



The synthesis of possible transition state analogue inhibitors of thymidine phosphorylase



Gary B. Evans^{a,*}, Graeme J. Gainsford^b, Vern L. Schramm^c, Peter C. Tyler^a

^a Ferrier Research Institute, Victoria University of Wellington, 69 Gracefield Rd, Lower Hutt 5040, New Zealand

^b Department of Biochemistry, Albert Einstein College of Medicine, 1300 Morris Park Ave., Bronx, NY 10461, USA

^c Callaghan Innovation Ltd, Gracefield Rd., PO Box 31-310, Lower Hutt 5040, New Zealand

ARTICLE INFO

Article history:

Received 31 August 2014

Revised 17 October 2014

Accepted 25 November 2014

Available online 29 November 2014

Keywords:

Thymidine phosphorylase

Enzyme

Transition state analogue

Cancer

Corey–Link

ABSTRACT

The synthetically challenging S_N2 transition state mimic for thymidine phosphorylase, along with its phosphonate analogue, were synthesised via a modified Corey–Link reaction in good overall yields and ensuring the correct stereochemical outcome.

© 2014 Elsevier Ltd. All rights reserved.

Enzymes comprise one of the major categories of drug targets pursued by the pharmaceutical and biotechnology industries.¹ Historically, small molecule inhibitors of enzyme function have afforded a significant proportion of FDA approved drugs.² Enzymes have been optimised by Nature to make and break covalent chemical bonds as opposed to ligand binding.¹ The methodology used to design enzyme inhibitors has progressed markedly over the past quarter of a century and shares many aspects of ligand design in general.³ These design aspects can include knowledge of the natural ligand or substrate of the target of interest or of another compound already known to interact with the target of interest, screening of both random and biased libraries against a target, fragment-based screening (which can involve a variety of physical methods of analysis but mostly centre around X-ray crystallography and NMR).^{3a}

As an alternative to these ground state methods of inhibitor design, transition state affinity⁴ of enzymes and the design of transition state analogs should be considered as the preeminent method by which to afford both potent and selective inhibitors of enzymes.^{1,3a,5} Pauling first posited that enzymes had evolved or were designed to recognise the activated state of reactants. This premise was restated by Leinhard^{5b} and Wolfenden⁴ that the transition state binds more tightly than the substrate(s) and so

accounts for catalysis. Schramm⁶ has developed methods that can “image” the activated state of reactants or the transition state, on-enzyme, and we can then develop a blueprint for the design of selective tight binding inhibitors. Some of these inhibitor designs have been realised over the past two decades⁷ and have been shown to possess the advantages of covalently bound inhibitors without the off-site activity that sometimes impacts on the utility of this class of enzyme inhibitor.⁸

Thymidine phosphorylase (TP) is an enzyme that catalyses the reversible phosphorolysis of thymidine into thymine and 2'-deoxy-D-ribose 1-phosphate. TP is an enzyme important in the salvage of pyrimidine nucleosides, and to promote angiogenesis.⁹ Improved vasculature is necessary for the growth of solid tumours and the expression of this enzyme in many solid tumours correlates with the aggressiveness and invasiveness of the cancer.^{9c,f,10} TP inhibitors affect the production of 2'-deoxy-D-ribose, and in turn suppress tumour growth. Consequently, inhibitors of TP are expected to be useful as anti-cancer compounds.¹¹

Schramm's initial design blueprint for a transition state analogue inhibitor of human TP, was derived using an enzyme construct with purification tags, and suggested an S_N2 -like transition state, with a bond order of 0.5 to the leaving group and 0.33 to the attacking oxygen nucleophile of the phosphate, which was unprecedented for *N*-ribosyl transferase. This transition state supports continuous electron density extending from N1 through C1 to the phosphate oxygen as expected for an S_N2 transition state, that is, with equal participation of both the incoming phosphate

* Corresponding author. Tel.: +64 4 463 0048; fax: +64 4 931 3055.

E-mail address: gary.evans@vuw.ac.nz (G.B. Evans).

