

Synthesis and antiviral activities of spacer-linked 1-thioglucuronide analogues of glycyrrhizin

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Full Research Paper

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Abstract

The influenza virus infection remains a significant threat to public health and the increase of antiviral resistance to available drugs generates an urgent need for new antiviral compounds. Starting from the natural, antivirally active compound glycyrrhizin, spacerbridged derivatives were generated with improved antiviral activity against the influenza A virus infection. Simplified analogues of the triterpene saponin glycyrrhizin containing 1-thio- β -D-glucuronic acid residues have been prepared in good yields by alkylation of 3-amino and 3-thio derivatives of glycyrrhetinic acid with a 2-iodoethyl 1-thio- β -D-glucopyranosiduronate derivative. The spacer-connected 3-amino derivatives were further transformed into N-acetylated and N-succinylated derivatives. The deprotected compounds containing these carboxylic acid appendices mimic the glycon part of glycyrrhizin as well as the hemisuccinate derivative of glycyrrhetinic acid, carbenoxolone. Antiviral activities of the compounds were determined in a biological test based on influenza A virus-infected cells, wherein the 3-(2-thioethyl)-*N*-acetylamino- and 3-(2-thioethyl)-thio-linked glucuronide derivatives were effective inhibitors with IC₅₀ values as low as 54 μ M.

Introduction

The triterpene saponin glycyrrhizin (GL) and its aglycon glycyrrhetinic acid (GA) are the main triterpene components of licorice roots and harbor various pharmacological activities, including antitumor, anti-inflammatory, antioxidant and antiviral properties [1,2]. The antiviral activities have been reported to be directed against a broad spectrum of viruses comprising herpes-, corona-, alpha-, and flaviviruses, HIV, Epstein–Barr virus, influenza A virus (IAV), vaccinia and



polio type I viruses as examples [3-8]. In particular antiinfluenza virus activities have been described, although the underlying mechanisms of action are diverse [9-12]. In order to evaluate the potential antiviral properties of glycyrrhizin derivatives, we set out to modify the chemical environment in the vicinity of ring A of glycyrrhetinic acid. The envisaged modifications should assess the impact of acidic groups, which reside at the glucuronic acid residues in the β -(1 \rightarrow 2)-linked disaccharide unit of **GL**, with respect to antiviral properties (Figure 1).

Previously, ring A modified derivatives of glycyrrhetinic acid were prepared containing 3-amino, 3-thio and 1-thio groups, allowing for an extension of the oleanolic acid unit with spacer groups as well as facile covalent attachment of carboxyalkyl groups by alkylation or acylation reactions [13,14]. For the compounds included in this investigation, the size of the spacer group was selected to closely match the formal distances between the A-ring and the carboxylic acid function present in the glucopyranosiduronic residues of the parent compound glycyrrhizin (GL) and in the hemisuccinate moiety of the glycyrrhetinic acid derivative carbenoxolone (CBX) [15] (Figure 1). For the sake of clarity, the nomenclature and numbering for the triterpene system as used throughout this report is illustrated in Figure 1. In order to enhance resistance against enzymatic cleavage by glucuronidases, the glucuronic acid residue was introduced through a stable thioglycosidic linkage.

Results and Discussion Synthesis of 3-amino derivatives

Commercially available methyl (2,3,4-tri-*O*-acetyl-D-glucopyranosyl)uronate bromide (1) was first reacted with potassium thioacetate in DMF to furnish the known 1-thioacetyl derivative 2 in 82% yield [16]. Subsequent treatment of 2 with sodium methoxide, under controlled conditions at low temperature (-60 °C \rightarrow -45 °C), provided the corresponding glucuronyl 1-thiol 3 in 89% yield [17,18]. Reaction of 3 with an excess of 1,2-dibromoethane (3-4 equiv) in DMF in the presence of sodium hydride, with the strict exclusion of oxygen, afforded the 2-bromoethyl 1-thioglycoside 4 in 80% yield. Under these conditions formation of the bis-substitution product 5 (3%) and the disulfide oxidation product of 1-thiol 3 was observed in only very minor quanities. The corresponding 2-iodoethyl derivative 6 was prepared by a Finkelstein reaction from the bromoethyl 1-thioglycoside 4 in 95% yield (Scheme 1).

