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Preparation of 3-benzyloxy-2-pyridinone functional linkers: tools for the synthesis of 3,2-hydroxypyridinone (HOPO) and HOPO/hydroxamic acid chelators



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ABSTRACT

In contrast to 2,3-dihydroxypyridine, the 3-benzyloxy protected derivative, **2**, undergoes facile alkylation at ambient temperatures with a variety of functionalized alkyl halides in good yields. This alkylation has been used to prepare a number of linkers that permit the attachment of 3,2-HOPO moieties onto various scaffolds using a wide range of coupling methods. The Mitsunobu reaction of **2** with representative al-cohols was found to be of limited value due to competing O-alkylation that led to product mixtures. The phthalimide **3j** can be converted in two steps to HOPO isocyanate **6** in excellent yields. Isocyanate **6** can be coupled to amines at room temperature or to alcohols in refluxing dichloroethane to obtain the corresponding urea or carbamate linked ligand systems. The coupling of isocyanate **6** with TREN followed by deprotection gave the tris-HOPO **10**, an interesting target as it has both cationic and anionic binding sites. The HOPO hydroxylamine linker **11** was shown to be especially valuable as its coupling with carboxylic acids proceeds with the concomitant generation of an additional hydroxamate ligand moiety in the framework. The utility of this linker was shown by the preparation of two mixed HOPO-hydroxamate chelators, **16** and **19**, based on the structure of desferrioxamine, a well-known trihydroxamate siderophore.

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1. Introduction

Siderophores are low molecular weight molecules with high affinity for iron (III) that are produced by bacteria and fungi to sequester iron from their environment. Structures of natural siderophores can be quite diverse and the ligand groups involved in the complexation of iron vary. Desferrioxamine, DFO is a trihydroxamate that was first isolated from Streptomyces pilosus in 1950 and is probably the most widely studied siderophore in terms of therapeutic potential. DFO is used for the treatment of iron overload in patients with β -thalassemia.¹ It is one of the rare siderophores to have a functional group (amine) available for subsequent modification.² This fact has facilitated the preparation of a number of DFOconjugates for various biochemical studies.³ Enterobactin is a siderophore of enterobacteria in which three catecholate groups anchored to a cyclic serine trimer backbone are used to bind the ferric iron in an hexadentate manner.⁴ Many analogs of both DFO and enterobactin have been synthesized and examined for iron coordination and potential biomedical applications (Fig. 1).⁵



Fig. 1. Structures of desferrioxamine, enterobactin and 3,2-hopobactin.

Hydroxypyridinones, HOPOs, are powerful chelating groups for hard metal ions (Fig. 2). As chelating groups, one could view HOPOs as monobasic pyridine analogs of catechols in which bidentate coordination occurs through both oxygens. There has been much interest in the incorporation of this ligand onto various platforms to prepare new generations of synthetic siderophores. The use of hydroxypyridinones as 'privileged' structures for the design of medicinal drugs has been recently reviewed.⁶ As therapeutic agents, the 3,4-HOPO, *deferiprone*, is used for in vivo iron clearance



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Fig. 2. Hydroxypyridinone chelators for Fe(III).¹⁶

(Fig. 2).⁷ HOPO compounds have also been evaluated for cancer treatment⁸ and as novel antibacterial compounds.^{9,10} HOPO side-rophores have been studied in a variety of diagnostic applications including as MRI contrast agents^{11,12} and in positron emission to-mography (PET).^{13,14} It is important to mention that hydroxypyr-idinones for selective actinide binding and remediation of transuranic waste is an active area of interest.¹⁵

Methods to introduce hydroxypyridinones onto scaffolds and for its conjugation into biomolecules are limited both in terms of number and flexibility. One popular approach to introduce the 3,2-HOPO ligand, developed by Raymond and co-workers, involves acylation of an activated HOPO carboxylic acid substrate (C, Scheme 1) with various amines. Reagent **C** is available in four steps from commercially available 2,3-dihydroxypyridine, DHP, 1. The key step in this sequence involves carboxylation at the 4 position of the pyridinone ring of **A** using Kolbe–Schmitt conditions to get **B**. This reaction is limited in terms of substrate tolerance and works best when there is either a methyl or ethyl group on the nitrogen of the pyridinone ring. The activated ester **C** is prepared in two steps from **B** and can be coupled to a polyamine such as tris(2-amino)ethylamine, TREN (Scheme 1) to obtain the hexadentate siderophore, **TRENHOPO**, after deprotection.¹⁷ The method developed by Hider and co-workers for tethering HOPO to amine backbones involves the use of an activated acyl derivative introduced on the nitrogen of the pyridinone ring. The sequence begins with the alkylation of **1** with excess ethyl bromoacetate at refluxing temperature to give **D**. The workup for the reaction involves removing the excess ethyl bromoacetate, a potent lachrymator, by distillation. Subsequently, ester **D** is converted in two steps to NHS ester, **E**, for acylation with amines. Coupling with an amine, for example, TREN, followed by deprotection gives CP-130.¹⁸ The serious drawback to this approach is that the alkylation of **1** is a poor reaction with most electrophiles. Both of the protocols described above, involve attachment of the HOPO moiety via an amide linkage.

Our group has published methods for the attachment of the 3,2-HOPO ligand onto scaffolds without concomitant formation of an amide bond. The goal has been to prepare less structurally rigid ligands with greater water solubility. In one method, ester **D** was converted into electrophilic imidate salt, **F**, which reacts readily with secondary amines and phenols. For example, salt **F**, was reacted with 1,4,7-triazacyclononane, to give tris HOPO ligand, **TACN-3,2-HOPO**, after deprotection.¹⁹

Given the interest in siderophores for various applications, synthetic methods that would allow the introduction/tethering of 3,2-HOPO onto a variety of scaffolds are of value. However, as discussed, current methods to introduce 3,2-HOPO ligands to prepare a variety of targets are limited. There is a need for functional 3,2-HOPO linkers, **3**, that can be readily conjugated/tethered to polymers, biomolecules and antibacterial drugs (Scheme 2). Further, functional linkers, **3**, could be used to prepare new classes of siderophores including those that have mixed ligands and analogs essential for structure activity relationships. It is important that the functional groups, Z, on the HOPO linker can be varied to accommodate the proposed coupling method to the substrate of interest.



The major impediment to prepare 3,2-HOPO linkers has been limitations on the alkylation of 2,3-dihydroxypyridine, **1**. In this paper, we report that *O*-benzyl protected HOPO, **2**, is a good nucleophile and can be readily alkylated to give access to a variety of functional HOPO linkers, **3**. The synthetic utility of the linkers prepared in this study has been demonstrated by the preparation of a urea linked tris-3,2-HOPO attached to a TREN backbone. It is also noteworthy that this study gives access to a new class of HOPO linkers that can be coupled to acids with concomitant generation of a hydroxamate bond. To demonstrate the value of this class of linkers, two new mixed ligand hydroxamate-HOPO siderophores, which are modeled on DFO, have been prepared.

We began with a systematic study of the alkylation of 3benzyloxy-2-pyridinone, **2**, which is readily prepared by O-benzylation of **1** (Scheme 2).²⁰ There have been some isolated examples in the literature of *N*-alkylation of protected 3-alkoxy-2-



Scheme 1.

pyridinones, such as **2**.²¹ However, the reaction has not been well studied. Further, the relative reactivity of the O-protected derivative of 2,3-dihydroxypyridine, **2**, relative to the parent, **1**, in alkylation reactions has not been evaluated. It is known that protection of the 3-hydroxy group of both 3,2 and 3,4-HOPOs has a large impact on the observed regioselectivity of aminomethylation (Mannich reaction) compared to the corresponding unprotected parents.²² Hence, a potential concern was the possibility of competing O-alkylation with the desired N-alkylation of the substrate and how that might impact the utility of this reaction.

2. Results and discussion

2.1. Alkylation of 2

It is known that the alkylation of **1** with ethyl bromoacetate can be carried out using large excess of the alkylating agent under reflux conditions. To determine the effect of protecting the 3hydroxy group, the alkylation of 2 with ethyl bromoacetate (1 equiv) using sodium hydride in DMF was investigated. The reaction was facile and the desired N-alkylated product was isolated in 93% yield after only 2 h (Table 1, entry 1). The ease of this reaction compared to the direct alkylation of **1** was remarkable. Further, the product from O-alkylation was not observed.

The positive results from this initial reaction led us to study the alkylation of 2 with tert-butyl bromoacetate (Table 1, entry 2) under a variety of conditions. Treatment of 2 with sodium hydride (1.1 equiv) in DMF followed by addition of tert-butyl bromoacetate (1.1 equiv), gave the desired product in 80% yield after chromatographic purification. The reaction could be carried out in either DMF or THF though the reaction was much faster in DMF as expected. When the alkylation was carried out using potassium fluoride on alumina as the base in acetonitrile solvent, the product was isolated in 73% yield after purification. Although the KF/alumina is somewhat expensive, the simple filtration work-up and avoiding DMF (easier for workup) is an advantage. The product from O-alkylation was not observed in either the sodium hydride or KF/alumina reactions. The reaction was also investigated using either DBU or K₂CO₃ as the base in DMF. In both cases, the product formation was very slow, making it an unattractive reaction.

The alkylation study was then expanded to include a variety of functionalized alkyl halides (Table 2). In the case of reactive allyl bromide (Table 2, entry 1), excellent yields of the desired N-alkylated product could be obtained using either sodium hydride in DMF or KF/alumina. In general, the yields of the alkylation reaction

Table 1

Alkylation of 3-benzyloxy-2-pyridinone, 2, with α-bromoesters



Conditions: A. NaH, DMF, 0 °C to rt, N2, 2 h. B. KF/alumina, CH₃CN, rt, 24 h.

were higher when sodium hydride in DMF was used instead of KF/ alumina. In a few cases, some of the corresponding O-alkylated product, 4, was also formed (Table 2, entries 2, 4, 8, 9, 11). The Oalkylation products were much less polar than the desired product of N-alkylation and were easily removed by chromatography.

