample was further purified by recrystallization with ethyl acetate-hexane to yield the title compound as an orange solid: mp 199-201 °C; ¹H NMR $(DMSO-d_6) \delta 6.89 (2H, d, J = 8.51 Hz), 7.33 (1H, d, J = 9.05)$ Hz), 7.62 (1H, d, *J* = 8.94 Hz), 7.63(2H, d, 8.48 Hz), 7.91 (1H, dd, J = 8.87 Hz, J = 1.59 Hz), 8.06 (1H, d, 9.18 Hz), 8.17 (1H, d, J = 1.31 Hz), 9.64(1H, s), 11.40 (1H, s); MS (ESI) m/z 280 $(M\,-\,H)^{-}.$ Anal. $(C_{16}H_{11}NO_{4})$ C, H, N.

6-Methoxy-2-naphthalenyl)boronic Acid. 2-Bromo-6methoxynaphthalene (8.02 g, 33.8 mmol, Aldrich) was reacted with *sec*-butyllithium (56 mL of 1.3 N solution in hexanes, 72.8 mmol) in THF at -78 °C, followed by the addition of triisopropyl borate (47 mL, 203.7 mmol). The mixture was then stirred with aqueous HCl (200 mL of 1 N solution) for 10 min and then extracted with ethyl acetate and concentrated, and crystallization was induced with hexanes to afford a solid which was filtered and dried to give 7.01 g (~100%) of 6-methoxy-2-naphthalenyl)boronic acid as a light orange powder: ¹H NMR (DMSO- d_6) δ 3.88 (3H, s), 7.15 (1H, dd, J = 8.9Hz, 2.7 Hz), 7.29 (1H, d, J = 2.5 Hz), 7.75 (1H, d, J = 8.3 Hz), 7.83 (2H, appt), 8.30 (1H, s); MS (EI) m/z 202.17 (M⁺).

2-(2-Fluoro-4-methoxyphenyl)-6-methoxynaphthalene (93). Reaction of trifluoromethanesulfonic acid 2-fluoro-4-methoxyphenyl ester [3.86 g, 14.0 mmol; prepared according to the procedure used for **74**, with 6-methoxy-2-naphthalenylboronic acid (2.97 g, 14.8 mmol)] according to method A yielded 2.27 g (58%) of white solid: mp 104–106 °C; ¹H NMR (DMSO d_6) δ 3.83 (3H, s), 3.89 (3H, s), 6.92 (1H, dd, J = 8.46 Hz, J = 2.64 Hz), 6.98 (1H, dd, J = 12.92, J = 2.48 Hz), 7.19 (1H, dd, J = 8.94 Hz, J = 2.46 Hz), 7.35 (1H, J = 2.38 Hz), 7.53–7.63 (2H, m), 7.90 (2H, d, J = 8.72 Hz), 7.97 (1H, s); MS (ESI) *m/z* 283 (M + H⁺). Anal. (C₁₈H₁₅FO₂) C, H.

2-(2,5-Difluoro-4-methoxyphenyl)-6-methoxynaphthalene (94). The title compound was prepared according to method A by reacting 4-bromo-2,5-difluoroanisole (4.06 g, 18.3 mmol) with 6-methoxy-2-naphthalenylboronic acid (4.81 g, 23.8 mmol, prepared above) to yield 5.18 g (94.2%) of white solid: mp 153–155 °C; ¹H NMR (DMSO- d_6) δ 3.90 (3H, s), 3.91 (3H, s), 7.20 (1H, dd, J = 8.94 Hz, J = 2.50 Hz), 7.28 (1H, dd, J =12.38 Hz, J = 7.47 Hz), 7.36 (1H, d, J = 2.42 Hz), 7.56 (1H, dd, J = 12.17 Hz, J = 7.53 Hz), 7.62–7.66 (1H, m), 7.89 (2 H, d, J = 8.67 Hz), 8.02 (1H, s); MS (ESI) *m/z* 301 (M – H)⁺; HRMS calcd for C₁₈H₁₄F₂O₂ 300.0962, found 300.0872. Anal. (C₁₈H₁₄F₂O₂) C, H, N: calcd, 71.99; found, 73.00.

3-Chloro-4-(6-methoxy-2-naphthyl)phenol (96). The title compound was prepared by reacting 4-bromo-3-chlorophenol (3.8 g, 18.3 mmol) and 6-methoxy-2-naphthalenylboronic acid (4.81 g, 23.8 mmol, prepared above) according to method A to yield 5.09 g (98%) of white solid: mp 139–142 °C; ¹H NMR (DMSO- d_6) δ 3.89 (3H,s), 6.87 (1H, dd, J = 8.39 Hz, J = 2.45 Hz), 6.98 (1H, J = 2.40 Hz), 7.19 (1H, dd, J = 8.98 Hz, J = 2.46 Hz), 7.32 (1H, d, J = 8.39 Hz), 7.35 (1H, d, J = 2.38

Hz), 7.50 (1H, dd, J=8.47 Hz, J=1.73 Hz), 7.83–7.88 (3H, m), 10.04 (1H, s); MS (ESI) $m\!/z$ 283/285 (M - H) $^-$. Anal. (C16H11CIO) C, H.

6-(4-Methoxy-2-methylphenyl)-2-naphthol (97). Reaction of 4-bromo-3-methylanisole (10 g, 0.050 mol) with *n*-butyllithium (24 mL of 2.5 M solution in hexane, 0.055 mol) followed by triisopropyl borate (57.7 mL, 47.02 g, 0.25 mol), according to the procedure described for 6-methoxy-2-naphthalenyl)boronic acid above, yielded 5.7 g (69%) of 4-methoxy-2-methylphenylboronic acid as a white solid: MS (ESI) *m/z* 313 (2M - H₂O - H)⁻.

Treatment of 6-bromo-2-naphthol (1.8 g, 5.4 mmol) with 4-methoxy-2-methylphenylboronic acid (1.74 g, 7.0 mmol) according to method A afforded 1.56 g (73%) of yellowish solid: mp 124–126 °C; ¹H NMR (DMSO- d_6) δ 2.25 (3H,s), 3.78 (3H, s), 6.85 (1H, dd, J = 8.35 Hz, J = 2.56 Hz), 6.90 (1H, d, J = 2.37 Hz), 7.09 (1H, dd, J = 8.75 Hz, J = 2.25 Hz), 7.13 (1H, s), 7.20 (1H, d, J = 8.33 Hz), 7.35 (1H, dd, J = 8.39 Hz, J = 1.37 Hz), 7.67 (1H, s), 7.70 (1H, d, J = 8.53 Hz), 7.78 (1H, d, J = 8.78 Hz), 9.74 (1H, s); MS (ESI) m/z 263 (M – H)[–]. Anal. (C₁₈H₁₆ O₂) C, H.

4-(6-Hydroxy-2-naphthyl)-3-methoxyphenyl 4-methylbenzenesulfonate (101). 4-Bromo-3-methoxyphenyl 4methylbenzenesulfonate. A mixture of 4-bromoresorcinol (4.92 g, 26.0 mmol), p-toluenesulfonic chloride (5.95 g, 31.2 mmol), potassium carbonate (23 g, 167 mmol), and acetone (300 mL) was refluxed for 16 h. Iodomethane (9.89 g, 70 mmol) was added and the mixture was refluxed for an additional 12 h. The mixture was cooled to room temperature, ether (200 mL) was added, and the suspension was filtered. The filtrate was concentrated and the product purified on a silica column (10% ethyl acetate-hexanes) to yield 6.49 g (70%) of 4-bromo-3-methoxyphenyl 4-methylbenzenesulfonate as a white solid: ¹H NMR (DMSO-d₆) δ 2.45 (3H, s), 3.78 (3H, s), 6.40 (1H, dd, J = 2.45 Hz, J = 8.60 Hz, 6.59 (1H, d, J = 2.48 Hz), 7.33 (2H, d, J = 8.30 Hz), 7.40 (1H, d, J = 8.69 Hz), 7.71 (2H, d, J =8.23 Hz); MS (ESI) m/z 355/357 (M - H)⁻. Anal. (C₁₄H₁₃BrO₄S) C, H.

tert-Butyl[(6-bromo-2-naphthyl)oxy]dimethylsilane. To a solution of 6-bromo-2-naphthol (13.68 g, 61.33 mmol) and TBDMS-Cl (11.09 g, 73.6 mmol) in DMF (50 mL) was added imidazole (10.2 g, 150 mmol). The solution was stirred for 3 h, mixed with sodium bicarbonate solution (250 mL), and extracted with 50% ethyl acetate-hexanes (3 × 250 mL). The combined organic layers were washed with water, dried over magnesium sulfate, and filtered, and the solvent was evaporated to yield an oil which was dried under vacuum to yield 19.9 g (97%) of the title compound as a white solid: ¹H NMR (CDCl₃) δ 0.24 (6H, s), 1.01 (9H, s), 7.08 (1H, dd, J = 2.26 Hz, J = 8.81 Hz), 7.14 (1H, d, J = 2.15 Hz), 7.46 (1H, dd, J = 1.80 Hz, J = 8.77 Hz), 7.54 (1H, d, J = 8.77 Hz), 7.61 (1H, d, J = 8.81 Hz), 7.90 (1H, s); MS (EI) m/z 336/338 (M•)⁺. Anal. (C₁₆H₂₁BrO₁Si·0.25H₂O) C, H, N.

tert-Butyl[(2-(6-naphthylboronic acid)oxy]dimethylsilane. To a solution of tert-butyl[(6-bromo-2-naphthyl)oxy]dimethylsilane (19.18 g, 56.9 mmol), prepared as described above, in THF (200 mL) at -78 °C was slowly added *n*butyllithium (25 mL of 2.5 N solution in hexanes). The solution was stirred for 30 min followed by the addition of triisopropyl borate (53.5 g, 285 mmol). The solution was stirred for 1 h at -78 °C and then allowed to warm to room temperature overnight. The solution was then cooled to 0 °C and stirred with HCl (200 mL of 1 N solution) for 10 min. The mixture was extracted with ethyl acetate (3 \times 250 mL). The combined organic layers were concentrated to a volume of 25 mL. Crystallization was induced with hexanes, and the solid product was collected by filtration and dried under vacuum to yield 13.5 g (79%) of an off-white solid: ¹H NMR (DMSO d_6) δ 0.25 (6H, s), 0.99 (9H, s), 7.10 (1H, dd, J = 2.56 Hz, J =8.97 Hz, 7.26 (1 H, d, J = 2.56 Hz), 7.73 (1 H, d, J = 8.54 Hz), 7.82 (1H, dd, J = 1.07 Hz, J = 8.33 Hz), 7.83 (1H, d, J = 8.97Hz), 8.30 (1H, s); MS (ESI) m/z 303 (M + H)⁺.

4-Bromo-3-methoxyphenyl 4-methylbenzenesulfonate (3.07 g, 8.60 mmol) and *tert*-butyl[(2-(6-naphthyl boronic acid)oxy]- dimethylsilane (2.86 g, 9.46 mmol) were reacted according to method A to yield 2.15 g (53%) of an orange solid: ¹H NMR (CDCl₃) δ 2.44 (3H, s), 3.65 (3H, s), 6.69 (1H, dd, J = 2.10 Hz, J = 8.29 Hz), 6.74 (1H, d, J = 2.10 Hz), 7.06–7.11 (2H, m), 7.36 (1H, d, J = 8.27 Hz), 7.45 69 (1H, dd, J = 1.32 Hz, J = 8.57 Hz), 7.51 (1H, d, J = 8.28 Hz), 7.67 (1H, d, J = 8.61 Hz), 7.77 (1H, d, J = 8.80 Hz), 7.80–7.85 (3H, m), 9.78 (1H, s); MS (ESI) *m/z* 419 (M – H)⁻. Anal. (C₂₄H₂₀O₅S·0.25H₂O) C, H.

6-(4-Hydroxy-2-methoxyphenyl)-2-naphthol (31). A solution of 101 (1.74 g, 4.05 mmol), potassium hydroxide (5 g), water (85 mL), and ethanol (85 mL) was stirred at 90 °C for 2 h. The reaction was cooled to room temperature and concentrated to 50% volume, neutralized with acetic acid, and extracted with ethyl acetate (3 \times 200 mL). The combined organic layers were washed with brine, dried over sodium sulfate, filtered, and concentrated, and the product was purified by silica column (25% ethyl acetate-hexanes) to yield 1.01 g (94%) of a tan solid. A sample was further purified by preparative reverse phase HPLC to yield a tan solid: mp 152-154 °C; ¹H NMR (DMSO- d_6) δ 3.72 (3H, s), 6.46 (1H, dd, J =1.52 Hz, J = 8.20 Hz), 6.52 (1H, d, J = 1.77 Hz), 7.04–7.09 (2H, m), 7.16 (1H, d, J = 8.23 Hz), 7.47 (1H, d, J = 8.50 Hz), 7.63 (1H, d, J = 8.57 Hz), 7.72–7.75 (2H, m), 9.56 (1H, bs), 9.68 (1H, bs); MS (ESI) m/z 265 (M - H)⁻. Anal. (C₁₇H₁₄O₃) C, H.N.

6-(2-Fluoro-4-hydroxyphenyl)-2-naphthol (23). Treatment of **93** (1.12 g, 3.97 mmol) with boron tribromide (23.8 mL of 1.0 M solution in CH₂Cl₂, 23.8 mmol) according to method C yielded 0.92 g (91%) of a white solid: mp 208–209 °C; ¹H NMR (DMSO- d_6) δ 6.66–6.75 (2H, m), 7.08–7.13 (2H, m), 7.41 (1H, dd, J = 9.37 Hz, J = 8.57 Hz), 7.49–7.53 (1H, m), 7.72 (1H, d, J = 8.64 Hz), 7.80 (1H, d, J = 8.76 Hz), 7.86 (1H, s), 9.79 (1H, s), 10.01 (1H, s); MS (ESI) *m/z* 255 (M + H)⁺, MS (ESI) *m/z* 253 (M – H)⁻. Anal. (C₁₆H₁₁FO₂·0.15H₂O) C, H, N.

6-(3,5-Difluoro-4-hydroxyphenyl)-2-naphthol (38). Treatment of 6-bromo-2-naphthol (0.165 g, 0.74 mmol) with 3,5-difluoro-4-*tert*-butyldimethylsilyoxyboronic acid (0.25 g, 0.87 mmol) according to method A yielded 0.14 g (70%) of a tan solid. This material was further purified by preparative reverse phase HPLC to yield the title compound as a white solid: mp 216–220 °C; ¹H NMR (DMSO-*d*₆) δ 7.09–7.13 (2h, m), 7.50 (2H, d, *J* = 10.00 Hz), 7.73 (2H, s), 7.78 (1H, d, *J* = 8.58 Hz), 8.10 (1H, s), 9.83 (1H, s), 10.28 (1H, s); MS (ESI) *m/z* 271 (M – H)⁻. Anal. (C₁₆H₁₀F₂O₂·0.25H₂O) C, H.