The previously reported 3β -amino derivative of the diphenylmethyl ester of glycyrrhetinic acid 7 [19] was then used for the introduction of the spacer-extended 1-thio-glucopyranosiduronate residue. Coupling of the 3β -amino derivative 7 with two equivalents of the 2-iodoethyl glycoside 6 in DMF in the presence of Hünig base proceeded smoothly to provide the alkylated amine 8 in 79% yield without the formation of a bisalkylated product (Scheme 2), whereas the use of the corresponding bromide 4 resulted in a significantly slower reaction.





Scheme 2: Synthesis of thioalkylglucuronide **GA** derivatives: (a) DMF, DIPEA, 45-50 °C, 16 h, 79%; (b) TEA, Ac₂O, DCM, 0 °C, 2 h, 96%; (c) anisole, TFA, DCM, rt, 3 h (75% for **10**, 89% for **11**); (d) 0.1 M NaOMe, MeOH, 0 °C (89% for **12**, 97% for **14**, 99% for **18**); (e) 0.2 M NaOH, MeOH, rt, 3 h (91% for **13**, ~quant. for **15**, 99% for **19**); (f) MeO₂C(CH₂)₂COCI in DCM, DIPEA, 0 °C to rt, 90 min, then anisole, TFA, DCM, 0 °C, 3 h, 83% for two steps.

At elevated temperature, elimination leading to the corresponding hexenuronic acid derivative was observed (structure not shown). The secondary amino group in compound **8** was subjected to further derivatization by N-acetylation (triethylamine, acetic anhydride), which gave compound **9** in 96% yield. Deprotection of the ethyl 1-thio-glucuronide derivatives **8** and **9** was achieved by acid-catalyzed cleavage of the diphenylmethyl ester group with TFA/anisole (as carbocation scavenger), giving the monoacid derivatives **10** and **11** in 75% and 89% yield, respectively. Subsequent transesterification of **10** and **11** with methanolic NaOMe afforded the deacetylated methyl ester glucuronide derivatives **12** and **14**, respectively, which were finally fully deprotected by hydrolysis of the methyl ester group with 0.2 M methanolic NaOH to furnish the diacid derivatives **13** and **15**, in high overall yield.

A second series of compounds was designed as glucuronideextended amide derivatives of the previously reported carbenoxolone analogue 16 [20,21]. Reaction of the 3-amino group of 8 with the monomethyl ester of succinic acid chloride in dichloromethane in the presence of Hünig base was followed by deblocking of the diphenylmethyl ester group to afford 17 in a combined yield of 83%. The acetyl groups of 17 were removed under Zemplén conditions to afford the dimethyl ester 18. Alkaline hydrolysis of the ester groups eventually gave the triacid derivative 19 in good yield. In contrast to the straightforward chemical transformations, analysis of the NMR spectra of this series of compounds was challenging and the N-acylated derivatives 9, 11, 14, 15 and 17-19 revealed complex spectra. Whereas the secondary amides of 3\beta-amino-glycyrrhetinic acid derivatives displayed coherent NMR signals [22,23], the tertiary amides frequently exist as mixtures of cis/trans rotamers leading to the duplication of signals [24,25]. The steric congestion at position 3 exerted by the two adjacent methyl groups may additionally contribute to the restricted rotation of the amide linkage. Thus, in the ¹H NMR spectra of the N-acetyl-



Figure 2: 400 MHz ¹H NMR expansion plots of the carbohydrate region of compound 11, recorded at various temperatures.