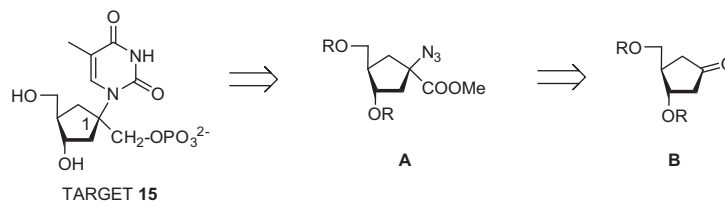


Figure 1. Retrosynthetic analysis of the transition state analogue target.

nucleophile and pyrimidine leaving group with little or no charge at the reaction centre of the ribosyl moiety.¹² With this background, we embarked on the synthesis of analogs geometrically and electronically similar to the S_N2 transition state. The novel chemistry of their synthesis is described here. They were not significant inhibitors of the enzyme, supporting a different transition state for TP.

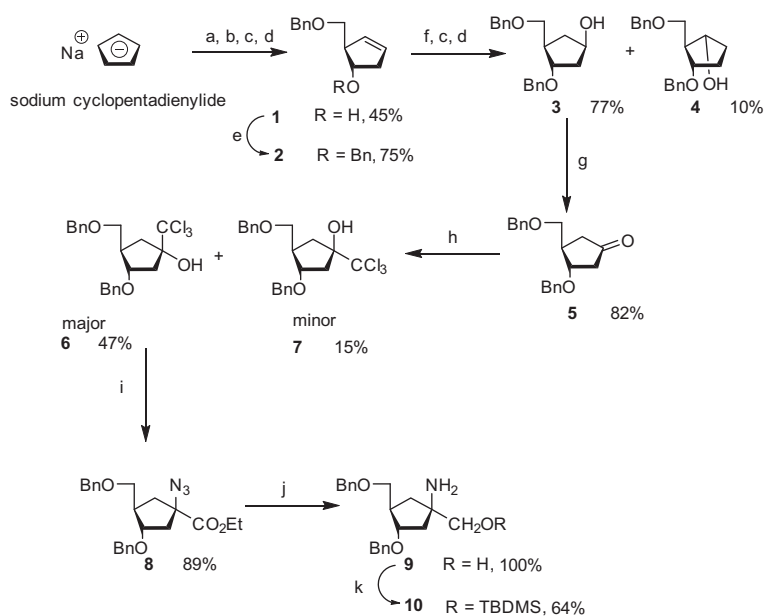
Subsequent hydrolytic¹³ and arsenolytic^{6d} transition state analysis with native human TP, and without the protein-modifying purification tags, supported a transition state with ribocationic character with little or no participation of the incoming nucleophile in an A_ND_N mechanism.

Regardless of these latter results, incorporating both a thymine or thymine mimic and a phosphate or phosphate mimic at a single carbon of a cyclopentane ring, with a methylene group between the phosphate and the ring as a 'spacer' provided a real synthetic challenge (Fig. 1). The resulting compounds proved useful in eliminating the earlier S_N2 transition state proposal for TP. The present chemistry provides a useful precedent for the synthesis of similar compounds for related enzyme chemistry.

Ludek and Meier¹⁴ previously described the synthesis of a carbocyclic thymidine analogue, and so we investigated ways in which this procedure could be modified to afford our desired target compound. The core cyclopentane structure could be realised via the alkene intermediate **1**, which utilised an enantioselective hydroboration of cyclopentadiene first described by Biggadike et al.,¹⁵ and inspired by Partridge et al. (Scheme 1).¹⁶ The physical

characteristics of alkene **1** were the same as those reported in the literature and it was benzylated to yield compound **2** and then hydroborated using borane to afford a mixture of alcohols **3** and **4**, which could be separated by chromatography.¹⁷ Oxidation of the alcohol **3** using PDC afforded the key ketone intermediate, compound **5**,¹⁷ in good overall yield from cyclopentadiene.