2.2. Alkylation of 2 with alcohols

Because functionalized alcohols are readily available starting materials, the Mitsunobu reaction of 2 using allyl alcohol as a model was investigated. Reaction of HOPO 2 with allyl alcohol in the

Table 2

1

2

3

5

6

7

8

Alkylation of 3-benzyloxy-2-pyridinone, 2, with alkyl halides



(continued on next page)

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Table 2 (continued)



Conditions: A. NaH, DMF, 0 °C to rt, N2, 24 h.

B. KF/Al₂O₃, CH₃CN, rt, 24 h.

^a 10–15 % yield of the corresponding O-alkylated product **4** was isolated.

presence of diisopropyl azodicarboxylate (DIAD) and polystyrenesupported triphenylphosphine in THF gave predominantly the product of N-alkylation, **3c**, along with 20% of the product from Oalkylation, **4c** (Table 3, entry 1) (Scheme 3). It is noteworthy that the O-allyl product, **4c**, can be easily rearranged to the *N*-allyl product, **3c**, in xylene at 80 °C in the presence of a catalytic amount of PdCl₂ (see 4.6). When the Mitsunobu reaction of **2** was carried out with 11-decen-1-ol under the same conditions, the desired product of Nalkylation, **3d**, was obtained in only 27% yield along with 20% yield of the corresponding O-alkylated product, **4d** (Table 3, entry 2). To



Table 3				
Reaction	of 2 with	alcohols under	Mitsunobu	conditions

Entry	ROH	Reagent	Product, yield %	Product, yield %
1	Allyl alcohol	PS-PPh ₃	3c , 47	4c , 20
2	11-Undecen-1-ol	PS-PPh₃	3d , 27	4d , 20
3	11-Undecen-1-ol	PPh_3	3d , 26	4d , 51

improve the reaction yield, the reaction was performed using triphenylphosphine in place of PS-PPh₃ (Table 3, entry 3). In this case, the product of O-alkylation, **4d**, was the major product and was isolated in 51% yield. While it is interesting to note that alkylation of **2** directly with alcohols is feasible under Mitsunobu conditions, there is significant and undesirable competition with O-alkylation.

2.3. Applications of functional HOPO linkers

2.3.1. Tethering of 3,2-HOPO via carbamate and urea linkages. The N-alkylation of **2** provides easy access to a variety of HOPO linkers differing in functional groups. The phthalimide products, **3j** and **3k**, are precursors to the corresponding amines, **5**. This was demonstrated by the conversion of phthalimide, **3j**, to amine, **5**, using aqueous methylamine in 90% yield (Scheme 4). Amine **5** has been prepared by an alternate route and successfully coupled with acid chlorides and sulfonyl chlorides.²³

The conversion of the HOPO amine **5** to the corresponding isocyanate provides a reactive linker for coupling with amines and alcohols. Treatment of amine **5** with a solution of phosgene in toluene in the presence of proton sponge in dichloromethane gave the corresponding isocyanate **6** in 82% yield. The isocyanate was of sufficient purity to use without further purification. The reaction of HOPO isocyanate was then examined with alcohols and amines. As an example, reaction of isocyanate, **6**, with 1-octanol proceeded cleanly in dichloroethane at 80 °C to give carbamate linked HOPO, **7**. As expected, the reaction of 1-octylamine with isocyanate, **6**, in dichloromethane at 0 °C was faster and gave excellent yield of the corresponding HOPO urea, **8**, after chromatographic purification.

2.3.2. Synthesis of urea-linked tris-HOPO chelator, **10**. Ion pair recognition is an emerging area in supramolecular chemistry.²⁴ There has been much interest in the design of ditopic chelators capable of simultaneous complexation of both anions and cations in their scaffolds. It has been hypothesized that the binding of the cation could preorganize the receptor for enhanced binding of the anion.²⁵ TREN based urea receptors have been shown to be effective for the binding of oxyanions and to facilitate their encapsulation.²⁶ TREN has been a highly used tripodal platform to prepare hydroxamates and hydroxypyridinones that show strong binding to iron.²⁷

A TREN tris-urea HOPO ditopic chelator was a tempting target to demonstrate the utility of the new HOPO isocyanate linker **6**. Reaction of TREN with a slight excess of isocyanate **6**, in dichloromethane at 0 °C gave the protected HOPO, **9**, in 67% yield after chromatographic purification (Scheme 5). Debenzylation of **9** using a hydrogen balloon and 5% Pd on carbon gave the desired urea-linked tris-HOPO chelator, **10**, in quantitative yield.

2.3.3. Synthesis of mixed HOPO/hydroxamate analogs of desferrioxamine, DFO. In DFO and enterobactin, iron complexation is achieved by three hydroxamate or three catecholate ligands, respectively. There are also many natural siderophores where the ferric ion is complexed by a mixed ligand system. For example, *Mycobacterium tuberculosis* produces a suite of siderophores called



Scheme 4.



Scheme 5.

carboxymycobactins to sequester iron to support its growth.²⁹ The coordination of iron is accomplished by a cyclic hydroxamic acid, a hydroxamic acid and a phenolate oxazoline. In aerobactin, one of the siderophores of pathogenic *Escherichia coli*, the complexation of iron is achieved by two hydroxamic acids and a citrate moiety.²⁸

While mixed ligand chelators occur widely in natural siderophores,²⁹ synthetic analogs have not been as widely studied perhaps due to greater difficulty in their syntheses. An interesting example of the use of mixed ligands is the recent publication of a biscatecholate—monohydroxamate conjugated to an antibiotic to generate a new drug candidate that is selective against *A. baumannii*.³⁰ A recent review on sequestering agents for actinides documents several synthetic mixed ligand 3,2-HOPO chelators.¹⁵ Our own group has published the synthesis of mixed ligand 3,2-HOPO/hydroxamic acid and 3,2-HOPO/carboxylic acid and 3,2-HOPO mixed chelators on polyphenol platforms.³¹

The HOPO linkers developed in this study are convenient intermediates for the synthesis of HOPO mixed ligand systems that have not been hitherto examined. We envisioned using the bifunctional linkers to prepare mixed HOPO/hydroxamic acid ligands inspired by the structure of the siderophore, desferrioxamine, DFO. The protected HOPO-hydroxylamine linker **11** is a useful tool as it provides a novel advantage. After deprotection of **3m** to the corresponding hydroxylamine **11**, it can be coupled to a carboxylic acid. This coupling reaction generates a hydroxamic acid linkage, resulting in a mixed hydroxamate-HOPO mixed ligand system.

The first DFO analog to be prepared was the HOPO/dihydroxamate mixed ligand, **15**, in which one hydroxamic acid of DFO is replaced with a 3,2-HOPO ligand (Scheme 6). The protected hydroxylamine HOPO, **11**, was coupled to the functionalized carboxylic acid, **12**, using HATU in the presence of triethylamine in dichloromethane to give **13**. It is pleasing to note that this key step, in which the hydroxamic acid bond is formed, proceeded in good yield. Removal of the Boc protecting group of **13** using TFA gave the free hydroxylamine **14**, in 77% yield after chromatographic purification. Hydroxylamine, **14**, was coupled with acetyl chloride in the presence of triethylamine in dichloromethane to generate the fully protected mixed ligand **15**. Global deprotection of the benzyl protecting groups using 5% Pd on carbon gave the mixed dihydroxamic acid/HOPO ligand, **16**, in excellent yield. It is important to point out that the chain lengths of both linkers **11** and **12** can be readily varied. Also, a variety of acid chlorides can substitute for acetyl chloride in the preparation of **15**, making this synthetic approach highly flexible.

The next target to be prepared was the diHOPO/hydroxamate mixed ligand, **19**, in which two hydroxamic acids of DFO are replaced with 3,2-HOPO ligands (Scheme 7). It is interesting to note that the chemistry uses two HOPO linkers produced from this work to create the mixed ligand siderophore. Hydrolysis of t-butyl ester, **3g**, using TFA in dichloromethane gave the HOPO acid, **17**, in 95% yield. HATU mediated coupling of HOPO acid, **17**, with HOPO hydroxylamine, **11**, generated the fully protected mixed ligand, **18**, in 83% yield after purification. Global deprotection of the benzyl groups was accomplished using 5% Pd/C in ethanol to give the diHOPO/hydroxamate mixed ligand DFO analog, **19**, in 87% yield.



3. Conclusions

The hydroxypyridinone ligand has become an integral part in the design of new classes of synthetic siderophores targeted towards therapeutic and diagnostic applications. In particular, HOPO and HOPO/terephthalamide (mixed ligand) chelators have been widely studied as MRI contrast agents.¹¹ Hence, synthetic methods that give access to new generations of HOPO chelators as well as those that allow the conjugation of this ligand to existing platforms are important. The functional HOPO linkers reported in this study allow its attachment to various platforms



Scheme 6.

via a number of coupling protocols. For instance, the HOPO alkynes from this study are potential partners for both click chemistry with azides³² and Sonogashira coupling³³ with aryl halides. The ability to alkylate 3-benzyloxy-2-pyridinone, **2**, with a variety of functionalized alkylating agents in good yields and under mild conditions (sodium hydride in DMF at room temperature) removes a serious hurdle faced by researchers in this area by greatly improving the availability of HOPO coupling agents. In the case of more active alkylating agents, such as allyl bromide, the alkylation can be performed using potassium fluoride on alumina. This protocol offers simplicity in the product isolation. In most cases, O-alkylation of **2** was a minor pathway and posed no problems.

It is interesting to note that **2** can function as a partner in Mitsunobu reactions with primary alcohols. To our knowledge, this reaction has not been studied before.³⁴ Unfortunately, under the reaction conditions both *O* and *N*-alkylated products of **2** are formed with low selectivity, limiting the usefulness of this approach.