1-Chloro-6-(2-fluoro-4-hydroxyphenyl)-2-naphthol (24). The title compound was prepared by reacting **23** (300 mg, 1.18 mmol) and NCS (191 mg, 1.43 mmol) in THF (30 mL) according to method E to yield 170 mg (50%) of yellowish solid: mp 179–180 °C; ¹H NMR (DMSO- d_6) δ 6.73 (1H, dd, J = 12.69 Hz, J = 2.44 Hz), 6.75 (1H, dd, J = 8.30 Hz, J = 2.44 Hz), 7.31 (1H, d, J = 8.79 Hz), 7.45 (1H, t, J = 9.28 Hz), 7.70–7.72 (1H, m), 7.83 (1H, d, J = 8.79 Hz), 7.97 (1H, s), 8.06 (1H, d, J = 7.79 Hz), 10.06 (1H, s), 10.48 (1H, s); MS (ESI) *m/z* 287/289 (M – H)⁻. Anal. (C₁₆H₁₀ClFO₂) C, H.

1-Chloro-6-(3,5-difluoro-4-hydroxyphenyl)-2-naphthol (39). The title compound was prepared by reacting **38** (300 mg, 1.10 mmol) with NCS (155 mg, 1.16 mmol) in THF (30 mL) according to method E to yield 264 mg (78%) of gray solid: mp 209–210 °C; ¹H NMR (DMSO-*d*₆) δ 7.32 (1H, d, *J* = 8.89 Hz), 7.50–7.61 (2H, m), 7.83 (1H, d, *J* = 8.96 Hz), 7.92 (1H, dd. *J* = 8.94 Hz, *J* = 1.59 Hz), 8.05 (1H, d, *J* = 8.88 Hz), 8.22 (1H, d, *J* = 1.19 Hz), 10.36 (1H, s), 10.54 (1H, s); MS (ESI) *m/z* 305/307 (M – H)⁻. Anal. (C₁₆H₉ClF₂O₂) C, H.

N-(7-Hydroxynaphthyl)acetamide (107). To a solution of commercially available 8-amino-2-naphthol (149.1 g, 0.937 mol) in methanol (1 L) was added acetic anhydride (93 mL, 0.984 mol). The reaction was refluxed for 90 min and cooled to room temperature. The solvent was removed and the residue was filtered through a plug of silica with ethyl acetate. The solvent was removed to yield 175.8 g (93%) of the desired product as a dark purple solid. An analytical sample was further purified by reverse phase preparative HPLC to yield

a pink solid: mp 161–162 °C; ¹H NMR (DMSO- d_6) δ 2.15 (3H, s), 7.09 (1H, dd, J = 1.95 Hz, J = 8.79 Hz), 7.20–7.26 (2H, m), 7.49 (1H, d, J = 7.31 Hz), 7.63 (1H, d, J = 8.11 Hz), 7.77 (1H, d, J = 8.81 Hz), 9.76 (1H, s), 9.79 (1H, s), MS (ESI) m/z 202 (M + H)⁺. Anal. (C₁₂H₁₁NO₂) C, H, N.

N-(7-Methoxynaphthyl)acetamide (108). To a mixture of 107 (175.4 g, 0.872 mol), potassium carbonate (301 g, 2.18 mol), and acetone (1 l) was added iodomethane (270 mL, 4.36 mol). The reaction mixture was heated to reflux for 6 h and then cooled to room temperature, and the solvent was removed under vacuum. The residue was filtered through silica with ethyl acetate and triturated with ethyl acetate to yield 161.6 g (86%) of a gray solid. An analytical sample was further purified by preparative reverse phase HPLC to yield the title compound as a white solid: mp 154–155 °C; ¹H NMR (DMSO-*d*₆) δ 2.19 (3H, s), 3.90 (3H, s), 7.19 (1H, dd, *J* = 2.39 Hz, *J* = 8.94 Hz), 7.29–7.34 (2H, m), 7.39 (1H, d, *J* = 1.86 Hz), 7.75–7.68 (2H, m), 7.87 (1H, d, *J* = 8.96 Hz), 9.82 (1H, s); MS (ESI) *m/z* 216 (M + H)⁺. Anal. (C₁₃H₁₃NO₂) C, H, N.

7-Methoxynaphthylamine (109). A mixture of **108** (160.6 g, 0.747 mol) and HCl (1.5 L of 1 N solution) was heated to reflux for 5 h. The reaction was allowed to cool to room temperature, neutralized with solid sodium bicarbonate, and extracted with dichloromethane. The combined organic layers were filtered through silica, and the solvent was removed under vacuum to yield 89.5 g (69%) of a brown solid. An analytical sample was prepared by preparative HPLC to yield the title compound as a white solid: mp 134–136 °C; ¹H NMR (CDCl₃) δ 3.94 (3H, s), 4.01 (2H, bs), 6.80 (1H, d, J = 7.26 Hz), 7.05 (1H, d, J = 2.20 Hz), 7.12–7.19 (2H, m), 7.28 (1H, d, J = 8.17 Hz), 7.71 (1H, d, J = 8.93 Hz); MS (ESI) *m/z* 174 (M + H)⁺. Anal. (C₁₁H₁₁NO·CF₃CO₂H) C, H, N.

1-Fluoro-7-methoxynaphthalene (110). To a mixture of 109 (10.94 g, 63.24 mmol), HCl (15.8 mL of 12 N solution, 190 mmol), and water (50 mL) cooled to 10 °C was added a solution of sodium nitrite (4.58 g, 66.40 mmol) in water (25 mL) over 10 min. The solution was stirred for 30 min and combined with fluoroboric acid (100 mL). The resulting green solid was collected by filtration and washed with water, with ethanol, and then with ether to yield 15.48 g (90%) of the diazonium fluoroborate salt intermediate as a yellow solid. This yellow solid was combined with xylenes (250 mL) and allowed to reflux for 1 h. The solvent was removed and the residue was partitioned between ethyl acetate and sodium bicarbonate solution. The organic layer was dried over sodium sulfate and filtered, the solvent was evaporated, and the product was purified on silica (hexanes) to yield a light yellow liquid: 1H NMR (CDCl₃) & 3.94 (3H, s), 7.10-7.28 (3H, m), 7.34 (1H, d, J = 2.50 Hz), 7.55 (1H, d, J = 8.12 Hz), 7.75 (1H, dd, J = 1.69Hz, J = 9.00 Hz); MS (EI) m/z 176 (M[•])⁺. Anal. (C₁₁H₉FO) C, H.

8-Fluoro-2-naphthol (111). The title compound was prepared by reacting **110** (7.99 g, 45.34 mmol) with boron tribromide (68 mL of 1 N solution, 68 mmol) according to method C to yield 3.99 g (54%) of a red solid. An analytical sample was prepared by preparative reverse phase HPLC to yield a white solid: mp 89–92 °C; ¹H NMR (DMSO- d_6) δ 7.16 (1H, dd, J = 2.42 Hz, J = 8.90 Hz), 7.21–7.25 (3H, m), 7.62–7.65 (1H, m), 7.85 (1H, dd, J = 1.72 Hz, J = 8.87 Hz), 10.08 (1H, s); MS (ESI) m/z 161(M – H)⁻. Anal. (C₁₀H₇FO) C, H.

6-Bromo-8-fluoro-2-naphthol (113). To a solution of **111** (3.24 g, 20.0 mmol) in glacial acetic acid (30 mL) was slowly added a solution of bromine (7.35 g, 46.0 mmol) in glacial acetic acid (30 mL). The solution was stirred at 100 °C for 1 h. The reaction was cooled to room temperature, poured into water (50 mL), and extracted with ethyl acetate (3×150 mL). The combined organic layers were washed with sodium bicarbonate solution, dried over sodium sulfate, and filtered; the solvent was evaporated; and the product was purified on a silica column (2.5% ethyl acetate—hexanes) to yield 3.96 g (12.4 mmol, 62%) of **112** as a yellow solid. This solid was combined with SnCl₂ (7.0 g, 31 mmol), glacial acetic acid (35 mL), and HCl (35 mL of 12 N) and heated to 100 °C for 1 h. The resulting solution was cooled to room temperature, poured into water

(100 mL), and extracted with ethyl acetate (3 × 200 mL). The combined organic layers were washed with water, dried over sodium sulfate, filtered, and concentrated, and the product was purified on silica 2.5% ethyl acetate–hexanes) to yield 1.97 g (41%) a white solid: mp 124–126 °C; ¹H NMR (DMSO- d_6) δ 7.20 (1H, s), 7.23 (1H, d, J = 2.39 Hz), 7.48 (1H, dd, J = 1.78 Hz, J = 10.52 Hz), 7.84–7.88 (1H, m), 7.94 (1H, d, J = 0.79 Hz), 10.28 (1H, s); MS (ESI) *m*/*z* 239/241 (M – H)[–]. Anal. (C₁₀H₆BrFO) C, H.

8-Fluoro-6-(4-methoxyphenyl)-2-naphthol (116). The title compound was prepared by reacting **113** (0.39 g, 1.62 mmol) with 4-methoxyphenyl boronic acid (0.34 g, 2.27 mmol) according to method A to yield 0.35 g (81%) of a white solid: mp 150–151 °C; ¹H NMR (DMSO- d_6) δ 3.81 (3H, s), 7.05 (2H, d, J = 8.75 Hz), 7.16–7.20 (2H, m), 7.57 (1H, dd, J = 1.21 Hz, J = 12.76 Hz), 7.74 (2H, d, J = 8.74 Hz), 7.89–7.92 (2H, m), 10.10 (1H, s); MS (ESI) m/z 269 (M – H)⁺. Anal. (C₁₇H₁₃FO₂· 0.1H₂O) C, H.

8-Fluoro-6-(3-fluoro-4-methoxyphenyl)-2-naphthol (117). The title compound was prepared by reacting **113** (0.66 g, 2.74 mmol) with 3-fluoro-4-methoxyphenyl boronic acid^{7,8} (0.56 g, 3.3 mmol) according to method A to yield 0.67 g (85%) of a white solid: mp 138–140 °C; ¹H NMR (DMSO-*d*₆) δ 3.90 (3H, s), 7.17–7.30 (3H, m), 7.60–7.65 (2H, m), 7.71 (1H, dd, J = 2.19 Hz), J = 13.14 Hz), 7.90 (1H, d, J = 8.37 Hz), 7.99 (1H, bs), 10.15 (1H, bs); MS (ESI) *m/z* 285 (M – H)[–]. Anal. (C₁₇H₁₂F₂O₂) C, H.

8-Fluoro-6-(4-hydroxyphenyl)-2-naphthol (40). The title compound was prepared by reacting **116** (0.10 g, 0.37 mmol) with boron tribromide (0.56 mL of 1 N solution, 0.56 mmol) according to method C to yield 0.10 g (100%) of a white solid: mp 236–238 °C; ¹H NMR (DMSO-*d*₆) δ 6.87 (2H, d, *J* = 8.57 Hz), 7.14–7.18 (2H, m), 7.52 (1H, dd, *J* = 1.18 Hz, *J* = 12.78 Hz), 7.61 (2H, d, *J* = 8.59 Hz, 7.86–7.89 (2H, m), 9.61 (1H, bs), 10.05 (1H, bs); MS (ESI) *m*/*z* 253 (M – H)⁻. Anal. (C₁₆H₁₁FO₂·0.1H₂O) C, H.

8-Fluoro-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (41). The title compound was prepared by reacting **117** (0.10 g, 0.35 mmol) with boron tribromide (0.7 mL of 1 N solution, 0.7 mmol) according to method C to yield 0.020 g (21%) of a white solid: mp 218–220 °C (dec); ¹H NMR (DMSO-*d*₆) δ 7.01–7.07 (1H, m), 7.16–7.19 (2H, m), 7.46 (1H, dd, *J* = 1.74 Hz, *J* = 8.40 Hz), 7.56–7.65 (2H, m), 7.88 (1H, d, *J* = 8.31 Hz), 7.93 (1H, bs), 10.04 (1H, s), 10.11 (1H, s); MS (ESI) *m/z* 271 (M – H⁺)⁻. Anal. (C₁₆H₁₀F₂O₂·0.25H₂O) C, H.

1-Chloro-8-fluoro-6-(4-methoxyphenyl)-2-naphthol (118). A solution of **116** (0.17 g, 0.63 mmol) and NCS (0.10 g, 0.76 mmol) in THF (20 mL) was stirred under nitrogen at room temperature overnight according to method E. The solution was concentrated onto Florosil and purified on a silica column (20% ethyl acetate-hexanes) to yield 0.16 g (84%) a yellow solid. An analytical sample was further prepared by preparative reverse phase HPLC to yield a light yellow solid: mp 120–124 °C; ¹H NMR (DMSO-*d*₆) δ 3.82 (3H, s), 7.06 (2H, d, J = 8.80 Hz), 7.34 (1H, d, J = 8.91 Hz), 7.67 (1H, dd, J = 1.69 Hz, J = 15.48 Hz), 7.78 (2H, d, J = 8.78 Hz), 8.01 (1H, d, J = 1.54 Hz), 10.62 (1H, s); MS (ESI) *m/z* 301/303 (M – H⁺)⁻. Anal. (C₁₇H₁₂ClFO₂) C, H.

1-Chloro-8-fluoro-6-(3-fluoro-4-methoxyphenyl)-2-naphthol (119). The title compound was prepared by reacting **41** (0.30 g, 1.05 mmol) and NCS (0.17 g, 1.26 mmol) according to method E to yield 0.21 g (62%) of a light orange solid: mp 126–128 °C; ¹H NMR (DMSO-*d*₆) δ 3.90 (3H, s), 7.25–7.31 (1H, m), 7.35 (1H, d, *J* = 8.91 Hz), 7.64–7.67 (1H, m), 7.70–7.78 (2H, m), 7.87 (1H, dd, *J* = 1.58 Hz, *J* = 9.04 Hz), 10.68 (1H, s); MS (ESI) *m/z* 319/321 (M – H⁺)⁻. Anal. (C₁₇H₁₁ClF₂O₂) C, H.

1-Chloro-8-fluoro-6-(4-hydroxyphenyl)-2-naphthol (42). The title compound was prepared by reacting **118** (0.14 g, 0.46 mmol) with boron tribromide (0.69 mL of 1 N solution, 0.69 mmol) according to method C to yield 0.050 g (38%) of a white solid mp 174–176 °C; ¹H NMR (DMSO-*d*₆) δ 6.88 (2H, d, *J* = 8.62 Hz), 7.33 (1H, d, *J* = 8.92 Hz), 7.60–7.67 (3H, m), 7.86 (1H, dd, *J* = 1.43 Hz, *J* = 9.02 Hz), 7.95 (1H, d, *J* = 1.37 Hz), 9.68 (1H, s), 10.59 (1H, s); MS (ESI) $m\!/z$ 287/289 (M - H^+)^-. Anal. (C16H10ClFO2*0.25H2O) C, H.

1-Chloro-8-fluoro-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (43). The title compound was prepared by reacting 119 (0.13 g, 0.41 mmol) with boron tribromide (0.8 mL of 1 N solution, 0.8 mmol) according to method C to yield 0.070 g (56%) of a white solid: mp 198–200 °C; ¹H NMR (DMSO-d₆) δ 7.03–7.09 (1H, m) 7.34 (1H, d, J = 8.92 Hz), 7.49–7.52 (1H, m), 7.68 (2H, d, J = 15.0 Hz), 7.85–7.88 (1H, m), 8.02 (1H, d, J = 0.70 Hz), 10.13 (1H, s), 10.66 (1H, s); MS (ESI) m/z 305/307 (M – H⁺)⁻. Anal. (C₁₆H₉ClF₂O₂) C, H.