Consideration was now given to the stereochemical outcome of methods appropriate for installing the nitrogen and carbon functionalities C1', where the nitrogen required to construct the thymine moiety is above the ring plane and the carbon, which will provide the methylene 'spacer' between the ring and the phosphate moiety, is below. We were influenced by the stereochemical analysis of Stick and co-workers on a carbohydrate ketone in their study and the likely outcomes of a Strecker reaction¹⁸ versus that of a modified Corey–Link reaction.¹⁹ Corey and Link first described the synthesis of α -amino acids via the reaction of a trichloromethylcarbinol with NaOH and NaN₃ in 1992.²⁰ This was followed by a modification of this procedure by Domínguez et al., exemplified in a ring system not dissimilar to our own, using DBU and MeOH to afford the desired α -azido methyl ester.²¹ Further literature precedent^{21,22} demonstrated the utility of the modified Corey–Link reaction and so we were keen to adopt this approach in the setting described above. We considered that the addition of a sterically, and hence, stereochemically demanding trichloromethylcarbanion would occur predominantly from the least hindered β face of the cyclopentanone ring and therefore, where the Corey–Link reaction with azide, base and the intermediate *gem*-dichlorooxirane



Scheme 1. Reagents and conditions: (a) BOMCl, THF, $-60\text{ }^{\circ}\text{C} \rightarrow -20\text{ }^{\circ}\text{C}$; (b) (–)-diisopinane-3-yl borane, THF, $-60\text{ }^{\circ}\text{C} \rightarrow -20\text{ }^{\circ}\text{C}$; (c) 3 M NaOH, $0\text{ }^{\circ}\text{C}$; (d) 30% H₂O₂, THF, $0\text{ }^{\circ}\text{C} \rightarrow \text{rt}$; (e) BnBr, NaH, DMF, $0\text{ }^{\circ}\text{C} \rightarrow \text{rt}$; (f) 9-BBN, THF, $0\text{ }^{\circ}\text{C} \rightarrow \text{rt}$; (g) PDC, Ac₂O, CH₂Cl₂, rt; (h) CHCl₃, LHMDS, THF, $-78\text{ }^{\circ}\text{C} \rightarrow \text{rt}$; (i) NaN₃, DBU, EtOH, $50\text{ }^{\circ}\text{C}$; (j) LAH, THF, $0\text{ }^{\circ}\text{C} \rightarrow \text{rt}$; (k) TBDMSCl, imidazole, DMF, rt.

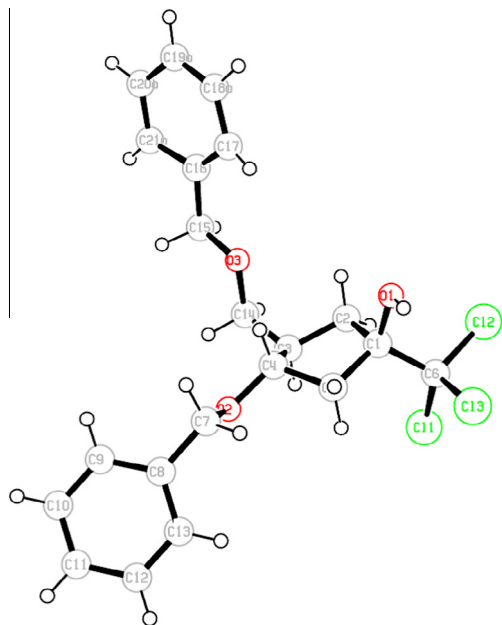


Figure 2. XRD of compound 7.

proceeds with complete stereochemical inversion,²⁰ our nitrogen, in the form of an azide, would ultimately be installed on the desired β face and the carbon functionality on the α face of C1'.

Initial addition of the trichloromethylcarbanion yielded compounds **6** (47%) and **7** (15%), the X-ray crystal structure of the minor product **7** (Fig. 2) confirming the stereochemistry of both isomers. As expected attack of the trichloromethylcarbanion occurred predominantly from the β face of the cyclopentane ring.

Treatment of **6** with NaN_3 and DBU in ethanol afforded the desired azido ethyl ester **8** in excellent yield. Concomitant reduction of both the azido and ester moieties of **8** using LAH gave the amino alcohol **9** and presumably due to the lability of any silylated amine, treatment of **9** with TBDMSCl, under standard conditions, yielded only the desired silyl ether **10** in good overall yield.