Particularly useful HOPO linkers to emerge from this work are the isocyanate 6 and the HOPO-hydroxylamine, 11. HOPO isocyanate, **6**, should be a highly desirable partner for bioconjugation where an amine is available for coupling. This reagent also allows the preparation of new classes of potential ditopic receptors as demonstrated by the preparation of TREN-UREA-3,2-HOPO, 10. The syntheses of mixed HOPO/hydroxamate ligands, 16 and 19, exemplifies the special advantages of linkers such as 11. The tethering of an HOPO moiety with concomitant generation of a hydroxamate binding site in the skeleton is novel. In addition, ligands **16** and **19** are the first examples of mixed hydroxamate/ HOPO analogs of DFO in which the HOPO moiety is part of the backbone and not just appended to the terminal amine of DFO. This new coupling protocol should greatly enhance the ability to synthesize mixed HOPO-hydroxamate mixed ligands for therapeutic and diagnostic studies.

4. Experimental

4.1. General methods

Melting points were measured in capillary tubes and are uncorrected. Infrared spectra were recorded on an FT-IR Spectrometer. NMR spectral samples were prepared using CDCl₃ unless otherwise indicated. All chemical shifts are reported in parts per million (ppm) relative to the deuterated solvent or an internal tetramethylsilane (TMS) reference. High resolution mass spectral analyses were obtained using electrospray ionization with the samples dissolved in either chloroform or methanol. Analytical thin layer chromatography was performed on silica 60/F254 plastic plates. Column chromatography was done using silica gel (60–200 mesh or 200-400 mesh) unless otherwise indicated. Radial chromatography was conducted using a centrifugal thin-layer chromatograph and plates were prepared using silica gel 60 containing gypsum. Polystyrene-triphenylphosphine (PS-PPh₃) had a loading capacity of 1.88 mmol/g. Sodium hydride (NaH) was obtained as a 57-63% mineral oil dispersion. Potassium fluoride on alumina (KF/Al₂O₃) was 40% by weight KF. Phosgene was obtained as a 20% solution in toluene. 3-(Benzyloxy)pyridin-2(1H)-one, 2, was prepared according to literature procedure.²⁰

4.2. Representative procedure A for the alkylation of 3-(ben-zyloxy)pyridin-2(1*H*)-one, 2, using sodium hydride

4.2.1. Synthesis of ethyl 2-(3-(benzyloxy)-2-oxopyridin-1(2H)-yl)acetate (**3a**). To a solution of **2** (0.20 g, 0.99 mmol) in anhydrous DMF (3 mL) under N_2 at 0 °C was added NaH (0.044 g,

1.09 mmol). The mixture was warmed to rt and stirred for 30 min. After cooling back to 0 °C, ethyl bromoacetate (0.12 mL, 1.09 mmol) was added and the mixture stirred at rt for 2 h. The reaction was guenched with saturated NH₄Cl (10 mL) and the product extracted into ethyl acetate (3×30 mL). The combined organic extract was dried (Na₂SO₄) and the solvent removed in vacuo. Excess DMF was removed by Kugelrohr distillation. The crude product was purified by radial chromatography (hexane/ ethyl acetate gradient) to give ester **3a** (0.26 g, 93%) as a white solid: mp 114-115 °C; IR (neat) 2986, 1751 (C=O), 1654 (HOPO C=O), 1598 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.43–7.36 (m, 2H), 7.34–7.21 (m, 3H), 6.87 (dd, *J*=6.9, 1.6 Hz, 1H), 6.64 (dd, *J*=7.4, 1.6 Hz, 1H), 5.99 (dd, *J*=7.4, 6.9 Hz, 1H), 5.04 (s, 2H), 4.62 (s, 2H), 4.18 (q, J=7.1 Hz, 2H), 1.23 (t, J=7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) § 167.7, 158.1, 148.6, 136.2, 129.6, 128.4, 127.9, 127.3, 116.1, 104.8, 70.6, 61.6, 50.6, 14.1, HRMS $[M+H^+]$ m/z calcd for C₁₆H₁₈NO₄ 288.1236, found 288.1230.

4.3. Representative procedure B for the alkylation of 2 using KF/Al_2O_3

4.3.1. Synthesis of tert-butyl 2-(3-(benzyloxy)-2-oxopyridin-1(2H)yl)acetate (**3b**). To a suspension of **2** (0.20 g, 0.994 mmol) in CH₃CN (17 mL) was added tert-butyl bromoacetate (0.22 mL, 1.49 mmol) and KF/Al₂O₃ (1.73 g, 11.9 mmol) and the mixture was stirred at rt for 24 h. The solids were filtered off and rinsed with ethyl acetate. The solvent was removed in vacuo and the crude product was purified by radial chromatography (hexane/ethyl acetate gradient) to give ester **3b** (0.23 g, 73%) as a white solid. Using representative procedure A, **3b** was isolated in 80% yield: mp 130–131 °C; IR (neat) 2975, 1743 (C=O), 1655 (HOPO C=O), 1602 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.44-7.37 (m, 2H), 7.36-7.23 (m, 3H), 6.84 (dd, J=7.2, 1.7 Hz, 1H), 6.64 (dd, J=7.2, 1.7 Hz, 1H), 6.02 (t, J=7.2 Hz, 1H), 5.09 (s, 2H), 4.57 (s, 2H), 1.47 (s, 9H); ¹³C NMR (50 MHz, CDCl₃) δ 166.8, 158.1, 148.8, 136.3, 129.5, 128.5, 127.9, 127.3, 115.9, 104.6, 82.6, 70.7, 51.0, 28.0. HRMS $[M+H^+]$ m/z calcd for $C_{18}H_{22}NO_4$ 316.1549, found 316.1537.

4.4. Synthesis of functional HOPO linkers, 3

4.4.1. 1-Allyl-3-(benzyloxy)pyridin-2(1H)-one (**3c**). Representative procedure A was followed using allyl bromide in place of ethyl bromoacetate and the reaction mixture was stirred 24 h to give alkene **3c** as a yellow solid in 88% yield. Using representative procedure B, **3c** was isolated in 90% yield. mp 51–52 °C; IR (neat) 3033, 1652 (HOPO C=O), 1602 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.47–7.40 (m, 2H), 7.40–7.24 (m, 3H), 6.88 (dd, *J*=6.9, 1.7 Hz, 1H), 6.64 (dd, *J*=7.4, 1.7 Hz, 1H), 6.03 (dd, *J*=7.4, 6.9 Hz, 1H), 5.99–5.87 (m, 1H), 5.27–5.15 (m, 2H), 5.12 (s, 2H), 4.61 (dt, *J*=5.9, 1.5 Hz, 2H); ¹³C NMR (50 MHz, CDCl₃) δ 157.9, 148.8, 136.3, 132.5, 128.5, 128.5, 127.9, 127.2, 118.3, 115.6, 104.7, 70.6, 51.0. HRMS [M+H⁺] *m/z* calcd for C₁₅H₁₆NO₂ 242.1181, found 242.1172.

4.4.2. 3 - (Benzyloxy) - 1 - (undec - 10 - enyl)pyridin - 2(1H) - one(**3d**). Representative procedure A was followed using 11bromoundec-1-ene in place of ethyl bromoacetate and the reaction mixture was stirred 24 h to give alkene **3d** as a yellow oil in 79% yield. The corresponding *O*-alkylated product **4d** was also isolated in 12% yield. **3d**: IR (film) 2926, 1655 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.38 (m, 2H), 7.36–7.22 (m, 3H), 6.87 (dd, *J*=7.1, 1.7 Hz, 1H), 6.62 (dd, *J*=7.1, 1.7 Hz, 1H), 5.97 (t, *J*=7.1 Hz, 1H), 5.87–5.70 (m, 1H), 5.08 (s, 2H), 5.04–4.86 (m, 2H), 3.92 (t, *J*=7.4 Hz, 2H), 2.03 (q, *J*=6.8 Hz, 2H), 1.81–1.65 (m, 2H), 1.43–1.19 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 158.01, 148.83, 139.04, 136.43, 129.06, 128.42, 127.81, 127.25, 115.52, 114.13, 104.34, 70.63, 49.80, 33.76, 29.37, 29.34, 29.19, 29.10, 29.04, 28.86, 26.61. HRMS [M+H⁺] m/z calcd for C₂₃H₃₂NO₂ 354.2433, found 354.2430. **4d**: IR (film) 2926, 1595, 1449, 1201 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.73 (dd, *J*=5.0, 1.6 Hz, 1H), 7.46–7.23 (m, 5H), 7.04 (dd, *J*=7.7, 1.6 Hz, 1H), 6.74 (dd, *J*=7.7, 5.0 Hz, 1H), 5.90–5.70 (m, 1H), 5.14 (s, 2H), 5.04–4.86 (m, 2H), 4.38 (t, *J*=6.9 Hz, 2H), 2.04 (q, *J*=6.8 Hz, 2H), 1.85 (quintet, *J*=7.2 Hz, 2H), 1.57–1.18 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 155.3, 143.1, 139.2, 138.0, 136.8, 128.6, 127.9, 127.2, 120.9, 116.4, 114.1, 71.0, 66.3, 33.8, 29.6, 29.5, 29.4, 29.2, 29.1, 29.0, 26.1. HRMS [M+H⁺] m/z calcd for C₂₃H₃₂NO₂ 354.2433, found 354.2433.

4.4.3. 3 - (Benzyloxy) - 1 - (prop - 2 - ynyl) pyridin - 2(1H) - one(**3e**). Representative procedure A was followed using propargyl bromide in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give alkyne **3e** as a light yellow oil in 81% yield. Using representative procedure B, **3e** was isolated in 63% yield: IR (neat) 3217, 2121 (C=C), 1652 (HOPO C=O), 1602 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.38 (m, 2H), 7.37–7.25 (m, 3H), 7.21 (dd, *J*=7.2, 1.7 Hz, 1H), 6.63 (dd, *J*=7.2, 1.7 Hz, 1H), 6.05 (t, *J*=7.2 Hz, 1H), 5.09 (s, 2H), 4.77 (d, *J*=2.6 Hz, 2H), 2.47 (t, *J*=2.6 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 157.6, 148.5, 136.2, 128.5, 127.9, 127.3, 127.2, 115.6, 105.0, 77.1, 75.0, 70.7, 37.5. HRMS [M+H⁺] *m/z* calcd for C₁₅H₁₄NO₂ 240.1025, found 240.1022.