8-Chloro-2-methoxynaphthalene (120). CuCl₂ (4.6 g, 24.6 mmol) and tert-butyl nitrite (4.46 g, 43.3 mmol) were added to acetonitrile (125 mL) at 0 °C. To this mixture was slowly added a solution of 109 (4.99 g, 28.8 mmol) in acetonitrile (25 mL). The reaction was allowed to warm to room temperature, stirred with HCl (400 mL of 2 N solution), and extracted with ethyl acetate (3 \times 200 mL). The combined organic layers were washed with 2 N HCl, dried over sodium sulfate, and filtered; the solvent was evaporated; and the product was purified on a silica column (hexanes) to yield 2.27 g (41%) of an orange liquid. An analytical sample was further purified by reverse phase preparative HPLC to yield the title compound as a yellow solid: mp 32-34 °C; ¹H NMR (CDCl₃) δ 3.98 (3H, s), 7.20 (1H, dd, J = 2.52 Hz, J = 8.96 Hz), 7.22-7.27 (1H, m), 7.52 (1H, d, J = 2.47 Hz), 7.55 (1h, d, J = 7.47Hz), 7.69 (1H, d, J = 8.15 Hz), 7.75 (1H, d, J = 8.95 Hz); MS (ESI) m/z 191/193 (M – H⁺)⁻. Anal. (C₁₁H₉ClO) C, H.

8-Chloro-2-naphthol (121). The title compound was prepared by reacting **120** (10.25 g, 53.4 mmol) with boron tribromide (67 mL of 1 N solution, 67 mmol) according to method C to yield 8.76 g (92%) of a yellow solid. An analytical sample was further purified by reverse phase preparative HPLC to yield the title compound as a white solid: mp 95–100 °C; ¹H NMR (DMSO-*d*₆) δ 7.18 (1H, dd, J = 2.37 Hz, J = 8.85 Hz), 7.23–7.28 (1H, m), 7.41 (1H, d, J = 2.28 Hz), 7.59 (1H, d, J = 7.37 Hz), 7.81 (1H, d, J = 8.15 Hz), 7.87 (1H, d, J = 8.86 Hz), 10.17 (1H, s); MS (ESI) *m/z* 177/179 (M – H⁺)⁻. Anal. (C₁₀H₇ClO) C, H.

3,6-Dibromo-8-chloro-2-naphthol (122). To a solution of 121 (4.92 g, 27.6 mmol) in glacial acetic acid (40 mL) was slowly added a solution of bromine (9.7 g, 60.7 mmol) in glacial acetic acid (40 mL). The solution was stirred at 100 °C for 1 h, cooled to room temperature, poured into water (250 mL), and extracted with ethyl acetate. The combined organic layers were washed with sodium bicarbonate, dried with sodium sulfate, and filtered; the solvent was evaporated; and the product was purified on a silica column (20% ethyl acetatehexanes) to yield 4.61 g (50%) of a yellow solid. An analytical sample was further purified by reverse phase preparative HPLC to yield the title compound as a white solid: mp 156-158 °C; ¹H NMR (DMSO- d_6) δ 7.57 (1H, s), 7.82 (1H, d, J =1.89 Hz), 8.11 (1H, d, J = 1.69 Hz), 8.31 (1H, s), 11.30 (1H, bs); MS (ESI) *m/z* 333/335/337 (M – H)⁻. Anal. (C₁₀H₅Br₂ClO) C, H.

In addition, 1.80 g of **123** was isolated (16%) as a light orange solid: mp 166–170 °C; ¹H NMR (DMSO- d_6) δ 7.90 (1H, d, J = 2.07 Hz), 8.23 (1H, d, J = 2.08 Hz), 8.40 (1H, s), 10.54 (1H, bs); MS (ESI) m/z 411/413/415/417/419 (M – H)⁻. Anal. (C₁₀H₄Br₃ClO) C, H calcd, 28.92; found: 30.15.

tert-Butyl[(3,6-dibromo-8-chloro-2-naphthyl)oxy]dimethylsilane (124). The title compound was prepared by reacting 122 (3.00 g, 8.92 mmol) with TBDMSCl (1.75 g, 15.07 mmol), according to the experimental for 101, to yield 3.36 g (84%) of a white solid: mp 58–64 °C; ¹H NMR (CDCl₃) δ 0.34 (6H, s), 1.08 (9H, s), 7.57 (1H, s), 7.62 (1H, d, J = 1.72 Hz), 7.75 (1H, d, J = 1.24 Hz), 7.97 (1H, s); MS (ESI) *m*/*z* 333/335/337 (M - H)⁻. Anal. (C₁₆H₁₉Br₂ClOSi) C, H.

3-Bromo-8-chloro-6-(4-methoxyphenyl)-2-naphthol (125). Naphthalene 124 (1.95 g, 4.32 mmol) was reacted with with 4-methoxyphenylmagnesium bromide (13.8 mL of 0.5 N solution, 6.9 mmol) and tetrakis(triphenylphosphine)palladium (0.25 g, 0.21 mmol) in THF (10 mL) according to method B to yield 1.42 g (69%) of *tert*-butyl[(3-bromo-8-chloro-(4-methoxyphenyl)-2-naphthyl)
oxy]dimethylsilane as a white solid:
 $^{1}\mathrm{H}$ NMR (CDCl₃) δ 0.36 (6H, s), 1.10 (9H, s), 3.87 (3H, s),
 7.01 (2H, d, J=8.78 Hz), 7.33 (1H, d, J=1.36 Hz), 7.58 (2H,
 d, J=8.75 Hz), 7.73 (1H, s), 7.78 (1H, d, J=1.62 Hz), 8.10 (1H, s).

tert-Butyl [(3-bromo-8-chloro-(4-methoxyphenyl)-2-naphthyl)-oxy] dimethylsilane (0.50 g, 1.06 mmol) was reacted with TBAF (5 mL of 1 N solution in THF, 55 mmol), according to the preparation of compound **22** (method G), to yield 0.34 g (88%) of a white solid: mp 138–139 °C; ¹H NMR (DMSO- d_6) δ 3.82 (3H, s), 7.06 (2H, d, J = 8.76 Hz), 7.58 (1H, s), 7.73 (2H, d, J = 8.73 Hz), 7.94 (1H, d, J = 1.56 Hz), 8.07 (H, bs), 8.34 (1H, s), 11.05 (1H, s); MS (ESI) m/z 361/363/365 (M – H)[–]. Anal. (C₁₇H₁₂BrClO₂·0.6H₂O) C, H.

8-Chloro-6-(4-methoxyphenyl)-2-naphthol (126). Method **H.** To a solution of **125** (0.254 g, 0.698 mmol) in THF (25 mL) at -78 °C was slowly added tert-butyllithium (1.65 mL of 1.7 N solution, 2.8 mmol). The resulting solution was stirred for 20 min at -78 °C, 2.5 mL of water was added, and the mixture was warmed to room temperature slowly with stirring. The reaction was poured into 50 mL of water and extracted with ethyl acetate (3 \times 100 mL). The combined organic layers were washed with water, dried with sodium sulfate, filtered; the solvent was removed; and the product was purified on a silica column (10% ethyl acetate-hexanes) to yield 0.15 g (88%) of a yellow solid. An analytical sample was further prepared by reverse phase preparative HPLC to yield a light yellow solid: mp 116-118 °C; ¹H NMR (DMSO-*d*₆) δ 3.81 (3H, s), 7.05 (2H, d, J = 8.75 Hz), 7.19 (1H, dd, J = 2.34 Hz, J = 8.84 Hz), 7.40 (1H, d, J = 2.25 Hz), 7.74 (2H, d, J = 8.73 Hz), 7.89 (1H, d, J = 1.65 Hz), 7.92 (1H, d, J = 8.93 Hz), 8.07 (1H, s), 10.19 (1H, s); MS (ESI) m/z 283/285 (M – H)⁻. Anal. (C₁₇H₁₃ClO₂•0.1H₂O) C, H.

8-Chloro-6-(4-hydroxyphenyl)-2-naphthol (44). The title compound was prepared by reacting **126** (0.10 g, 0.35 mmol) with boron tribromide (0.9 mL of 1 N solution, 0.9 mmol) according to method C to yield 0.080 g (85%) of product that was further purified by reverse phase preparative HPLC to yield a white solid: mp 204–206 °C; ¹H NMR (DMSO- d_6) δ 6.87 (2H, d, J = 8.61 Hz), 7.17 (1H, dd, J = 2.34 Hz, J = 8.83 Hz), 7.38 (1H, d, J = 2.26 Hz), 7.62 (2H, d, J = 8.59 Hz), 7.85 (1H, d, J = 1.64 Hz), 7.90 (1H, d, J = 8.92 Hz), 8.01 (1H, bs), 9.62 (1H, s), 10.14 (1H, s); MS (ESI) *m/z* 269 (M – H)⁻. Anal. (C₁₆H₁₁ClO₂·0.25H₂O) C, H.

3-Bromo-1,8-dichloro-6-(4-methoxyphenyl)-2-naphthol (127). The title compound was prepared by reacting **125** (0.69 g, 1.90 mmol) and NCS (0.31 g, 2.3 mmol) in THF (25 mL) according to method E to yield 0.61 g (81%) of a yellow solid. An analytical sample was further prepared by reverse phase preparative HPLC to yield a white solid: mp 176–178 °C (dec); ¹H NMR (DMSO-*d*₆) δ 3.82 (3H, s), 7.08 (2H, d, *J* = 8.77 Hz), 7.77 (2H, d, *J* = 8.75 Hz), 8.00 (1H, d, *J* = 1.84 Hz), 8.19 (1H, d, *J* = 1.84 Hz), 8.39 (1H, s), 10.53 (1H, bs); MS (ESI) *m/z* 395/397/399 (M – H)⁻. Anal. (C₁₇H₁₁BrCl₂O₂) C, H.

1,8-Dichloro-6-(4-methoxyphenyl)-2-naphthol (128). The title compound was prepared by reacting **127** (0.30 g, 0.754 mmol) with *tert*-butyl lithium (1.75 mL of 1.7 N solution, 3.0 mmol) and quenching with water according to the procedure used to prepare **126** (method H) to yield 0.19 g (79%) of a yellow solid. An analytical sample was further prepared by reverse phase preparative HPLC to yield a light yellow solid: mp 132–134 °C; ¹H NMR (DMSO-*d*₆) δ 3.82 (3H, s), 7.06 (2H, d, J = 8.77 Hz), 7.37 (1H, d, J = 8.90 Hz), 7.77 (2H, d, J = 8.75 Hz), 7.89–7.93 (2H, m), 8.15 (1H, d, J = 1.85 Hz), 10.68 (1H, s); MS (ESI) *m/z* 317/319/321 (M – H)⁻. Anal. (C₁₇H₁₂Cl₂O₂) C, H.

1,8-Dichloro-6-(4-hydroxyphenyl)-2-naphthol (45). The title compound was prepared by reacting **128** (0.12 g, 0.38 mmol) with boron tribromide (0.75 mL of 1 N solution, 0.75 mmol) according to method C to yield 0.10 g (86%) which was further purified by reverse phase preparative HPLC to yield the title compound as a white solid: mp 172–174 °C (dec); ¹H NMR (DMSO-*d*₆) δ 6.89 (2H, d, J = 8.54 Hz), 7.36 (1H, d, J = 8.90 Hz), 7.65 (2H, d, J = 8.56 Hz), 7.88–7.91 (2H, m), 8.10

(1H, d, J = 1.63 Hz), 9.69 (1H, s), 10.64 (1H, s); MS (ESI) m/z 303/305/307 (M - H)⁻. Anal. (C₁₆H₁₀Cl₂O₂•0.25H₂O) C, H.

7-Methoxy-3,4-dihydronaphthalene-1-carbonitrile (130). To a mixture of commercially available 7-methoxy-1-tetralone (129) (39.65 g, 0.23 mol), zinc iodide (1.73 g, 5.4 mmol), and benzene (100 mLs) was added trimethylsilyl cyanide (25.0 g. 0.25 mol). The mixture was stirred overnight at room temperature. Pyridine (350 mL) was added and phosphorus oxychloride (100 mL) was then added dropwise. The mixture was heated to reflux for 6 h and then allowed to stir overnight at room temperature. The mixture was carefully poured onto ice/hydrochloric acid (1.5 L of 10% HCl) for a total volume of 2 L. This mixture was extracted with ethyl acetate (3 \times 500 mL), and the organics were combined and washed with water $(2 \times 500 \text{ mL})$ and then brine (500 mL). The ethyl acetate layer was dried over magnesium sulfate and the solvent evaporated to provide a liquid that solidified upon standing. This crude solid was triturated in hexane and filtered to give 31.3 g (74%) of a tan solid. A portion of this material was purified by silica gel chromatography (10% ethyl acetate-hexanes) to provide a solid: mp 47-49 °C; ¹H NMR (DMSO-d₆) & 2.44-2.50 (2H, m), 2.72 (2H, t, J = 8.4 Hz), 6.81 (1 H, d, J = 2.6 Hz), 6.90 (1H, dd, J = 2.9 Hz, J = 8.2 Hz), 7.17 - 7.19 (2H, m); MS (ESI)m/z 168 (M – H)⁻. Anal. (C₁₂H₁₁NO) C, H, N.

7-Methoxy-1-cyanonaphthalene (131). The title compound was prepared by reacting **130** (9.95 g, 53.1 mmol) with 10% Pd/C according to the procedure used to prepare naphthol **6** to yield a white solid (4.3 g, 44%). A portion was purified further by silica gel (10% ethyl acetate-hexanes) to produce a white solid: mp 77–78 °C; ¹H NMR (DMSO-*d*₆) δ 3.97 (3H, s), 7.36 (1H, s), 7.39 (1 H, d, J = 2.5 Hz), 7.52 (1H, t, J = 7.4 Hz), 8.04–8.13 (2H, m), 8.24 (1H, d, J = 8.2 Hz). Anal. (C₁₂H₉NO) C, H, N.

7-Hydroxy-1-naphthonitrile (132). The title compound was prepared by reacting **131** (1.19 g, 6.50 mmol) with pyridinium HCl (9 g) according to method D to yield 0.88 g (80%) of a white solid: mp 184–188 °C; ¹H NMR (DMSO-*d*₆) δ 7.25 (1H, dd, J = 2.30 Hz, J = 8.88 Hz), 7.38 (1H, d, J = 2.20 Hz), 7.39–7.44 (1H, m), 7.98 (1H, d, J = 8.91 Hz), 8.04 (1H, d, J = 7.14 Hz), 8.17 (1H, d, J = 8.19 Hz), 10.48 (1H, s); MS (ESI) *m/z* 170 ([M + H]⁺); MS (ESI) *m/z* 168 (M – H)⁻. Anal. (C₁₁H₇NO) C, H, N.