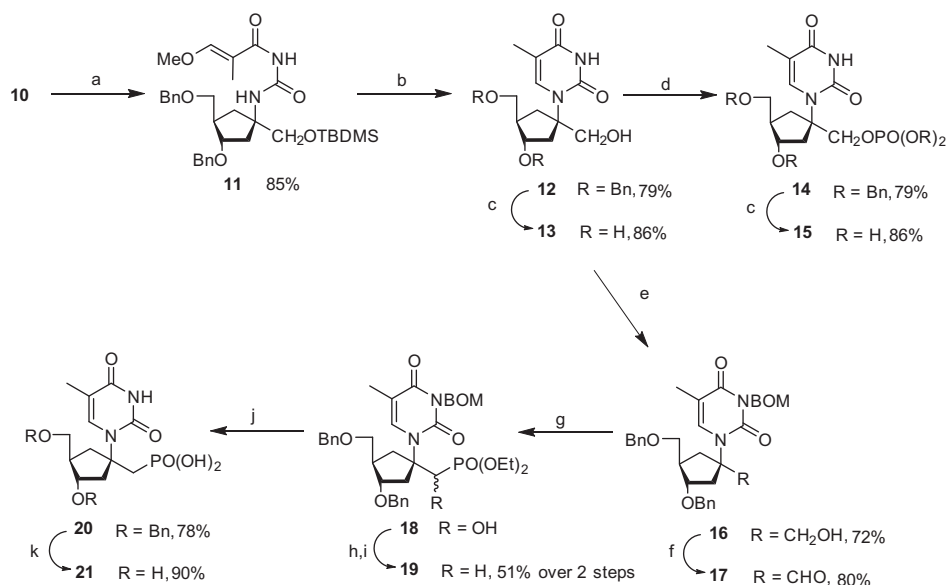
Installation of the thymine moiety was achieved via amine **10**, which was reacted with 3-methoxy-2-methylacryloyl isocyanate²³ to afford intermediate compound **11** and then cyclisation and desilylation with HCl proceeded in good yield to afford thymine analogue **12** in excellent yield over the two steps. Debenzylation of **12** gave compound **13** and this compound was put aside to be assayed against TP as a potential inhibitor. Compound **13** was subsequently investigated as a source of the target phosphate **15**. Treatment of alcohol **13** with dibenzyl diisopropylphosphoramidite yielded the tetrabenzyl compound **14** in good yield which, on hydrogenolysis, afforded the putative transition state analogue target phosphate **15** (see Scheme 2).²⁴

We also proposed that the phosphonate analogue **21** of phosphate **15** would also afford a putative inhibitor of TP. Compound **13** was used as a starting point for its construction. We began by N-protection of **13** with BOMCl followed by oxidation of the resulting alcohol **16** with Dess Martin periodinane to afford aldehyde **17** in good overall yield. The anion of diethyl phosphate was generated in situ with sodium hydride²⁵ and added to aldehyde **17**, and the intermediate alcohol was converted into the protected phosphonate **19** using a Barton–McCombie deoxygenation procedure.²⁶ Sequential deprotection of compound **19** using TMSBr and then BBr_3 yielded the required phosphonate **21**²⁷ in excellent overall yield.

In summary, we have realised for the first time a carbocyclic thymidine analogue with a pendant α -carbon at the C1 position in order to incorporate either a methylene group linked to a phosphate or phosphonate group. The design was based on the premise that the transition state of thymidine phosphorylase had $\text{S}_{\text{N}}2$, but no ribocation character, and the key synthetic steps involved a modified Corey–Link reaction which ultimately yielded the target compounds in good overall yields.

Acknowledgments

This work was supported by funds from the NIH GM 41916 (VLS) and the New Zealand Foundation for Research, Science, and Technology.



Scheme 2. Reagents and conditions: (a) $\text{MeOCH}=\text{C}(\text{CH}_3)\text{CONCO}$, MeCN, rt; (b) 10% HCl, 1,4-dioxane, reflux; (c) H_2 (1 atm), Pd/C, EtOH, rt; (d) dibenzyl diisopropylphosphoramidite, tetrazole, CH_2Cl_2 , rt; (e) BOMCl, DBU, DMF, rt; (f) Dess Martin periodinane, CH_2Cl_2 , rt; (g) diethyl phosphate, NaH, THF, $0^\circ\text{C} \rightarrow \text{rt}$; (h) CS_2 , NaH, MeI, DMF, rt; (i) $n\text{Bu}_3\text{SnH}$, benzene, reflux; (j) TMSBr, Et_3N , CH_2Cl_2 , rt; (k) BBr_3 , CH_2Cl_2 , Ar, -78°C .

Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.tetlet.2014.11.113>.

References and notes

- Robertson, J. G. *Curr. Opin. Struct. Biol.* **2007**, *17*, 674.
- Robertson, J. G. *Biochemistry* **2005**, *44*, 5561.
- (a) Banner, D. W. *Enzyme Inhibitor Design. Protein Science Encyclopedia*; Wiley-VCH Verlag GmbH & Co: KGaA, 2008. <http://dx.doi.org/10.1002/9783527610754.pl03>; (b) Sandler, M.; Smith, H. J. *Design of Enzyme Inhibitors as Drugs*; Oxford University Press: USA, 1989.
- Wolfenden, R. *Ann. Rev. Biophys. Bioeng.* **1976**, *5*, 271.
- (a) Schramm, V. L. *Annu. Rev. Biochem.* **2011**, *80*, 703; (b) Lienhard, G. E. *Science* **1973**, *180*, 149; (c) Lindquist, R. *Drug Des.* **1975**, *5*, 23.
- (a) Berti, P. J.; Blanke, S. R.; Schramm, V. L. *J. Am. Chem. Soc.* **1997**, *119*, 12079; (b) Lewandowicz, A.; Schramm, V. L. *Biochemistry* **2004**, *43*, 1458; (c) Singh, V.; Lee, J. E.; Nunez, S.; Howell, P. L.; Schramm, V. L. *Biochemistry* **2005**, *44*, 11647; (d) Schwartz, P. A.; Veticatt, M. J.; Schramm, V. L. *Biochemistry* **2011**, *50*, 1412; (e) Schramm, V. L.; Hirsch, B. M.; Burgos, E. S. *ACS Chem. Biol.* **2014**, *9*, 2255–2262.
- (a) Kicska, G. A.; Tyler, P. C.; Evans, G. B.; Furneaux, R. H.; Kim, K.; Schramm, V. L. *J. Biol. Chem.* **2002**, *277*, 3219; (b) Evans, G. B.; Furneaux, R. H.; Lewandowicz, A.; Schramm, V. L.; Tyler, P. C. *J. Med. Chem.* **2003**, *46*, 5271; (c) Singh, V.; Shi, W.; Evans, G. B.; Tyler, P. C.; Furneaux, R. H.; Almo, S. C.; Schramm, V. L. *Biochemistry* **2004**, *43*, 9; (d) Evans, G. B.; Furneaux, R. H.; Lenz, D. H.; Painter, G. F.; Schramm, V. L.; Singh, V.; Tyler, P. C. *J. Med. Chem.* **2005**, *48*, 4679; (e) Singh, V.; Evans, G. B.; Lenz, D. H.; Mason, J. M.; Clinch, K.; Mee, S.; Painter, G. F.; Tyler, P. C.; Furneaux, R. H.; Lee, J. E.; Howell, P. L.; Schramm, V. L. *J. Biol. Chem.* **2005**, *280*, 18265. <http://dx.doi.org/10.1074/jbc.M414472200>; (f) Taylor Ringia, E. A.; Schramm, V. L. *Curr. Top. Med. Chem.* **2005**, *5*, 1237; (g) Clinch, K.; Evans, G. B.; Frohlich, R. F.; Kelly, P. M.; Murkin, A. S.; Li, L.; Schramm, V. L.; Tyler, P. C. *J. Med. Chem.* **2009**, *52*, 1126; (h) Gutierrez, J. A.; Crowder, T.; Rinaldo-Matthis, A.; Ho, M. C.; Almo, S. C.; Schramm, V. L. *Nat. Chem. Biol.* **2009**, *5*, 251; (i) Longshaw, A. I.; Adanitsch, F.; Gutierrez, J. A.; Evans, G. B.; Tyler, P. C.; Schramm, V. L. *J. Med. Chem.* **2010**, *53*, 6730; (j) Wang, S.; Haapalainen, A. M.; Yan, F.; Du, Q.; Tyler, P. C.; Evans, G. B.; Brown, R. L.; Norris, G. E.; Almo, S. C. *Biochemistry* **2012**, *51*, 6892; (k) Zhang, Y.; Evans, G. B.; Clinch, K.; Crump, D. R.; Harris, L. D.; Tyler, P. C.; Hazelton, K. Z.; Cassera, M. B.; Schramm, V. L. *J. Biol. Chem.* **2013**, *288*, 34746.