4.4.4. 3-(Benzyloxy)-1-(hex-5-ynyl)pyridin-2(1H)-one (3f). Representative procedure A was followed using 6-iodo-1hexyne in place of ethyl bromoacetate and the reaction was stirred for 24 h to give alkyne **3f** as a light brown solid in 75% yield. The corresponding O-alkylated product **4f** was isolated in 10% yield. Representative procedure B was followed using 6-iodo-1-hexyne in place of tert-butyl bromoacetate to give 3f in 49% yield. 3f: mp 41-43 °C; IR (neat) 3232, 2952, 2111 (C=C), 1651 (HOPO C=O), 1592 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.46-7.38 (m, 2H), 7.38-7.22 (m, 3H), 6.89 (dd, J=6.9, 1.7 Hz, 1H), 6.63 (dd, J=7.4, 1.7 Hz, 1H), 6.01 (dd, J=7.4, 6.9 Hz, 1H), 5.10 (s, 2H), 3.98 (t, J=7.2 Hz, 2H), 2.24 (td, J=7.0, 2.7 Hz, 2H), 1.96 (t, J=2.7 Hz, 1H), 1.91-1.83 (m, 2H), 1.62–1.50 (m, 2H); 13 C NMR (50 MHz, CDCl₃) δ 158.1, 148.9, 136.4, 128.9, 128.5, 127.9, 127.3, 115.5, 104.6, 83.7, 70.7, 68.9, 49.1, 28.2, 25.4, 18.1. HRMS [M+H⁺] *m*/*z* calcd for C₁₈H₂₀NO₂ 282.1494, found 282.1484.

4.4.5. *tert-Butyl* 6-(3-(*benzyloxy*)-2-*oxopyridin*-1(2H)-*yl*)*hexanoate* (**3***g*). Representative procedure A was followed using *tert*-butyl 6-iodohexanoate in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give ester **3***g* as a light brown oil in 75% yield: IR (film) 2935, 1729 (C=O), 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.36 (m, 2H), 7.36–7.20 (m, 3H), 6.88 (dd, *J*=7.1, 1.7 Hz, 1H), 6.63 (dd, *J*=7.1, 1.7 Hz, 1H), 5.98 (t, *J*=7.1 Hz, 1H), 5.08 (s, 2H), 3.93 (t, *J*=7.3 Hz, 2H), 2.20 (t, *J*=7.4 Hz, 2H), 1.75 (quintet, *J*=7.5 Hz, 2H), 1.61 (quintet, *J*=7.5 Hz, 2H), 1.42 (s, 9H), 1.38–1.29 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 158.0, 148.8, 136.4, 129.1, 128.4, 127.8, 127.2, 115.5, 104.4, 79.9, 70.6, 49.5, 35.2, 28.7, 28.1, 26.0, 24.6. HRMS [M+H⁺] *m/z* calcd for C₂₂H₃₀NO₄ 372.2175, found 372.2166.

4.4.6. Dibenzyl 3-(3-(benzyloxy)-2-oxopyridin-1(2H)-yl)propylphosphonate (**3h**). Representative procedure A was followed using dibenzyl 3-iodopropylphosphonate in place of ethyl bromoacetate to give **3h** as a yellow oil in 77% yield IR (film) 3033, 1656 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.48–7.21 (m, 15H), 6.77 (dd, *J*=7.2, 1.6 Hz, 1H), 6.60 (dd, *J*=7.2, 1.6 Hz, 1H), 5.93 (t, *J*=7.2 Hz, 1H), 5.07 (s, 2H), 5.06–4.90 (m, 4H), 3.96 (t, *J*=7.0 Hz, 2H), 2.10–1.94 (m, 2H), 1.80–1.68 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 158.0, 148.8, 136.4, 136.2 (d, *J*_{PC}=5.6 Hz), 129.0, 128.6, 128.5, 128.5, 128.0, 127.9, 127.3, 115.4, 104.6, 70.7, 67.3 (d, *J*_{PC}=6.7 Hz), 49.4 (d, J_{PC} =15.7 Hz), 22.9 (d, J_{PC} =141.9 Hz), 22.0 (d, J_{PC} =5.1 Hz). HRMS [M+H⁺] m/z calcd for C₂₉H₃₁NO₅P 504.1940, found 504.1927.

4.4.7. Diethyl 3-(3-(benzyloxy)-2-oxopyridin-1(2H)-yl)propylphosphonate (**3i**). Representative procedure A was followed using diethyl 3-bromopropylphosphonate in place of ethyl bromoacetate to give **3i** as a yellow oil in 80% yield: IR (film) 2983, 1654 (HOPO C=O), 1604 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.48–7.23 (m, 5H), 6.95 (dd, *J*=7.2, 1.8 Hz, 1H), 6.65 (dd, *J*=7.2, 1.8 Hz, 1H), 6.0 (t, *J*=7.2 Hz, 1H), 5.11 (s, 2H), 4.20–3.95 (m, 6H), 2.21–1.98 (m, 2H), 1.88–1.64 (m, 2H), 1.32 (t, *J*=7.0 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 158.1, 148.9, 136.3, 129.0, 128.5, 128.0, 127.3, 115.5, 104.8, 70.7, 61.7 (d, *J*_{PC}=6.5 Hz), 49.6 (d, *J*_{PC}=6.1 Hz). HRMS [M+H⁺] *m/z* calcd for C₁₉H₂₇NO₅P 380.1627, found 380.1628.

4.4.8. 2-(3-(3-(Benzyloxy)-2-oxopyridin-1(2H)-yl)propyl)isoindoline-1,3-dione (**3***j*). Representative procedure A was followed using *N*-(3-bromopropyl)phthalimide in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give phthalimide **3***j* as a light yellow solid in 82% yield. The corresponding O-alkylated product **4***j* was also isolated in 10% yield. **3***j*: mp 102–103 °C; IR (neat) 2937, 1702 (C=O), 1650 (HOPO C=O), 1595 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.87–7.77 (m, 2H), 7.76–7.64 (m, 2H), 7.46–7.39 (m, 2H), 7.38–7.22 (m, 3H), 7.03 (dd, *J*=7.1, 1.6 Hz, 1H), 6.64 (dd, *J*=7.1, 1.6 Hz, 1H), 6.03 (t, *J*=7.1 Hz, 1H), 5.09 (s, 2H), 4.02 (t, *J*=6.9 Hz, 2H), 3,75 (t, *J*=6.9 Hz, 2H), 2.17 (quintet, *J*=6.9 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 168.2, 157.9, 148.8, 136.3, 134.0, 131.9, 129.2, 128.4, 127.9, 127.3, 123.2, 115.5, 104.7, 70.6, 47.5, 35.3, 28.2. HRMS [M+H⁺] *m/z* calcd for C₂₃H₂₁N₂O₄ 389.1501, found 389.1490.

4.4.9. 2-(5-(3-(*Benzyloxy*)-2-oxopyridin-1(2H)-yl)pentyl)isoindoline-1,3-dione (**3k**). Representative procedure A was followed using *N*-(5-bromopentyl)phthalimide in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give phthalimide **3k** as a yellow solid in 69% yield. The corresponding *O*-alkylated product **4k** was also isolated in 12% yield. **3k**: mp 123–124 °C; IR (neat) 2938, 1704 (C=O), 1651 (HOPO C=O), 1595 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.85–7.75 (m, 2H), 7.73–7.62 (m, 2H), 7.46–7.38 (m, 2H), 7.37–7.21 (m, 3H), 6.89 (dd, *J*=7.1, 1.7 Hz, 1H), 6.63 (dd, *J*=7.1, 1.7 Hz, 1H), 5.98 (t, *J*=7.1 Hz, 1H), 5.08 (s, 2H), 3.94 (t, *J*=7.4 Hz, 2H), 3.67 (t, *J*=7.2 Hz, 2H), 1.81 (quintet, *J*=7.4 Hz, 2H), 1.17 (quintet, *J*=7.2 Hz, 2H), 1.47–1.30 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 168.3, 158.0, 148.8, 136.4, 133.9, 132.0, 129.0, 128.4, 127.8, 127.3, 123.1, 115.5, 104.5, 70.6, 49.5, 37.6, 28.5, 28.1, 23.8. HRMS [M+H⁺] *m*/z calcd for C₂₅H₂₅N₂O₄ 417.1814, found 417.1816.

4.4.10. tert-Butyl benzyloxy(3-(3-(benzyloxy)-2-oxopyridin-1(2H)yl)propyl)carbamate (**3l**). Representative procedure A was followed using tert-butyl benzyloxy(3-bromopropyl)carbamate³⁵ in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give **3l** as a light brown oil in 86% yield: IR (film) 2980, 1695 (C=O), 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.37 (m, 4H), 7.37–7.22 (m, 6H), 6.85 (dd, *J*=6.9, 1.7 Hz, 1H), 6.61 (dd, *J*=7.3, 1.7 Hz, 1H), 5.95 (dd, *J*=7.3, 6.9 Hz, 1H), 5.06 (s, 2H), 4.85 (s, 2H), 3.96 (t, *J*=6.8 Hz, 2H), 3.48 (t, *J*=6.8 Hz, 2H), 2.05 (quintet, *J*=6.8 Hz, 2H), 1.49 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 158.0, 156.4, 148.8, 136.4, 135.4, 129.4, 129.3, 128.5, 128.5, 128.4, 127.9, 127.3, 115.5, 104.5, 81.5, 77.0, 70.7, 47.7, 47.0, 28.3, 26.8. HRMS (M+Na⁺) *m*/z calcd for C₂₇H₃₂N₂NaO₅ 487.2209, found 487.2207.