3-Bromo-7-hydroxy-1-naphthonitrile (133). To a mixture of **132** (26.03 g, 154.0 mmol) and glacial acetic acid (400 mL) was added bromine (51.8 g, 323 mmol). The mixture was stirred at 100 °C for 6 h, HCl (400 mL of 12 N solution) and SnCl₂ (69 g, 308 mmol) were added, and the mixture was allowed to stir at 100 °C for 1 h. The resulting solution was cooled to room temperature and poured into water (1 l). The resulting yellow precipitate was collected by filtration, dried under vacuum, and triturated with ethyl acetate to yield 14.68 g (38%) of an off-white solid. An analytical sample was prepared by preparative reverse phase HPLC to yield the desired product as a white solid: mp 222–224 °C; ¹H NMR (DMSO-*d*₆) δ 7.28–7.35 (2H, m), 7.97 (1H, d, *J* = 8.73 Hz), 8.26 (1H, s), 8.47 (1H, s), 10.65 (1H, s); MS (ESI) *m/z* 246/247 (M – H)⁻. Anal. (C₁₁H₆BrNO): C, H, N, calcd, 5.65; found, 4.82.

7-Hydroxy-3-(4-methoxyphenyl)-1-naphthonitrile (134). The title compound was prepared by reacting **133** (0.249 g, 1.00 mmol) with 4-methoxyphenylboronic acid (0.21 g, 1.4 mmol) according to method A to yield 0.19 (69%) of a light yellow solid: mp 226 °C; ¹H NMR (DMSO-*d*₆) δ 3.82 (3H, s), 7.07 (2H, d, J = 8.82 Hz), 7.26 (1H, dd, J = 2.32 Hz, J = 8.90 Hz), 7.37 (1H, d, J = 2.27 Hz), 7.80 (2H, d, J = 8.80 Hz), 8.02 (1H, d, J = 8.96 Hz), 8.37 (1H, d, J = 1.43 Hz), 8.44 (1H, d, J = 1.43 Hz), 10.48 (1H, bs); MS (ESI) *m/z* 274 (M – H)⁻. Anal. (C₁₈H₁₃NO₂•0.1H₂O) C, H, N.

3-(3-Fluoro-4-methoxyphenyl)-7-hydroxy-1-naphthonitrile (135). The title compound was prepared by reacting **133** (0.208 g, 0.839 mmol) with 3-fluoro-4-methoxyphenylboronic acid^{7,8} (0.18 g, 1.1 mmol) according to method A to yield 0.16 g (65%) of a light yellow solid. An analytical sample was prepared by preparative reverse phase HPLC to yield the title compound as a white solid: mp 214–216 °C; ¹H NMR (DMSO- $d_6)$ δ 3.90 (3H, s), 7.23–7.32 (2H, m), 7.37 (1H, d, J=2.24 Hz), 7.65–7.70 (1H, m), 7.79 (1H, dd, J=2.22 Hz, J=13.10 Hz), 8.03 (1H, d, J=8.96 Hz), 8.41 (1H, d, J=1.86 Hz), 8.50 (1H, d, J=1.43 Hz), 10.55 (1H, s); MS (ESI) m/z 292 (M-H)⁻. Anal. (C18H12FNO2·0.1H2O) C, H, N.

7-Hydroxy-3-(4-hydroxyphenyl)-1-naphthonitrile (46) The title compound was prepared by reacting **134** (0.14 g, 0.51 mmol) with pyridinium HCl (3 g) according to method D to yield 0.10 g (75%) of a white solid: mp 254–257 °C; ¹H NMR (DMSO- d_6) δ 6.89 (2H, d, J = 8.53 Hz), 7.25 (1H, dd, J = 2.15 Hz, J = 8.88 Hz), 7.36 (1H, d, J = 1.86 Hz), 7.67 (2H, d, J = 8.55 Hz), 8.00 (1H, d, J = 8.94 Hz), 8.31–8.38 (2H, m), 9.70 (1H, bs), 10.44 (1H, bs); MS (ESI) *m*/*z* 260 (M – H)[–]. Anal. (C₁₇H₁₁NO₂•0.25H₂O) C, H, N.

3-(3-Fluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (47). The title compound was prepared by reacting **135** (0.12 g, 0.41 mmol) with pyridinium HCl (3 g) according to method D to yield 0.085 g (74%) of a white solid: mp 265–269 °C; ¹H NMR (DMSO- d_6) δ 7.04–7.09 (1H, m), 7.26 (1H, dd, J = 2.31 Hz, J = 8.88 Hz), 7.36 (1H, d, J = 2.19 Hz), 6.52 (1H, dd, J = 1.76 Hz, J = 8.40 Hz), 6.71 (1H, dd, J = 2.22 Hz, J = 12.88 Hz), 8.01 (1H, d, J = 8.97 Hz), 8.37 (1H, d, J = 1.83 Hz), 8.45 (1H, d, J = 1.41 Hz), 10.33 (2H, bs); MS (ESI) m/z 278 (M – H)[–]. Anal. (C₁₇H₁₀FNO₂) C, H, N.

8-Bromo-7-hydroxy-3-(4-methoxyphenyl)-1-naphthonitrile (136). The title compound was prepared by reacting **134** (0.160 g, 0.58 mmol) with NBS (0.12 g, 0.70 mmol) in THF (10 mL) according to the procedure used to prepare **16** to yield 0.080 g (39%) of a yellow solid: mp 145–146 °C; ¹H NMR (DMSO-*d*₆) δ 3.83 (3H, s), 7.08 (2H, d, *J* = 8.79 Hz), 7.42 (1H, d, *J* = 8.79 Hz), 7.84 (2H, d, *J* = 8.79 Hz), 8.04 (1H, d, *J* = 8.79 Hz), 8.42 (1H, d, *J* = 1.95 Hz), 8.53 (1H, d, *J* = 1.95 Hz), 11.21 (1H, s); MS (ESI) *m/z* 354/356 (M + H⁺); MS (ESI) *m/z* 352/354 (M - H)⁻. Anal. (C₁₈H₁₂BrNO₂·0.25H₂O) C, H, N.

8-Bromo-7-hydroxy-3-(4-hydroxyphenyl)-1-naphthonitrile (48). The title compound was prepared by reacting **136** (0.13 g, 0.37 mmol) with boron tribromide (1.1 mL of 1 N solution, 1.1 mmol) according to method C to yield 0.050 g (40%) of an off white solid: mp 204–208 °C (dec); ¹H NMR (DMSO- d_6) δ 6.90 (2H, d, J = 8.30 Hz), 7.41 (1H, d, J = 8.79 Hz), 7.72 (2H, d, J = 8.79 Hz), 8.02 (1H, d, J = 9.23 Hz), 8.37 (1H, d, J = 1.95 Hz), 8.47 (1H, d, J = 2.44 Hz), 9.72 (1H, s), 11.16 (1H, s); MS (ESI) *m/z* 338/340 (M – H)⁻. Anal. (C₁₇H₁₀BrNO₂) C, H, N.

3-Bromo-8-chloro-7-hydroxy-1-naphthonitrile (137). The title compound was prepared by reacting **133** (0.32 g, 1.28 mmol) with NCS (0.24 g, 1.8 mmol) in THF (25 mL) at 45 °C for 3 h according to method E to yield 0.30 g (83%) of a yellow solid. An analytical sample was prepared by preparative reverse phase HPLC to yield a white solid: mp 148–150 °C; ¹H NMR (DMSO-*d*₆) δ 7.47 (1H, d, *J* = 8.98 Hz), 7.95 (1H, d, *J* = 9.02 Hz), 8.32 (1H, d, *J* = 2.08 Hz), 8.55 (1H, d, *J* = 2.08 Hz), 11.33 (1H, s); MS (ESI) *m/z* 280/282/284 (M – H)⁻. Anal. (C₁₁H₅BrClNO·0.25H₂O) C, H, N.

8-Chloro-3-(4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (49). The title compound was prepared by reacting **137** (0.10 g, 0.37 mmol) with 4-*tert*-butyldimethylsilyloxyphenylboronic acid⁸ (0.13 g, 0.52 mmol) according to method A to yield 0.036 g (33%) of a white solid: mp 254–256 °C; ¹H NMR (DMSO-*d*₆) δ 6.90 (2H, d, J = 8.64 Hz), 7.43 (1H, d, J = 8.95 Hz), 7.72 (2H, d, J = 8.64 Hz), 7.99 (1H, d, J = 8.97 Hz), 8.38 (1H, d, J = 1.95 Hz), 8.47 (1H, d, J = 1.92 Hz), 9.73 (1H, s), 11.08 (1H, s); MS (ESI) *m/z* 294/296 (M – H)⁻. Anal. (C₁₇H₁₀ClNO₂) C, H, N.

8-Chloro-3-(3-fluoro-4-methoxyphenyl)-7-hydroxy-1-naphthonitrile (138). The title compound was prepared by reacting **137** (0.20 g, 0.71 mmol) with 3-fluoro-4-methoxyphenylboronic acid^{7,8} (0.17 g, 1.0 mmol) according to method A to yield 0.14 g (60%) of a yellow solid: mp 198–200 °C; ¹H NMR (DMSO- d_6) δ 3.91 (3H, s), 7.27–7.33 (1H, m), 7.45 (1H, d, J = 8.93 Hz), 7.71–7.74 (1H, m), 7.85 (1H, dd, J = 2.23 Hz, J = 13.09 Hz), 8.00 (1H, d, J = 9.08 Hz), 8.48 (1H, d, J = 1.97

Hz), 8.57 (1H, d, J = 1.95 Hz), 11.18 (1H, s); MS (ESI) m/z 326/328 (M - H)⁻. Anal. (C₁₈H₁₁ClFNO₂) C, H, N.

8-Chloro-3-(3-fluoro-4-hydroxyphenyl)-7-hydroxy-1naphthonitrile (50). The title compound was prepared by reacting 138 (0.042 g, 0.13 mmol) with pyridinium HCl (1.8 g) according to method D to yield 0.033 g (78%) of a tan solid. This material was further purified by preparative reverse phase HPLC to yield the title compound as a white solid: mp 246-252 °C; ¹H NMR (DMSO- d_6) δ 7.04-7.10 (1H, m), 7.44 (1H, d, J = 8.94 Hz), 7.56 (1H, dd, J = 1.66 Hz, J = 8.37 Hz), 7.76 (1H, dd, J = 2.20 Hz, J = 12.88 Hz), 7.99 (1H, d, J =9.09 Hz), 8.43 (1H, d, J = 1.98 Hz), 8.52 (1H, d, J = 1.94 Hz), 10.19 (1H, bs), 11.05 (1H, bs); MS (ESI) m/2 312/ 314 (M – H)⁻. Anal. (C₁₇H₉ClFNO₂·0.25H₂O) C, H, N.

3-Bromo-7-methoxy-1-naphthonitrile (139). The title compound was prepared by reacting 133 (3.01 g, 12.2 mmol) with iodomethane (8.62 g, 61 mmol) and K₂CO₃ (5.1 g, 37 mmol) in acetone (100 mL) overnight at reflux temperature with stirring. The solution was cooled to room temperature and poured into 300 mL of water. The resulting mixture was made slightly acidic and extracted with ethyl acetate (3 × 150 mL). The combined organic layers were washed with water, dried over sodium sulfate, and filtered, and the solvent was evaporated. Purification by silica column (95% hexanes/5% ethyl acetate) yielded 2.41 g (75%) of a white solid: mp 140–143 °C; ¹H NMR (DMSO-*d*₆) δ 3.97 (3H, s), 7.33 (1H, d, J = 2.37 Hz), 7.42 (1H, dd, J = 9.03 Hz, J = 2.45 Hz), 8.04 (1H, d, J = 9.08 Hz), 8.35 (1H, d, J = 2.00 Hz), 8.55 (1H, d, J = 1.86 Hz); MS (EI) *m/z* 261 (M•)⁺. Anal. (C₁₂H₈BrNO) C, H, N.

3-Bromo-8-fluoro-7-methoxy-1-naphthonitrile (140). To a mixture of **139** (0.93 g, 4.27 mmol) and Selectfluor (1.66 g, 4.70 mmol) in a sealed flask under nitrogen was added acetonitrile (30 mL) via syringe. The resulting solution was stirred at room temperature overnight and then poured into 200 mL of water. The resulting mixture was extracted with ethyl acetate (2 × 100 mL). The combined organic layers were washed with brine, dried over sodium sulfate, and filtered; the solvent was evaporated; and the product waspurified on silica (80% hexanes/20% ethyl acetate) to yield 0.15 g (13%) of a white solid: mp 167–170 °C; ¹H NMR (CDCl₃) δ 4.07 (3H, d, J = 0.64 Hz), 7.41–7.45 (1H, m), 7.61 (1H, dd, J = 9.12 Hz, J = 1.62 Hz), 8.00 (1H, d, J = 1.81 Hz), 8.15–8.16 (1H, m); MS (EI) m/z 279. Anal. (C₁₂H₇BrFNO·0.1H₂O) C, H, N.

8-Fluoro-3-(3-fluoro-4-methoxyphenyl)-7-methoxy-1naphthonitrile (141). The title compound was prepared by reacting **140** (0.13 g, 0.46 mmol) with 3-fluoro-4-methoxyphenylboronic acid^{7,8} (0.11 g, 0.65 mmol) according to method A to yield 0.11 g (74%) of a tan solid: mp 170–172 °C; ¹H NMR (CDCl₃) δ 3.97 (3H, s), 4.09 (3h, s), 7.07–7.11 (1H, m), 7.37– 7.46 (3H, m), 7.73 (1H, dd, J = 9.46 Hz, J = 1.70 Hz), 8.09– 8.10 (1H, m), 8.14 (1H,d, J = 1.68 Hz); MS (EI) *m*/*z* 325.1 (M[•])⁺. Anal. (C₁₉H₁₃F₂NO₂·0.2H₂O) C, H, N.

8-Fluoro-3-(3-fluoro-4-hydroxyphenyl)-7-hydroxy-1naphthonitrile (51). Treatment of **141** (0.090 g, 0.28 mmol) with BBr₃ (1.1 mL of a 1 N solution, 1.1 mmol) according to method C yielded 0.074 g (90%) of a light yellow solid: mp >250 °C; ¹H NMR (DMSO-d₆) δ 7.05–7.09 (1H, m), 7.40–7.42 (1H, m), 7.53–7.58 (1H, m), 7.74 (1H, dd, J = 12.87 Hz, J = 2.27 Hz), 7.84 (1H, dd, J = 9.06 Hz, J = 1.04 Hz), 8.42 (1H, d, J = 1.81 Hz), 8.50–8.51 (1H, m), 10.15 (1H, bs), 10.69 (1H, bs); MS (ESI) m/z 296 (M – H⁺)⁻. Anal. (C₁₇H₉F₂NO₂) C, H, N: calcd, C: 68.69 N: 4.71; found, C: 64.07 N: 4.21.

7-[[tert-Butyl(dimethyl)silyl]oxy]-3-(4-[[tert-butyl(dimethyl)silyl]oxy]-3-fluorophenyl)-1-naphthonitrile (142). The title compound was prepared by reacting **47** (4.24 g, 15.20 mmol) with TBDMS-Cl (5.74 g, 38.0 mmol), according to the procedure used to prepare compound **101**, to yield 7.68 g (15.12 mmol) of a white solid: mp 104-105 °C; ¹H NMR (CDCl₃) δ 0.24 (3H, s), 0.25 (3H, s), 0.31 (6H, s), 1.03 (9H, s), 1.04 (9H, s), 7.02-7.05 (1H, m), 7.22 (1H, dd, J = 8.79 Hz, J = 2.33 Hz), 7.27-7.30 (1H, m), 7.36 (1H, dd, J = 11.64 Hz, J = 2.32 Hz), 7.56 (1H, d, J = 0.32 Hz), 7.84 (1H, d, J = 8.79 Hz), 8.04 (1H, d, J = 1.81 Hz), 8.10 (1H, s); MS (EI) *m*/z 507.4 (M•)⁺. Anal. (C₂₉H₃₈FNO₂Si₂) C, H: calcd, 68.59; found, 67.84.