and *N*-succinyl derivatives, recorded at room temperature, two sets of signals in an approximate 6:4 ratio were observed. Reversible coalescence of these signal groups was observed upon heating a solution of the *N*-acetyl derivative **11** up to 100 °C in DMSO- d_6 and subsequent cooling to room temperature (Figure 2), supporting the assignment of two signal data sets for two rotamers. The orientation of the amide carbonyl group in the rotamers also had a pronounced effect on the NMR chemical shifts of carbon and proton signals in ring A, which led to a shift of H-3 to lower field (δ 4.47) and to a shielding of C-3 (δ 59.9) for the minor isomer, whereas the inverse effects were seen for the major rotamer (δ for H-3 at 3.36 and δ for C-3 at 66.7). Similar, but less-pronounced effects were observed for the respective ¹H and ¹³C signals at position 5 of ring A (see Supporting Information File 1).

Synthesis of 3-thio derivatives

Complementing the series of the 3β -amino series, related 3-thio derivatives were employed for the spacer elongation [14]. The previously reported 1,2-dehydro-3-thiol derivative **20** was subjected to alkylation with the 2-iodoethyl 1-thioglucuronide compound **6** in the presence of K₂CO₃ to furnish the thioetherbridged glucuronide triterpene **21** in 60% yield. Deprotection of

21 was performed in two steps as described for **10**, which furnished the methyl ester derivative **22** and the glucuronic acid compound **23**, respectively, in good yields (Scheme 3).

Antiviral activities

For biological testing the compounds were dissolved in DMSO and diluted in cell culture medium to a maximum final DMSO concentration of 1%. For determination of the cytotoxicity of the compounds, MDCK cells were treated with compound concentrations ranging from 3.1 µM to 250 µM for 48 h and then cell viability was determined as a surrogate endpoint for cytotoxicity. Except for the aglycons of GL, namely glycyrrhetinic acid (GA) and the hemisuccinate carbenoxolone (CBX), which showed CC50 values (half-maximal cytotoxic concentration) of 7.4 µM and 17.8 µM, respectively, none of the compounds tested were toxic at concentrations up to 250 µM. GL was not toxic at 2500 µM (Table 1). The antiviral activity of the compounds was determined by infecting MDCK cells with influenza A H3N2 virus and treating them with different concentrations of compounds at the same time. Most compounds prevented virus-induced cytopathicity and restored cell viability compared to virus-infected control cells. The antiviral activity of glycyrrhizin (GL) against influenza and other viruses



reported in the literature was confirmed in this study resulting in an IC₅₀ of 1026 μ M (Table 1). **GA** and **CBX** did not show any antiviral activity at nontoxic concentrations. Compounds **12**, **13**, **14**, **15**, **18** and **19**, which possess a (1-thio- β -D-glucopyranosyluronic acid)ethylamino substituent at position 3 of **GA** did not show toxicity at concentrations up to 250 μ M, but showed enhanced antiviral activity compared to **GL**. Good activities were found for compounds **12** and **18** with IC₅₀ values of 220.4 μ M and 125 μ M, respectively, being between 4.7-fold to 8-fold more active compared to the lead compound **GL**. The methyl ester **14** containing the N-acetylated spacer group was approximately 14-fold more active compared to **GL**, reflected by an IC₅₀ value of 72.1 μ M. The free acid **15** displayed an IC₅₀ value in the same range as the methyl ester **14**. Replacement of the *N*-acetyl group of **14** with a methylsuccinyl group (18) did not improve the activity; however, the compound was still active with an IC_{50} value of 125 μ M. The cleavage of both methyl ester groups of this compound resulted in compound 19 and led to the loss of antiviral activity up to the highest concentration tested (250 μ M). Compound 16 containing only the *N*-succinyl substituent is a close analogue to CBX bearing an amide instead of the ester group. Unlike CBX, which is toxic at low concentrations, 16 was not toxic but inactive, indicating that the glucuronic acid moiety is associated with the antiviral properties. This was further supported by the data obtained with compound 23 containing a 1-thioethyl-linked glucuronic acid residue at position 3 of 1,2-dehydro-glycyrrhetinic acid. Compound 23 showed good activity with an even lower IC_{50} value of 54 μ M compared to 15, and exhibited also no cytotoxicity at the concentrations tested.