4.4.11. tert-Butyl benzyloxy(5-(3-(benzyloxy)-2-oxopyridin-1(2H)yl)pentyl)carbamate (**3m**). Representative procedure A was followed using tert-butyl benzyloxy(5-iodopentyl)carbamate³⁵ in place of ethyl bromoacetate and the reaction mixture was stirred for 24 h to give **3m** as a yellow oil in 81% yield. The corresponding O-alkylated product **4m** was also isolated in 12% yield. **3m**: IR (film) 2938, 1694 (C=O), 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.19 (m, 10H), 6.81 (dd, *J*=7.1, 1.7 Hz, 1H), 6.60 (dd, *J*=7.1, 1.7 Hz, 1H), 5.94 (t, *J*=7.1 Hz, 1H), 5.05 (s, 2H), 4.80 (s, 2H), 3.89 (t, *J*=7.3 Hz, 2H), 3.40 (t, *J*=7.1 Hz, 2H), 1.72 (quintet, *J*=7.5 Hz, 2H), 1.61 (quintet, *J*=7.4 Hz, 2H), 1.49 (s, 9H), 1.38–1.23 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 157.9, 156.4, 148.7, 136.4, 135.6, 129.3, 129.0, 128.4, 128.3, 127.8, 127.2, 115.5, 104.4, 81.0, 76.7, 70.5, 49.5, 49.2, 28.7, 28.3, 26.6, 23.7. HRMS (M+Na⁺) *m/z* calcd for C₂₉H₃₆N₂NaO₅ 515.2516, found 515.2521.

4.5. Representative procedure for the Mitsunobu reaction of 2 with alcohols

4.5.1. Synthesis of **3c**. To a suspension of **2** (0.20 g, 0.994 mmol) in anhydrous THF (9 mL) under N₂ was added PS-PPh₃ (1.06 g, 1.99 mmol) followed by DIAD (0.39 mL, 1.99 mmol). After 5 min, allyl alcohol (0.068 mL, 0.994 mmol) was added and the mixture stirred at rt for 24 h. The PS-PPh₃ was filtered off and the solvent removed in vacuo. The crude product was purified by radial chromatography (hexane/ethyl acetate gradient) to give **3c** (0.11 g, 47%) (see 4.4.1) and **4c** (0.047 g, 20%) as a light yellow oil. **4c**: IR (film) 3066, 2933, 1595 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.72 (dd, *J*=5.0, 1.5 Hz, 1H), 7.45–7.23 (m, 5H), 7.05 (dd, *J*=7.8, 1.5 Hz, 1H), 6.75 (dd, *J*=7.8, 5.0 Hz, 1H), 6.23–6.07 (m, 1H), 5.42 (dq, *J*=17.2, 1.5 Hz, 1H), 5.25 (dq, *J*=10.4, 1.5 Hz, 1H), 5.14 (s, 2H), 4.95 (dt, *J*=5.4, 1.5 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 154.4, 143.0, 137.7, 136.6, 133.6, 128.6, 128.0, 127.2, 120.7, 117.3, 116.8, 70.9, 66.6. HRMS [M+H⁺] *m/z* calcd for C₁₅H₁₆NO₂ 242.1181, found 242.1180.

4.5.2. Synthesis of **3d**. Using representative procedure for the Mitsunobu alkylation, the *N*-alkylated product **3d** was obtained in 27% yield and the *O*-alkylated product **4d** was obtained in 20% yield. When triphenylphosphine was used in place of PS-PPh₃, and the crude reaction mixture purified by column chromatography on neutral alumina (hexane/ethyl acetate gradient), **3d** was obtained in 26% yield and **4d** was obtained in 51% yield.

4.6. Synthesis of 3c via rearrangement of 4c

To a solution of **4c** (0.039 g, 0.162 mmol) in xylene (1 mL) under N₂ was added PdCl₂ (0.0028 g, 0.0162 mmol) and the mixture was heated at 80 °C for 3 h. After cooling, the catalyst was removed by filtration through a short silica gel column rinsing with CHCl₃. The solvent was removed in vacuo and the crude product was purified by radial chromatography to give **3c** (0.032 g, 82%) (see 4.4.1).

4.7. 1-(3-Aminopropyl)-3-(benzyloxy)pyridin-2(1H)-one (5)

The phthalimide **3j** (1.83 g, 4.71 mmol) was suspended in 40% aqueous methylamine (23 mL, 0.30 mol) and the mixture stirred for 2 days until all of the phthalimide had dissolved. The reaction mixture was diluted with water (20 mL) and the product extracted into CHCl₃ (3×30 mL), dried (MgSO₄) and the solvent removed in vacuo to give the known HOPO amine **5**.^{31b} (1.07 g, 87.7%). ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.33 (m, 5H), 6.92 (dd, *J*=6.9, 1.8 Hz, 1H), 6.63 (dd, *J*=6.9, 1.8 Hz, 1H), 6.03 (t, *J*=7.3 Hz, 1H), 5.12 (s, 2H), 4.09 (t, *J*=6.9 Hz, 2H), 2.72 (t, *J*=6.5 Hz, 2H), 1.89 (quintet, *J*=6.8 Hz, 2H).

4.8. 3-(Benzyloxy)-1-(3-isocyanatopropyl)pyridin-2(1H)-one(6)

A solution of **5** (0.20 g, 0.77 mmol) and 1,8-bis(dimethyl)aminonaphthalene (0.36 g, 1.54 mmol) in CH_2Cl_2 (5 mL) was added dropwise to a solution of phosgene (0.79 mL, 1.54 mmol) in toluene at 0 °C and the reaction mixture was stirred for 15 min. The solvents

were removed in vacuo and the residue was dissolved in CH₂Cl₂ (15 mL) and washed with 1M HCl (15 mL). The aqueous layer was again extracted with CH₂Cl₂ (15 mL×2) and the combined organic layers were dried (Na₂SO₄) and the solvent was removed in vacuo. The product was further dried under high vacuum to give the isocyanate **6** as a pale yellow oil (0.18 g, 82%), which was used without further purification: IR (neat) 2952, 2278 (NCO), 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.18–7.45 (m, 5H), 6.89 (dd, *J*=6.8, 1.5 Hz, 1H), 6.65 (dd, *J*=7.4, 1.4 Hz, 1H), 6.05 (t, *J*=7.1 Hz, 1H), 5.11 (s, 2H), 4.07 (t, *J*=6.8 Hz, 2H), 3.40 (t, *J*=6.3 Hz, 2H), 2.04–2.13 (quintet, *J*=6.6 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 158.3, 149.2, 136.4, 128.7, 128.6, 128.2, 127.5, 122.0, 115.7, 105.2, 70.9, 47.5, 40.4, 30.2. Anal. Calcd C₁₆H₁₆N₂O₃: C, 67.59; H, 5.67; N, 9.85; Found: C, 67.81, H, 5.84, N, 9.86.

4.9. Octyl-3-(3-(benzyloxy)-2-oxopyridin-1(2*H*)-yl)pro-pylcarbamate (7)

A solution of **6** (0.34 g, 0.18 mmol) and 1-octanol (0.043 mL, 0.27 mmol) in anhydrous 1,2-dichloroethane (2.5 mL) was heated at reflux for 1 day under N₂. After cooling, the reaction mixture was diluted with CH₂Cl₂ (5 mL) and washed with saturated NH₄Cl (15 mL), saturated NaCl (15 mL), dried (Na₂SO₄) and the solvents removed in vacuo. Excess octanol was removed by Kugelrohr distillation to give the carbamate **7** as a pale yellow oil (0.065 g, 87%): IR (neat) 3317 (NH), 2928, 2856, 1714 (C=O), 1652 (HOPO C=O), 1601 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.30–7.44 (m, 5H), 6.89 (dd, *J*=6.8, 1.2 Hz, 1H), 6.67(dd, *J*=7.5, 1.6 Hz, 1H), 6.07 (t, *J*=7.1 Hz, 1H), 5.76 (br s, 1H), 5.12 (s, 2H), 4.00–4.09 (m, 4H), 3.13 (dt, *J*=6.4, 5.7 Hz, 2H), 1.86–1.95 (m, 2H), 1.52–1.65 (m, 2H), 1.27 (br s, 10H), 0.88 (t, *J*=6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 158.9, 157.4, 149.0, 136.4, 128.9, 128.8, 128.3, 127.6, 115.7, 105.7, 71.1, 65.1, 46.4, 37.2, 32.1, 30.2, 29.46, 29.3, 26.1, 22.9, 14.4.

4.10. 1-(3-(3-(Benzyloxy)-2-oxopyridin-1(2H)-yl)propyl)-3-octylurea (8)

A solution of 1-octylamine (0.03 mL, 0.21 mmol) in CH₂Cl₂ (2 mL) was added to isocyanate **6** (0.04 g, 0.141 mmol) at 0 °C and stirred for 10 min. The reaction mixture was diluted with CH₂Cl₂ (5 mL) and washed with saturated NH₄Cl (15 mL), saturated NaCl (15 mL), dried (Na₂SO₄) and the solvents removed in vacuo. The crude product was purified using radial chromatography (ethyl acetate) to give urea **8** as a pure white solid (0.05 g, 90%): mp 93–94 °C; IR (KBr) 3335 (NH), 2925, 2854, 1656 (HOPO C=O), 1621 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.33–7.43 (m, 5H), 6.90 (dd, *J*=6.7, 1.7 Hz, 1H), 6.67 (dd, *J*=7.5, 1.6 Hz, 1H), 6.11 (t, *J*=7.2 Hz, 1H), 5.76 (t, *J*=5.7 Hz, 1H), 5.13 (s, 2H), 4.37 (m, 1H), 4.09 (t, *J*=6.1 Hz, 2H), 3.12–3.20 (m, 4H), 1.85–1.93 (m, 2H), 1.43–1.49 (m, 2H), 1.27 (br s, 10H), 0.88 (t, *J*=6.8 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 158.9, 149.0, 136.3, 129.1, 128.9, 128.4, 127.6, 115.9, 106.0, 71.1, 46.7, 40.8, 36.4, 32.1, 30.6, 30.5, 29.7, 27.2, 23.0, 14.4.