- (a) Johnson, D. S.; Weerapana, E.; Cravatt, B. F. *Future Med. Chem.* **2010**, *2*, 949; (b) Schramm, V. L. *ACS Chem. Biol.* **2013**, *8*, 71.
- (a) Miyazono, K.; Usuki, K.; Heldin, C. H. *Prog. Growth Factor Res.* **1991**, *3*, 207; (b) Usuki, K.; Saras, J.; Waltenberger, J.; Miyazono, K.; Pierce, G.; Thomason, A.; Heldin, C. H. *Biochem. Biophys. Res. Commun.* **1992**, *184*, 1311; (c) Fox, S. B.; Westwood, M.; Moghaddam, A.; Comley, M.; Turley, H.; Whitehouse, R. M.; Bicknell, R.; Gatter, K. C.; Harris, A. L. *Br. J. Cancer* **1996**, *73*, 275; (d) Haraguchi, M.; Akiyama, S. *Seikagaku* **1996**, *68*, 143; (e) Harris, A. L.; Zhang, H.; Moghaddam, A.; Fox, S.; Scott, P.; Pattison, A.; Gatter, K.; Stratford, I.; Bicknell, R. *Breast Cancer Res. Treat.* **1996**, *38*, 97; (f) Zhang, X.; Zheng, Z.; Shin, Y. K.; Kim, K. Y.; Rha, S. Y.; Noh, S. H.; Chung, H. C.; Jeung, H. C. *Pathology* **2014**, *46*, 316.
- (a) Mitselou, A.; Ioachim, E.; Skoufi, U.; Tsironis, C.; Tsimogiannis, K. E.; Skoufi, C.; Vougiouklakis, T.; Briasoulis, E. *In Vivo* **2012**, *26*, 1057; (b) Deves, C.; Rostirolla, D. C.; Martinelli, L. K.; Bizarro, C. V.; Santos, D. S.; Basso, L. A. *Mol. Biosyst.* **2014**, *10*, 592; (c) Ko, J. C.; Chiu, H. C.; Syu, J. J.; Jian, Y. J.; Chen, C. Y.; Jian, Y. T.; Huang, Y. J.; Wo, T. Y.; Lin, Y. W. *Biochem. Pharmacol.* **2014**, *88*, 119; (d) Lindskog, E. B.; Derwinger, K.; Gustavsson, B.; Falk, P.; Wettergren, Y. *BMC Clin. Pathol.* **2014**, *14*, 25; (e) Takebayashi, Y.; Yamada, K.; Ohmoto, Y.; Sameshima, T.; Miyadera, K.; Yamada, Y.; Akiyama, S.; Aikou, T. *Cancer Lett.* **1995**, *95*, 57.
- (a) Cha, S. M. *Yonsei Med. J.* **1989**, *30*, 315; (b) Klein, R. S.; Lenzi, M.; Lim, T. H.; Hotchkiss, K. A.; Wilson, P.; Schwartz, E. L. *Biochem. Pharmacol.* **2001**, *62*, 1257; (c) Pomeisl, K.; Votruba, I.; Holy, A.; Pohl, R. *Nucleosides Nucleotides Nucleic Acids* **2007**, *26*, 1025; (d) Liekens, S.; Bronckaers, A.; Belleri, M.; Bugatti, A.; Sienaert, R.; Ribatti, D.; Nico, B.; Gigante, A.; Casanova, E.; Opdenakker, G.; Pérez-Pérez, M.; Balzarini, J.; Presta, M. *Mol. Cancer Ther.* **2012**, *11*, 817; (e) Bera, H.; Tan, B. J.; Sun, L.; Dolzhenko, A. V.; Chui, W. K.; Chiu, G. N. *Eur. J. Med. Chem.* **2013**, *67*, 325; (f) Shahzad, S. A.; Yar, M.; Bajda, M.; Jadoon, B.; Khan, Z. A.; Naqvi, S. A.; Shaikh, A. J.; Hayat, K.; Mahmood, A.; Mahmood, N.; Filipek, S. *Bioorg. Med. Chem.* **2014**, *22*, 1008.