7-[[tert-Butyl(dimethyl)silyl]oxy]-3-(4-[[tert-butyl(dimethyl)silyl]oxy]-3-fluorophenyl)-1-naphthaldehyde (143). To a solution of 142 (7.46 g, 14.69 mmol) in toluene (150 mL) at -78 °C was added DIBAL (14.7 mL of a 1 N solution, 14.7 mmol). The solution was allowed to stir for 1 h at -78 °C and then allowed to warm to room temperature slowly overnight. Methanol (5 mL) and water (5 mL) were added and the mixture was stirred for 30 min. The resulting suspension was filtered and the solid was washed well with ethyl acetate. The combined filtrates were washed with water and with brine and dried over sodium sulfate, the solvent was evaporated, and the product was purified on silica (1-2% ethyl acetate/ hexanes) to yield the product as a yellow solid: mp 105-107 °C; ¹H NMR (CDCl₃) δ 0.24 (3H, s), 0.25 (3H, s), 0.34 (6H, s), 1.04 ((9H, s), 1.05 (9H, s), 7.01-7.06 (1H, m), 7.20 (H, dd, J =8.79 Hz, J = 2.46 Hz), 7.34–7.37 (1H, m), 7.43 (1H, dd, J =11.77 Hz, J = 2.20 Hz), 7.84 (1H, d, J = 8.93 Hz), 8.12 (1H, dd, J = 7.50 Hz, J = 1.81 Hz), 8.72 (1H, d, J = 2.33 Hz), 10.39(1H, s); MS (EI) m/z 510.26 (M[•])⁺. Anal. (C₂₉H₃₉FO₃Si₂) C, H.

3-(3-Fluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthaldehyde (52). The title compound was prepared by reacting **143** (0.34 g, 0.67 mmol) with TBAF, according to method G, to yield 0.12 g (64%) of a yellow solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 7.07-7.11 (1H, m), 7.20 (1H, dd, J = 8.92 Hz, J= 2.46 Hz), 7.54-7.56 (1H, m), 7.72 (1H, dd, J = 12.80, J = 2.20 Hz), 7.97 (1H, d, J = 9.06 Hz), 8.43-8.44 (1H, m), 8.54 (H, d, J = 2.46 Hz), 10.07 (1H, bs), 10.22 (1H, bs), 10.34 (1H, s); MS (ESI) m/z 281 (M - H⁺)⁻. Anal. (C₁₇H₁₁FO₃·0.1H₂O) C, H.

6-(3-Fluoro-4-hydroxyphenyl)-8-vinyl-2-naphthol (53). To a suspension of methyltriphenylphosphonium bromide (2.1 g, 5.88 mmol) in THF (30 mL) at 0 °C was added nbutyllithium (2.35 mL of 2.5 N, 5.88 mmol). The mixture was stirred for 15 min at 0 °C. A solution of 143 (1.50 g, 2.94 mmol) in THF (15 mL) was added and stirred for 30 min at 0 °C. Water (5 mL) was added and the mixture was stirred for 10 min. The reaction was diluted with 100 mL of ethyl acetate and washed with 100 mL of 1 N HCl. The organic layer was concentrated and treated with TBAF in THF, according to method G, to yield 0.33 g (66%) of a tan solid: mp 132-136°C; ¹H NMR (DMSO- d_6) δ 5.50 (1H, dd, J = 11.00 Hz, J =1.96 Hz), 5.93-5.98 (1H, m), 7.03-7.07 (1H, m), 7.12 (1H, dd, J = 8.86 Hz, J = 2.26 Hz), 7.33-7.40 (2H, m), 7.45-7.47 (1H, m), 7.62 (1H, dd, J = 12.94 Hz, J = 2.20 Hz), 7.82–7.85 (2H, m), 8.00 (1H, d, J = 1.43 Hz), 9.93 (1H, s), 10.20 (1H, s); MS (ESI) m/z 279 (M – H⁺)⁻. Anal. (C₁₈H₁₃FO₂·0.20H₂O) C, H.

8-Ethyl-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (54). A mixture of **53** (0.16 g, 0.57 mmol) and 10% Pd/C (60 mg) in ethyl acetate (20 mL) was stirred under a balloon of hydrogen for 4 h. The mixture was filtered through Celite and the solvent evaporated. Purification by silica column (25% ethyl acetate/ hexanes) yielded 0.13 g (71%) of the desired product as a white solid: mp 126–128 °C; ¹H NMR (DMSO-*d*₆) δ 1.33 (3H, t, *J* = 7.43 Hz), 2.99 (2H, q, *J* = 7.43 Hz), 7.02–7.09 (2H, m), 7.24 (1H, d, *J* = 8.40 Hz, *J* = 1.60 Hz), 7.53–7.57 (2H, m), 7.80 (1H, d, *J* = 8.97 Hz), 7.86 (H, d, *J* = 1.40 Hz), 9.74 (1H, s), 9.89 (1H, s); MS (ESI) *m*/*z* 283; MS (ESI) *m*/*z* 281 (M – H⁺)⁻; HRMS (ESI–FT, [M + H]⁺) calcd for C₁₈H₁₅FO₂ +H⁺ 283.11288, found 283.11253. Anal. (C₁₈H₁₅FO₂· 0.1H₂O) C, H.

8-Ethynyl-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (55). To a solution of CBr_4 (3.89 g, 11.7 mmol) in dichloromethane (30 mL) at 0 °C were added **143** (1.50 g, 2.94 mmol), zinc dust (0.76 g, 11.7 mmol), and triphenylphosphine (3.06 g, 11.7 mmol). The mixture was stirred at room temperature for 3 h and then filtered through silica. The silica was washed with dichloromethane, and the combined filtrates were concentrated and purified on silica (1.5% ethyl acetate-hexanes) to yield the intermediate **144** as a yellow oil.

To a solution of 144 (1.85 g, 2.77 mmol) in THF (25 mL) at -78 °C was slowly added *n*-butyllithium (4.4 mL of 2.5 N, 11.1 mmol). The resulting solution was stirred at -78 °C for 30 min and then allowed to warm to room temperature. Water (5 mL) was added, and the mixture was stirred at room

temperature for 3 h. TLC analysis indicated a mixture of products with one or both silyl groups removed. The reaction was poured into 150 mL of water. The resulting mixture was made slightly acidic and extracted with ethyl acetate $(2 \times 100$ mL). The combined organic layers were washed with water and with brine, dried over sodium sulfate, filtered, and concentrated under vacuum. The resulting oil was treated with TBAF in THF according to method G to yield 0.61 g (86%) of the title compound as a white solid: mp 140-141 °C; ¹H NMR $(DMSO-d_6) \delta 4.60 (1H, s), 7.02-7.07 (1H, m), 7.15 (1H, dd, J)$ = 8.86 Hz, J = 2.40 Hz), 7.43–7.45 (1H, m), 7.50 (1H, d, J = 2.46 Hz), 7.60 (1H, dd, J = 12.82 Hz, J = 2.32 Hz), 7.88 (1H, d, J = 8.79 Hz), 7.90 (1H, d, J = 1.94 Hz), 8.12 (1H, d, J =1.55 Hz), 9.99 (1H, s), 10.06 (1H, s); MS (ESI) $m\!/\!z$ 279 (M + $H^+)^+$; MS (ESI) *m/z* 277 (M - H^+)^-. Anal. (C₁₈H₁₁FO₂·1.0H₂O) C, H.

6-(3-Fluoro-4-hydroxyphenyl)-8-prop-1-ynyl-2-naphthol (56). Treatment of 55 (0.38 g, 1.36 mmol) with TBDMSCl (0.52 g, 3.4 mmol), according to the procedure used to prepare 101, afforded 0.53 g (77%) of 145 as a yellow oil. To a solution of 145 (0.53 g, 1.05 mmol) in THF (20 mL) at -78 °C was added *n*-butyllithium (0.8 mL of 2.5 N, 2.1 mmol). The resulting solution was allowed to stir at -78 °C for 30 min. Iodomethane (2 mL, filtered through basic alumina) was added, and the reaction was allowed to warm to room temperature overnight. The solvent was removed by evaporation (in order to remove excess iodomethane) and the resulting oil was treated with TBAF in THF, according to method G, to yield 0.30 g (98%) of a yellow solid: mp 83-86 °C; ¹H NMR (DMSO- d_6) δ 2.22 (3H, s), 7.01–7.06 (1H, m), 7.12 (1H, dd, J = 8.80 Hz, J = 2.46Hz), 7.42 (1H, dd, J = 8.34 Hz, J = 1.62 Hz), 7.50 (1H, d, J =2.33 Hz), 7.58 (1H, dd, J = 12.93 Hz, J = 2.20 Hz), 7.79 (1H, d, J = 1.82 Hz), 7.84 (1H, d, J = 8.80 Hz), 8.02 (1H, d, J =1.42 Hz), 9.96 (2H s); MS (ESI) m/z 293 (M + H⁺)⁺; MS (ESI) m/z 291 (M – H)⁻.

3-(2-Fluoro-4-methoxyphenyl)-7-methoxy-1-naphthonitrile (146). A mixture of 139 (0.73 g, 2.79 mmol), bis-(pinacolato)diboron (1.06 g, 4.18 mmol), [1,1'-bis(diphenylphosphino)ferrocene]dichloropalladium(II) complex with dichloromethane (1:1) (0.23 g, 0.28 mmol), and potassium acetate (0.55 g, 5.6 mmol) in DMF (15 mL) was stirred at 85 °C for 2 h. The mixture was cooled to room temperature; 2-fluoro-4-methoxyphenyl trifluoromethanesulfonate (1.52 g, 5.6 mmol, synthesized as described previously), sodium carbonate (2.8 mL of 2 N aqueous solution), and an additional 0.23 g of Pd(II) were added; and the mixture was heated to 85 °C for 6 h. The reaction was cooled to room temperature, dissolved in 100 mL of water, and extracted with ethyl acetate (3 \times 100 mL). The combined organic layers were washed with water, dried over sodium sulfate, and concentrated, and the product was purified on silica to yield 0.60 g (70%) a white solid: mp 144-146 °C; ¹H NMR (CDCl₃) δ 3.87 (3H, s), 4.01 (3H, s), 6.77 (1H, dd, J =12.53 Hz, J = 2.47 Hz), 6.83 (1H, dd, J = 8.60 Hz, J =2.69 Hz), 7.27 (1H, dd, J = 8.99 Hz, J = 2.43 Hz), 7.39-7.45 (1H, m), 7.47 (1H, d, J = 2.36), 7.84 (1H, d, J = 9.02 Hz), 8.05-8.06 (1H, m), 8.11 (1H, s); MS (EI) m/z 307 (M[•])⁺. Anal. (C₁₉H₁₄FNO₂) C, H, N.

3-(2,5-Difluoro-4-methoxyphenyl)-7-methoxy-1-naph-thonitrile (147). The title compound was prepared by reacting **139** (0.67 g, 2.56 mmol) with 1-bromo-2,5-difluoro-4-methoxybenzene as described for **146** to yield 0.55 g (66%) of a white solid: mp 190–193 °C; ¹H NMR (CDCl₃) δ 3.95 (3H, s), 4.02 (3H, s), 6.84 (1H, dd, J = 11.68 Hz, 7.15 Hz), 7.20–7.30 (2H, m), 8.75 (1H, d, J = 2.36 Hz), 7.85 (1H, d, J = 9.02 Hz), 8.03–8.05 (1H, m), 8.10 (1H, s); MS (EI) m/z 325 (M[·])⁺. Anal. (C₁₉H₁₃F₂NO₂) C, H, N.

3-(2,6-Difluoro-4-methoxyphenyl)-7-methoxy-1-naphthonitrile (148). The title compound was prepared by reacting 139 (0.51 g, 1.94 mmol) with commercially available 2-bromo-1,3-difluoro-5-methoxybenzene as described for 146 to yield 0.44 g (74%) of a white solid: mp 203–208 °C; ¹H NMR (CDCl₃) δ 3.86 (3H, s), 4.02 (3H, s), 6.58–6.63 (2H, m), 7.26–7.30 (1H, m), 7.48 (1H, d, J = 2.35 Hz), 8.78 (1H, d, J = 9.02 Hz), 7.96 (1H, d, J = 1.38 Hz), 8.07 (1H, s); MS (EI) m/z 325 (M[•])⁺. Anal. (C₁₉H₁₃F₂NO₂) C, H, N.

3-(2-Fluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (57). Treatment of **146** (0.55 g, 1.80 mmol) with pyridinium HCl according to method D yielded 0.35 g (67%) of a white solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 6.70 (1H, dd, J = 12.83 Hz, J = 2.29 Hz), 6.75 (1H, d, J = 8.40 Hz, J = 2.29 Hz), 7.25 (1H, dd, J = 8.86 Hz, J = 2.29 Hz), 7.36 (1H, d, J = 2.14 Hz), 7.46–7.50 (1H, m), 8.00 (1H, d, J = 9.01 Hz), 10.17 (1H, bs), 10.44 (1H, bs); MS (ESI) m/z 278 (M – H)⁻. Anal. (C₁₇H₁₀FNO₂·0.2H₂O) C, H, N.

3-(2,5-Difluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (58). Treatment of **147** (0.48 g, 1.48 mmol) with BBr₃ according to method C yielded 0.18 g (41%) of a white solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 6.90 (1H, dd, J = 11.83 Hz, J = 7.41 Hz), 7.26 (1H, dd, J = 8.93 Hz, J = 2.21 Hz), 7.37 (1H, d, J = 1.99 Hz), 7.55 (1Hk dd, J = 11.76 Hz, J = 7.63 Hz), 8.01 (1H, d, J = 8.85 Hz), 1 8.16 (1H, s), 8.30 (1H, s), 10.57 (2H, bs); MS (ESI) m/z 296 (M - H)⁻. Anal. (C₁₇H₉F₂NO₂·0.25H₂O) C, H, N.

3-(2,6-Difluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (59). Treatment of **148** (0.32 g, 0.98 mmol) with BBr₃ according to method C yielded 0.15 g (51%) of a white solid: mp >250 °C; ¹H NMR (DMSO-*d*₆) δ 6.59–6.64 (2H, m), 7.27 (1H, dd, J = 8.85 Hz, J = 2.29 Hz), 7.39 (1H, d, J = 2.13 Hz), 8.01 (1H, dJ = 9.01 Hz), 8.04 (1H, s), 8.19 (1H, s), 10.59 (1H, bs); MS (ESI) *m*/*z* 296 (M – H)⁻. Anal. (C₁₇H₉F₂NO₂· 0.25H₂O) C, H, N.