4.11. 1,1',1"-(2,2',2"-Nitrilotris(ethane-2,1-diyl))tris(3-(3-(benzyloxy)-2-oxopyridin-1(2H)-yl)propyl)urea) (9)

A solution of tris-(2-aminoethyl)amine (0.040 g, 0.24 mmol) in CH₂Cl₂ (2 mL) was added to isocyanate **6** (0.21 g, 0.80 mmol) at 0 °C and stirred for 2 h at 0 °C. The solvent was removed in vacuo and the crude product was purified using silica gel chromatography (methanol) to give urea **9** as a white solid (0.16 g, 67%): mp 65–67 °C; IR (KBr) 3366 (NH), 3063, 2922, 2852, 1651 (HOPO C=O), 1598 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.29–7.39 (m, 15H), 6.94 (dd, *J*=6.9, 1.6 Hz, 3H), 6.64 (dd, *J*=7.4, 1.4 Hz, 3H), 6.16 (br s, 3H), 6.04 (t, *J*=7.1 Hz, 3H), 5.95–6.05 (m, 3H), 5.02 (s, 6H), 3.99 (t, *J*=6.5 Hz, 6H), 3.06–3.14 (m, 12H), 2.49–2.52 (m, 6H), 1.79–1.88 (m,

6H); ¹³C NMR (75 MHz, CDCl₃) δ 159.6, 158.5, 148.8, 136.3, 129.5, 128.8, 128.4, 127.7, 115.7, 105.6, 71.0, 55.8, 47.1, 39.0, 36.8, 30.3; HRMS [M+H⁺] *m*/*z* calcd for C₅₄H₆₇N₁₀O₉ 999.5087, found 999.5105.

4.12. 1,1',1"-(2,2',2"-nitrilotris(ethane-2,1-diyl))tris(3-(3-(3-hydroxy-2-oxopyridin-1(2*H*)-yl)propyl)urea (10)

A suspension of **9** (0.07 g, 0.07 mmol) and 5% Pd/C (0.08 g) in methanol was stirred under H₂ balloon for four days. The catalyst was removed by the filtration through a 0.45 μ m nylon filter rinsing with methanol. The solvents were removed in vacuo and the product was dried under high vacuum to give urea **10** as a viscous brown oil (0.059 g, quant.): IR (neat) 3334 (NH), 1646 (br) (C=O) cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.09–7.11 (d, *J*=5.13 Hz, 3H), 6.79–6.82 (d, *J*=7.2 Hz, 3H), 6.20–6.25 (m, 3H), 4.04 (unres t, 6H), 3.17 (unres t, 12H), 2.61 (t, *J*=5 MHz, 6H), 1.89 (unres t, 6H); ¹³C NMR (75 MHz, CD₃OD) δ 161.3, 160.1, 148.4, 129.2, 117.0, 108.5, 56.1, 48.5, 39.6, 38.2, 31.3; HRMS [M+H⁺] *m/z* calcd for C₃₃H₄₉N₁₀O₉ 729.3679, found 729.3709.

4.13. Representative procedure for hydrolysis of Boc protecting group

Synthesis of 3-(benzyloxy)-1-(5-(benzyloxyamino)pentyl)pyridin-2(1H)-one (**11**). A solution of **3m** (0.695 g, 1.41 mmol) in 1:1 TFA/ CH₂Cl₂ (12 mL) was stirred at rt for 2.5 h. The solvent was removed in vacuo and the residue was adjusted to pH 10 using 1 N NaOH. The product was extracted into CH₂Cl₂ (3×50 mL) and the extract was dried (Na₂SO₄). The solvent was removed in vacuo to give **11** (0.442 g, 98%) as a brown oil, which was used without purification: IR (neat) 3249, 2937, 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.37 (m, 2H), 7.37–7.19 (m, 8H), 6.82 (dd, *J*=7.1, 1.3 Hz, 1H), 6.60 (dd, *J*=7.1, 1.3 Hz, 1H), 5.96 (t, *J*=7.1 Hz, 1H), 5.52 (br s, 1H), 5.06 (s, 2H), 4.66 (s, 2H), 3.90 (t, *J*=7.3 Hz, 2H), 2.88 (t, *J*=7.0 Hz, 2H), 1.73 (quintet, *J*=7.5 Hz, 2H), 1.52 (quintet, *J*=7.3 Hz, 2H), 1.40–1.25 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 158.0, 148.8, 138.0, 136.4, 129.0, 128.4, 128.3, 128.3, 127.8, 127.7, 127.2, 115.4, 104.4, 76.1, 70.6, 51.8, 49.6, 28.9, 27.0, 24.2.

4.14. 6-(Benzyloxy(*tert*-butoxycarbonyl)amino)hexanoic acid (12)

To a solution of *tert*-butyl benzyloxycarbamate (4.02 g, 18.0 mmol) in anhydrous DMF (60 mL) under N2 at 0 °C was added NaH (1.51 g, 37.8 mmol). The mixture was warmed to rt and stirred for 30 min. After cooling back to 0 °C, 6-bromohexanoic acid (3.86 g, 19.8 mmol) was added and the mixture was stirred at rt for 1 h and then heated at 70 °C for 18 h. The reaction mixture was cooled and guenched with saturated NH₄Cl (50 mL) and the aqueous layer acidified to pH 2 with conc. HCl. The product was extracted into ethyl acetate (3×50 mL), dried (Na₂SO₄) and the solvent was removed in vacuo. The crude product was dissolved in 1M NaOH and impurities removed by extraction with ethyl acetate (2×20 mL). The aqueous later was then acidified with conc. HCl, and the product extracted into ethyl acetate (3×30 mL), dried (Na₂SO₄) and the solvent removed in vacuo to give acid **12** (4.27 g, 70%) as an oil, which was used in the next step without further purification. A portion of 12 was purified by preparative chromatography for characterization: IR (film) 2937, 1705 (C=O) cm $^{-1}$; 1 H NMR (300 MHz, CDCl₃) δ 11.2–10.85 (br s, 1H), 7.46–7.28 (m, 5H), 4.82 (s, 2H), 3.40 (t, *J*=7.1 Hz, 2H), 2.32 (t, *J*=7.5 Hz, 2H), 1.73–1.54 (m, 4H), 1.50 (s, 9H), 1.40–1.23 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 179.7, 156.6, 135.7, 129.4, 128.5, 128.4, 81.3, 76.9, 49.4, 34.0, 28.3, 26.7, 26.2, 24.4. HRMS (M+Na⁺) *m/z* calcd for C₁₈H₂₇NO₅Na 360.1781, found 360.1777.

4.15. *tert*-Butyl benzyloxy(6-(benzyloxy)5-(3-(benzyloxy)-2oxopyridin-1(2*H*)-yl)pentyl)amino)-6-oxohexyl)carbamate (13)

To a solution of acid 12 (0.134 g, 0.398 mmol) in anhydrous CH₂Cl₂ (1.5 mL) under N₂ at 0 °C was added triethylamine (0.076 mL, 0.543 mmol) followed by HATU (0.151 g, 0.398 mmol) and **11** (0.142 g. 0.362 mmol). The mixture was stirred at rt for 3 h. The reaction mixture was diluted with CH₂Cl₂ (30 mL) and washed with 2 N HCl (2×15 mL), saturated NaHCO₃ (2×15 mL), saturated NaCl (15 mL) and dried (Na₂SO₄). The solvent was removed in vacuo and the crude product was purified by radial chromatography (hexane/ethyl acetate gradient) to give 13 (0.195 g, 76%) as a colorless oil: IR (film) 2937, 1698, 1655 (HOPO C=O), 1607 cm^{-1; 1}H NMR (300 MHz, CDCl₃) δ 7.47–7.25 (m, 15H), 6.84 (dd, *J*=7.1, 1.4 Hz, 1H), 6.62 (dd, *J*=7.1, 1.4 Hz, 1H), 5.97 (t, *J*=7.1 Hz, 1H), 5.08 (s, 2H), 4.81 (s, 2H), 4.76 (s, 2H), 3.92 (t, J=7.3 Hz, 2H), 3.62 (t, J=6.8 Hz, 2H), 3.40 (t, J=7.2 Hz, 2H), 2.36 (t, J=7.5 Hz, 2H), 1.83-1.69 (m, 2H), 1.69-1.53 (m, 6H), 1.49 (s, 9H), 1.40–1.23 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 174.6, 158.0, 156.5, 148.9, 136.4, 135.6, 134.5, 129.3, 129.1, 129.0, 128.9, 128.7, 128.5, 128.4, 128.4, 127.8, 127.3, 115.5, 104.5, 81.0, 76.8, 76.3, 70.7, 49.6, 49.4, 45.0, 32.2, 28.6, 28.3, 26.9, 26.6, 26.4, 24.3, 23.7. HRMS [M+H⁺] m/z cald for C₄₂H₅₄N₃O₇ 712.3962, found 712.3965.

4.16. *N*-(Benzyloxy)-*N*-(5-(3-(benzyloxy)-2-oxopyridin-1(2*H*)yl)pentyl)-6-(benzyloxyamino)hexanamide (14)

Following the representative procedure for hydrolysis of Boc group, and substituting **13** for **3m**, the protected hydroxylamine **14** was obtained in 77% yield as a yellow oil after radial chromatography (hexane/ethyl acetate gradient then 10% methanol/ethyl acetate). IR (film) 3436 (NH), 2939, 1653 (HOPO C=0), 1605 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.38 (m, 2H), 7.38–7.20 (m, 13H), 6.82 (dd, *J*=6.9, 1.6 Hz, 1H), 6.60 (dd, *J*=7.4, 1.6 Hz, 1H), 6.01–5.91 (m, 1H), 5.09 (s, 2H), 4.76 (s, 2H), 4.68 (s, 2H), 3.90 (t, *J*=7.3 Hz, 2H), 3.69–3.55 (m, 2H), 2.90 (t, *J*=7.0 Hz, 2H), 2.36 (t, *J*=7.4 Hz, 2H), 1.79–1.70 (m, 2H), 1.68–1.55 (m, 4H), 1.55–1.44 (m, 2H), 1.37–1.25 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 174.7, 158.0, 148.8, 138.0, 136.4, 134.5, 129.1, 129.0, 128.8, 128.7, 128.4, 128.3, 127.8, 127.7, 127.2, 115.6, 104.6, 76.2, 76.1, 70.7, 51.9, 49.6, 45.0, 32.2, 28.5, 27.1, 26.9, 26.4, 24.4, 23.7. HRMS [M+H⁺] *m*/*z* calcd for C₃₇H₄₆N₃O 612.3437, found 612.3443.