 Table 1: Cytotoxic concentration 50% (CC_{50}) of compound-treated uninfected cells and antiviral activities, shown as half-maximal inhibitory concentration (IC_{50}) values of glycyrrhizin (**GL**) analogues.

Compound	Substructure	CC ₅₀ [µM]	IC ₅₀ [μM]
GL	$HO_2C O O O O O O O O O O O O O O O O O O O$	>2500	1026 ± 181
GA	HO	7.4 ± 0.05	nd ^a
СВХ	HO	17.8 ± 2.3	nd ^a
12	HO OH S H	>250	220.4 ± 41

Table 1: Cytotoxic condtration (IC_{50}) values of g	entration 50% (CC ₅₀) of compound-treated uninfecte llycyrrhizin (GL) analogues. (continued)	ed cells and antiviral activities, show	n as half-maximal inhibitory concen-
13	HO ₂ C HO HO OH S N H	>250	nd (>250) ^b
14	MeO ₂ C HO HO OH OH	>75	72.1 ± 19.1 ^c
15	HO ₂ C HO HO OH OH	>250	87 ± 7.9
16	HOUND	>250	nd (>250) ^b
18	MeO ₂ C HO HO OH MeO ₂ C	>250	125 ± 15.9
19	HO ₂ C HO HO OH HO ₂ C	>250	nd (>250) ^b
23	HO ₂ C HO HO OH S S ^{rat}	>250	54 ± 13

^and not determined due to cytotoxicity; ^bnd (>250) no detectable antiviral activity up to 250 μ M; ^capproximate value, since the compound precipitates at concentrations >75 μ M.

Conclusion

3-Amino and 3-thio derivatives of glycyrrhetinic acid served as versatile scaffolds for the attachment of glycosyl extensions through high-yielding alkylation reactions. The glucuronide derivatives linked via a thioethyl spacer group to the oleanolic acid unit exhibited no toxicity at concentrations up to 250 μ M and significantly enhanced the anti-influenza virus activity of the natural triterpene glycoside glycyrrhizin.

Supporting Information

Supporting Information File 1

Experimental details for the preparation of compounds **2–19** and **21–23** as well as the biological assays. [http://www.beilstein-journals.org/bjoc/content/ supplementary/1860-5397-8-79-S1.pdf]

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References

- 1. Baltina, L. A. Curr. Med. Chem. 2003, 10, 155–171.
- Schwarz, S.; Csuk, R. *Bioorg. Med. Chem.* 2010, *18*, 7458–7474. doi:10.1016/j.bmc.2010.08.054
- Lampis, G.; Deidda, D.; Pinza, M.; Pompei, R. Antiviral Chem. Chemother. 2001, 12, 125–131.
- Cinatl, J.; Morgenstern, B.; Bauer, G.; Chandra, P.; Rabenau, H.; Doerr, H. W. *Lancet* 2003, *361*, 2045–2046. doi:10.1016/S0140-6736(03)13615-X