3-(3,5-Difluoro-4-hydroxyphenyl)-7-hydroxy-1-naphthonitrile (60). 4-Bromo-2,6-difluorophenol (10.5 g, 50.4 mmol) was reacted with TBDMS-Cl (9.88 g, 150.7 mmol) according to the procedure used to prepare **101**, yielding 14.6 g (90%) of *tert*-butyl(4-bromo-2,6-difluorophenyloxy)-dimethylsilane a clear colorless, oil: ¹H NMR (CDCl₃) δ 0.17 (6H, s), 0.99 (9H, s), 7.02 (H, d, J = 7.13 Hz). This material (12.98 g, 40.19 mmol) was treated with *n*-butyllithium (27.6 mL of 1.6 N solution, 44.2 mmol) followed by triisopropyl borate (37.8 g, 200 mmol) according to the procedure used prepare **101** to afford 4.38 g (38%) of 3,5-difluoro-4-*tert*-butyldimethylsilyoxyboronic acid as a white solid: ¹H NMR (DMSO- d_6) δ 0.25 (6H, s), 1.06 (9H, s), 7.54 (2H, d, J = 7.93 Hz).

The title compound was prepared by reacting **133** (0.124 g, 0.50 mmol) with 3,5-difluoro-4-*tert*-butyldimethylsilyloxyphenylboronic acid (0.17 g, 0.59 mmol, prepared above) according to method A to yield 0.090 g (61%) of a yellow solid. An analytical sample was prepared by preparative reverse phase HPLC to produce a light yellow solid: mp 300–304 °C (dec); ¹H NMR (DMSO-*d*₆) δ 7.27 (1H, dd, J = 2.26 Hz, J = 8.88 Hz), 7.36 (1H, d, J = 2.12 Hz), 7.63–7.66 (2H, m), 8.00 (1H, d, J = 8.95 Hz), 8.43 (1H, d, J = 1.77 Hz), 8.52 (1H, s), 10.44 (1H, s); MS (ESI) *m/z* 296 (M – H)⁻. Anal. (C₁₇H₉F₂NO₂•0.3H₂O) C, H, N.

7-Hydroxy-3-methoxy-1-naphthonitrile (149). The title compound was prepared by reacting **133** (4.04 g, 16.3 mmol) with sodium methoxide in the presence of copper(I) bromide according to method F to yield 2.17 g (67%) of a yellow solid: mp 185–188 °C; ¹H NMR (DMSO-*d*₆) δ 3.88 (3H, s), 7.21 (1H, dd, J = 8.87 Hz, J = 2.39 Hz), 7.31 (1H, d, J = 2.34 Hz), 7.68 (1H, d, J = 2.52 Hz), 7.77 (1H, d, J = 0.60 Hz), 7.86 (1H, d, J = 8.96 Hz), 10.16 (1H, s); MS (ESI) *m/z* 198 (M – H)⁻. Anal. (C₁₂H₉NO₂) C, H, N.

8-Cyano-6-methoxy-2-naphthyl Trifluoromethanesulfonate (150). The title compound was prepared by reacting 149 (2.07 g, 10.4 mmol) with trifluoromethanesulfonic anhydride according to the procedure used to prepare 74 to yield 3.25 g (94%) of a light yellow solid: mp 91–94 °C; ¹H NMR (CDCl₃) δ 3.97 (3H, s), 7.42 (1H, d, J = 2.40 Hz), 7.49 (1H, dd, J = 9.05 Hz, J = 2.46 Hz), 7.68 (1H, d, J = 2.47 Hz), 7.90 (1H, d, J = 9.04 Hz), 8.03 (1H, d, J = 2.39 Hz); MS (ESI) *m/z* 332 (M + H⁺)⁺. Anal. (C₁₃H₈F₃NO₄S) C, H, N.

3-Methoxy-7-(4-methoxyphenyl)-1-naphthonitrile (151). Treatment of **150** (0.71 g, 2.14 mmol) with 4-methoxyphenylboronic acid according to method A yielded 0.50 g (81%) a white solid: mp 163–165 °C; ¹H NMR (CDCl₃) δ 3.88 (3H, s), 3.96 (3H, s), 7.01–7.06 (2H, m), 7.39 (1H, d, J = 2.40 Hz), 7.59 (1H, d, J = 2.50 Hz), 7.65–7.70 (2H, m), 7.81 (1H, dd, J = 8.59 Hz, J = 1.64 Hz), 7.85 (1H, dJ = 8.51 Hz), 8.26 (1H, s). Anal. (C₁₉H₁₅NO₂) C, H, N.

7-(3-Fluoro-4-methoxyphenyl)-3-methoxy-1-naphthonitrile (152). Treatment of **150** (0.71 g, 2.14 mmol) with 3-fluoro-4-methoxyphenylboronic acid^{7,8} according to method A afforded 0.48 g (73%) a white solid: mp 204–207 °C; ¹H NMR (CDCl₃) δ 3.97 (6H, s), 7.06–7.12 (1H, m), 7.40 (1H, d, J = 2.44 Hz), 7.45–7.50 (2H, m), 7.60 (1H, d, J = 2.55 Hz), 7.77 (1H, dd, J = 8.58 Hz, J = 1.80 Hz), 7.76 (1H, d, J = 9.57 Hz), 8.24–8.25 (1H, m). Anal. (C₁₉H₁₄FNO₂) C, H, N.

3-Hydroxy-7-(4-hydroxyphenyl)-1-naphthonitrile (61). Compound **151** (0.42 g, 1.45 mmol) was reacted with pyridinium HCl according to method D to yield 0.30 g (80%) of a light yellow solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 6.90–6.94 (2H, m), 7.53 (1H, d, J = 2.27 Hz), 7.60–7.64 (2H, m), 7.68 (1H, d, J = 2.39 Hz), 7.85 (1H, dd, J = 8.67 Hz, J = 1.72 Hz), 7.95 (1H, d, J = 8.68 Hz), 8.02 (1H, d, J = 0.68 Hz), 9.69 (1H, s), 10.43 (1H, s); MS (ESI) *m*/*z* 260 (M – H)⁻. Anal. (C₁₇H₁₁NO₂) C, H, N.

7-(3-Fluoro-4-hydroxyphenyl)-3-hydroxy-1-naphthonitrile (62). Compound **152** (0.34 g, 1.11 mmol) was treated with pyridine HCl according to method D to afford 0.27 g (88%) of a light yellow solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 7.07–7.13 (1H, m), 7.45 (1H, dd, J = 8.38 Hz, J = 1.64 Hz), 7.54 (1H, d, J = 2.26 Hz), 7.61 (1H, dd, J = 12.70 Hz, J = 2.20 Hz), 7.69 (1H, d, J = 2.39 Hz), 7.87 (1H, dd, J = 8.68 Hz, J = 1.73 Hz), 7.96 (1H, d, J = 8.70 Hz), 8.04 (1H, d, J = 0.63 Hz), 10.26 (2H, bs); MS (ESI) m/z 278 (M – H)⁻. Anal. (C₁₇H₁₀FNO₂· 0.3H₂O) C, H, N.

4-Bromo-7-(3-fluoro-4-hydroxyphenyl)-3-hydroxy-1naphthonitrile (63). A solution of **62** (0.82 g, 2.94 mmol) and NBS (0.55 g, 3.1 mmol) in THF (30 mL) was stirred overnight at room temperature, according to the procedure used to prepare **16**, to yield 0.82 g (78%) of a white solid: mp >250 °C; ¹H NMR (DMSO-*d*₆) δ 7.110–7.14 (1H, m), 7.48 (1H, d, *J* = 7.94 Hz), 7.64 (1H, d, *J* = 12.30 Hz), 7.79 (1H, s), 8.04–8.09 (2H, m), 8.21 (1H, d *J* = 8.84 Hz), 10.17 (1H, s), 11.32 (1H, s); MS (ESI) *m/z* 356 (M – H)⁻. Anal. (C₁₇H₉BrFNO₂·0.1H₂O) C, H, N.

7-(3-Fluoro-4-hydroxyphenyl)-3-hydroxy-4-methyl-1naphthonitrile (65). A solution of 63 (0.22 g, 0.61 mmol), tetramethyltin (0.16 g, 0.9 mmol), and dichlorobis(tri-otolylphosphine)palladium(II) (0.048 g, 0.006 mmol) in DMF (3 mL) was stirred overnight at 80 °C. The reaction was cooled to room temperature, poured into 50 mL of 1 N NH₄Cl, and extracted with ethyl acetate (2 \times 75 mL). The combined organic layers were washed with water, dried over sodium sulfate, filtered, concentrated, and the product was purified on silica (30% ethyl acetate/hexanes) to yield 0.10 g (56%) of a white solid: mp >250 °C; ¹H NMR (acetone- d_6) δ 2.63 (3H, s), 7.15–7.19 (1H, m), 7.50–7.53 (1H, m), 7.60 (1H, dd, J = 12.43 Hz, J = 2.18 Hz), 7.72 (1H, s), 7.94 (1H, dd, J = 8.84Hz, J = 1.92 Hz), 8.17 (1H, dd, J = 8.97 Hz, J = 0.51 Hz), 8.21 (1H, d, J = 1.54 Hz), 8.95 (2H, bs); MS (ESI) m/z 292 (M - H)⁻. Anal. (C₁₈H₁₂FNO₂•0.1H₂O) C, H, N.

4-Chloro-3-hydroxy-7-(4-hydroxyphenyl)-1-naphthonitrile (66). Treatment of **61** (0.24 g, 0.92 mmol) with NCS (0.15 g, 1.10 mmol) in THF (10 mL) at room temperature according to method E yielded 0.15 g (56%) of a yellow solid: mp >250 °C; ¹H NMR (DMSO- d_6) δ 6.94 (2H, d, J = 8.59 Hz), 7.77 (2H, d, J = 8.61 Hz), 7.83 (1H, s), 8.04 (1H, dd, J = 8.89 Hz, J = 1.70 Hz), 8.08 (1H, d, J = 1.16 Hz), 8.22 (1H, d, J = 8.87 Hz), 9.75 (1H, bs), 11.24 (1H, bs); MS (ESI) *m/z* 294 (M – H)⁻. Anal. (C₁₇H₁₀ClNO₂·0.3H₂O) C, H, N.

4-Chloro-7-(3-fluoro-4-hydroxyphenyl)-3-hydroxy-1naphthonitrile (67). Reacting **62** (0.13 g, 0.47 mmol) with NCS (0.079 g, 0.6 mmol) in THF (10 mL) at room temperature according to method E produced 0.063 g (43%) of a white solid: mp >250 °C; ¹H NMR (DMSO-*d*₆) δ 7.10–7.12 (1H, m), 7.48 (1H, dd, *J* = 8.39 hz, *J* = 1.60 Hz), 7.64 (1H, dd, *J* = 12.62 Hz, *J* = 1.98 Hz), 7.83 (1H, s), 8.05 (1H, dd, *J* = 8.97 Hz, J = 1.41 Hz), 8.09 (1H, s), 8.21 (1H, d, J = 8.97 Hz), 10.18 (1H, bs), 11.25 (1H, bs); MS (ESI) m/z 312 (M – H)[–]. Anal. (C₁₇H₉ClFNO₂•0.1H₂O) C, H, N.

4-Bromo-7-(3-fluoro-4-methoxyphenyl)-3-methoxy-1naphthonitrile (153). A solution of **152** (0.98 g, 3.19 mmol) and bromine (0.56 g, 3.5 mmol) in acetic acid (10 mL) was heated to 100 °C overnight in a pressure flask. The flask was cooled to room temperature and the reaction was poured into 300 mL of water. The resulting dark yellow solid was collected by filtration and triturated with THF to yield 0.83 g (69%) of the product as a yellow solid: mp 192–194 °C; ¹H NMR (CDCl₃) δ 3.97 (3H, s), 4.07 (3H, s), 7.07–7.12 (1H, m), 7.46– 7.50 (2H, m), 7.64 (1H, s), 7.87 (1H, dd, J = 8.92 Hz, J = 1.81Hz), 8.26 (1H, d, J = 1.43 Hz), 8.35 (1H, d, J = 8.92 Hz); MS (EI) m/z 385 (M•)⁺. Anal. (C₁₉H₁₃BrFNO₂) C, H, N.

6-(3-Fluoro-4-methoxyphenyl)-2-methoxynaphthalene-1,4-dicarbonitrile (154). A mixture of **153** (0.48 g, 1.24 mmol), Zn(CN)₂ (0.10 g, 0.87 mmol), tris(dibenzylideneacetone-)dipalladium(0) (0.11 g, 0.12 mmol), zinc dust (0.039 g, 0.62 mmol), and dppf (0.069 g, 0.12 mmol) in DMF (15 mL) was stirred for 3 h at 120 °C. The reaction was cooled to room temperature and poured into 200 mL of 1 N NaOH. The resulting dark gray solid was collected by filtration. The crude product was filtered through a silica pad (50%THF/hexanes) and the filtrate was concentrated. The product was further purified by trituration to yield 0.33 g (80%) of a yellow solid: mp >220 °C; ¹H NMR (CDCl₃) δ 3.98 (3H, s), 4.14 (3H, s), 7.09–7.13 (1H, m), 7.46–7.49 (2H, m), 7.69 (1H, s), 7.98 (1H, dd, J = 8.73 Hz, J = 1.36 Hz), 8.24 (1H, d, J = 8.79 Hz), 8.30 (1H, s); MS (EI) m/z 332 (M•)⁺. Anal. (C₂₀H₁₃FN₂O₂) calcd, C: 72.28 N: 8.43; found, C: 70.26 N: 6.66.

6-(3-Fluoro-4-hydroxyphenyl)-2-hydroxynaphthalene-1,4-dicarbonitrile (64). Treatment of **154** (0.10 g, 0.31 mmol) with pyridinium HCl according to method D yielded 0.070 g (74%) of a yellow solid: ¹H NMR (DMSO- d_6) δ 7.10–7.14 (1H, m), 7.47–7.49 (1H, m), 7.65 (1H, dd, J = 12.60 Hz, J = 2.21 Hz), 7.81 (1H, s), 8.03 (1H, dd, J = 7.74 Hz, J = 1.76 Hz), 8.13–8.15 (2H, m), 10.19 (1H, s), 12.41 (1H, s); MS (ESI) *m/z* 303 (M – H)⁻. Anal. (C₁₈H₉FN₂O₂) C, H, N: calcd, C: 71.05 N: 9.21; found, C: 66.20 N: 8.00.

3-[[tert-Butyl(dimethyl)silyl]oxy]-7-(4-[[tert-butyl(dimethyl)silyl]oxy]-3-fluorophenyl)-1-naphthonitrile (155). The title compound was prepared by reacting **62** (2.28 g, 8.17 mmol) with TBDMSCI (4.9 g, 32.7 mmol) according to the procedure used to prepared compound **101** to yield 2.44 g (59%) of a white solid: mp 112–114 °C; ¹H NMR (CDCl₃) δ 0.24 (3H, s), 0.25 (3H, s), 0.28 (6H, s), 1.03 (9H, s), 1.04 (9H, s), 7.00–7.05 (1H, m), 7.35–7.39 (1H, m), 7.41–7.46 (2H, m), 7.51 (1H,d, J = 2.38 Hz), 7.75 (1H, dd, J = 8.61 Hz, J = 1.66 Hz), 7.81 (1H, d, J = 8.61 Hz), 8.25 (1H, s); MS (EI) *m/z* 507.2 (M•)⁺. Anal. (C₂₉H₃₈FNO₂Si₂) C, H, N: calcd, 68.59; found, 68.00.