4.17. *N*-(Benzyloxy)-*N*-(5-(3-(benzyloxy)-2-oxopyridin-1(2*H*)yl)pentyl)-6-(*N*-(benzyloxy)acetamido)hexanamide (15)

To a solution of **14** (0.305 g, 0.499 mmol) in anhydrous CH₂Cl₂ (2 mL) under N₂ at 0 °C was added triethylamine (0.10 mL, 0.748 mmol) followed by acetyl chloride (0.039 mL, 0.548 mmol). The reaction mixture was stirred at rt for 24 h. The reaction mixture was diluted with CH₂Cl₂ (30 mL) and washed with 2 N HCl (2×15 mL), saturated NaHCO₃ (2×15 mL), saturated NaCl (15 mL) and dried (Na₂SO₄). The solvent was removed in vacuo and the crude product was purified by radial chromatography (hexane/ ethyl acetate gradient then 10% methanol/ethyl acetate) to give 15 (0.279 g, 86%) as a yellow oil: IR (film) 2939, 1651 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.22 (m, 15H), 6.85 (dd, J=7.2, 1.7 Hz, 1H), 6.62 (dd, J=7.2, 1.7 Hz, 1H), 5.97 (t, J=7.2 Hz, 1H), 5.07 (s, 2H), 4.78 (s, 2H), 4.76 (s, 2H), 3.91 (t, J=7.3 Hz, 2H), 3.61 (t, J=6.8 Hz, 4H), 2.36 (t, J=7.4 Hz, 2H), 2.07 (s, 3H), 1.75 (quintet, J=7.5 Hz, 2H), 1.69–1.54 (m, 6H), 1.39–1.22 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 174.6, 172.0, 158.0, 148.8, 136.4, 134.5, 129.1, 128.8, 128.6, 128.4, 127.8, 127.2, 115.5, 104.4, 76.2, 76.1, 70.6, 49.5, 45.1, 32.1, 28.6, 26.7, 26.5, 26.4, 24.2, 23.7, 20.5. HRMS [M+H⁺] m/z calcd for C₃₉H₄₈N₃O₆ 654.3543, found 654.3553.

4.18. *N*-Hydroxy-*N*-(5-(3-hydroxy-2-oxopyridin-1(2*H*)-yl)pentyl)-6-(*N*-hydroxyacetamido)hexanamide (16)

To a solution of **15** (0.377 g, 0.577 mmol) in EtOH (5 mL) was added 5% Pd on carbon (0.123 g, 0.0577 mmol) and the mixture was stirred at rt under a H₂ balloon for 24 h. The catalyst was removed by centrifugation followed by filtration. The filtrate was concentrated in vacuo to give **16** (0.215 g, 97%) as a light brown oil: IR (neat) 3176, 2937, 1600 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.10 (d, *J*=6.8 Hz, 1H), 6.83 (d, *J*=6.8 Hz, 1H), 6.24 (d, *J*=6.8 Hz, 1H), 4.96 (br s, 3H), 4.00 (t, *J*=7.0 Hz, 2H), 3.64–3.56 (m, 4H), 2.47 (t, *J*=7.2 Hz, 2H), 2.09 (s, 3H), 1.82–1.72 (m, 2H), 1.71–1.55 (m, 6H), 1.40–1.27 (m, 4H); ¹³C NMR (75 MHz, CD₃OD) δ 175.8, 173.4, 159.8, 148.2, 129.2, 117.1, 108.4, 50.7, 48.8, 48.7, 33.2, 29.9, 27.5, 27.2, 25.6, 24.5, 20.4. HRMS [M+H⁺] *m/z* calcd for C₁₈H₃₀N₃O₆ 354.2129, found 384.2138.

4.19. 6-(3-(Benzyloxy)-2-oxopyridin-1(2H)-yl)hexanoic acid (17)

Following the representative procedure for hydrolysis of Boc group, **3g** was hydrolyzed to give acid **17** in 95% yield as a light brown solid: mp 104–105 °C; IR (neat) 2943, 1723 (C=O), 1639 (HOPO C=O), 1579 cm-1; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.39 (m, 2H), 7.39–7.24 (m, 3H), 6.89 (dd, *J*=7.2, 1.6 Hz, 1H), 6.67 (dd, *J*=7.2, 1.6 Hz, 1H), 6.50 (t, *J*=7.2 Hz, 1H), 5.06 (s, 2H), 3.95 (t, *J*=7.3 Hz, 2H), 2.28 (t, *J*=7.3 Hz, 2H), 1.75 (quintet, *J*=7.5 Hz, 2H), 1.62 (quintet, *J*=7.5 Hz, 2H), 1.45–1.28 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 177.6, 158.2, 148.8, 136.2, 128.9, 128.5, 127.9, 127.4, 115.6, 105.3, 70.7, 49.8, 33.9, 28.7, 26.0, 24.3. HRMS [M+H⁺] *m/z* calcd for C₁₈H₂₂NO₄ 316.1549, found 316.1540.

4.20. *N*-(Benzyloxy)-6-(3-(benzyloxy)-2-oxopyridin-1(2*H*)-yl)-*N*-(5-(3-(benzyloxy)-2-oxopyridin-1(2*H*)-yl)pentyl)hexanamide (18)

To a solution of acid **17** (0.127 g, 0.404 mmol) in anhydrous CH_2Cl_2 (1.5 mL) under N_2 at 0 °C was added triethylamine (0.077 mL, 0.550 mmol) followed by HATU (0.153 g, 0.404 mmol) and 11 (0.144 g, 0.367 mmol). The reaction mixture was stirred at rt for 4 h. The reaction mixture was diluted with CH₂Cl₂ (30 mL) and washed with 2 N HCl (2×15 mL), saturated NaHCO₃ (2×15 mL), saturated NaCl (15 mL) and dried (Na₂SO₄). The solvent was removed in vacuo and the crude product was purified by radial chromatography (hexane/ethyl acetate gradient then 10% methanol/ethyl acetate) to give 18 (0.209 g, 83%) as a yellow oil: IR (film) 2942, 1660, 1652 (HOPO C=O), 1606 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.45–7.40 (m, 4H), 7.37–7.23 (m, 11H), 6.89–6.82 (m, 2H), 6.63 (d, J=7.5 Hz, 1H), 6.62 (d, J=7.5 Hz, 1H), 5.98 (t, J=7.1 Hz, 1H), 5.97 (t, J=7.1 Hz, 1H), 5.08 (s, 4H), 4.78 (s, 2H), 3.93 (t, J=7.3 Hz, 2H), 3.92 (t, *J*=7.3 Hz, 2H), 3.61 (br t, *J*=6.8 Hz, 2H), 2.36 (t, *J*=7.4 Hz, 2H), 1.85–1.70 (m, 4H), 1.69–1.54 (m, 4H), 1.41–1.23 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 174.5, 158.0, 148.9, 136.4, 134.5, 129.1, 129.1, 128.9, 128.7, 128.5, 127.9, 127.3, 115.49, 115.48, 104.5, 104.4, 76.3, 70.7, 49.6, 45.1, 32.1, 28.9, 28.6, 26.5, 26.3, 24.1, 23.7. HRMS [M+H⁺] m/z calcd for C₄₂H₄₈N₃O₆ 690.3543, found 690.3526.

4.21. *N*-Hydroxy-6-(3-hydroxy-2-oxopyridin-1(2*H*)-yl)-*N*-(5-(3-hydroxy-2-oxopyridin-1(2*H*)-yl)pentyl)hexanamide (19)

To a solution of **18** (0.208 g, 0.302 mmol) in EtOH (3 mL) was added 5% Pd on carbon (0.064 g, 0.0302 mmol) and the mixture stirred at rt under a H₂ balloon for 24 h. The catalyst was filtered off using diatomaceous earth and the filtrate was concentrated in vacuo to give **19** (0.110 g, 87%) as a light brown solid: mp 40–46 °C (del); IR (neat) 3130, 2932, 1647, 1585 cm⁻¹; ¹H NMR (300 MHz, CD₃OD) δ 7.08 (d, *J*=6.8 Hz, 2H), 6.82 (d, *J*=6.8 Hz, 2H), 6.22 (d,

J=6.8 Hz, 2H), 4.86 (br s, 3H), 4.00 (t, *J*=7.0 Hz, 4H), 3.60 (t, *J*=6.7 Hz, 2H), 2.47 (d, *J*=7.2 Hz, 2H), 1.82–1.70 (m, 4H), 1.70–1.56 (m, 4H), 1.44–1.26 (m, 4H); ¹³C NMR (75 MHz, CD₃OD) δ 175.8, 160.0, 159.9, 148.3, 129.3, 129.2, 116.9, 116.9, 108.3, 108.3, 50.9, 50.7, 48.7, 33.1, 30.1, 30.0, 27.4, 27.3, 25.6, 24.6. HRMS [M+H⁺] *m/z* calcd for C₂₁H₃₀N₃O₆ 420.2135, found 420.2133.

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Supplementary data

Supplementary data associated with this article can be found in the online version, at http://dx.doi.org/10.1016/j.tet.2015.10.028. These data include MOL files and InChiKeys of the most important compounds described in this article.