- Crance, J. M.; Scaramozzino, N.; Jouan, A.; Garin, D. Antiviral Res. 2003, 58, 73–79. doi:10.1016/S0166-3542(02)00185-7
- Hattori, T.; Ikematsu, S.; Koito, A.; Matsushita, S.; Maeda, Y.;
 Hada, M.; Fujimaki, M.; Takatsuki, K. *Antiviral Res.* **1989**, *11*, 255–261. doi:10.1016/0166-3542(89)90035-1
- 7. Harada, S. Biochem. J. 2005, 392, 191-199. doi:10.1042/BJ20051069
- Lin, J.-C. Antiviral Res. 2003, 59, 41–47. doi:10.1016/S0166-3542(03)00030-5
- Pompei, R.; Paghi, L.; Ingianni, A.; Uccheddu, P. *Microbiologica* 1983, 6, 247–250.
- 10. Utsunomiya, T.; Kobayashi, M.; Pollard, R. B.; Suzuki, F. *Antimicrob. Agents Chemother.* **1997**, *41*, 551–556.
- Wolkerstorfer, A.; Kurz, H.; Bachhofner, N.; Szolar, O. Antiviral Res. 2009, 83, 171–178. doi:10.1016/j.antiviral.2009.04.012
- Michaelis, M.; Geiler, J.; Naczk, P.; Sithisam, P.; Leutz, A.; Doerr, H. W.; Cinatl, J., Jr. *PLoS One* **2011**, *6*, e19705. doi:10.1371/journal.pone.0019705
- Claßen-Houben, D.; Kueenburg, B.; Kosma, P.; Jordis, U.; Stanetty, C.; Szolar, O.; Wolkerstorfer, A. Antiviral Triterpene Derivatives. EP 2 258 713 A1, Dec 8, 2010.
- Amer, H.; Mereiter, K.; Stanetty, C.; Hofinger, A.; Czollner, L.;
 Beseda, I.; Jordis, U.; Kueenburg, B.; Claßen-Houben, D.; Kosma, P.
 Tetrahedron 2010, 66, 4390–4402. doi:10.1016/j.tet.2010.03.098
- Jellinck, P. H.; Monder, C.; McEwen, B. S.; Sakai, R. R. J. Steroid Biochem. Mol. Biol. **1993**, 46, 209–213. doi:10.1016/0960-0760(93)90296-9
- Florio, P.; Thomson, R. J.; von Itzstein, M. Carbohydr. Res. 2000, 328, 445–448. doi:10.1016/S0008-6215(00)00228-7
- MacDougall, J. M.; Zhang, X.-D.; Polgar, W. E.; Khroyan, T. V.; Toll, L.; Cashman, J. R. *J. Med. Chem.* **2004**, *47*, 5809–5815. doi:10.1021/jm049554t
- 18. Cao, H.; Yu, B. *Tetrahedron Lett.* **2005**, *46*, 4337–4340. doi:10.1016/j.tetlet.2005.04.088
- Beseda, I.; Czollner, L.; Shah, P. S.; Khunt, R.; Gaware, R.; Kosma, P.; Stanetty, C.; del Ruiz-Ruiz, M. C.; Amer, H.; Mereiter, K.; Da Cunha, T.; Odermatt, A.; Claßen-Houben, D.; Jordis, U. *Bioorg. Med. Chem.* **2010**, *18*, 433–454. doi:10.1016/j.bmc.2009.10.036
- Ijichi, S.; Tamagaki, S. Chem. Lett. 2005, 34, 356–357. doi:10.1246/cl.2005.356
- Brieskorn, C. H.; Eschelbach, H. Arch. Pharm. 1979, 312, 752–762. doi:10.1002/ardp.19793120907
- Stanetty, C.; Czollner, L.; Koller, I.; Shah, P.; Gaware, R.; Da Cunha, T.; Odermatt, A.; Jordis, U.; Kosma, P.; Claßen-Houben, D. *Bioorg. Med. Chem.* 2010, *18*, 7522–7541. doi:10.1016/j.bmc.2010.08.046
- Csuk, R.; Schwarz, S.; Siewert, B.; Kluge, R.; Ströhl, D. *Eur. J. Med. Chem.* **2011**, *46*, 5356–5369. doi:10.1016/j.ejmech.2011.08.038
- Zhou, C. C.; Stewart, K. D.; Dhaon, M. K. Magn. Reson. Chem. 2005, 43, 41–46. doi:10.1002/mrc.1501
- 25. Stewart, W. E.; Siddall, T. H., III. Chem. Rev. **1970**, *70*, 517–551. doi:10.1021/cr60267a001

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