3-[[tert-Butyl(dimethyl)silyl]oxy]-7-(4-[[tert-butyl(dimethyl)silyl]oxy]-3-fluorophenyl)-1-naphthaldehyde (156). The title compound was prepared by reacting **155** (2.27 g, 4.47 mmol) with DIBAL (4.9 mL of 1 N solution, 4.9 mmol) according to the procedure used to prepare compound **143** to yield 1.13 g (49%) of a yellow solid: mp 67–68 °C; ¹H NMR (CDCl₃) δ 0.24 (3H, s), 0.25 (3H, s), 0.30 (6H, s), 1.04 (9H, s), 1.05 (9H, s), 6.99–7.04 (1H, m), 7.38–7.41 (1H, m), 7.43–7.48 (2H, m), 7.58 (1H, d, J = 2.52 Hz), 7.74 (1H, dd, J = 8.59 Hz, J = 1.78 Hz), 7.81 (1H, d, J = 8.60 Hz), 9.32 (1H, d, J = 1.20 Hz), 10.36 (1H, s). Anal. (C₂₉H₃₉FO₃Si₂) C, H: calcd, 68.19; found, C: 67.65.

7-(3-Fluoro-4-hydroxyphenyl)-3-hydroxy-1-naphthaldehyde (70). The title compound was prepared by reacting **156** (0.21 g, 0.41 mmol) with TBAF in THF according to method G to yield 0.080 g (68%) of a yellow solid: mp >250 °C; ¹H NMR (DMSO-*d*₆) δ 7.07–7.11 (1H, m), 7.43–7.46 (1H, m), 7.50 (1H, d, *J* = 2.46 Hz), 7.58 (1H, dd, *J* = 12.81 Hz, *J* = 2.20 Hz), 7.74 (1H, d, *J* = 2.58 Hz), 7.82 (1H, dd, *J* = 8.61 Hz, *J* = 1.88 Hz), 7.91 (1H, d, *J* = 8.67 Hz), 9.17–9.18 (1H, m), 10.03 (1H, s), 10.24 (1H, s), 10.45 (1H, s); MS (ESI) *m/z* 281 (M – H)⁻. Anal. (C₁₇H₁₁FO₃•0.5H₂O) C, H. **6-(3-Fluoro-4-hydroxyphenyl)-4-vinyl-2-naphthol (71).** The title compound was prepared by reacting **156** (1.74 g, 3.41 mmol) with methyl triphenylphosphonium bromide (2.08 g, 5.8 mmol) according to the procedure for **53** to yield 0.68 g (71%) of a yellow solid: ¹H NMR (DMSO- d_6) δ 5.48 (1H, dd, J = 10.92 Hz, J = 1.43 Hz), 5.82 (1H, dd, J = 17.22 Hz, J = 1.50 Hz), 7.02–7.07 9H, m), 7.12 (1H, d, J = 2.34 Hz), 7.26 (1H, d, J = 2.20 Hz), 7.66–7.70 (2H, m), 7.75 (1H, d, J = 8.57 Hz), 8.17 (1H, s), 9.78 (1H, s), 9.93 (1H, s); MS (ESI) *m/z* 279 (M – H)⁻. Anal. (C₁₈H₁₃FO₂·0.25H₂O) C, H.

4-Ethyl-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (72). The title compound was prepared by reacting **71** (0.57 g, 2.03 mmol) with hydrogen and Pd/C according to the procedure for **54** to yield 0.42 g (73%) of a white solid: mp 126–127 °C; ¹H NMR (DMSO- d_6) δ 1.31 (2H, t, J = 7.54 Hz), 3.08 (3H, q, J = 7.41 hz), 6.96–6.98 (2H, m), 7.03–7.08 (1H, m), 7.41–7.43 (1H, m), 7.58 (1H, dd, J = 12.87 Hz, J = 2.20 Hz), 7.64 (1H, dd, J = 8.58 Hz, J = 1.82 Hz), 7.72 (1H, d, J = 8.58 Hz), 8.04 (1H, d, J = 1.42 Hz), 9.63 (1H, s), 9.92 (1H, s); MS (ESI) *m*/*z* 283 (M – H)⁻; MS (ESI) *m*/*z* 281. Anal. (C₁₈H₁₅FO₂·0.1H₂O) C, H.

4-Ethynyl-6-(3-fluoro-4-hydroxyphenyl)-2-naphthol (68). 3-(tert-Butyldimethylsilanyloxy)-7-[4-(tert-butyldimethylsilanyloxy)-3-fluorophenyl]-1-(2,2-dibromovinyl)naphthalene. A mixture of **156** (0.51 g, 1.00 mmol) CBr₄, zinc dust, and triphenylphosphine was reacted according to the procedure used to prepare **55** to yield 0.61 g (92%) of the intermediate 3-(*tert*-butyldimethylsilanyloxy)-7-[4-(*tert*-butyldimethylsilanyloxy)-3-fluorophenyl]-1-(2,2-dibromovinyl)naphthalene as a white solid.

Treatment of this 2,2-dibromovinyl intermediate (0.40 g, 0.61 mmol) with *n*-butyllithium followed by deprotection as described in the preparation of **55** afforded 0.060 g (35%) of the desired product as a tan solid: mp 83–85 °C; ¹H NMR (DMSO-*d*₆) δ 4.63 (1H, s), 7.06–7.10 (1H, m), 7.24 (1H, d, *J* = 2.33 Hz), 7.30 (1H, d, *J* = 2.32 Hz), 7.38–7.40 (1H, m), 7.52 (1H, dd, *J* = 12.67 Hz, *J* = 2.20 Hz), 7.74 (1H, dd, *J* = 8.60 Hz, *J* = 0.88 Hz), 7.82 (1H, d, *J* = 8.53 Hz), 8.20 (1H, m), 10.01 (2H, s); MS (ESI) *m*/*z* 279 (M – H)⁻; MS (ESI) *m*/*z* 277. Anal. (C₁₈H₁₁FO₂·0.75H₂O) C, H.

6-(3-Fluoro-4-hydroxyphenyl)-4-prop-1-ynyl-2-naphthol (69). Treatment of **68** (0.25 g, 0.915 mmol) with TBDMS– Cl according to the procedure used to prepare compound **101** yielded 0.40 g (83%) of **157** as a white solid. The title compound was prepared by reacting **157** (0.40 g, 0.79 mmol) with *n*-butyllithium according to the procedure used to prepare **56** to yield 0.16 g (69%) of a gray solid: mp 124–126 °C; ¹H NMR (DMSO-*d*₆) δ 2.23 (3H, s), 7.05–7.10 (1H, m), 7.14 (1H, d, *J* = 2.20 Hz), 7.19 (1H, d, *J* = 2.46 Hz), 7.38–7.41 (1H, m), 7.52 (1H, dd, *J* = 12.68 Hz, *J* = 2.20 hz), 7.71 (1H, dd, *J* = 8.53 Hz, *J* = 1.94 Hz), 7.78 (1H, d, *J* = 8.53 Hz), 8.20 (1H), 9.89 (1H, s), 9.99 (1H, s); MS (ESI) *m/z* 291 (M – H)⁻. Anal. (C₁₉H₁₃FO₂·0.75H₂O) C, H.

Biological Assay Methods. Competitive Radioligand Binding Assay. This assay was performed as previously described.³⁰ Briefly, Human ER α and ER β ligand binding domains were expressed in *Escherichia coli*, and a crude cell lysate of these cells was used in a solid-phase binding assay. The radioligand was [³H]-17 β -estradiol, a ligand known to bind equally well to ER α and ER β .

Cell-Based Transcriptional Assay. Regulation of IG-FBP-4 mRNA was monitored in SAOS-2 cells engineered with an adenovirus to express human $\text{ER}\beta$ as previously described.³¹ Briefly, cells were treated for 24 h with test compound (typically 1 μ M) or 10 nM 17 β -estradiol and mRNA levels measured by real-time quantitative RT-PCR.

Rat Uterotrophic Assay. This assay was performed as previously described.³¹ Briefly, sexually immature rats (19 days old) were treated with **47** daily for 3 days and euthanized 24 h after the last dose. The vehicle was 50% DMSO/50% $1 \times$ Dulbecco's phosphate-buffered saline, and doses were administered subcutaneously in 0.2 mL. The doses given are expressed in milligram or microgram per rat per day, because

the rats typically grow from 38 to 55 g during the course of the study. Thus the dose of 2 mg/rat translates into ${\sim}36{-}53$ mg/kg and the 17 α -ethynyl-17 β -estradiol dose of 0.06 μ g/rat ranges between 1.1 and 1.6 μ g/kg.

HLA-B27 Transgenic Rat Model. Assessment of **47** effects on intestinal endpoints was performed as previously described.¹⁵ To assess the ability of **47** to prevent joint swelling, 8–10 week old male rats were used. These rats were treated with daily oral doses of **47** (10 mg/kg) in a vehicle of 2% Tween-80/0.5% methylcellulose for 46 days. Gross evaluation of joint swelling was scored as previously described.¹⁵

Lewis Rat Adjuvant-Induced Arthritis Model. This model was performed as previously described.¹⁵ Briefly, male Lewis rats were injected intradermally with Freunds Complete Adjuvant at the base of the tail. After full joint inflammation developed (8 days after adjuvant injection), vehicle or 47 was administered in daily oral doses. Joint redness and swelling were evaluated daily, and after euthanasia, joint histology was evaluated (synovitis and Mankin scores). At necropsy, blood samples were collected and serum prepared. Haptoglobin concentrations were assessed using a commercially available kit (TriDelta Diagnostics), which measures the inherent peroxidase activity of hemoglobin that is preserved at low pH when haptoglobin is present. The assay was performed according to the manufacturer's directions, except that a kinetic (V_{max}) rather than endpoint measurement was used. Plates were read in a microplate reader at A_{595} (Molecular Devices).

Cell-Based Selectivity Assays. These assays were performed as previously described. 35

SHBG Binding Assay. Human blood was collected in SST VACUTAINER tubes (Becton-Dickenson) and kept at room temperature for 30 min before centrifugation at 1000g for 10 min (4 °C). The serum was aliquoted and stored at -80 °C. Human serum was diluted with TG buffer (10 mM Tris and 10% glycerol, pH 7.4, at room temperature) $20{-}100{\text{-}fold}$ to achieve a total binding signal of about 3000 cpm. Seven hundred microliters of dextran-coated charcoal (1% charcoal, 0.05% dextran 69K in Tris/EDTA buffer, pH 7.4) was added to every 1000 μ L of diluted serum, and the mixture was incubated on ice for 1 h, followed by centrifugation at 2000g for 20 min. The SHBG binding assay was performed with 100 μ L of this supernatant solution, 25 μ L [1,2-N-³H]-5 α -dihydrotestosterone ($[^{3}H]$ DHT; final concentration = 8 nM), and 25 μ L of various concentrations of 47. Following overnight incubation at 4 °C, free and bound [3H]DHT were separated by using 100 μ L of 0.5% dextran-coated charcoal followed by centrifugation at 1000g for 5 min. Bound [³H]DHT (150 μ L) was counted in a Beckman LS6500 scintillation counter (Beckman Instruments, Inc).

Crystallography. Human ER β LBD was cloned, expressed, and purified as previously described.^{14,26} The ER β complexes were concentrated to approximately 10 mg/mL in a buffer containing 0.2 M NaCl, 1 mM EDTA, 5 mM DTT, and 10 mM Tris-HC1 at pH 7.5 and mixed with a nuclear receptor box peptide Biotin-SGSHKLVQLLTTT-COOH (derived from steroid receptor coactivator-1)^{36,37} at a molar ratio of 1.5:1 peptide to protein–ligand complex. Screening of crystallization conditions was performed at 18 °C using the hanging drop vapor diffusion method.³⁸ Crystals were grown from a drop containing a mixture of protein–ligand solution and reservoir solution of 20% PEG3350 (v/v), 0.2 M Mg formate, pH 5.9.

X-ray data were collected at 100 K using Quantum-4 CCD area detector at the Advanced Light Source (ALS, Berkeley, CA) and processed using DENZO and Scalepack.³⁹ Crystal structures were solved by molecular replacement using the program AMORE.⁴⁰ All complex structures were solved using ER β complexed with genistein as a search model. This ER β / genistein structure was in turn solved in-house by molecular replacement, using ER α complexed with E2 as a model⁴¹ (pdb code 1A52), and was later found to be in good agreement with the published structure¹⁷ (pdb code 1QKM). To avoid model bias, the ligand, the loop connecting H8–H9, the C and N terminal helices, and the coactivator peptide were omitted from the search models. Structures were refined using the program

Table 12.X-ray Data Collection and Refinement Statistics forLigands 15 and 47

	15	47
Data Collection	ı	
space group	$P2_{1}2_{1}2_{1}$	
wavelength (Å)	1.10	
unit cell dimensions		
a (Å)	51.970	52.075
b (Å)	87.677	88.504
c (Å)	99.760	100.139
maximal resolution (Å)	2.7	2.03
observations	12673	30401
completeness (%)	96.1	98.5
$R_{ m merge}{}^a$	0.081	0.073
mean I/σ (I)	17.38	18.21
reflections used	12278	29584
reflections in working set	11652	28130
reflections in test set	626	1454
highest resolution bin (Å)	2.8 - 2.7	2.10 - 2.03
completeness (%)	89.5	95.7
$R_{ m merge}{}^a$	0.357	0.489
mean I/σ (I)	3.38	2.80
mosaicity (deg)	0.947	0.467
Refinement		
no. of molecules per asymmetric unit	2	2
protein atoms ^b	3546	3575
other atoms		
ligand ^b	38	42
coactivator peptide b	142	162
water	102	219
resolution range (Å)	15 - 2.2	15 - 2.0
$R_{ m work}{}^c$	0.208	0.222
$R_{\rm free}^{d}$	0.271	0.281
rms bond length ^e (Å)	0.006	0.0067
rms bond angles ^e (deg)	1.033	1.094
mean <i>B</i> -factor (Å ²)		
complex	36.2	26.1
main-chain atoms	35.9	24.9
side-chain atoms	37.0	27.8
ligand	35.7	21.0
water	31.2	28.7
rms backbone ΔB (Å $^2)^f$	1.00	1.15
%A, B, L (a, b, l, p) ^g	100	100

 $^{a}R_{\rm merge} = \sum_{n} \sum_{hkl} |I_{hkl}{}^{n} - \langle I_{hkl}{}^{n} \rangle | / \sum_{n} \langle I_{hkl}{}^{n} \rangle$, where $I_{hkl}{}^{n}$ is the nth observation of reflection hkl, and $\langle I_{hkl}{}^{n} \rangle$ denotes an average of reflection hkl over n observations. b Per monomer unit. c Crystallographic R-factors were computed using $R_{\rm work} = \sum_{hkl} |F_{hkl}{}^{\rm obs} - F_{hkl}{}^{\rm calc} | / \sum_{hkl} F_{hkl}{}^{\rm obs} \cdot d R_{\rm free}$ values were calculated in the same manner as $R_{\rm work}$, except over approximately 4% of the data excluded from the refinement. e Root-mean-square deviation in bond length and bond angle distances from the Engh and Huber ideal values. f Root-mean-square deviation between B-factors for bonded main chain atoms. g Percentage of residues located in most favored (additional) regions of the Ramachandran plot as determined by PROCHECK.^{44}

CNS.⁴² The resulting difference electron density maps show clear electron density for the compounds, residues within the binding site, helix 12, and the coactivator peptide. Cysteine modifications and some flexible loop residues were not included in the models due to poor electron density. Table 12 gives the data collection and refinement details for all of the complexes studied. Atomic coordinates for ER β complexed with compounds **15** and **47** have been deposited in the Protein Data Bank, with accession codes 1YY4 and 1YYE, respectively.