- Birck, M. R.; Schramm, V. L. *J. Am. Chem. Soc.* **2004**, *126*, 2447.
- Schwartz, P. A.; Veticatt, M. J.; Schramm, V. L. *J. Am. Chem. Soc.* **2010**, *132*, 13425.
- Ludek, O. R.; Meier, C. *Nucleosides Nucleotides Nucleic Acids* **2003**, *22*, 683.
- Biggadike, K.; Borthwick, A. D.; Exall, A. M.; Kirk, B. E.; Roberts, S. M.; Youds, P.; Slawin, A. M. Z.; Williams, D. J. *J. Chem. Soc., Chem. Commun.* **1987**, 255.
- Partridge, J. J.; Chadha, N. K.; Uskokovic, M. R. *J. Am. Chem. Soc.* **1973**, *95*, 532.
- Huang, H.; Greenberg, M. M. *J. Org. Chem.* **2008**, *73*, 2695.
- Gröger, H. *Chem. Rev.* **2003**, *103*, 2795.
- Scaffidi, A.; Skelton, B. W.; Stick, R. V.; White, A. H. *Aust. J. Chem.* **2006**, *59*, 426.
- Corey, E. J.; Link, J. O. *J. Am. Chem. Soc.* **1992**, *114*, 1906.
- Dominguez, C.; Ezquerro, J.; Richard Baker, S.; Borrelly, S.; Prieto, L.; Espada, M.; Pedregal, C. *Tetrahedron Lett.* **1998**, *39*, 9305.
- (a) Snider, J. R.; Entekin, J. T.; Snowden, T. S.; Dolliver, D. *Synthesis* **2013**, 45, 1899; (b) Lee, C.-W.; Lira, R.; Dutra, J.; Ogilvie, K.; O'Neill, B. T.; Brodney, M.; Helal, C.; Young, J.; Lachapelle, E.; Sakya, S.; Murray, J. C. *J. Org. Chem.* **2013**, *78*, 2661.
- Lee, Y. H.; Kim, H. K.; Youn, I. K.; Chae, Y. B. *Bioorg. Med. Chem. Lett.* **1991**, *1*, 287.
- Compound **15**. ¹H NMR (300 MHz, D₂O): δ 7.45 (s, 1H), 4.23 (dd, *J* = 10.9, 5.6 Hz, 1H), 4.07 (m, 2H), 3.71 (dd, *J* = 11.3, 5.3 Hz, 1H), 3.62 (dd, *J* = 11.3, 6.2 Hz, 1H), 2.58 (m, 2H), 2.23 (m, 1H), 2.14 (m, 1H), 1.84 (s, 3H), 1.77 (t, *J* = 12.1 Hz, 1H); ¹³C NMR (75 MHz, D₂O): δ 167.3, 152.4, 142.4, 109.9, 72.1, 70.0, 66.5, 63.1, 47.3, 43.1, 35.9, 11.8; HRMS calcd for C₁₂H₁₉N₂O₈P (MH)⁺ 350.0879, found 350.0885.
- Klepacz, A.; Zwierzak, A. *Tetrahedron Lett.* **2002**, *43*, 1079.
- Barton, D. H.; McCombie, S. W. *J. Chem. Soc., Perkin Trans. 1* **1975**, 1574.
- Compound **21**. ¹H NMR (300 MHz, D₂O): δ 7.52 (s, 1H), 4.06 (q, *J* = 6.0 Hz, 1H), 3.66 (m, 2H), 2.82 (dd, *J* = 12.1, 6.9 Hz, 1H), 2.60–2.17 (m, 5H), 1.94 (s, 3H), 1.80 (t, *J* = 11.7 Hz, 1H); ¹³C NMR (75 MHz, D₂O): δ 167.3, 152.5, 142.4, 108.9, 72.3, 68.0 (d, *J*_{CP} = 5.9 Hz), 63.4, 48.6 (d, *J*_{CP} = 9.4 Hz), 47.8, 39.6, 35.3 (d, *J*_{CP} = 132 Hz), 11.9; ³¹P NMR (D₂O): δ 19.1; HRMS calcd for C₁₂H₁₈N₂O₇P (MH)[−] 333.0857, found 333.0853.