References and notes

- (a) Tam, T. F.; Leung-Toung, R.; Li, W.; Wang, Y.; Kariman, K.; Spino, M. Curr. Med. Chem. 2003, 10, 938–995; (b) Kalinowski, D. S.; Richardson, D. R. Pharmacol. Rev. 2005, 57, 547–583.
- (a) Ji, C.; Miller, M. J. Bioorg. Med. Chem. 2012, 20, 3828–3836; (b) Poreddy, A. R.; Schall, O. F.; Osiek, T. A.; Wheatley, J. R.; Beusen, D. D.; Marshall, G. R.; Slomczynska, U. J. Comb. Chem. 2004, 6, 239–254.
- (a) Möllmann, U.; Heinisch, L.; Bauernfeind, A.; Köhler, T.; Ankel-Fuchs, D. Biometals 2009, 22, 615–624; (b) Girijavallabhan, V.; Miller, M. J. "Therapeutic Uses of Iron (III) Chelators and Their Antimicrobial Conjugates" in Iron Transport in Bacteria; American Society for Microbiology: Washington, DC, 2004; pp 413–433.
- 4. Budzikiewicz, H. Mini-Reviews Org. Chem. 2004, 1, 163-168.
- (a) Weizman, H.; Shanzer, A. Chem. Commun. 2000, 2013–2014; (b) Meyer, M.; Telford, J. R.; Cohen, S. M.; White, D. J.; Xu, J.; Raymond, K. N. J. Am. Chem. Soc. 1997, 119, 10093–10103.
- Santos, M. A.; Marques, S. M.; Chaves, S. Coord. Chem. Rev. 2012, 256, 240–259.
 (a) Hider, R. C. Thalass. Rep. 2014, 4, 19–27; (b) Crisponi, G.; Remelli. Coord.
- (a) Hider, R. C. Thalass. Rep. 2014, 4, 19–27; (b) Crisponi, G.; Remelli. Coord. Chem. Rev. 2008, 252, 1225–1240.
- (a) Saghaie, L.; Sadeghi-aliabadi, H.; Ashaehshoar, M. Res. Pharm. Sci. 2012, 8, 185–195; (b) Kontoghiorghes, G. J.; Efstathiou, A.; Ioannou-Loucaides, S.; Kolnagou, A. Hemoglobin 2008, 32, 217–227.
- Zhou, Y.-J.; Liu, M.-S.; Osamah, A. R.; Kong, X.-L.; Alsam, S.; Battah, S.; Xie, Y.-Y.; Hider, R. C.; Zhou, T. Eur. J. Med. Chem. 2015, 94, 8–21.
- (a) Mitton-Fry, M. J.; Arcari, J. T.; Brown, M. F.; Casavant, J. M.; Finegan, S. M.; Flanagan, M. E.; Gao, H.; George, D. M.; Gerstenberger, B. S.; Han, S.; Hardink, J. R.; Harris, T. M.; Hoang, T.; Huband, M. D.; Irvine, R.; Lall, M. S.; Lemmon, M. M.; Li, C.; Lin, J.; McCurdy, S. P.; Mueller, J. P.; Mullins, L.; Niosi, M.; Noe, M. C.; Pattavina, D.; Penzien, J.; Plummer, M. S.; Risley, H.; Schuff, B. P.; Shanmugasundaram, V.; Starr, J. T.; Sun, J.; Winton, J.; Young, J. A. *Bioorg. Med. Chem. Lett.* 2012, *22*, 5989–5994; (b) Page, M. G. P.; Dantier, C.; Desarbre, E. *Antimicrob. Agents Chemother.* 2010, *54*, 2291–2302.
- 11. Datta, A.; Raymond, K. N. Acc. Chem. Res. 2009, 42, 938-947.
- (a) Pailloux, S. L.; Nguyen, S.; Zhou, S.; Hom, M. E.; Keyser, M. N.; Smiles, D.; Raymond, K. N. H.. Heterocycl. Chem. Early View, DOI 10.1002/jhet.2372. (b) Klemm, P. J.; Floyd, W. C., III; Andolina, C. M.; Fréchet, J. M. J.; Raymond, K. N. *Eur. J. Inorg. Chem.* **2012**, 2108–2114.
- 13. (a) Berry, D. J.; Ma, Y.; Ballinger, J. R.; Tavaré, R.; Koers, A.; Sunassee, K.; Zhou, T.; Nawaz, S.; Mullen, G. E. D.; Hider, R. C.; Blower, P. J. *Chem. Commun.* 2011, 7068–7070; (b) Chaves, S.; Mendonca, A. C.; Marques, S. M.; Prata, M. I.; Santos, A. C.; Martins, A. F.; Geraldes, C. F. G. C.; Santos, M. A. *J. Inorg. Biochem.* 2011, 105, 31–38.
- (a) Ma, M. T.; Meszaros, L. K.; Paterson, B. M.; Berry, D. J.; Cooper, M. S.; Ma, Y.; Hider, R. C.; Blower, P. J. *Dalton Trans.* **2015**, *44*, 4884–4900; (b) Deri, M. A.; Ponnala, S.; Zeglis, B. M.; Pohl, G.; Dannenberg, J. J.; Lewis, J. S.; Francesconi, L. C. *J. Med. Chem.* **2014**, *57*, 4849–4860.
- 15. Gorden, A. E. V.; Xu, J.; Raymond, K. N. Chem. Rev. 2003, 103, 4207–4282.
- 16. Liu, Z. D.; Hider, R. C. Coord. Chem. Rev. 2002, 232, 151–171.
- Xu, J.; Franklin, S. J.; Whisenhunt, D. W., Jr.; Raymond, K. N. J. Am. Chem. Soc. 1995, 117, 7245–7246.
- Streater, M.; Taylor, P. D.; Hider, R. C.; Porter, J. J. Med. Chem. 1990, 33, 1749–1755.
- Harrington, J. M.; Chittamuru, S.; Dhungana, S.; Jacobs, H. K.; Gopalan, A. S.; Crumbliss, A. L. *Inorg. Chem.* 2010, 49, 8208–8221.
- Meunier, S.; Siaugue, J.-M.; Sawicki, M.; Calbour, F.; Dézard, S.; Taran, F.; Mioskowski, C. J. Comb. Chem. 2003, 5, 201–204.
- (a) Chen, L.; Ling, L.; Xu, X. Polym. Prepr. 2008, 49, 889–890; (b) Zhang, Y.-M.; Fan, X.; Yang, S.-M.; Scannevin, R. H.; Burke, S. L.; Rhodes, K. J.; Jackson, P. F. Bioorg. Med. Chem. Lett. 2008, 18, 405–408; (c) Katoh, A.; Kudo, H.; Saito, R.

Heterocycles 2005, 66, 285–297; (d) Jankowska, K. Ph. D. Thesis, Tubingen, 2013.

- 22. Patel, M. K.; Fox, R.; Taylor, P. D. Tetrahedron 1996, 52, 1835–1840.
- 23. Martinez, G.; Arumugam, J.; Jacobs, H. K.; Gopalan, A. S. Tetrahedron Lett. 2013, 54, 630-634.
- 24. Kim, S. K.; Sessler, J. L. Chem. Soc. Rev. 2010, 39, 3784-3809.
- 25. Amendola, V.; Fabbrizzi, L.; Mosca, L. Chem. Soc. Rev. 2010, 39, 3889-3915.
- 26. Custelcean, R. *Chem. Commun.* **2013**, 2173–2182.
- (a) d'Hardemare, A. M.; Torelli, S.; Serratrice, G.; Pierre, J.-L. *BioMetals* 2006, *19*, 349–366; (b) Xu, J.; O'Sullivan, B.; Raymond, K. N. *Inorg. Chem.* 2002, *41*, 6731–6742; (c) Fernando, R.; Shirley, J. M.; Torres, E.; Jacobs, H. K.; Gopalan, A. S. *Tetrahedron Lett.* 2012, *53*, 6367–6371.
- 28. Correnti, C.; Strong, R. K. J. Biol. Chem. 2012, 287, 13524–13531.
- 29. Sandy, M.; Butler, A. Chem. Rev. 2009, 109, 4580–4595.
- Wencewicz, T. A.; Miller, M. J. J. Med. Chem. 2013, 56, 4044–4052.
 (a) Young, J. A.; Karmakar, S.; Jacobs, H. K.; Gopalan, A. S. Tetrahedron 2012, 68,
- (a) Young, J. A.; Karmakar, S.; Jacobs, H. K.; Gopalan, A. S. *Tetrahedron* 2012, 68, 10030–10039;
 (b) Arumugam, J.; Brown, H. A.; Jacobs, H. K.; Gopalan, A. S.

Synthesis **2011**, 57–64; (c) Liu, Y.; Jacobs, H. K.; Gopalan, A. S. *Tetrahedron* **2011**, 67, 2206–2214; (d) Liu, Y.; Jacobs, H. K.; Gopalan, A. S. *J. Org. Chem.* **2009**, 74, 782–788.

- (a) Kluba, C. A.; Mindt, T. L. Molecules 2013, 18, 3206–3226; (b) Goswami, L. N.; Ma, L.; Kueffer, P. J.; Jalisatgi, S. S.; Hawthorne, M. F. Molecules 2013, 18, 9034–9048; (c) Suchý, M.; Milne, M.; Li, A. X.; McVicar, N.; Dodd, D. W.; Bartha, R.; Hudson, R. H. E. Eur, J. Org. Chem. 2011, 6532–6543; (d) Nwe, K.; Brechbiel, M. W. Cancer Biotherapy Radiopharm. 2009, 24, 289–301.
- (a) Yus, M.; Pastor, I. M. Chem. Lett. 2013, 42, 94–108; (b) Bakherad, M. Appl. Organometal. Chem. 2013, 27, 125–149; (c) Chinchilla, R.; Nájera, C. Chem. Soc. Rev. 2011, 40, 5084–5121.
- 34. Torhan, M. C.; Peet, N. P.; Williams, J. D. *Tetrahedron Lett.* **2013**, 54, 3926–3928.
- 35. Protected hydroxamic acid precursors were prepared by alkylation of *tert*-butyl benzyloxycarbamate with the desired alkyl halide using sodium hydride in DMF. See Ref. 31d.