Computational Methods. Docking calculations were performed using the QXP software package.⁴³ The QXP Monte Carlo docking algorithm mcdock was used to generate potential binding modes in the active site. In general, 1000 Monte Carlo steps was sufficient for the poses and their energy scores to converge. Constrained residue flexibility was utilized to take subtle movements of the pocket into account when necessary. Visualization of X-ray structures and docking results was performed using the InsightII and Quanta software packages (Accelrys, Inc., San Diego, CA). Protein overlays were performed using the homology module of the InsightII software package. After performing a preliminary structural alignment based on all alpha-carbons, the superimpose_aln command was used to generate matches based on fitting only α -carbons closer than 2.00 Å. This ensured that conformational changes in highly flexible regions such as loops did not exert an undesired influence on the overlays.

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Supporting Information Available: Experimental, analytical, and spectroscopic data for compounds 9–14, 25–30, 32–37, 77, 78, 80, 82, 84, 95, 98, 100, 102–105 and elemental analysis data. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- Green, S.; Walter, P.; Kumar, V.; Krust, A.; Bornert, J. M.; Argos, P.; Chambon, P. Human oestrogen receptor cDNA: Sequence, expression and homology to v-erb-A. *Nature* **1986**, *320*, 134– 139.
- (2) Kuiper, G. G. J. M.; Enmark, E.; Pelto Huikko, M.; Nilsson, S.; Gustafsson, J. A. Cloning of a novel estrogen receptor expressed in rat prostate and ovary. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 5925–5930.
- (3) Mosselman, S.; Polman J.; Dijkema, R. ER beta: Identification and characterization of a novel human estrogen receptor. *FEBS Lett.* **1996**, *392*, 49–53.
- (4) Couse, J. F.; Lindzey, J.; Grandien, K.; Gustafsson, J. A.; Korach, K. S. Tissue distribution and quantitative analysis of estrogen receptor-alpha (ERalpha) and estrogen receptor-beta (ERbeta) messenger ribonucleic acid in the wild-type and ERalpha-knockout mouse. *Endocrinology* **1997**, *138*, 4613–4621.
- (5) Fitzpatrick, S. L.; Funkhouser, J. M.; Sindoni, D. M.; Stevis, P. E.; Deecher, D. C.; Bapat, A. R.; Merchenthaler, I.; Frail, D. E. Expression of estrogen receptor-beta protein in rodent ovary. *Endocrinology* **1999**, *140*, 2581–2591.
 (6) Kuiper, G. G. J. M.; Carlsson, B.; Grandian, K.; Enmark, E.;
- (6) Kuiper, G. G. J. M.; Carlsson, B.; Grandian, K.; Enmark, E.; Haggblad, J.; Nilsson, S.; Gustafsson, J. A. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptor alpha and beta. *Endocrinology* **1997**, *138*, 863– 870.
- (7) Yang, C. J.; Edsall, R.; Harris, H. A.; Zhang, X. C.; Manas, E. S.; Mewshaw, R. E. ERβ ligands. Part 2: Synthesis and structure-activity relationships of a series of 4-hydroxy-biphenyl-carbaldehyde oxime derivatives. *Bioorg. Med. Chem.* 2004, 12, 2553-2570.
- (8) Edsall, R. J.; Harris, H. A.; Manas, E. S.; Mewshaw, R. E. ER beta ligands. Part 1: The discovery of ER β selective ligands which embrace the 4-hydroxy-biphenyl template. *Bioorg. Med. Chem.* **2003**, *11*, 3457–3474.
- (9) Meyers, M. J.; Sun, J.; Carlson, K. E.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. Estrogen receptor subtype-selective ligands: Asymmetric synthesis and biological evaluation of cisand *trans*-5, 11-dialkyl-5,6,11,12-tetrahydrochrysenes. J. Med. Chem. 1999, 42, 2456-2468.
- (10) Meyers, M. J.; Sun, J.; Carlson, K. E.; Marriner, G. A.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. Estrogen receptor-beta potency-selective ligands: Structure-activity relationship studies of diarylpropionitriles and their acetylene and polar analogues. J. Med. Chem. 2001, 44, 4230-4251.
- (11) Schopfer, U.; Schoeffter, P.; Bischoff, S. F.; Nozulak, J.; Feuerbach, D.; Floersheim, P. Toward selective ER beta Agonists for central nervous system disorders: Synthesis and characterization of aryl benzthiophenes. J. Med. Chem. 2002, 45, 1399–1401.
- (12) Henke, B. R.; Consler, T. G.; Go, N.; Hale, R. L.; Hohman, D. R.; Jones, S. A.; Lu, A. T.; Moore, L. B.; Moore, J. T.; Orband-Miller, L. A.; Robinett, R. G.; Shearin, J.; Spearing, P. K.; Stewart, E. L.; Turnbull, P. S.; Weaver, S. L.; Williams, S. P.; Wisely, G. B.; Lambert, M. H. A new series of estrogen receptor modulators that display selectivity for estrogen receptor beta. J. Med. Chem. 2002, 45, 5492-5505.
- (13) Wallace, O. B.; Richardson, T. I.; Dodge, J. A. Estrogen receptor modulators: Relationships of ligand structure, receptor affinity and functional activity. *Curr. Top. Med. Chem.* **2003**, *3*, 1663– 1682.

- (14) Malamas, M. S.; Manas, E. S.; McDevitt, R. E.; Gunawan, I.; Zhang, X. B.; Collini, M. D.; Miller, C. P.; Dinh, T.; Henderson, R. A.; Keith, J. C., Jr.; Harris, H. A. Design and synthesis of aryl di-phenolic azoles as potent and selective estrogen receptor-b
- aryı ur-pnenonc azores as potent and selective estrogen receptor-b ligands. J. Med. Chem. 2004, 47, 5021-5040.
 (15) Harris, H. A.; Albert, L. M.; Leathurby, Y.; Malamas, M. S.; Mewshaw, R. E.; Miller, C. P.; Kharode, Y. P.; Marzolf, J.; Komm, B. S.; Winneker, R. C.; Frail, D. E.; Henderson, R. A.; Zhu, Y.; Keith, J. C., Jr. Evaluation of an estrogen receptor-bate agonist in animal models of humon discoge formation. beta agonist in animal models of human disease [comment]. Endocrinology 2003, 144, 4241-4249.
- (16) Kuiper, G.; Lemmen, J. G.; Carlsson, B.; Corton, J. C.; Safe, S. H.; Vandersaag, P. T.; Vanderburg, P.; Gustafsson, J. A. Interac-tion Of Estrogenic Chemicals and Phytoestrogens With Estrogen Receptor Beta. Endocrinology 1998, 139, 4252-4263.
- Pike, A. C. W.; Brzozowski, A. M.; Hubbard, R. E.; Bonn, T.; Thorsell, A. G.; Engstrom, O.; Ljunggren, J.; Gustafsson, J. K.; (17)Carlquist, M. Structure of the ligand-binding domain of oestrogen receptor beta in the presence of a partial agonist and a full
- antagonist. *EMBO J.* **1999**, *18*, 4608–4618. Fang, H.; Tong, W.; Shi, L. M.; Blair, R.; Perkins, R.; Branham, W.; Hass, B. S.; Xie, Q.; Dial, S. L.; Moland, C. L.; Sheehan, D. (18)M. Structure-Activity Relationships for a Large Diverse Set of Natural, Synthetic, and Environmental Estrogens. Chem. Res. Toxicol. 2001, 14, 280-294.
- (19) Ferte, J.; Kuhnel, J. M.; Chapuis, G.; Rolland, Y.; Lewin, G.; Schwaller, M. A. Flavonoid-related modulators of multidrug resistance: Synthesis, pharmacological activity, and structureactivity relationships. J. Med. Chem. 1999, 42, 478-489.
- (20) Kozerski, L.; Kamienski, B.; Kawecki, R.; Urbanczyk-Lipkowska, Z.; Bocian, W.; Bednarek, E.; Sitkowski, J.; Zakrzewska, K.; Nielsen, K. T.; Hansen, P. E. Solution and solid state 13C NMR and X-ray studies of genistein complexes with amines. Potential biological function of the C-7, C-5, and C-4'-OH groups. Org. Biomol. Chem. 2003, 1, 3578–3585.
 (21) Nishiyama, T.; Ogura, K.; Nakano, H.; Kaku, T.; Takahashi, E.;
- Ohkubo, Y.; Sekine, K.; Hiratsuka, A.; Kadota, S.; Watabe, T. Sulfation of environmental estrogens by cytosolic human sulfotransferases. Drug Metab. Pharmacokinet. 2002, 17, 221-228.
- (22) Michalak, K.; Hendrich, A. B.; Wesolowska, O.; Pola, A. Compounds That Modulate Multidrug Resistance In Cancer Cells.
- pounds That Modulate Multidrug Resistance In Cancer Cells. Cell. Biol. Mol. Lett. 2001, 6, 362-368.
 (23) Mazurek, A. P.; Kozerski, L.; Sadlej, J.; Kawecki, R.; Bednarek, E.; Sitkowski, J.; Dobrowolski, J. C.; Maurin, J. K.; Biniecki, K.; Witowska, J.; Fiedor, P.; Pachecka, J. Genistein complexes with amines: Structure and properties. J. Chem. Soc., Perkin Trans 2: Phys. Org. Chem. 1998, 1223-1230.
 (24) Anstead, G. M.; Carlson, K. E.; Katzenellenbogen, J. A. The estradiol pharmacophore: Ligand structure-estrogen receptor
- estradiol pharmacophore: Ligand structure-estrogen receptor binding affinity relationships and a model for the receptor
- binding site. Steroids 1997, 62, 268–303.
 (25) Dodge, J. A.; Glasebrook, A. L.; Lugar, C. W. Preparation of 5-(4aminoalkoxybenzyl)-6-(4-hydroxyphenyl)-2-naphthols and analogues as antiestrogens. In Eur. Pat. Appl. (Eli Lilly and Company, USA). EP 895989 A1, 1999; 35 pp.
 (26) Manas, E. S.; Unwalla, R. J.; Xu, Z. B.; Malamas, M. S.; Miller,
- C. P.; Harris, H. A.; Hsiao, C.; Akopian, T.; Hum, W.-T.; Malakian, K.; Wolfrom, S.; Bapat, A.; Bhat, R. A.; Stahl, M. L.; Somers, W. S.; Alvarez, J. C. Structure-Based Design Of Estrogen Receptor-b Selective Ligands. J. Am. Chem. Soc. 2004, 126, 15106-15119.
- (27) Brzozowski, A. M.; Pike, A. C. W.; Dauter, Z.; Hubbard, R. E.; Bonn, T.; Engstrom, O.; Ohman, L.; Greene, G. L.; Gustafsson, J.-A.; Carlquist, M. Molecular basis of agonism and antagonism in the oestrogen receptor. Nature 1997, 389, 753-758.

- (28) Warnmark, A.; Treuter, E.; Gustafsson, J. A.; Hubbard, R. E.; Brzozowski, A. M.; Pike, A. C. W. Interaction of transcriptional intermediary factor 2 nuclear receptor box peptides with the coactivator binding site of estrogen receptor alpha. J. Biol. Chem. 2002, 277, 21862-21868.
- Shiau, A. K.; Barstad, D.; Radek, J. T.; Meyers, M. J.; Nettles, K. W.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A.; Agard, (29)D. A.; Greene, G. L. Structural characterization of a subtypeselective ligand reveals a novel mode of estrogen receptor antagonism. *Nature Struct. Biol.* **2002**, *9*, 359–364. (30) Harris, H. A.; Bapat, A. R.; Gonder, D. S.; Frail, D. E. The ligand
- binding profiles of estrogen receptors alpha and beta are species dependent. *Steroids* **2002**, *67*, 379–384.
- (31) Harris, H. A.; Katzenellenbogen, J. A.; Katzenellenbogen, B. S. Characterization of the biological roles of the estrogen receptors, ER alpha and ER beta, in estrogen target tissues in vivo through the use of an ER alpha-selective ligand. Endocrinology 2002,
- 143, 4172–4177. (32) Bodine, P. V. N.; Harris, H. A.; Lyttle, C. R.; Komm, B. S. Estrogenic effects of 7 alpha-methyl-17 alpha-ethynylestradiol: A newly discovered tibolone metabolite. Steroids 2002, 67, 681 686
- (33) Miller, C. P.; Komm, B. S. Targeting the estrogen receptor with SERMs. Ann. Rep. Med. Chem. 2001, 36, 149-158.
- (34)Taurog, J. D.; Maika, S. D.; Satumtira, N.; Dorris, M. L.; McLean, I. L.; Yanagisawa, H.; Sayad, A.; Stagg, A. J.; Fox, G. M.; Le O'Brien, A.; Rehman, M.; Zhou, M.; Weiner, A. L.; Splawski, J. B.; Richardson, J. A.; Hammer, R. E. Inflammatory disease in HLA-B27 transgenic rats. Immunolog. Rev. 1999, 169, 209-223.
- (35) Zhang, Z. M.; Lundeen, S. G.; Zhu, Y.; Carver, J. M.; Winneker, R. C. In vitro characterization of trimegestone: A new potent and selective progestin. Steroids 2000, 65, 637-643.
- Heery, D. M.; Hoare, S.; Hussain, S.; Parker, M. G.; Sheppard, H. Core LXXLL motif sequences in CREB-binding protein, SRC1 and RIP140 define affinity and selectivity for steroid and retinoid words, C. W.; Komm, B.; Cheskis, B. J. Structure-function
- evaluation of ER alpha and beta interplay with SRC family coactivators. ER selective ligands. Biochemistry 2001, 40, 6756-6765.
- (38)McPherson, A., Jr. The growth and preliminary investigation of protein and nucleic acid crystals for X-ray diffraction analysis. Methods. Biochem. Anal. 1976, 23, 249-345.
- Otwinowski, Z.; Minor, W. Processing of X-ray diffraction data (39)collected in oscillation mode. Methods. Enzymol. 1994, 276, 307
- Bailey, S. The CCP4 suite: Programs for protein crystallography. Acta Crystallogr., Sect. D 1994, 50, 760–763. Tanenbaum, D. M.; Wang, Y.; Williams, S. P.; Sigler, P. B. (40)
- (41)Crystallographic comparison of the estrogen and progesterone receptor's ligand binding domains. Proc. Natl. Acad. Sci. U.S.A. **1998**, *95*, 5998–6003.
- (42) Brunger, A. Crystallography and NMR system: A new sofware suite for macromolecular structure determination. Acta Crystallogr., Sect. D 1998, 55, 941-944.
- McMartin, C.; Bohacek, R. S. QXP: Powerful, rapid computer (43)algorithms for structure-based drug design. J. Comput.-Aided Mol. Des. 1997, 11, 333-344.
- Laskowski, R. A.; MacArthur, M. W.; Moss, D. S.; Thornton, J. (44)M. PROCHECK: A program to check the stereochemical quality of protein structures. J. Appl. Crystallogr. 1993, 26, 283-291.

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